Plant Physiol. (1998) 118: 1101–1103

The Electronic Plant Gene Register

Plant Gene Register titles for PGR 98–175 to PGR 98–189 appear below. The sequences have been deposited in GenBank and the articles listed online through the World Wide Web.

To cite an electronic Plant Gene Register article as a bibliographic reference, follow the style given below:

Weers B, Thornburg R (1998) Characterization of the cDNA and gene for the Arabidopsis GDP-mannose pyrophosphorylase (accession no. AF076484) (PGR 98–175). Plant Physiol **118:** 1101.

To access the Plant Gene Register through the World Wide Web, use the URL:

http://www.tarweed.com/pgr/

Plant Gene Register PGR 98–175

Characterization of the cDNA and Gene for the Arabidopsis GDP-Mannose Pyrophosphorylase (Accession No. AF076484).

Brock Weers and Robert Thornburg*.
Department of Biochemistry and Biophysics, Iowa State University, Ames, Iowa 50011.
* Corresponding author; e-mail thorn@iastate.edu; fax 1–515–294–0453.

Plant Gene Register PGR 98–176

Nucleotide Sequences of cDNAs (Accession Nos. AJ010160, AJ010161, and AJ010162) Encoding a Type-2 Metallothionein-Like Protein from Water Hyacinth.

Ka Fai Kong and Jimmy S.H. Tsang*.
Department of Botany, University of Hong Kong, Pokfulam Road, Hong Kong.
* Corresponding author; e-mail jshtsang@hkucc.hku.hk; fax 852–2858–3477.

Plant Gene Register PGR 98–177

Molecular Characterization of a cDNA Encoding a Ubiquitin-Carrier Protein (UBC7) Isolated from Egg Cells of Maize (Accession No. AJ002959).

Arne de Vries, Simone Cordts, and Thomas Dresselhaus*.
Applied Plant Molecular Biology II, University of Hamburg, Ohnhorststrasse 18, D-22609 Hamburg, Germany.
* Corresponding author; e-mail dresselh@botanik.uni-hamburg.de; fax 49–40–8228–2229.

Plant Gene Register PGR 98–178

Structure of Rice Genes Encoding Three Class-I Low-Molecular-Mass Heat-Shock Proteins (Accession Nos. U83669, U83670, and U83671).

Jiahn-Chou Guan, Fa-Cheng Chang, Tong-Shun Tseng, Pi-Fang L. Chang, Kai-Wun Yeh, Yih-Ming Chen, and Chu-Yung Lin*.
Department of Botany, National Taiwan University, Taipei, Taiwan.
* Corresponding author; e-mail chuyung@ccms.ntu.edu.tw; fax 886–2–2363–8598.

Plant Gene Register PGR 98–179

Cloning and Characterization of a Cellulose Synthase cDNA (Accession No. AF081534) from Xylem of Hybrid Poplar.

Hongyan Wang and Carol A. Loopstra*.
Department of Forest Science, Texas A&M University, College Station, Texas 77843-2135.
* Corresponding author; e-mail c-loopstra@ tamu.edu; fax 1–409–862–4790.

Plant Gene Register PGR 98–180

The Genomic Organization of the Arabidopsis 6–4 Photolyase Gene (Accession No. AB017331).

Ayako Sakamoto*, Atsushi Tanaka, Shigemitsu Tano, Saatoshi Nakajima, Kazuo Yamamoto, and Hiroshi Watanabe.
Plant Resources Laboratory, Japan Atomic Energy Research Institute, Watanuki-machi 1233, Takasaki 370-1292, Japan (A.S., A.T., S.T., H.W.); and Biological Institute, Graduate School of Science, Tohoku University, Sendai 98077, Japan (S.N., K.Y.).
* Corresponding author; e-mail sakamoto@taka.jaeri.go.jp; fax 81–27–346–9696.

Plant Gene Register PGR 98–181

Nucleotide Sequence of a 3-Oxoacyl-(Acyl-Carrier-Protein) Synthase Gene (3-Ketoacyl-ACP Synthase) (Accession No. AF085148) from Habanero Chile.

Maneesha Aluru, Jeanne Curry, and Mary O'Connell*.
Molecular Biology Program, Department of Agronomy and Horticulture, New Mexico State University, Las Cruces, New Mexico 88003.
* Corresponding author; e-mail moconnel@nmsu.edu; fax 1–505–646–6041.

Plant Gene Register PGR 98–182

Nucleotide Sequence of a Probable Aminotransferase Gene (Accession No. AF085149) from Habanero Chile.

Maneesha Aluru, Jeanne Curry, and Mary O'Connell*.
Molecular Biology Program, Department of Agronomy and Horticulture, New Mexico State University, Las Cruces, New Mexico 88003.
* Corresponding author; e-mail moconnel@nmsu.edu; fax 1–505–646–6041.

Plant Gene Register PGR 98–183

Cloning and Characterization of the Arabidopsis Germin-Like Protein (GLP1) Gene (Accession No. AF090733).

Clay Carter and Robert Thornburg*.
Department of Biochemistry and Biophysics, Iowa State University, Ames, Iowa 50011.
* Corresponding author; e-mail thorn@iastate.edu; fax 1–515–294–0453.

Plant Gene Register PGR 98–184

A Full-Length cDNA for Phenylalanine Ammonia-Lyase Cloned from Ripe Sweet Cherry Fruit (Accession No. AF036948).

Paul A. Wiersma* and Zhencai Wu.
Agriculture and Agri-Food Canada, PARC Summerland, Summerland, British Columbia, Canada V0H 1Z0.
* Corresponding author; e-mail wiersmap@em.agr.ca; fax 1–250–494–0755.

Plant Gene Register PGR 98–185

Nucleotide Sequence of a cDNA Encoding a Glutathione Peroxidase (Accession No. AF053311) from *Zantedeschia aethiopica* (L.) Spreng.

Teresa Lino-Neto*, Rui M. Tavares, Klaus Palme, and Maria Salomé S. Pais.
Plant Molecular Biology Laboratory, Department of Biology, University of Minho, Campus de Gualtar, 4710 Braga, Portugal (T.L.-N., R.M.T.); Max Delbrück Laboratorium in der Max Planck Gesellschaft, Carl von Linné Weg, 10, D-50829 Köln, Germany (K.P.); and Department of Plant Biology, Science Faculty of Lisbon, Bloco C2, Campo Grande, 1700 Lisboa, Portugal (M.S.S.P.).
* Corresponding author; e-mail tlneto@bio.uminho.pt; fax 351–53–678–980.

Plant Gene Register PGR 98–186

DNA Sequence (Accession No. AF076166) of the *Brassica campestris* Mitochondrial *atpA* Gene.

Lisa A. Hammett* and Brent L. Nielsen.
Research Genetics, Huntsville, Alabama 35801 (L.A.H.); and Department of Botany and Microbiology, Auburn University, Auburn, Alabama 36849 (B.L.N.).
* Corresponding author; e-mail hammett@resgen.com; fax 1–256–551–1021.

Plant Gene Register PGR 98–187

Molecular Cloning of a cDNA Encoding a Novel UDP-Glucose Glucosyltransferase Homolog from Arabidopsis (Accession No. AB016819).

Emiko Yamagishi, Zhizhong Gong, Mami Yamazaki, and Kazuki Saito*.
Laboratory of Molecular Biology and Biotechnology, Research Center of Medicinal Resources, Faculty of Pharmaceutical Sciences, Chiba University, Chiba 2 63–8522, Japan.
* Corresponding author; e-mail ksaito@p.chiba-u.ac.jp; fax 81–43–290–2905.

Plant Gene Register PGR 98–188

Identification of Two Isoenzymes (Accession Nos. AF090444 and AF090445) of Phospholipase D from Cabbage.

Ines Pannenberg, Johanna Mansfeld, and Renate Ulbrich-Hofmann*.
Martin-Luther University Halle-Wittenberg, Department of Biochemistry/Biotechnology, Institute of Biotechnology, Kurt-Mothes-Strasse 3, D-06120 Halle, Germany.
* Corresponding author; e-mail Ulbrich-hofmann@biochemtech.uni-halle.de; fax 49–345–552–7013.

Plant Gene Register PGR 98–189

A Full-Length β-Galactosidase cDNA Sequence (Accession No. AF064786) from Ripening Papaya.

Roohaida Othman, Tze-Siang Choo, Zainon Mohd Ali, Zamri Zainal, and Hamid Lazan*.
Plant Molecular Physiology Group, Departments of Biochemistry (R.O., T.-S.C., Z.M.A.) and Botany (Z.Z., H.L.), Faculty of Life Sciences, Universiti Kebangsaan Malaysia, 43600 Bangi, Selangor, Malaysia.
* Corresponding author; e-mail hlazan@pkrisc.cc.ukm.my; fax 6–3–829–3244.

CORRECTION

Volume 117: 1401–1410, 1998

Fleischer, A., Titel, C., and Ehwald, R. The Boron Requirement and Cell Wall Properties of Growing and Stationary Suspension-Cultured *Chenopodium album* L. Cells.

Several inaccuracies were printed in this article and they are corrected below.

On page 1401, the correct e-mail address for Rudolf Ehwald is: rudolf-ehwald@rz.hu-berlin.de.

On page 1402 under the heading *Propagation Culture*, the second to last sentence should read: The final biomass concentration (c_t) at subcultivation time and the initial biomass concentration (c_o) were constant (see Fig. 1b) and, therefore, the mean specific-growth rate ($\mu = ½\ d\ [\ln c_t - \ln c_o]$) equals the mean dilution rate ($r = ½\ d\ [\ln 5 - \ln 2] = 0.46\ d^{-1}$).

On page 1403, the second and third sentences in the legend to Figure 2 should read: a, Particle-size distribution curves of suspension-cultured *C. album* cells after disaggregating treatment. Cells grown without boron (−B) or with 100 μM boron (+B).

On page 1404, the first sentence in the legend to Figure 3 should read: Size distribution profiles of the polydisperse dextran-probing solutions after equilibration with denatured growth-phase cells.

On page 1404, the last complete sentence in the right column should read: The elution profiles of the cell-exposed dextran solutions contained "steps," or more gradual changes attributable to the size-dependent diffusion of the dextrans into the cell lumina (Fig. 3).

On page 1407, the entire legend to Figure 7 should read: Size distribution profiles of the polydisperse dextran-probing solutions after equilibration with denatured stationary cells. Denatured stationary cells derived from cells grown in medium containing 100 μM boron (top) or 7 μM boron (bottom). The size distribution profile of the untreated dextran-probing solution is shown in Figure 3. c, Dextran concentration.