



Published in final edited form as:

Eur J Nucl Med Mol Imaging. 2018 January ; 45(1): 132–150. doi:10.1007/s00259-017-3839-5.

PET and SPECT Imaging of Melanoma: State of the Art

WeiJun Wei^{1,2,#}, Emily B. Ehlerding^{3,#}, Xiaoli Lan^{4,*}, Quanyong Luo^{1,*}, and Weibo Cai^{2,3,5,*}

¹Department of Nuclear Medicine, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, 600[#] Yishan Road, 200233, Shanghai, China

²Department of Radiology, University of Wisconsin - Madison, Wisconsin 53705, United States

³Department of Medical Physics, University of Wisconsin - Madison, Wisconsin 53705, United States

⁴Department of Nuclear Medicine, Union Hospital, Tongji Medical College, Wuhan, China

⁵University of Wisconsin Carbone Cancer Center, Madison, Wisconsin 53705, United States

Abstract

Melanoma is the most aggressive form of skin cancer and incidences continue to rise worldwide. ¹⁸F-FDG PET imaging has transformed diagnostic nuclear medicine and become an essential component in the management of melanoma, but still has its drawbacks. With the rapid growth in the field of nuclear medicine and molecular imaging, a variety of promising probes that enable early diagnosis and detection of melanoma have been developed. The substantial preclinical success of melanin- and peptide-based probes has recently resulted in translation of several radiotracers to clinical settings for noninvasive imaging and/or treatment of melanoma in human patients. In this review, we have focused on the latest developments in radiolabelled molecular imaging probes for melanoma in preclinical and clinical settings and discussed the challenges and opportunities for future development.

Keywords

Melanoma; molecular imaging; positron emission tomography (PET); single photon emission computer tomography (SPECT); cancer; theranostics; immunotherapy

1. Introduction

Although melanoma accounts for a small percentage of all skin cancer cases, it is estimated that melanoma accounted for 76,380 new cases and 10,130 deaths in the United States in 2016 [1]. Unsatisfactory 5-year survival rates for patients with distant metastases (less than

* Address correspondence to: Xiaoli Lan, LXL30724@hotmail.com, Address: Department of Nuclear Medicine, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology; Hubei Key Laboratory of Molecular Imaging, No. 1277 Jiefang Ave, Wuhan, 430022, China. Quanyong Luo, lqyn@sh163.net, Address: Department of Nuclear Medicine, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, 600[#] Yishan Road, 200233, Shanghai, China. Weibo Cai, wcai@uwhealth.org, Address: Room 7137, 1111 Highland Avenue, University of Wisconsin – Madison, Madison, WI 53705-2275, United States.

[#]Co-first authors contributed equally to this work

Conflict of interest

The authors have declared no competing interest.

10%) render early detection and accurate assessment of metastatic melanoma crucial for improved outcome and disease-free survival [2]. With the gradual increase in understanding of the molecular pathogenesis of melanoma, the treatment landscape for advanced melanoma has changed markedly during the past 10 years. Novel agents such as molecularly-targeted therapies specifically inhibiting carcinogenic pathways and immunotherapies augmenting the antitumor immunity have emerged as new standards of care for patients with melanoma [3].

As elegantly reviewed by Wong et al, ^{18}F -fluorodeoxyglucose (FDG) positron emission tomography (PET) could play a pivotal role in the interpretation of therapeutic response following BRAF inhibition and immunotherapy in patients with melanoma, facilitating early assessment of drug resistance and detection of life-threatening autoimmune side effects [4]. ^{18}F -FDG PET was also recommended for staging and detecting recurrent melanoma [5, 6]. However, since ^{18}F -FDG PET monitors cellular metabolism and immunotherapy elicits a natural inflammatory response, traditional PET imaging using ^{18}F -FDG has proven inadequate in examining responses to immunotherapy in certain cancer types [7]. Moreover, owing to increased glucose metabolism in inflammatory tissues, ^{18}F -FDG displays relatively poor selectivity for distinguishing tumor from inflammatory tissue. Importantly, in one study, ^{18}F -FDG PET failed to detect melanoma in patients showing a positive sentinel lymph node biopsy for cancer, and no recurrent melanoma occurred where the ^{18}F -FDG PET scans were positive or suspicious [8]. ^{18}F -FDG PET scans also failed to detect pulmonary and brain metastases, indicating its limited value when staging patients with more advanced melanoma [9]. Currently, gadolinium-enhanced MRI is the most sensitive and reproducible method available to measure brain metastases or to assess treatment response [10, 11].

In the era of precision medicine and molecular imaging [12], many radiolabeled probes for imaging different molecular targets or biochemical processes have been designed and evaluated for melanoma imaging. Although clinically-available ^{111}In -DOTA-lanreotide and ^{111}In -DOTA-Tyr3-octreotide may image melanoma, the detection rates may not meet the clinical requirement and their mechanisms remain unknown [13]. In the past, we and others organized reviews regarding anatomical and molecular imaging of skin cancer [14, 15]; since then, substantial amounts of molecular imaging probes for melanoma have been developed. Although nanoparticle-based multimodality imaging systems have been intensively applied to melanoma detection and therapy, it is beyond the scope of our current paper. Therefore, with an emphasis on the small molecule- and peptide- based PET imaging probes, we aim to systematically review the recent advances in the field and extend an outlook on future development. Given the multidisciplinary nature of this field, our present review is by no means exhaustive, but we intend to summarize the most recent advances for scientists to further refine these probes and for clinicians to push the clinical transition of those which are promising, facilitating better management of patients with melanoma in the foreseeable future.

2. Small Molecule-based Probes Targeting Melanin

Melanin, an amorphous, irregular, functional biopolymer and a ubiquitous natural pigment in many organs including human skin, is a source of novel research opportunities in the

fields of biomedicine, nanotechnology, and materials science [16]. In malignant melanoma, melanin formation is highly increased because tyrosinase activity is significantly elevated [17, 18], making it a very attractive theranostic target. Many drugs, including methylene blue (MTB), chloroquine, acridine orange, benzamide (BZA) and its analogs, and other aromatic compounds, have been found to bind to melanin both in vivo and in vitro [19–24]. Melanin-targeting prodrugs also have been developed and used for in vivo melanoma imaging [25, 26].

2.1. Radiolabeled benzamide and benzamide derivatives

Various versions of radiolabeled benzamide and its analogs have been developed for targeting melanin in clinical practice [27–33]. Specifically, as evidenced by a recent prospective and multicenter phase III clinical study, ^{123}I -BZA₂ (Fig. 1A) [30, 34], a benzamide derivative able to bind to melanin pigment in melanoma cells, had statistically higher specificity than ^{18}F -FDG for diagnosis of melanoma metastases in a lesion-based analysis [35]. Katsifis and co-authors substituted the benzamide moiety with a nicotinamide moiety and labeled the compound with no-carrier-added ^{123}I -iodine via iododestannylation reactions. Biodistribution showed that 66% of the injected ^{123}I -1 (^{123}I -MEL008, Fig. 1B) was excreted from the urinary system at 1 h postinjection, also displaying the highest tumor uptake at that time point mainly because of its enhanced hydrophilicity and rapid clearance via renal excretion [36]. Chezal et al. then examined biological properties of aromatic or heteroaromatic BZA analogs in B16 melanoma-bearing mice and found that melanin-specific binding compound ICF01012, when labeled with ^{131}I or ^{125}I , could be applied in radionuclide diagnosis and therapy of disseminated melanoma [20, 37–39]. Furthermore, the authors developed a new multimodal approach by designing iodinated and fluorinated analogs of ICF01012, of which ^{18}F -8 emerged as the most promising compound and demonstrated high tumor uptake, high contrast, and rapid clearance [40]. These above-mentioned efforts also led to the development of ^{123}I -MEL037 and ^{123}I -53 (Fig. 1C, D); while the former demonstrated high and prolonged tumor uptake, the latter was derived from structural modification of ^{123}I -MEL037 and performed even better than ^{123}I -ICF01012, as the tumor-to-background uptake ratios of ^{123}I -53 and ^{123}I -ICF01012 were 31.9 and 18.5, respectively [41, 42]. While the above probes accumulated at high levels in the eyes and thyroid, radioiodinated iochlonicotinamide and radioiodinated phenylacetamides (^{131}I -IHPA and ^{131}I -IHPP) had lower uptake in most normal organs and highly specific uptake in the melanotic tumor (Fig. 1E, F) [43, 44].

Since ^{18}F -FBZA exhibited high tumor uptake and emerged as a promising candidate for clinical study (Fig. 2A) [45], Wu et al. modified the phenol moiety of benzamide with a short chain PEG and then labeled with ^{18}F to obtain ^{18}F -FPBZA (Fig. 2B). In vivo, it showed excellent tumor-to-background contrast, and this radiotracer was able to distinguish tumor from inflammatory tissue as turpentine-induced inflammation revealed low radioactivity accumulation [46]. Garg and co-authors developed 4- ^{11}C -MBZA and reported that this probe displayed advantages over its ^{18}F analogs while delivering a lower radiation dose to the subject (Fig. 2C). In vitro binding studies showed specific binding of 4- ^{11}C -MBZA to B16/F1 cells; however, it also accumulated to a high level in the kidneys [47]. Chang et al. developed ^{18}F -NOTA-BZA, and the melanin-specific binding ability, low bone

uptake, sustained tumor retention, high hydrophilicity ($\log P = -1.96$), fast normal tissue clearance and low radiation burden indicated that ^{18}F -NOTA-BZA is a promising PET probe for melanin-specific imaging of melanin-positive melanoma [48]. Based on the inspiring and promising preclinical results of ^{18}F -MEL050 (Fig. 2D) [49, 50], Liu et al. successfully synthesized a series of ^{18}F -MEL050 analogs for melanoma imaging (Fig. 2E, F), of which ^{18}F -2 showed superior tumor-targeting efficacy and imaging properties [51]. Interestingly, recent research proposed that N-(2-diethylaminoethyl) rather than the aromatic ring structure in benzamide analogs is a plausible pharmacophore responsible for melanin targeting, as the synthesized probe ^{18}F -FPDA exhibited relatively high B16/F10 tumor-targeting efficacy and favorable in vivo pharmacokinetics (Fig. 2G) [52]. Besides the most commonly used PET radionuclides, ^{68}Ga ($t_{1/2} = 68$ min) is an outstanding radioisotope for molecular imaging due to its significant 89% positron yield and its availability without the establishment of expensive cyclotron and synthesis models [53]. Trencsényi et al. conjugated PCA with two different chelators, HBED-CC and NODAGA, then labeled the compounds using Ga-68 and investigated the diagnostic value of ^{68}Ga -HBED-CC-PCA and ^{68}Ga -NODAGA-PCA in vitro and in vivo. The authors found that uptake of ^{68}Ga -NODAGA-PCA by melanin-containing melanoma was significantly higher than the accumulation of the ^{68}Ga -HBED-CC-conjugated PCA [54, 55].

2.2. Imaging of melanoma metastases

Considering that the presence of distant metastases, especially brain metastases, confers worse prognosis for patients with melanoma, their early detection is critical [56]. In a study comparing diagnostic values of ^{18}F -FDG PET/CT and MRI in melanoma patients with palpable lymph node metastases, Aukema et al. found that ^{18}F -FDG PET/CT changed the intended regional node dissection in 26 patients (37%) and resulted in a superior diagnostic accuracy of 93%, but missed 5 patients with brain metastases which were detected by MRI [57]. Other study also demonstrated that ^{18}F -FDG PET failed to detect metastatic lesions of less than 1 cm located in the lung, liver or brain [58]. Currently only contrast-enhanced MRI and ^{18}F -FET PET seem to be reliable methods to detect brain metastases from melanoma but still lack specificity [10, 59]. Moreover, in patients with surgically treatable IIIC and IV metastatic melanoma following targeted/immunotherapy, PET/CT can detect unexpected metastases that are missed with conventional imaging, and can be considered as part of preoperative workup [4, 60, 61]. Thus it is of great importance to develop novel radiotracers to identify occult lesions or distant small metastases from melanoma with high specificity and a low false positive rate. Notably, the ability of an imaging agent to cross the blood-brain barrier (BBB) is considered critical to effectively target metastatic lesions in the brain. Of the reported probes, 4- ^{11}C -MBZA was able to cross the BBB and the corresponding uptake was moderate in the normal brain [47]. As observed from biodistribution and PET studies, 4- ^{11}C -MBZA uptake in normal tissues was noticeably lower than that for several other ^{18}F -benzamides like ^{18}F -FPBZA [46] and ^{18}F -DAFBA [62]. In addition, newly developed radiotracers, such as ^{18}F -FBZA, ^{18}F -5-FPN, ^{18}F -MEL050, ^{18}F -FITM and ^{18}F -ICF01006 (Fig. 2H), may have better performance in the delineation of small lymph node and lung metastases from melanoma than that of ^{18}F -FDG PET/CT [45, 46, 63–66]. ^{18}F -5-FPN, a probe identical to ^{18}F -2, successfully detected pigmented B16/F10 tumors as early as 1 min after injection of the tracer. The uptake increased over time and the tracer was rapidly

excreted via the kidneys. This and later studies from the same group further validated the potential of ^{18}F -5-FPN PET for the early detection of metastatic melanoma lesions (Fig. 2I) [63, 67].

^{18}F -MEL050 had excellent retention in melanin-containing tumors and rapid background clearance [49]; however it is notable that the route of administration of ^{18}F -MEL050 matters when imaging regional lymph node metastasis from melanoma. While ^{18}F -MEL050 PET correctly identified 100% of the lymph node metastases after subcutaneous administration of the tracer, only 60% of those metastases were found after systemic administration of the tracer in the lateral tail vein [50].

3. Peptide-based imaging probes

Peptides are emerging as potent and selective ligands that can be designed to bind with high affinity and specificity to cell surface receptors on a wide range of tumors [68]. Three major types of peptides, namely α -Melanocyte-stimulating hormone (α -MSH), tumor angiogenesis associated integrins, and peptides targeting both MC1R and integrin, are under intensive development for molecular imaging of melanoma.

3.1. α -Melanocyte-stimulating hormone (α -MSH)-based probes

α -MSH, a ligand specific for melanocortin receptor subtype 1 (MC1R), has been reported to be overexpressed in both melanotic and amelanotic human melanoma cases and has been widely used as a vehicle for melanoma-targeted imaging and therapy [69–73]. As native α -MSH (a linear 13 amino acid peptide, Ac-Ser-Tyr-Ser-Met-Glu-His-Phe-Arg-Trp-Gly-Lys-Pro-Val-NH₂) has a biological half-life of less than 3 minutes in vivo [74], tremendous work has been done in the past 20 years and several modified analogs and synthesis strategies have been developed in an effort to add biological stability and improve targeting. For example, substitution of Met⁴ with Nle⁴ and Phe⁷ with D-Phe⁷ yields NDP- α -MSH [75]. Since His⁶-Phe⁷-Arg⁸-Trp⁹ had been identified as the “essential core” of native α -MSH peptide [76], both linear and transition metal rhenium cyclized α -MSH such as NAPamide [77], DOTA-NAPamide [78], ReCCMSH [77], MTII [79], analogs of MTII [80], DOTA-CycMSH and DOTA-GlyGlu-CycMSH [81], DOTA-Nle-CycMSH_{hex} [82], were constructed. Recently new highly-specific and selective ligands against MC1R for melanomas were also developed and have been explored as platforms for molecular imaging of melanoma [83, 84].

Using the previously synthesized ReCCMSH which has nanomolar affinity for MC1R, McQuade et al. labeled the peptide with PET isotopes ^{64}Cu and ^{86}Y and assessed their diagnostic efficacy in melanoma tumors. Biodistribution studies revealed that the tumor concentration of ^{86}Y -DOTA-ReCCMSH and ^{64}Cu -DOTA-ReCCMSH was two times higher compared to that of the metabolic agent ^{18}F -FDG [85]. Additionally, the use of the chelator CBTE2A provided improved stability, as uptake of ^{64}Cu -CBTE2A-ReCCMSH was significantly lower than that for ^{64}Cu -DOTA-ReCCMSH in normal organs such as liver, lung, heart, and spleen [86]. Wei et al. then successfully labeled DOTA-ReCCMSH (Arg¹¹) with ^{68}Ga and reported that the tumor uptake of ^{68}Ga -DOTA-ReCCMSH reached a maximum after 30 min and remained stable 2 h postinjection. A pretargeting strategy

employing D-lysine administration was able to significantly reduce kidney retention of the tracer [87]. ^{18}F -FB-RMSH-1 and ^{18}F -FP-RMSH-1, another two metallopeptide Ac-D-Lys-ReCCMSH(Arg¹¹) probes [88, 89], showed less optimal imaging properties than these three probes illustrated above.

Since ^{111}In -DOTA-GlyGlu-CycMSH was first developed to target MC1 receptors for primary and metastatic melanoma imaging [81, 90], recent studies determined that reduction of the ring size of ^{111}In -DOTA-GlyGlu-CycMSH, L-lysine co-injection, introduction of a -GG-linker, substitution of DOTA with NOTA, and $^{99\text{m}}\text{Tc}$ radiolabeling via new chelators may further increase high melanoma tumor uptake while reducing nonspecific kidney and liver uptake [82, 91–94].

In addition, MC1-R specific NAPamide analogs have been labeled with various radiometals and non-metallic radionuclides (such as ^{64}Cu , ^{68}Ga , ^{18}F , ^{111}In , $^{99\text{m}}\text{Tc}$ and ^{44}Sc) and have been intensively used to detect melanomas or to evaluate the expression level of MC1-R [95–101]. While ^{64}Cu -DOTA-NAPamide showed mild tumor uptake in B16/F10 xenografted melanoma (Fig. 3A) [96], ^{64}Cu -NOTA-GGNle-CycMSH_{hex} showed dramatic uptake at 2 h postinjection [93]. The latter study also elucidated that the substitution of DOTA with NOTA dramatically increased the melanoma uptake and decreased the renal and liver uptake of ^{64}Cu -NOTA-GGNle-CycMSH_{hex}.

Due to its positron emission with a high branching ratio ($I = 94.27\%$, $E_{\text{mean}}(\beta^+) = 0.63$ MeV), convenient production by the $^{44}\text{Ti}/^{44}\text{Sc}$ generator and satisfactory physical half-life (3.97 h), ^{44}Sc is a novel radiometal which has gained significant interest as a potential radioisotope for PET imaging [102–104]. A proof-of-concept study investigated the biological properties of the ^{44}Sc -labeled DOTA-NAPamide and found that this probe showed excellent binding properties to MC1-R positive melanoma cell and tumors, slightly superior to that of ^{68}Ga -DOTA-NAPamide [95].

Most recently, three new MC1R-targeting peptides (CCZ01047, CCZ01048, and CCZ01056) were successfully developed and all the three ^{68}Ga -labeled tracers produced high contrast PET images in B16/F10 tumors, of which ^{68}Ga -CCZ01048 exhibited the most ideal tumor uptake (Fig. 3B). This study indicated that introduction of a cationic Pip linker to Nle-CycMSH_{hex} could improve tumor uptake and tumor-to-normal tissue contrast in detecting melanoma [105].

3.2. Peptide-based probes targeting the integrin family

Integrins are heterodimeric $\alpha\beta$ transmembrane receptors that connect the extracellular matrix (ECM) to the cytoskeleton, always forming dimers by combining 1 of 18 α -chains with 1 of 8 β -chains [106]. Integrin $\alpha_v\beta_3$, and less commonly integrin $\alpha_5\beta_1$, have been attractive molecular targets for developing melanoma imaging probes [107–111]. Clinically, recent studies have validated that ^{18}F -Fluciclatide (formerly known as ^{18}F -AH111585) and ^{18}F -Galacto-RGD were able to measure $\alpha_v\beta_3$ expression in melanoma and may be useful radiotracers to assess the response to the antiangiogenic therapy in melanoma patients [112–116]. Nevertheless, low concentration of these clinically-available probes in $\alpha_v\beta_3$

positive tumors and less than optimal PET/CT imaging quality warrants many sensitive and specific probes to be developed.

In the preclinical setting, as we previously reviewed [117], a large variety of imaging strategies have been successfully employed for imaging of integrin expression in various cancer types, including melanoma. One study compared the diagnostic efficacy of ^{18}F -Galacto-RGD with that of ^{68}Ga -DOTA-RGD and ^{111}In -DOTA-RGD and found that ^{18}F -Galacto-RGD remained superior for imaging $\alpha_v\beta_3$ expression [108]. Even though ^{68}Ga -Oxo-DO3A-RGD and ^{68}Ga -NS3-RGD turned out to have inferior characteristics compared to the already existing ^{68}Ga -labeled RGD peptides [118], ^{68}Ga -NODAGA-RGD possessed improved imaging properties compared to ^{68}Ga -DOTA-RGD [119], even performing similarly to ^{18}F -Galacto-RGD (Fig. 3C) [120]. To achieve an easier and more rapid radiosynthesis, fusarinine C (FSC) and SarAr have been reported to be promising Ga-68 and Zr-89 binding bifunctional chelators [110, 121–124]. The most recent study from Zhai et al. reported that ^{68}Ga -FSC(succ-RGD)₃ exhibited improved properties compared to ^{68}Ga -NODAGA-RGD. The half-life of the radionuclide used (^{68}Ga , 68 min) was compatible with the pharmacokinetics of RGD peptides [125].

When compared to DOTA, the bifunctional chelator NODAGA has gained popularity because of its significant advantages in terms of labeling chemistry [119, 126]. However, the TRAP chelator possesses even better ^{68}Ga labeling properties and enables high yields and excellent reproducibility [127–129]. Notni et al. synthesized ^{68}Ga -avebetrin (formerly known as ^{68}Ga -TRAP(RGD)₃) and performed a comparison of biodistribution and PET data of ^{68}Ga -TRAP(RGD)₃ with those of ^{68}Ga -NODAGA-c(RGDyK) and ^{18}F -Galacto-RGD. Different from ^{68}Ga -NODAGA-RGD and ^{18}F -Galacto-RGD, ^{68}Ga -TRAP(RGD)₃ showed a very rapid blood clearance and renal excretion while maintaining activity concentration in tumor tissue, indicating the potential value of ^{68}Ga -TRAP(RGD)₃ as a next generation $\alpha_v\beta_3$ imaging agent (Fig. 3D) [130]. Considering that the TRAP-type chelator NOPO showed excellent ^{68}Ga labeling properties even in the presence of high concentrations of competing metal cations (Fig. 3E) [131], researchers further developed ^{68}Ga -NOPO-c(RGDfK) and found that this $\alpha_v\beta_3$ targeting probe exhibited a higher degree of hydrophilicity than similar conjugates with other chelators, resulting in rapid and specific uptake in M21 tumor xenografts, very rapid pharmacokinetics and renal clearance (Fig. 3F, G) [132].

There are a few reports on the development of $\alpha_5\beta_1$ specific radiotracers [111, 133–135]. Although there was one candidate peptide with a high specificity and affinity for $\alpha_5\beta_1$ in vitro, in vivo biodistribution studies demonstrated this radiotracer was not suitable for in vivo imaging due to its considerably high and constant radioactivity accumulation in the blood and other major organs [136]. The reported $\alpha_5\beta_1$ specific probes ^{18}F -PR_b and ^{68}Ga -NODAGAFR366 showed specific but low binding to $\alpha_5\beta_1$ positive murine melanoma tumors, and as a result, the poor tumor concentration and high kidney uptake may hinder these probe from further clinical transition [111, 137]. Recently Notni and colleagues obtained ^{68}Ga -aquibepirin by click-chemistry (CuAAC) trimerization of a $\alpha_5\beta_1$ pseudo-peptide on the TRAP chelator, followed by automated ^{68}Ga labeling. Surprisingly, the trimer ^{68}Ga -aquibepirin possessed approximately 16-times-higher $\alpha_5\beta_1$ affinity than the previously reported ^{68}Ga -labeled pseudo-peptide monomer. Although ^{68}Ga -aquibepirin

showed lower uptake than ^{68}Ga -avebetrin, low background activity and high target-to-nontarget contrast of the probe may offer great potential for elucidating biologic functions of $\alpha_5\beta_1$ and for detecting melanoma [138, 139].

RGD mimetic integrin inhibitors, like Cilengitide and Cilengitide-like RGD peptidomimetics, have also been investigated as chemical probes for the molecular imaging of angiogenesis in the literature [133, 140, 141]. In the near future, other strategies like sulfonation of tyrosine moieties in RGD peptides, which can modify the hydrophilicity of RGD peptides to increase renal clearance and to improve overall biodistribution [142], can also be applied to design novel probes for mapping $\alpha_v\beta_3$ expression in melanoma. Notably, RGD can be used as an agent for surface engineering in other imaging systems to enhance melanoma targeting efficacy and therapeutic effects [143–146].

3.3. Dual-targeted peptide-based probes

To improve the in vivo pharmacokinetics and stability of the above mentioned molecular probes, many hybrid peptides targeting both MC1R and integrin $\alpha_v\beta_3$ have been developed [147–153]. Initial synthesis and evaluation of $^{99\text{m}}\text{Tc}$ -RGD-Lys-(Arg¹¹)CCMSH showed MC1R-mediated cellular uptake of the tracer, higher tumor uptake, and prolonged tumor retention in B16/F1 melanoma-bearing mice [148]. Substitution of Gly with Ala in the hybrid peptide not only dramatically increased the MC1R binding affinity of RAD-Lys-(Arg¹¹)CCMSH compared to RGD-Lys-(Arg¹¹)CCMSH (0.3 vs. 2.0 nM) but also enhanced the melanoma uptake [149]. Further studies demonstrated that $^{99\text{m}}\text{Tc}$ -RTD-Lys-(Arg¹¹)CCMSH and $^{99\text{m}}\text{Tc}$ -RVD-Lys-(Arg¹¹)CCMSH exhibited similar imaging properties to RAD-Lys-(Arg¹¹)CCMSH, but $^{99\text{m}}\text{Tc}$ -RVD-Lys-(Arg¹¹)CCMSH reached its highest concentration in melanoma lesions at a later time point [150].

Flook et al. replaced the Gly with another four amino acids (Ser, Nle, Phe, and D-Phe) and found that $^{99\text{m}}\text{Tc}$ -RSD-Lys-(Arg¹¹)CCMSH displayed the strongest MC1R binding affinity in vitro and exhibited the most optimal melanoma uptake in vivo [151]. Importantly, linkers and charge status of the linkers between hybrid peptides may affect the renal uptake of the tracer significantly, as $^{99\text{m}}\text{Tc}$ -RGD-(Arg¹¹)-CCMSH without a linker dramatically enhanced the tumor-to-kidney uptake ratio [154], and substitution of the Lys linker with Aoc, PEG2, β Ala or Ahx linker reduced the renal uptake of the relevant probe by 58%~63% at 2 h post-injection [155–157].

4. Other PET/SPECT probes for melanoma imaging

Besides radiolabeled melanin, MCR1, and integrin based probes, here we would like to summarize other potential radiolabelled molecular imaging probes that have been found effective in detecting not only melanoma but also other solid tumors. These probes can be divided into several categories as discussed in the following sections.

4.1. Probes targeting the metabotropic glutamate 1 receptor

Metabotropic glutamate 1 (mGlu1) receptor is a G protein-coupled receptor normally expressed in the central nervous system, essential for learning and modulating the excitatory synaptic transmission in the central nervous system [158]. Recent reports have elucidated

that mGlu1 has oncogenic characteristics in melanoma by driving constitutive activation of mitogen activated protein kinase and phosphatidylinositol-3-kinase/protein kinase B pathways [159–163]. Xie et al. initially developed ^{18}F -FITM for quantifying mGlu1 expression in the brain [164, 165], and then successfully extended ^{18}F -FITM PET imaging in melanoma [65]. However, considerable uptake and slow clearance of radioactivity in the brain undermined its usage in clinical applications. The same group elaborately introduced halogen atoms (chlorine, bromine, or iodine) rather than ^{18}F into ^{18}F -FITM and labeled these compounds using ^{11}C (half-life: 20.2 min). Of the reported compounds, the iodine analogue ^{11}C -6 showed the highest ratio of radioactivity of tumor to brain and may act as a useful PET tracer for imaging mGlu1 in melanoma (Fig. 4A, B) [166]. Notably, future studies can feasibly label ^{11}C -6 using the isotopes of ^{124}I , ^{123}I or ^{131}I for mGlu1-based theranostics of melanoma without altering their chemical structures or pharmacological profiles.

4.2. Probes targeting the very late antigen-4

In the past several years, very late antigen-4 (VLA-4; also called integrin $\alpha_4\beta_1$) has been found in cancers including melanoma [167]. LLP2A, a high-affinity peptidomimetic ligand for VLA-4, was identified from a 1-bead 1-compound library and has been used for cancer imaging and therapy [168–170]. Specifically, Jiang and co-authors conjugated LLP2A with two different chelators and assessed their imaging properties in melanoma models after labeling the conjugated compounds with ^{64}Cu . From the biodistribution and in vivo imaging data, both ^{64}Cu -CB-TE1A1P-LLP2A and ^{64}Cu -CB-TE2A-LLP2A clearly visualized the tumors with better contrast than was observed for ^{64}Cu -CB-TE1A1P-LLP2A [171]. When compared to ^{68}Ga -labeled NODAGA-LLP2A, ^{64}Cu -CB-TE1A1P-PEG4-LLP2A trended toward higher uptake and better tumor-to-nontarget tissue ratios (Fig. 4C, D) [172].

Recently Gai et al. designed a new probe ^{64}Cu -NE3TA-PEG4-LLP2A using the newly discovered chelator p-SCN-PhPr-NE3TA and assessed the diagnostic efficacy of the probe in melanoma xenografts. Small animal PET/CT imaging with ^{64}Cu -NE3TA-PEG4-LLP2A demonstrated high uptake with a superior tumor-to-muscle ratio at 4 h postinjection [173]. In addition, a study from Beaino et al. reported that ^{177}Lu -DOTA-PEG₄-LLP2A could accumulate not only in primary melanoma but also in the metastatic lesions in the lung and brain, demonstrating the potential of ^{177}Lu -DOTA-PEG₄-LLP2A as an alternative treatment strategy for metastatic VLA-4 expressing melanoma [174].

4.3. Probes targeting indoleamine 2,3-dioxygenase (IDO)

In recent years, immunotherapy has dramatically changed the landscape of melanoma treatment [175]. However, recent findings revealed that indoleamine 2,3-dioxygenase (IDO) can be triggered by innate responses during tumorigenesis, and also by attempted T cell activation (either spontaneous or due to immunotherapy), contributing to restrain immunity and establish immunotolerance. IDO inhibitors act as a novel class of immunomodulators with broad application in the treatment of advanced human cancer [176, 177]. In an effort to map tryptophan (Trp, a substrate of IDO) metabolism [178, 179], one group found that radionuclide labelled IDO inhibitor, 5-[^{18}F]F-AMT, may hold great potential for melanoma imaging [180]. By using B16/F10 melanoma model, the authors found that tumors were clearly visible and this probe was possibly excreted through the urinary system as the

kidneys showed initial high uptake (Fig. 4E, F). These preliminary results implied potential usage of this series of probes because the superior tumor-to-background ratio may precisely delineate primary and metastatic melanomas, but these probes are not melanoma-specific because other tumor types like breast cancer concentrate this kind of probe as well [178].

4.4. Probes targeting the immune checkpoints

With the field of cancer immunotherapy has undergone tremendous growth during the past decade, as we recently pointed out, noninvasive molecular imaging strategies have been used to map the biodistribution of immune checkpoint molecules, monitor the efficacy and potential toxicities of the treatments, and identify potential patients who will benefit from immunotherapies [181]. Studies have shown that PET may be used for the noninvasive imaging of PD-1/PD-L1 expression in melanoma and for determining the extent of tumor-infiltration of lymphocytes [182, 183]. Recently Nedrow et al. developed a new PD-L1-targeted imaging agent, ^{111}In -DTPA-anti-PD-L1-BC and demonstrated that tumors had the greatest uptake at 24 h p.i. with a tumor-to-muscle ratio of 4.6 in mice bearing B16/F10 tumors. Whole body SPECT images demonstrated that the tumor as well as the spleen and liver were clearly defined at the later time points [184].

4.5. Probes targeting the human copper transporter 1 (CTR1)

Human copper transporter 1 (CTR1), a 190-amino acid protein of 28 kDa with three transmembrane domains [185], has been proven to be overexpressed in melanoma. The usefulness of $^{64}\text{Cu}^{2+}$ ions as PET probes is based on the fact that Cu is an essential element which plays an important role in cell proliferation and angiogenesis [186]. Therefore, copper radionuclide-based imaging of cancers have been investigated by several studies [187, 188]. $^{64}\text{CuCl}_2$ has been reported to be a novel and promising PET probe for imaging melanoma [189]. However, there have been reports that CTR1 is the specific influx copper transporter for $^{64}\text{Cu(I)}$ rather than $^{64}\text{Cu(II)}$. Using antioxidants, Jiang et al. prepared $^{64}\text{Cu(I)}$ and evaluated cellular uptake of $^{64}\text{Cu(I)}$ and $^{64}\text{Cu(II)}$ by melanoma cells *in vitro* and *in vivo*. The authors demonstrated that although $^{64}\text{Cu(I)}$ exhibited higher cellular uptake, no significant difference between $^{64}\text{Cu(I)}$ and $^{64}\text{Cu(II)}$ was observed through *in vivo* PET images and biodistribution [190]. However, future studies are still needed to evaluate whether or not $^{64}\text{Cu(I)}$ can act as a feasible PET imaging radiotracer for melanoma detection.

4.6. Antibody-based imaging probes

Radiolabeled monoclonal antibodies (mAb), antibody fragments, and engineered antibody derivatives are increasingly utilized as agents in diagnosis and therapy because of developments in antibody engineering and *in vivo stability* and high specificity of these probes [191, 192]. Twenty years have passed since the initial attempts to detect melanoma using radiolabeled antibodies [193, 194]. Besides melanin-specific antibodies [195–199], GD2 is highly expressed on the cell surface of a broad spectrum of human cancers including melanoma and has been successfully exploited as a molecular target for therapy and molecular imaging [200]. Voss et al. successfully developed a SarAr-conjugated, ^{64}Cu -labeled, anti-GD2 antibody construct, ^{64}Cu -SarAr-GD2 mAb ch14.18, and performed *in vivo* studies using GD2-expressing melanoma xenografts. Their results showed that about

20.5% of the injected dose accumulated in the M21 melanoma tumors [201], and further studies from the same group confirmed this finding [202, 203]. These preliminary results may indicate the feasibility of the ^{64}Cu -SarAr antibody as a platform for imaging melanoma.

4.7. Probes targeting CXCR4

C-X-C chemokine receptor type 4 (CXCR4, also called fusin, CD184) is a 7-transmembrane G-coupled receptor belonging to the chemokine receptor family and has been found to be overexpressed in various human cancers including lymphoma, neuroendocrine tumors, malignant glioma, lung cancer and multiple myeloma [204–208]. CXCR4-based imaging has also been investigated to detect melanoma recently [209]. In a clinical trial dedicated to estimate CXCR4 overexpression by using the novel CXCR4-specific probe ^{68}Ga -Pentixafor, Vag et al. included 21 patients (2 of them melanoma patients) and demonstrated the feasibility of ^{68}Ga -Pentixafor for PET imaging of solid malignancies, although the detectability of solid cancers by ^{68}Ga -Pentixafor seemed to be lower than with ^{18}F -FDG PET [210].

5. Image-guided therapy of melanoma

In addition to melanoma imaging, molecular imaging-guided therapy of melanoma has long been a hot topic in the field, as recently reviewed by Norain et al. [211]. Radionuclide-based therapy of melanoma can be realized through radiolabeled antibodies, peptides or small molecules.

5.1. Radiolabeled antibodies and peptides

Lutetium-177 (^{177}Lu) is a low energy β -emitter (497 keV, 90%) with a half-life of 6.7 days and a maximum tissue penetration of 1.6 mm. ^{177}Lu also emits γ -rays (113 and 208 keV, 6% and 11%) suitable for image-guided drug delivery using SPECT. Antibodies and peptides radiolabeled with ^{177}Lu are attractive therapeutic agents due to localized deposition of beta decay energy and a radioactive half-life which matches in vivo pharmacokinetics of targeting antibodies quite well [212–214]. Vascular endothelial growth factor (VEGF) has been extensively studied as one of the most important proteins involved in the development of physiological and pathological angiogenesis [215, 216]. Recently, ^{177}Lu -DOTA-bevacizumab (a recombinant humanized monoclonal antibody that binds to all VEGF isoforms) has shown preliminary potential as a novel radioimmunotherapy and molecular imaging agent for melanoma [217]. Building upon the success of the lactam bridge-cyclized a-MSH peptides for melanoma imaging discussed above, Guo et al. assessed the image-guided therapeutic effect of ^{177}Lu -DOTAGGNIe-CycMSH_{hex} in B16/F1 melanoma-bearing mice and found high melanoma uptake and fast urinary clearance of the probe, underscoring its potential as a theranostic agent for metastatic melanoma [218].

^{188}Re , a high-energy β -emitter (maximal energy: 2.12 MeV), has considerable range (several millimeters) in tissue and a relatively short half-life of 16.9 h. ^{188}Re -labeled melanin-specific antibodies for melanoma therapy have also been studied. ^{188}Re -labeled 6D2 [197], a melanin-binding IgM antibody, has been validated to be effective in treating pigmented human melanoma tumors and in augmenting the efficacy of the chemotherapeutic

agent dacarbazine (DTIC) [198]. Notably, by administering the targeting vector and radioisotope separately, in vivo pretargeting seems to be a promising approach to enhance tumor-targeting properties of radiolabeled antibodies while simultaneously skirting their pharmacokinetic limitations.

5.2. Radiolabeled small molecules

For melanin-targeted imaging-guided radionuclide therapy, many radiolabeled benzamide derivatives such as ^{131}I -MIP-1145 and ^{131}I -ICF01012 exhibited strong efficacy in murine and human melanoma xenografts [17, 219, 220]. ICF01012 was labeled with ^{123}I for melanoma imaging and with ^{131}I for melanoma treatment (Fig. 5A,B). The preliminary results not only showed a correlation between radiotracer uptake and melanin content but also demonstrated significantly reduced tumor growth and prolonged the median survival of the melanoma-bearing mice after administration of ^{131}I -ICF01012 [221]. The combination of ^{131}I -ICF01012 and coDbait, a DNA repair inhibitor, could overcome melanoma radioresistance and increase the efficacy of targeted radionuclide therapy (TRT) without increasing side effects [222]. Clinically, in a study which enrolled 26 patients with metastatic melanoma, Mier et al. reported that, out of five patients who received higher doses of treatment by administration of the melanin-binding ^{131}I -BA52 (Fig. 5C,D), three of them survived more than two years after therapy. In contrast, the mean overall survival of the untreated and insufficiently dosed patients was approximately three months [223]. Interestingly, iodinated and fluorinated radiotracers targeting melanin and offering potential for both diagnosis (SPECT and PET imaging) and therapy (iodine-131) have also been developed [224–226].

6. Conclusion and future perspectives

In this review, as summarized in Fig. 6, we provided an informative survey of the recent progress on radiolabelled molecular imaging probes for imaging different molecular targets or processes in malignant melanoma. While melanin, MCR1 and integrins are the traditional targets extensively used for melanoma detection and therapy, tumor metabolism and immune microenvironment-based molecular imaging probes have also been developed in recent years. In the era of precision medicine, devotion and efforts on radiolabelled molecular probes for mapping and treating melanoma is extremely important, and new imaging probes with optimal imaging properties are still highly demanded.

Looking into the future, we would like to put forward three proposals from our perspective. First, with the rapid development of the molecular imaging field, novel molecular imaging probes with high sensitivity and specificity enable us to characterize melanoma at the molecular level. These new probes will provide powerful platforms for early diagnosis of both primary and metastatic melanoma, monitoring of therapeutic response, accurate staging and restaging of melanoma, stratification of patients for antiangiogenesis therapy, immunotherapy and/or radionuclide therapy, and facilitation of new drug discovery for melanoma treatment. Second, melanin, MCR1, and integrins have been intensively investigated as targets for selective imaging and therapeutic agents against melanoma, and though the potential of many of these probes could be effectively demonstrated in preclinical

settings, very few of them could actually be translated to the clinics. Future studies should be dedicated to push the clinical transition of some of these most promising probes. Third, usage of humanized mice, rather than cultured cell lines and mouse xenografts [227], could examine the molecular imaging probes in the context of the human immune system and tumor microenvironment and accelerate the clinical transition of these molecular imaging probes.

To conclude, malignant melanoma represents a serious public health problem and is a deadly disease when diagnosed at late stage. With the elucidation of many important oncogenic signaling pathways and biomarkers involved in malignant melanoma pathogenesis, we believe that PET and SPECT imaging as well as image-guided therapy can undoubtedly provide better visualization and management of malignant melanoma in the forthcoming future.

Acknowledgments

This work was sponsored by Ph.D. Innovation Fund of Shanghai Jiao Tong University School of Medicine to Weijun Wei (No. BXJ201736), National Institutes of Health (1R01CA205101, P30CA014520, and T32GM008505) and the American Cancer Society (125246-RSG-13-099-01-CCE).

References

1. American Cancer Society. [Internet]<http://www.cancer.org/cancer/skincancer-melanoma/detailedguide/melanoma-skin-cancer-key-statistics>
2. Trinh VA. Current management of metastatic melanoma. *American journal of health-system pharmacy : AJHP : official journal of the American Society of Health-System Pharmacists*. 2008; 65:S3–8. DOI: 10.2146/ajhp080460
3. Luke JJ, Flaherty KT, Ribas A, Long GV. Targeted agents and immunotherapies: optimizing outcomes in melanoma. *Nature reviews Clinical oncology*. 2017; doi: 10.1038/nrclinonc.2017.43
4. Wong AN, McArthur GA, Hofman MS, Hicks RJ. The Advantages and Challenges of Using FDG PET/CT for Response Assessment in Melanoma in the Era of Targeted Agents and Immunotherapy. *European journal of nuclear medicine and molecular imaging*. 2017; doi: 10.1007/s00259-017-3691-7
5. Krug B, Crott R, Lonneux M, Baurain JF, Pirson AS, Vander Borgh T. Role of PET in the initial staging of cutaneous malignant melanoma: systematic review. *Radiology*. 2008; 249:836–44. DOI: 10.1148/radiol.2493080240 [PubMed: 19011184]
6. Danielsen M, Kjaer A, Wu M, Martineau L, Nosrati M, Leong SP, et al. Prediction of positron emission tomography/computed tomography (PET/CT) positivity in patients with high-risk primary melanoma. *American journal of nuclear medicine and molecular imaging*. 2016; 6:277–85. [PubMed: 27766186]
7. Gilles R, de Geus-Oei LF, Mulders PF, Oyen WJ. Immunotherapy response evaluation with (18)F-FDG-PET in patients with advanced stage renal cell carcinoma. *World journal of urology*. 2013; 31:841–6. DOI: 10.1007/s00345-011-0723-y [PubMed: 21739122]
8. Acland KM, Healy C, Calonje E, O'Doherty M, Nunan T, Page C, et al. Comparison of positron emission tomography scanning and sentinel node biopsy in the detection of micrometastases of primary cutaneous malignant melanoma. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2001; 19:2674–8. DOI: 10.1200/JCO.2001.19.10.2674 [PubMed: 11352959]
9. Fletcher JW, Djulbegovic B, Soares HP, Siegel BA, Lowe VJ, Lyman GH, et al. Recommendations on the use of 18F-FDG PET in oncology. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2008; 49:480–508. DOI: 10.2967/jnumed.107.047787

10. Lin NU, Lee EQ, Aoyama H, Barani IJ, Barboriak DP, Baumert BG, et al. Response assessment criteria for brain metastases: proposal from the RANO group. *The Lancet Oncology*. 2015; 16:e270–8. DOI: 10.1016/S1470-2045(15)70057-4 [PubMed: 26065612]
11. Schellinger PD, Meinck HM, Thron A. Diagnostic accuracy of MRI compared to CCT in patients with brain metastases. *Journal of neuro-oncology*. 1999; 44:275–81. [PubMed: 10720207]
12. Ghasemi M, Nabipour I, Omrani A, Alipour Z, Assadi M. Precision medicine and molecular imaging: new targeted approaches toward cancer therapeutic and diagnosis. *American journal of nuclear medicine and molecular imaging*. 2016; 6:310–27. [PubMed: 28078184]
13. Valencak J, Heere-Ress E, Traub-Weidinger T, Raderer M, Schneeberger A, Thalhammer T, et al. Somatostatin receptor scintigraphy with ¹¹¹In-DOTA-lanreotide and ¹¹¹In-DOTA-Tyr3-octreotide in patients with stage IV melanoma: in-vitro and in-vivo results. *Melanoma research*. 2005; 15:523–9. [PubMed: 16314738]
14. Hong H, Sun J, Cai W. Anatomical and molecular imaging of skin cancer. *Clinical, cosmetic and investigational dermatology*. 2008; 1:1–17.
15. Ren G, Pan Y, Cheng Z. Molecular probes for malignant melanoma imaging. *Current pharmaceutical biotechnology*. 2010; 11:590–602. [PubMed: 20497118]
16. d'Ischia M, Wakamatsu K, Cicoira F, Di Mauro E, Garcia-Borron JC, Commo S, et al. Melanins and melanogenesis: from pigment cells to human health and technological applications. *Pigment cell & melanoma research*. 2015; 28:520–44. DOI: 10.1111/pcmr.12393 [PubMed: 26176788]
17. Dadachova E, Casadevall A. Melanin as a potential target for radionuclide therapy of metastatic melanoma. *Future oncology*. 2005; 1:541–9. DOI: 10.2217/14796694.1.4.541 [PubMed: 16556030]
18. Thompson JF, Scolyer RA, Kefford RF. Cutaneous melanoma. *Lancet*. 2005; 365:687–701. DOI: 10.1016/S0140-6736(05)17951-3 [PubMed: 15721476]
19. Ings RM. The melanin binding of drugs and its implications. *Drug metabolism reviews*. 1984; 15:1183–212. DOI: 10.3109/03602538409033561 [PubMed: 6396056]
20. Chezal JM, Papon J, Labarre P, Lartigue C, Galmier MJ, Decombat C, et al. Evaluation of radiolabeled (hetero)aromatic analogues of N-(2-diethylaminoethyl)-4-iodobenzamide for imaging and targeted radionuclide therapy of melanoma. *Journal of medicinal chemistry*. 2008; 51:3133–44. DOI: 10.1021/jm701424g [PubMed: 18481842]
21. Link E, Lukiewicz S. A new radioactive drug selectively accumulating in Melanoma cells. *European journal of nuclear medicine*. 1982; 7:469–73. [PubMed: 7140781]
22. Link EM, Blower PJ, Costa DC, Lane DM, Lui D, Brown RS, et al. Early detection of melanoma metastases with radioiodinated methylene blue. *European journal of nuclear medicine*. 1998; 25:1322–9. [PubMed: 9724383]
23. Link EM, Carpenter RN. ²¹¹At-methylene blue for targeted radiotherapy of human melanoma xenografts: treatment of cutaneous tumors and lymph node metastases. *Cancer research*. 1992; 52:4385–90. [PubMed: 1643635]
24. Michelot JM, Moreau MF, Labarre PG, Madelmont JC, Veyre AJ, Papon JM, et al. Synthesis and evaluation of new iodine-125 radiopharmaceuticals as potential tracers for malignant melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 1991; 32:1573–80.
25. El Aissi R, Chezal JM, Tarrit S, Chavignon O, Moreau E. Melanoma-targeted delivery system (part 1): design, synthesis and evaluation of releasable disulfide drug by glutathione. *European journal of medicinal chemistry*. 2015; 101:668–80. DOI: 10.1016/j.ejmech.2015.06.055 [PubMed: 26210505]
26. El Aissi R, Miladi I, Chezal JM, Chavignon O, Miot-Noirault E, Moreau E. Melanoma-targeted delivery system (part 2): Synthesis, radioiodination and biological evaluation in B16F0 bearing mice. *European journal of medicinal chemistry*. 2016; 120:304–12. DOI: 10.1016/j.ejmech.2016.05.019 [PubMed: 27214141]
27. Sillaire-Houtmann I, Bonafous J, Veyre A, Mestas D, D'Incan M, Moins N, et al. Phase 2 clinical study of ¹²³I-N-(2-diethylaminoethyl)-2-iodobenzamide in the diagnostic of primary and metastatic ocular melanoma. *Journal francais d'ophtalmologie*. 2004; 27:34–9.
28. Michelot JM, Moreau MF, Veyre AJ, Bonafous JF, Bacin FJ, Madelmont JC, et al. Phase II scintigraphic clinical trial of malignant melanoma and metastases with iodine-123-N-(2-

- diethylaminoethyl 4-iodobenzamide). *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 1993; 34:1260–6.
29. Brandau W, Niehoff T, Pulawski P, Jonas M, Dutschka K, Sciuk J, et al. Structure distribution relationship of iodine-123-iodobenzamides as tracers for the detection of melanotic melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 1996; 37:1865–71.
 30. Moins N, D'Incan M, Bonafous J, Bacin F, Labarre P, Moreau MF, et al. 123I-N-(2-diethylaminoethyl)-2-iodobenzamide: a potential imaging agent for cutaneous melanoma staging. *European journal of nuclear medicine and molecular imaging*. 2002; 29:1478–84. DOI: 10.1007/s00259-002-0971-6 [PubMed: 12397467]
 31. Dittmann H, Coenen HH, Zolzer F, Dutschka K, Brandau W, Streffer C. In vitro studies on the cellular uptake of melanoma imaging aminoalkyl-iodobenzamide derivatives (ABA). *Nuclear medicine and biology*. 1999; 26:51–6. [PubMed: 10096501]
 32. Bacin F, Michelot J, Bonafous J, Veyre A, Moreau MF, Kemeny JL, et al. Clinical study of [123I] N-(2-diethylaminoethyl)-4-iodobenzamide in the diagnosis of primary and metastatic ocular melanoma. *Acta ophthalmologica Scandinavica*. 1998; 76:56–61. [PubMed: 9541435]
 33. Larisch R, Schulte KW, Vosberg H, Ruzicka T, Muller-Gartner HW. Differential accumulation of iodine-123-iodobenzamide in melanotic and amelanotic melanoma metastases in vivo. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 1998; 39:996–1001.
 34. Labarre P, Papon J, Moreau MF, Moins N, Veyre A, Madelmont JC. Evaluation in mice of some iodinated melanoma imaging agents using cryosectioning and multi-wire proportional counting. *European journal of nuclear medicine*. 1999; 26:494–8. [PubMed: 10382093]
 35. Cachin F, Miot-Noirault E, Gillet B, Isnardi V, Labeille B, Payoux P, et al. (123I)I-BZA2 as a melanin-targeted radiotracer for the identification of melanoma metastases: results and perspectives of a multicenter phase III clinical trial. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2014; 55:15–22. DOI: 10.2967/jnumed.113.123554
 36. Liu X, Pham TQ, Berghofer P, Chapman J, Greguric I, Mitchell P, et al. Synthesis and evaluation of novel radioiodinated nicotinamides for malignant melanoma. *Nuclear medicine and biology*. 2008; 35:769–81. DOI: 10.1016/j.nucmedbio.2008.05.011 [PubMed: 18848662]
 37. Bonnet-Duquennoy M, Papon J, Mishellany F, Labarre P, Guerquin-Kern JL, Wu TD, et al. Targeted radionuclide therapy of melanoma: anti-tumoural efficacy studies of a new 131I labelled potential agent. *International journal of cancer Journal international du cancer*. 2009; 125:708–16. DOI: 10.1002/ijc.24413 [PubMed: 19437532]
 38. Billaud EM, Maisonia-Besset A, Rbah-Vidal L, Vidal A, Besse S, Bequignat JB, et al. Synthesis, radiolabeling and preliminary in vivo evaluation of multimodal radiotracers for PET imaging and targeted radionuclide therapy of pigmented melanoma. *European journal of medicinal chemistry*. 2015; 92:818–38. DOI: 10.1016/j.ejmech.2015.01.034 [PubMed: 25637883]
 39. Degoul F, Borel M, Jacquemot N, Besse S, Communal Y, Mishellany F, et al. In vivo efficacy of melanoma internal radionuclide therapy with a 131I-labelled melanin-targeting heteroarylcarboxamide molecule. *International journal of cancer Journal international du cancer*. 2013; 133:1042–53. DOI: 10.1002/ijc.28103 [PubMed: 23404099]
 40. Billaud EM, Rbah-Vidal L, Vidal A, Besse S, Tarrat S, Askienazy S, et al. Synthesis, radiofluorination, and in vivo evaluation of novel fluorinated and iodinated radiotracers for PET imaging and targeted radionuclide therapy of melanoma. *Journal of medicinal chemistry*. 2013; 56:8455–67. DOI: 10.1021/jm400877v [PubMed: 24044531]
 41. Pham TQ, Berghofer P, Liu X, Greguric I, Dikic B, Ballantyne P, et al. Preparation and biologic evaluation of a novel radioiodinated benzylpiperazine, 123I-MEL037, for malignant melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2007; 48:1348–56. DOI: 10.2967/jnumed.107.041673
 42. Roberts MP, Nguyen V, Ashford ME, Berghofer P, Wyatt NA, Krause-Heuer AM, et al. Synthesis and in Vivo Evaluation of [123I]Melanin-Targeted Agents. *Journal of medicinal chemistry*. 2015; 58:6214–24. DOI: 10.1021/acs.jmedchem.5b00777 [PubMed: 26177000]
 43. Chang CC, Chang CH, Shen CC, Chen CL, Liu RS, Lin MH, et al. Synthesis and evaluation of (1)(2)(3)/(1)(3)(1)I-Iochlornicotinamide as a novel SPECT probe for malignant melanoma. *Bioorganic & medicinal chemistry*. 2015; 23:2261–9. DOI: 10.1016/j.bmc.2015.02.017 [PubMed: 25800432]

44. Chang CC, Chang CH, Shen CC, Chen CL, Liu RS, Lin MH, et al. Synthesis and characterization of a novel radioiodinated phenylacetamide and its homolog as theranostic agents for malignant melanoma. *European journal of pharmaceutical sciences : official journal of the European Federation for Pharmaceutical Sciences*. 2016; 81:201–9. DOI: 10.1016/j.ejps.2015.10.019 [PubMed: 26517961]
45. Ren G, Miao Z, Liu H, Jiang L, Limpa-Amara N, Mahmood A, et al. Melanin-targeted preclinical PET imaging of melanoma metastasis. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2009; 50:1692–9. DOI: 10.2967/jnumed.109.066175
46. Wu SY, Huang SP, Lo YC, Liu RS, Wang SJ, Lin WJ, et al. Synthesis and preclinical characterization of [18F]FPBZA: a novel PET probe for melanoma. *BioMed research international*. 2014; 2014:912498.doi: 10.1155/2014/912498 [PubMed: 25254219]
47. Garg PK, Nazih R, Wu Y, Singh R, Garg S. 4-11C-Methoxy N-(2-Diethylaminoethyl) Benzamide: A Novel Probe to Selectively Target Melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2017; 58:827–32. DOI: 10.2967/jnumed.116.184564
48. Chang CC, Chang CH, Lo YH, Lin MH, Shen CC, Liu RS, et al. Preparation and characterization of a novel Al(18)F-NOTA-BZA conjugate for melanin-targeted imaging of malignant melanoma. *Bioorganic & medicinal chemistry letters*. 2016; 26:4133–9. DOI: 10.1016/j.bmcl.2016.06.022 [PubMed: 27445169]
49. Greguric I, Taylor SR, Denoyer D, Ballantyne P, Berghofer P, Roselt P, et al. Discovery of [18F]N-(2-(diethylamino)ethyl)-6-fluoronicotinamide: a melanoma positron emission tomography imaging radiotracer with high tumor to body contrast ratio and rapid renal clearance. *Journal of medicinal chemistry*. 2009; 52:5299–302. DOI: 10.1021/jm9008423 [PubMed: 19691348]
50. Denoyer D, Potdevin T, Roselt P, Neels OC, Kirby L, Greguric I, et al. Improved detection of regional melanoma metastasis using 18F-6-fluoro-N-[2-(diethylamino)ethyl] pyridine-3-carboxamide, a melanin-specific PET probe, by perilesional administration. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2011; 52:115–22. DOI: 10.2967/jnumed.110.078154
51. Liu H, Liu S, Miao Z, Deng Z, Shen B, Hong X, et al. Development of 18F-labeled picolinamide probes for PET imaging of malignant melanoma. *Journal of medicinal chemistry*. 2013; 56:895–901. DOI: 10.1021/jm301740k [PubMed: 23301672]
52. Liu H, Liu S, Miao Z, Jiang H, Deng Z, Hong X, et al. A novel aliphatic 18F-labeled probe for PET imaging of melanoma. *Molecular pharmaceutics*. 2013; 10:3384–91. DOI: 10.1021/mp400225s [PubMed: 23927458]
53. Zhernosekov KP, Filosofov DV, Baum RP, Aschoff P, Bihl H, Razbash AA, et al. Processing of generator-produced 68Ga for medical application. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2007; 48:1741–8. DOI: 10.2967/jnumed.107.040378
54. Trencsenyi G, Denes N, Nagy G, Kis A, Vida A, Farkas F, et al. Comparative preclinical evaluation of 68Ga-NODAGA and 68Ga-HBED-CC conjugated procainamide in melanoma imaging. *Journal of pharmaceutical and biomedical analysis*. 2017; 139:54–64. DOI: 10.1016/j.jpba.2017.02.049 [PubMed: 28273651]
55. Kertesz I, Vida A, Nagy G, Emri M, Farkas A, Kis A, et al. In Vivo Imaging of Experimental Melanoma Tumors using the Novel Radiotracer 68Ga-NODAGA-Procainamide (PCA). *Journal of Cancer*. 2017; 8:774–85. DOI: 10.7150/jca.17550 [PubMed: 28382139]
56. Wang J, Wei C, Noor R, Burke A, McIntyre S, Bedikian AY. Surveillance for brain metastases in patients receiving systemic therapy for advanced melanoma. *Melanoma research*. 2014; 24:54–60. DOI: 10.1097/CMR.000000000000022 [PubMed: 24121189]
57. Aukema TS, Valdes Olmos RA, Wouters MW, Klop WM, Kroon BB, Vogel WV, et al. Utility of preoperative 18F-FDG PET/CT and brain MRI in melanoma patients with palpable lymph node metastases. *Annals of surgical oncology*. 2010; 17:2773–8. DOI: 10.1245/s10434-010-1088-y [PubMed: 20422454]
58. Belhocine TZ, Scott AM, Even-Sapir E, Urbain JL, Essner R. Role of nuclear medicine in the management of cutaneous malignant melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2006; 47:957–67.
59. Unterrainer M, Galldiks N, Suchorska B, Kowalew LC, Wenter V, Schmid-Tannwald C, et al. 18F-FET PET Uptake Characteristics in Patients with Newly Diagnosed and Untreated Brain

- Metastasis. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2017; 58:584–9. DOI: 10.2967/jnumed.116.180075
60. Valdes Olmos RA, Vidal-Sicart S, Manca G, Mariani G, Leon-Ramirez LF, Rubello D, et al. Advances in radioguided surgery in oncology. *The quarterly journal of nuclear medicine and molecular imaging : official publication of the Italian Association of Nuclear Medicine (AIMN) [and] the International Association of Radiopharmacology (IAR), [and] Section of the So*. 2017; 61:247–70. DOI: 10.23736/S1824-4785.17.02995-8
61. Bronstein Y, Ng CS, Rohren E, Ross MI, Lee JE, Cormier J, et al. PET/CT in the management of patients with stage IIIC and IV metastatic melanoma considered candidates for surgery: evaluation of the additive value after conventional imaging. *AJR American journal of roentgenology*. 2012; 198:902–8. DOI: 10.2214/AJR.11.7280 [PubMed: 22451559]
62. Garg S, Kothari K, Thopate SR, Doke AK, Garg PK. Design, synthesis, and preliminary in vitro and in vivo evaluation of N-(2-diethylaminoethyl)-4-[18F]fluorobenzamide ([18F]-DAFBA): a novel potential PET probe to image melanoma tumors. *Bioconjugate chemistry*. 2009; 20:583–90. DOI: 10.1021/bc8005094 [PubMed: 19222206]
63. Wang Y, Li M, Zhang Y, Zhang F, Liu C, Song Y, et al. Detection of melanoma metastases with PET-Comparison of 18F-5-FPN with 18F-FDG. *Nuclear medicine and biology*. 2017; 50:33–8. DOI: 10.1016/j.nucmedbio.2017.03.005 [PubMed: 28433794]
64. Denoyer D, Greguric I, Roselt P, Neels OC, Aide N, Taylor SR, et al. High-contrast PET of melanoma using (18)F-MEL050, a selective probe for melanin with predominantly renal clearance. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2010; 51:441–7. DOI: 10.2967/jnumed.109.070060
65. Xie L, Yui J, Fujinaga M, Hatori A, Yamasaki T, Kumata K, et al. Molecular imaging of ectopic metabotropic glutamate 1 receptor in melanoma with a positron emission tomography radioprobe (18) F-FITM. *International journal of cancer Journal international du cancer*. 2014; 135:1852–9. DOI: 10.1002/ijc.28842 [PubMed: 24643962]
66. Rbah-Vidal L, Vidal A, Besse S, Cachin F, Bonnet M, Audin L, et al. Early detection and longitudinal monitoring of experimental primary and disseminated melanoma using [(1) (0)F]ICF01006, a highly promising melanoma PET tracer. *European journal of nuclear medicine and molecular imaging*. 2012; 39:1449–61. DOI: 10.1007/s00259-012-2168-y [PubMed: 22707183]
67. Feng H, Xia X, Li C, Song Y, Qin C, Liu Q, et al. Imaging malignant melanoma with (18)F-5-FPN. *European journal of nuclear medicine and molecular imaging*. 2016; 43:113–22. DOI: 10.1007/s00259-015-3134-2 [PubMed: 26260649]
68. Sun X, Li Y, Liu T, Li Z, Zhang X, Chen X. Peptide-based imaging agents for cancer detection. *Advanced drug delivery reviews*. 2017; 110–111:38–51. DOI: 10.1016/j.addr.2016.06.007
69. Singh M, Mukhopadhyay K. Alpha-melanocyte stimulating hormone: an emerging anti-inflammatory antimicrobial peptide. *BioMed research international*. 2014; 2014:874610.doi: 10.1155/2014/874610 [PubMed: 25140322]
70. Chen J, Cheng Z, Hoffman TJ, Jurisson SS, Quinn TP. Melanoma-targeting properties of (99m)technetium-labeled cyclic alpha-melanocyte-stimulating hormone peptide analogues. *Cancer research*. 2000; 60:5649–58. [PubMed: 11059756]
71. Siegrist W, Oestreicher M, Stutz S, Girard J, Eberle AN. Radioreceptor assay for alpha-MSH using mouse B16 melanoma cells+ *Journal of receptor research*. 1988; 8:323–43. [PubMed: 2838620]
72. Garg PK, Alston KL, Welsh PC, Zalutsky MR. Enhanced binding and inertness to dehalogenation of alpha-melanotropic peptides labeled using N-succinimidyl 3-iodobenzoate. *Bioconjugate chemistry*. 1996; 7:233–9. DOI: 10.1021/bc960001+ [PubMed: 8983345]
73. Vaidyanathan G, Zalutsky MR. Fluorine-18-labeled [Nle4,D-Phe7]-alpha-MSH, an alpha-melanocyte stimulating hormone analogue. *Nuclear medicine and biology*. 1997; 24:171–8. [PubMed: 9089709]
74. Cowell SM, Balse-Srinivasan PM, Ahn JM, Hruby VJ. Design and synthesis of peptide antagonists and inverse agonists for G protein-coupled receptors. *Methods in enzymology*. 2002; 343:49–72. [PubMed: 11665587]

75. Sawyer TK, Sanfilippo PJ, Hruby VJ, Engel MH, Heward CB, Burnett JB, et al. 4-Norleucine, 7-D-phenylalanine-alpha-melanocyte-stimulating hormone: a highly potent alpha-melanotropin with ultralong biological activity. *Proceedings of the National Academy of Sciences of the United States of America*. 1980; 77:5754–8. [PubMed: 6777774]
76. Hruby VJ, Wilkes BC, Hadley ME, Al-Obeidi F, Sawyer TK, Staples DJ, et al. alpha-Melanotropin: the minimal active sequence in the frog skin bioassay. *Journal of medicinal chemistry*. 1987; 30:2126–30. [PubMed: 2822931]
77. Giblin MF, Wang N, Hoffman TJ, Jurisson SS, Quinn TP. Design and characterization of alpha-melanotropin peptide analogs cyclized through rhenium and technetium metal coordination. *Proceedings of the National Academy of Sciences of the United States of America*. 1998; 95:12814–8. [PubMed: 9788997]
78. Froidevaux S, Calame-Christe M, Tanner H, Eberle AN. Melanoma targeting with DOTA-alpha-melanocyte-stimulating hormone analogs: structural parameters affecting tumor uptake and kidney uptake. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2005; 46:887–95.
79. Bednarek MA, Silva MV, Arison B, MacNeil T, Kalyani RN, Huang RR, et al. Structure-function studies on the cyclic peptide MT-II, lactam derivative of alpha-melanotropin. *Peptides*. 1999; 20:401–9. [PubMed: 10447101]
80. Grieco P, Cai M, Liu L, Mayorov A, Chandler K, Trivedi D, et al. Design and microwave-assisted synthesis of novel macrocyclic peptides active at melanocortin receptors: discovery of potent and selective hMC5R receptor antagonists. *Journal of medicinal chemistry*. 2008; 51:2701–7. DOI: 10.1021/jm701181n [PubMed: 18412316]
81. Miao Y, Gallazzi F, Guo H, Quinn TP. ¹¹¹In-labeled lactam bridge-cyclized alpha-melanocyte stimulating hormone peptide analogues for melanoma imaging. *Bioconjugate chemistry*. 2008; 19:539–47. DOI: 10.1021/bc700317w [PubMed: 18197608]
82. Guo H, Yang J, Gallazzi F, Miao Y. Reduction of the ring size of radiolabeled lactam bridge-cyclized alpha-MSH peptide, resulting in enhanced melanoma uptake. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2010; 51:418–26. DOI: 10.2967/jnumed.109.071787
83. Barkey NM, Tafreshi NK, Josan JS, De Silva CR, Sill KN, Hruby VJ, et al. Development of melanoma-targeted polymer micelles by conjugation of a melanocortin 1 receptor (MC1R) specific ligand. *Journal of medicinal chemistry*. 2011; 54:8078–84. DOI: 10.1021/jm201226w [PubMed: 22011200]
84. Tafreshi NK, Huang X, Moberg VE, Barkey NM, Sondak VK, Tian H, et al. Synthesis and characterization of a melanoma-targeted fluorescence imaging probe by conjugation of a melanocortin 1 receptor (MC1R) specific ligand. *Bioconjugate chemistry*. 2012; 23:2451–9. DOI: 10.1021/bc300549s [PubMed: 23116461]
85. McQuade P, Miao Y, Yoo J, Quinn TP, Welch MJ, Lewis JS. Imaging of melanoma using ⁶⁴Cu- and ⁸⁶Y-DOTA-ReCCMSH(Arg11), a cyclized peptide analogue of alpha-MSH. *Journal of medicinal chemistry*. 2005; 48:2985–92. DOI: 10.1021/jm0490282 [PubMed: 15828837]
86. Wei L, Butcher C, Miao Y, Gallazzi F, Quinn TP, Welch MJ, et al. Synthesis and biologic evaluation of ⁶⁴Cu-labeled rhenium-cyclized alpha-MSH peptide analog using a cross-bridged cyclam chelator. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2007; 48:64–72.
87. Wei L, Miao Y, Gallazzi F, Quinn TP, Welch MJ, Vavere AL, et al. Gallium-68-labeled DOTA-rhenium-cyclized alpha-melanocyte-stimulating hormone analog for imaging of malignant melanoma. *Nuclear medicine and biology*. 2007; 34:945–53. DOI: 10.1016/j.nucmedbio.2007.07.003 [PubMed: 17998097]
88. Ren G, Liu Z, Miao Z, Liu H, Subbarayan M, Chin FT, et al. PET of malignant melanoma using ¹⁸F-labeled metalloptides. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2009; 50:1865–72. DOI: 10.2967/jnumed.109.062877
89. Ren G, Liu S, Liu H, Miao Z, Cheng Z. Radiofluorinated rhenium cyclized alpha-MSH analogues for PET imaging of melanocortin receptor 1. *Bioconjugate chemistry*. 2010; 21:2355–60. DOI: 10.1021/bc100391a [PubMed: 21073170]

90. Guo H, Shenoy N, Gershman BM, Yang J, Sklar LA, Miao Y. Metastatic melanoma imaging with an (111)In-labeled lactam bridge-cyclized alpha-melanocyte-stimulating hormone peptide. *Nuclear medicine and biology*. 2009; 36:267–76. DOI: 10.1016/j.nucmedbio.2009.01.003 [PubMed: 19324272]
91. Guo H, Gallazzi F, Miao Y. Gallium-67-labeled lactam bridge-cyclized alpha-MSH peptides with enhanced melanoma uptake and reduced renal uptake. *Bioconjugate chemistry*. 2012; 23:1341–8. DOI: 10.1021/bc300191z [PubMed: 22621181]
92. Guo H, Miao Y. Introduction of an 8-aminooctanoic acid linker enhances uptake of 99mTc-labeled lactam bridge-cyclized alpha-MSH peptide in melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2014; 55:2057–63. DOI: 10.2967/jnumed.114.145896
93. Guo H, Miao Y. Cu-64-labeled lactam bridge-cyclized alpha-MSH peptides for PET imaging of melanoma. *Molecular pharmaceutics*. 2012; 9:2322–30. DOI: 10.1021/mp300246j [PubMed: 22780870]
94. Guo H, Gallazzi F, Miao Y. Design and evaluation of new Tc-99m-labeled lactam bridge-cyclized alpha-MSH peptides for melanoma imaging. *Molecular pharmaceutics*. 2013; 10:1400–8. DOI: 10.1021/mp3006984 [PubMed: 23418722]
95. Nagy G, Denes N, Kis A, Szabo JP, Berenyi E, Garai I, et al. Preclinical evaluation of melanocortin-1 receptor (MC1-R) specific 68Ga- and 44Sc-labeled DOTA-NAPamide in melanoma imaging. *European journal of pharmaceutical sciences : official journal of the European Federation for Pharmaceutical Sciences*. 2017; 106:336–44. DOI: 10.1016/j.ejps.2017.06.026 [PubMed: 28625749]
96. Cheng Z, Xiong Z, Subbarayan M, Chen X, Gambhir SS. 64Cu-labeled alpha-melanocyte-stimulating hormone analog for microPET imaging of melanocortin 1 receptor expression. *Bioconjugate chemistry*. 2007; 18:765–72. DOI: 10.1021/bc060306g [PubMed: 17348700]
97. Froidevaux S, Calame-Christe M, Schuhmacher J, Tanner H, Saffrich R, Henze M, et al. A gallium-labeled DOTA-alpha-melanocyte-stimulating hormone analog for PET imaging of melanoma metastases. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2004; 45:116–23.
98. Cheng Z, Zhang L, Graves E, Xiong Z, Dandekar M, Chen X, et al. Small-animal PET of melanocortin 1 receptor expression using a 18F-labeled alpha-melanocyte-stimulating hormone analog. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2007; 48:987–94. DOI: 10.2967/jnumed.107.039602
99. Cheng Z, Chen J, Miao Y, Owen NK, Quinn TP, Jurisson SS. Modification of the structure of a metalloprotein: synthesis and biological evaluation of (111)In-labeled DOTA-conjugated rhenium-cyclized alpha-MSH analogues. *Journal of medicinal chemistry*. 2002; 45:3048–56. [PubMed: 12086490]
100. Miao Y, Benwell K, Quinn TP. 99mTc- and 111In-labeled alpha-melanocyte-stimulating hormone peptides as imaging probes for primary and pulmonary metastatic melanoma detection. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2007; 48:73–80.
101. Quinn T, Zhang X, Miao Y. Targeted melanoma imaging and therapy with radiolabeled alpha-melanocyte stimulating hormone peptide analogues. *Giornale italiano di dermatologia e venereologia : organo ufficiale, Societa italiana di dermatologia e sifilografia*. 2010; 145:245–58.
102. Price TW, Greenman J, Stasiuk GJ. Current advances in ligand design for inorganic positron emission tomography tracers 68Ga, 64Cu, 89Zr and 44Sc. *Dalton transactions*. 2016; 45:15702–24. DOI: 10.1039/c5dt04706d [PubMed: 26865360]
103. Chakravarty R, Goel S, Valdovinos HF, Hernandez R, Hong H, Nickles RJ, et al. Matching the decay half-life with the biological half-life: ImmunoPET imaging with (44)Sc-labeled cetuximab Fab fragment. *Bioconjugate chemistry*. 2014; 25:2197–204. DOI: 10.1021/bc500415x [PubMed: 25389697]
104. Hernandez R, Valdovinos HF, Yang Y, Chakravarty R, Hong H, Barnhart TE, et al. (44)Sc: an attractive isotope for peptide-based PET imaging. *Molecular pharmaceutics*. 2014; 11:2954–61. DOI: 10.1021/mp500343j [PubMed: 25054618]
105. Zhang C, Zhang Z, Lin KS, Pan J, Dude I, Hundal-Jabal N, et al. Preclinical Melanoma Imaging with 68Ga-Labeled alpha-Melanocyte-Stimulating Hormone Derivatives Using PET. *Theranostics*. 2017; 7:805–13. DOI: 10.7150/thno.17117 [PubMed: 28382155]

106. Margadant C, Monsuur HN, Norman JC, Sonnenberg A. Mechanisms of integrin activation and trafficking. *Current opinion in cell biology*. 2011; 23:607–14. DOI: 10.1016/j.ceb.2011.08.005 [PubMed: 21924601]
107. Decristoforo C, Faintuch-Linkowski B, Rey A, von Guggenberg E, Rupprich M, Hernandez-Gonzales I, et al. [99mTc]JHYNIC-RGD for imaging integrin alphavbeta3 expression. *Nuclear medicine and biology*. 2006; 33:945–52. DOI: 10.1016/j.nucmedbio.2006.09.001 [PubMed: 17127166]
108. Decristoforo C, Hernandez Gonzalez I, Carlsen J, Rupprich M, Huisman M, Virgolini I, et al. 68Ga- and 111In-labelled DOTA-RGD peptides for imaging of alphavbeta3 integrin expression. *European journal of nuclear medicine and molecular imaging*. 2008; 35:1507–15. DOI: 10.1007/s00259-008-0757-6 [PubMed: 18369617]
109. Hultsch C, Schottelius M, Auernheimer J, Alke A, Wester HJ. (18)F-Fluoroglucosylation of peptides, exemplified on cyclo(RGDfK). *European journal of nuclear medicine and molecular imaging*. 2009; 36:1469–74. DOI: 10.1007/s00259-009-1122-0 [PubMed: 19350236]
110. Wei L, Ye Y, Wadas TJ, Lewis JS, Welch MJ, Achilefu S, et al. (64)Cu-labeled CB-TE2A and diamsar-conjugated RGD peptide analogs for targeting angiogenesis: comparison of their biological activity. *Nuclear medicine and biology*. 2009; 36:277–85. DOI: 10.1016/j.nucmedbio.2008.12.008 [PubMed: 19324273]
111. Jin ZH, Furukawa T, Kumata K, Xie L, Yui J, Wakizaka H, et al. Development of the Fibronectin-Mimetic Peptide KSSPHSRN(SG)5RGDSP as a Novel Radioprobe for Molecular Imaging of the Cancer Biomarker alpha5beta1 Integrin. *Biological & pharmaceutical bulletin*. 2015; 38:1722–31. DOI: 10.1248/bpb.b15-00344 [PubMed: 26277991]
112. Mena E, Owenius R, Turkbey B, Sherry R, Bratslavsky G, Macholl S, et al. [(1)(8)F]fluciclatide in the in vivo evaluation of human melanoma and renal tumors expressing alphavbeta 3 and alpha vbeta 5 integrins. *European journal of nuclear medicine and molecular imaging*. 2014; 41:1879–88. DOI: 10.1007/s00259-014-2791-x [PubMed: 24973039]
113. Sharma R, Kallur KG, Ryu JS, Parameswaran RV, Lindman H, Avril N, et al. Multicenter Reproducibility of 18F-Fluciclatide PET Imaging in Subjects with Solid Tumors. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2015; 56:1855–61. DOI: 10.2967/jnumed.115.158253
114. Beer AJ, Haubner R, Goebel M, Luderschmidt S, Spilker ME, Wester HJ, et al. Biodistribution and pharmacokinetics of the alphavbeta3-selective tracer 18F-galacto-RGD in cancer patients. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2005; 46:1333–41.
115. Beer AJ, Haubner R, Sarbia M, Goebel M, Luderschmidt S, Grosu AL, et al. Positron emission tomography using [18F]Galacto-RGD identifies the level of integrin alpha(v)beta3 expression in man. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2006; 12:3942–9. DOI: 10.1158/1078-0432.CCR-06-0266 [PubMed: 16818691]
116. Gaertner FC, Kessler H, Wester HJ, Schwaiger M, Beer AJ. Radiolabelled RGD peptides for imaging and therapy. *European journal of nuclear medicine and molecular imaging*. 2012; 39(Suppl 1):S126–38. DOI: 10.1007/s00259-011-2028-1 [PubMed: 22388629]
117. Cai W, Chen X. Multimodality molecular imaging of tumor angiogenesis. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2008; 49(Suppl 2):113S–28S. DOI: 10.2967/jnumed.107.045922
118. Knetsch PA, Petrik M, Rangger C, Seidel G, Pietzsch HJ, Virgolini I, et al. [(6)(8)Ga]NS(3)-RGD and [(6)(8)Ga] Oxo-DO3A-RGD for imaging alpha(v)beta(3) integrin expression: synthesis, evaluation, and comparison. *Nuclear medicine and biology*. 2013; 40:65–72. DOI: 10.1016/j.nucmedbio.2012.09.006 [PubMed: 23102540]
119. Knetsch PA, Petrik M, Griessinger CM, Rangger C, Fani M, Kesenheimer C, et al. [68Ga]NODAGA-RGD for imaging alphavbeta3 integrin expression. *European journal of nuclear medicine and molecular imaging*. 2011; 38:1303–12. DOI: 10.1007/s00259-011-1778-0 [PubMed: 21487838]
120. Pohle K, Notni J, Bussemer J, Kessler H, Schwaiger M, Beer AJ. 68Ga-NODAGA-RGD is a suitable substitute for (18)F-Galacto-RGD and can be produced with high specific activity in a

- cGMP/GRP compliant automated process. *Nuclear medicine and biology*. 2012; 39:777–84. DOI: 10.1016/j.nucmedbio.2012.02.006 [PubMed: 22444238]
121. Knetsch PA, Zhai C, Rangger C, Blatzer M, Haas H, Kaeopookum P, et al. [(68)Ga]FSC-(RGD)₃ a trimeric RGD peptide for imaging alphavbeta3 integrin expression based on a novel siderophore derived chelating scaffold-synthesis and evaluation. *Nuclear medicine and biology*. 2015; 42:115–22. DOI: 10.1016/j.nucmedbio.2014.10.001 [PubMed: 25459110]
122. Zhai C, Summer D, Rangger C, Haas H, Haubner R, Decristoforo C, Fusarinine C, a novel siderophore-based bifunctional chelator for radiolabeling with Gallium-68. *Journal of labelled compounds & radiopharmaceuticals*. 2015; 58:209–14. DOI: 10.1002/jlcr.3286 [PubMed: 25874571]
123. Zhai C, Summer D, Rangger C, Franssen GM, Laverman P, Haas H, et al. Novel Bifunctional Cyclic Chelator for (89)Zr Labeling-Radiolabeling and Targeting Properties of RGD Conjugates. *Molecular pharmaceutics*. 2015; 12:2142–50. DOI: 10.1021/acs.molpharmaceut.5b00128 [PubMed: 25941834]
124. Petrik M, Zhai C, Novy Z, Urbanek L, Haas H, Decristoforo C. In Vitro and In Vivo Comparison of Selected Ga-68 and Zr-89 Labelled Siderophores. *Molecular imaging and biology : MIB : the official publication of the Academy of Molecular Imaging*. 2016; 18:344–52. DOI: 10.1007/s11307-015-0897-6 [PubMed: 26424719]
125. Zhai C, Franssen GM, Petrik M, Laverman P, Summer D, Rangger C, et al. Comparison of Ga-68-Labeled Fusarinine C-Based Multivalent RGD Conjugates and [(68)Ga]NODAGA-RGD-In Vivo Imaging Studies in Human Xenograft Tumors. *Molecular imaging and biology : MIB : the official publication of the Academy of Molecular Imaging*. 2016; 18:758–67. DOI: 10.1007/s11307-016-0931-3 [PubMed: 26905697]
126. Eisenwiener KP, Prata MI, Buschmann I, Zhang HW, Santos AC, Wenger S, et al. NODAGATOC, a new chelator-coupled somatostatin analogue labeled with [67/68Ga] and [111In] for SPECT, PET, and targeted therapeutic applications of somatostatin receptor (hsst2) expressing tumors. *Bioconjugate chemistry*. 2002; 13:530–41. [PubMed: 12009943]
127. Simecek J, Schulz M, Notni J, Plutnar J, Kubicek V, Havlickova J, et al. Complexation of metal ions with TRAP (1,4,7-triazacyclononane phosphinic acid) ligands and 1,4,7-triazacyclononane-1,4,7-triacetic acid: phosphinate-containing ligands as unique chelators for trivalent gallium. *Inorganic chemistry*. 2012; 51:577–90. DOI: 10.1021/ic202103v [PubMed: 22221285]
128. Notni J, Simecek J, Hermann P, Wester HJ. TRAP, a powerful and versatile framework for gallium-68 radiopharmaceuticals. *Chemistry (Weinheim an der Bergstrasse, Germany)*. 2011; 17:14718–22. DOI: 10.1002/chem.201103503
129. Notni J, Pohle K, Wester HJ. Comparative gallium-68 labeling of TRAP-, NOTA-, and DOTA-peptides: practical consequences for the future of gallium-68-PET. *EJNMMI research*. 2012; 2:28.doi: 10.1186/2191-219X-2-28 [PubMed: 22682112]
130. Notni J, Pohle K, Wester HJ. Be spoilt for choice with radiolabelled RGD peptides: preclinical evaluation of (6)(8)Ga-TRAP(RGD)(3). *Nuclear medicine and biology*. 2013; 40:33–41. DOI: 10.1016/j.nucmedbio.2012.08.006 [PubMed: 22995902]
131. Simecek J, Hermann P, Wester HJ, Notni J. How is (68)Ga labeling of macrocyclic chelators influenced by metal ion contaminants in (68)Ge/(68)Ga generator eluates? *ChemMedChem*. 2013; 8:95–103. DOI: 10.1002/cmde.201200471 [PubMed: 23136062]
132. Simecek J, Notni J, Kapp TG, Kessler H, Wester HJ. Benefits of NOPO as chelator in gallium-68 peptides, exemplified by preclinical characterization of (68)Ga-NOPO-c(RGDfK). *Molecular pharmaceutics*. 2014; 11:1687–95. DOI: 10.1021/mp5000746 [PubMed: 24669840]
133. Neubauer S, Rechenmacher F, Beer AJ, Curnis F, Pohle K, D'Alessandria C, et al. Selective imaging of the angiogenic relevant integrins alpha5beta1 and alphavbeta3. *Angewandte Chemie (International ed in English)*. 2013; 52:11656–9. DOI: 10.1002/anie.201306376 [PubMed: 24115324]
134. Heckmann D, Meyer A, Laufer B, Zahn G, Stragies R, Kessler H. Rational design of highly active and selective ligands for the alpha5beta1 integrin receptor. *ChemBiochem : a European journal of chemical biology*. 2008; 9:1397–407. DOI: 10.1002/cbic.200800045 [PubMed: 18481343]

135. Koivunen E, Wang B, Ruoslahti E. Isolation of a highly specific ligand for the alpha 5 beta 1 integrin from a phage display library. *The Journal of cell biology*. 1994; 124:373–80. [PubMed: 7507494]
136. Haubner R, Maschauer S, Prante O. PET radiopharmaceuticals for imaging integrin expression: tracers in clinical studies and recent developments. *BioMed research international*. 2014; 2014:871609.doi: 10.1155/2014/871609 [PubMed: 25013808]
137. D'Alessandria C, Pohle K, Rechenmacher F, Neubauer S, Notni J, Wester HJ, et al. In vivo biokinetic and metabolic characterization of the (6)(8)Ga-labelled alpha5beta1-selective peptidomimetic FR366. *European journal of nuclear medicine and molecular imaging*. 2016; 43:953–63. DOI: 10.1007/s00259-015-3218-z [PubMed: 26497698]
138. Notni J, Steiger K, Hoffmann F, Reich D, Kapp TG, Rechenmacher F, et al. Complementary, Selective PET Imaging of Integrin Subtypes alpha5beta1 and alphavbeta3 Using 68Ga-Aquibepirin and 68Ga-Avebetrin. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2016; 57:460–6. DOI: 10.2967/jnumed.115.165720
139. Notni J, Steiger K, Hoffmann F, Reich D, Schwaiger M, Kessler H, et al. Variation of Specific Activities of 68Ga-Aquibepirin and 68Ga-Avebetrin Enables Selective PET Imaging of Different Expression Levels of Integrins alpha5beta1 and alphavbeta3. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2016; 57:1618–24. DOI: 10.2967/jnumed.116.173948
140. Bianchini F, Fabbri P, Menchi G, Raspanti S, Bottoncetti A, Passeri A, et al. Radiosynthesis and micro-SPECT analysis of triazole-based RGD integrin ligands as non-peptide molecular imaging probes for angiogenesis. *Bioorganic & medicinal chemistry*. 2015; 23:1112–22. DOI: 10.1016/j.bmc.2014.12.065 [PubMed: 25637121]
141. Mas-Moruno C, Fraioli R, Rechenmacher F, Neubauer S, Kapp TG, Kessler H. alphavbeta3- or alpha5beta1-Integrin-Selective Peptidomimetics for Surface Coating. *Angewandte Chemie (International ed in English)*. 2016; 55:7048–67. DOI: 10.1002/anie.201509782 [PubMed: 27258759]
142. Haskali MB, Denoyer D, Noonan W, Culinane C, Rangger C, Pouliot N, et al. Sulfonation of Tyrosine as a Method To Improve Biodistribution of Peptide-Based Radiotracers: Novel 18F-Labeled Cyclic RGD Analogues. *Molecular pharmaceutics*. 2017; 14:1169–80. DOI: 10.1021/acs.molpharmaceut.6b01062 [PubMed: 28191977]
143. Park SH, Zheng JH, Nguyen VH, Jiang SN, Kim DY, Szardenings M, et al. RGD Peptide Cell-Surface Display Enhances the Targeting and Therapeutic Efficacy of Attenuated Salmonella-mediated Cancer Therapy. *Theranostics*. 2016; 6:1672–82. DOI: 10.7150/thno.16135 [PubMed: 27446500]
144. Melemenidis S, Jefferson A, Ruparelia N, Akhtar AM, Xie J, Allen D, et al. Molecular magnetic resonance imaging of angiogenesis in vivo using polyvalent cyclic RGD-iron oxide microparticle conjugates. *Theranostics*. 2015; 5:515–29. DOI: 10.7150/thno.10319 [PubMed: 25767618]
145. Fan Q, Cheng K, Hu X, Ma X, Zhang R, Yang M, et al. Transferring biomarker into molecular probe: melanin nanoparticle as a naturally active platform for multimodality imaging. *Journal of the American Chemical Society*. 2014; 136:15185–94. DOI: 10.1021/ja505412p [PubMed: 25292385]
146. Li C, Wang W, Wu Q, Ke S, Houston J, Sevic-Muraca E, et al. Dual optical and nuclear imaging in human melanoma xenografts using a single targeted imaging probe. *Nuclear medicine and biology*. 2006; 33:349–58. DOI: 10.1016/j.nucmedbio.2006.01.001 [PubMed: 16631083]
147. Yang J, Guo H, Gallazzi F, Berwick M, Padilla RS, Miao Y. Evaluation of a novel Arg-Gly-Asp-conjugated alpha-melanocyte stimulating hormone hybrid peptide for potential melanoma therapy. *Bioconjugate chemistry*. 2009; 20:1634–42. DOI: 10.1021/bc9001954 [PubMed: 19552406]
148. Yang J, Guo H, Miao Y. Technetium-99m-labeled Arg-Gly-Asp-conjugated alpha-melanocyte stimulating hormone hybrid peptides for human melanoma imaging. *Nuclear medicine and biology*. 2010; 37:873–83. DOI: 10.1016/j.nucmedbio.2010.05.006 [PubMed: 21055617]
149. Yang J, Miao Y. Substitution of Gly with Ala enhanced the melanoma uptake of technetium-99m-labeled Arg-Ala-Asp-conjugated alpha-melanocyte stimulating hormone peptide. *Bioorganic &*

medicinal chemistry letters. 2012; 22:1541–5. DOI: 10.1016/j.bmcl.2012.01.003 [PubMed: 22297112]

150. Flook AM, Yang J, Miao Y. Evaluation of new Tc-99m-labeled Arg-X-Asp-conjugated alpha-melanocyte stimulating hormone peptides for melanoma imaging. *Molecular pharmaceuticals*. 2013; 10:3417–24. DOI: 10.1021/mp400248f [PubMed: 23885640]
151. Flook AM, Yang J, Miao Y. Effects of amino acids on melanoma targeting and clearance properties of Tc-99m-labeled Arg-X-Asp-conjugated alpha-melanocyte stimulating hormone peptides. *Journal of medicinal chemistry*. 2013; 56:8793–802. DOI: 10.1021/jm4012356 [PubMed: 24131154]
152. Yang J, Flook AM, Feng C, Miao Y. Linker modification reduced the renal uptake of technetium-99m-labeled Arg-Ala-Asp-conjugated alpha-melanocyte stimulating hormone peptide. *Bioorganic & medicinal chemistry letters*. 2014; 24:195–8. DOI: 10.1016/j.bmcl.2013.11.042 [PubMed: 24316121]
153. Yang J, Guo H, Padilla RS, Berwick M, Miao Y. Replacement of the Lys linker with an Arg linker resulting in improved melanoma uptake and reduced renal uptake of Tc-99m-labeled Arg-Gly-Asp-conjugated alpha-melanocyte stimulating hormone hybrid peptide. *Bioorganic & medicinal chemistry*. 2010; 18:6695–700. DOI: 10.1016/j.bmc.2010.07.061 [PubMed: 20728365]
154. Yang J, Lu J, Miao Y. Structural modification on the Lys linker enhanced tumor to kidney uptake ratios of 99mTc-labeled RGD-conjugated alpha-MSH hybrid peptides. *Molecular pharmaceuticals*. 2012; 9:1418–24. DOI: 10.1021/mp2006642 [PubMed: 22452443]
155. Xu J, Yang J, Miao Y. Dual receptor-targeting (9)(9)mTc-labeled Arg-Gly-Asp-conjugated Alpha-Melanocyte stimulating hormone hybrid peptides for human melanoma imaging. *Nuclear medicine and biology*. 2015; 42:369–74. DOI: 10.1016/j.nucmedbio.2014.11.002 [PubMed: 25577037]
156. Yang J, Hu CA, Miao Y. Tc-99m-labeled RGD-conjugated alpha-melanocyte stimulating hormone hybrid peptides with reduced renal uptake. *Amino acids*. 2015; 47:813–23. DOI: 10.1007/s00726-014-1911-z [PubMed: 25557051]
157. Flook AM, Yang J, Miao Y. Substitution of the Lys linker with the beta-Ala linker dramatically decreased the renal uptake of 99mTc-labeled Arg-X-Asp-conjugated and X-Ala-Asp-conjugated alpha-melanocyte stimulating hormone peptides. *Journal of medicinal chemistry*. 2014; 57:9010–8. DOI: 10.1021/jm501114v [PubMed: 25290883]
158. Kunishima N, Shimada Y, Tsuji Y, Sato T, Yamamoto M, Kumasaka T, et al. Structural basis of glutamate recognition by a dimeric metabotropic glutamate receptor. *Nature*. 2000; 407:971–7. DOI: 10.1038/35039564 [PubMed: 11069170]
159. Namkoong J, Shin SS, Lee HJ, Marin YE, Wall BA, Goydos JS, et al. Metabotropic glutamate receptor 1 and glutamate signaling in human melanoma. *Cancer research*. 2007; 67:2298–305. DOI: 10.1158/0008-5472.CAN-06-3665 [PubMed: 17332361]
160. Pollock PM, Cohen-Solal K, Sood R, Namkoong J, Martino JJ, Koganti A, et al. Melanoma mouse model implicates metabotropic glutamate signaling in melanocytic neoplasia. *Nat Genet*. 2003; 34:108–12. DOI: 10.1038/ng1148 [PubMed: 12704387]
161. Ohtani Y, Harada T, Funasaka Y, Nakao K, Takahara C, Abdel-Daim M, et al. Metabotropic glutamate receptor subtype-1 is essential for in vivo growth of melanoma. *Oncogene*. 2008; 27:7162–70. DOI: 10.1038/onc.2008.329 [PubMed: 18776920]
162. Shin SS, Namkoong J, Wall BA, Gleason R, Lee HJ, Chen S. Oncogenic activities of metabotropic glutamate receptor 1 (Grm1) in melanocyte transformation. *Pigment cell & melanoma research*. 2008; 21:368–78. DOI: 10.1111/j.1755-148X.2008.00452.x [PubMed: 18435704]
163. Gelb T, Pshenichkin S, Rodriguez OC, Hathaway HA, Grajkowska E, DiRaddo JO, et al. Metabotropic glutamate receptor 1 acts as a dependence receptor creating a requirement for glutamate to sustain the viability and growth of human melanomas. *Oncogene*. 2015; 34:2711–20. DOI: 10.1038/onc.2014.231 [PubMed: 25065592]
164. Yamasaki T, Fujinaga M, Maeda J, Kawamura K, Yui J, Hatori A, et al. Imaging for metabotropic glutamate receptor subtype 1 in rat and monkey brains using PET with [18F]FITM. *European journal of nuclear medicine and molecular imaging*. 2012; 39:632–41. DOI: 10.1007/s00259-011-1995-6 [PubMed: 22113620]

165. Yamasaki T, Fujinaga M, Kawamura K, Yui J, Hatori A, Ohya T, et al. In vivo measurement of the affinity and density of metabotropic glutamate receptor subtype 1 in rat brain using 18F-FITM in small-animal PET. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2012; 53:1601–7. DOI: 10.2967/jnumed.112.105908
166. Fujinaga M, Xie L, Yamasaki T, Yui J, Shimoda Y, Hatori A, et al. Synthesis and evaluation of 4-halogeno-N-[4-[6-(isopropylamino)pyrimidin-4-yl]-1,3-thiazol-2-yl]-N-[11C]methylbenzamide for imaging of metabotropic glutamate 1 receptor in melanoma. *Journal of medicinal chemistry*. 2015; 58:1513–23. DOI: 10.1021/jm501845n [PubMed: 25602363]
167. Kuphal S, Bauer R, Bosserhoff AK. Integrin signaling in malignant melanoma. *Cancer metastasis reviews*. 2005; 24:195–222. DOI: 10.1007/s10555-005-1572-1 [PubMed: 15986132]
168. Peng L, Liu R, Marik J, Wang X, Takada Y, Lam KS. Combinatorial chemistry identifies high-affinity peptidomimetics against alpha4beta1 integrin for in vivo tumor imaging. *Nature chemical biology*. 2006; 2:381–9. DOI: 10.1038/nchembio798 [PubMed: 16767086]
169. Walker D, Li Y, Roxin A, Schaffer P, Adam MJ, Perrin DM. Facile synthesis and 18F-radiolabeling of alpha4beta1-specific LLP2A-aryltrifluoroborate peptidomimetic conjugates. *Bioorganic & medicinal chemistry letters*. 2016; 26:5126–31. DOI: 10.1016/j.bmcl.2016.08.011 [PubMed: 27623550]
170. Shokeen M, Zheleznyak A, Wilson JM, Jiang M, Liu R, Ferdani R, et al. Molecular imaging of very late antigen-4 (alpha4beta1 integrin) in the premetastatic niche. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2012; 53:779–86. DOI: 10.2967/jnumed.111.100073
171. Jiang M, Ferdani R, Shokeen M, Anderson CJ. Comparison of two cross-bridged macrocyclic chelators for the evaluation of 64Cu-labeled-LLP2A, a peptidomimetic ligand targeting VLA-4-positive tumors. *Nuclear medicine and biology*. 2013; 40:245–51. DOI: 10.1016/j.nucmedbio.2012.10.010 [PubMed: 23265977]
172. Beaino W, Anderson CJ. PET imaging of very late antigen-4 in melanoma: comparison of 68Ga- and 64Cu-labeled NODAGA and CB-TE1A1P-LLP2A conjugates. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2014; 55:1856–63. DOI: 10.2967/jnumed.114.144881
173. Gai Y, Sun L, Hui W, Ouyang Q, Anderson CJ, Xiang G, et al. New Bifunctional Chelator p-SCN-PhPr-NE3TA for Copper-64: Synthesis, Peptidomimetic Conjugation, Radiolabeling, and Evaluation for PET Imaging. *Inorganic chemistry*. 2016; 55:6892–901. DOI: 10.1021/acs.inorgchem.6b00395 [PubMed: 27347690]
174. Beaino W, Nedrow JR, Anderson CJ. Evaluation of (68)Ga- and (177)Lu-DOTA-PEG4-LLP2A for VLA-4-Targeted PET Imaging and Treatment of Metastatic Melanoma. *Molecular pharmaceutics*. 2015; 12:1929–38. DOI: 10.1021/mp5006917 [PubMed: 25919487]
175. Redman JM, Gibney GT, Atkins MB. Advances in immunotherapy for melanoma. *BMC medicine*. 2016; 14:20.doi: 10.1186/s12916-016-0571-0 [PubMed: 26850630]
176. Mondanelli G, Ugel S, Grohmann U, Bronte V. The immune regulation in cancer by the amino acid metabolizing enzymes ARG and IDO. *Current opinion in pharmacology*. 2017; 35:30–9. DOI: 10.1016/j.coph.2017.05.002 [PubMed: 28554057]
177. Munn DH, Mellor AL. IDO in the Tumor Microenvironment: Inflammation, Counter-Regulation, and Tolerance. *Trends in immunology*. 2016; 37:193–207. DOI: 10.1016/j.it.2016.01.002 [PubMed: 26839260]
178. Xin Y, Cai H. Improved Radiosynthesis and Biological Evaluations of L- and D-1-[18F]Fluoroethyl-Tryptophan for PET Imaging of IDO-Mediated Kynurenine Pathway of Tryptophan Metabolism. *Molecular imaging and biology : MIB : the official publication of the Academy of Molecular Imaging*. 2016; doi: 10.1007/s11307-016-1024-z
179. Huang X, Xiao X, Gillies RJ, Tian H. Design and automated production of 11C-alpha-methyl-l-tryptophan (11C-AMT). *Nuclear medicine and biology*. 2016; 43:303–8. DOI: 10.1016/j.nucmedbio.2016.02.001 [PubMed: 27150033]
180. Giglio BC, Fei H, Wang M, Wang H, He L, Feng H, et al. Synthesis of 5-[18F]Fluoro-alpha-methyl Tryptophan: New Trp Based PET Agents. *Theranostics*. 2017; 7:1524–30. DOI: 10.7150/thno.19371 [PubMed: 28529635]

181. Ehlerding EB, England CG, McNeel DG, Cai W. Molecular Imaging of Immunotherapy Targets in Cancer. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2016; 57:1487–92. DOI: 10.2967/jnumed.116.177493
182. Natarajan A, Mayer AT, Xu L, Reeves RE, Gano J, Gambhir SS. Novel Radiotracer for ImmunoPET Imaging of PD-1 Checkpoint Expression on Tumor Infiltrating Lymphocytes. *Bioconjugate chemistry*. 2015; 26:2062–9. DOI: 10.1021/acs.bioconjchem.5b00318 [PubMed: 26307602]
183. Hettich M, Braun F, Bartholoma MD, Schirmbeck R, Niedermann G. High-Resolution PET Imaging with Therapeutic Antibody-based PD-1/PD-L1 Checkpoint Tracers. *Theranostics*. 2016; 6:1629–40. DOI: 10.7150/thno.15253 [PubMed: 27446497]
184. Nedrow JR, Josefsson A, Park S, Ranka S, Roy S, Sgouros G. Imaging of programmed death ligand-1 (PD-L1): impact of protein concentration on distribution of anti-PD-L1 SPECT agent in an immunocompetent melanoma murine model. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2017; doi: 10.2967/jnumed.117.193268
185. Sharp PA. Ctr1 and its role in body copper homeostasis. *The international journal of biochemistry & cell biology*. 2003; 35:288–91. [PubMed: 12531240]
186. Chakravarty R, Chakraborty S, Dash A. ⁶⁴Cu²⁺ Ions as PET Probe: An Emerging Paradigm in Molecular Imaging of Cancer. *Molecular pharmaceutics*. 2016; 13:3601–12. DOI: 10.1021/acs.molpharmaceut.6b00582 [PubMed: 27709959]
187. Kim KI, Jang SJ, Park JH, Lee YJ, Lee TS, Woo KS, et al. Detection of increased ⁶⁴Cu uptake by human copper transporter 1 gene overexpression using PET with ⁶⁴CuCl₂ in human breast cancer xenograft model. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2014; 55:1692–8. DOI: 10.2967/jnumed.114.141127
188. Peng F, Lu X, Janisse J, Muzik O, Shields AF. PET of human prostate cancer xenografts in mice with increased uptake of ⁶⁴CuCl₂. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2006; 47:1649–52.
189. Qin C, Liu H, Chen K, Hu X, Ma X, Lan X, et al. Theranostics of malignant melanoma with ⁶⁴CuCl₂. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2014; 55:812–7. DOI: 10.2967/jnumed.113.133850
190. Jiang L, Tu Y, Hu X, Bao A, Chen H, Ma X, et al. Pilot Study of ⁶⁴Cu(I) for PET Imaging of Melanoma. *Scientific reports*. 2017; 7:2574. doi: 10.1038/s41598-017-02691-3 [PubMed: 28566692]
191. England CG, Rui L, Cai W. Lymphoma: current status of clinical and preclinical imaging with radiolabeled antibodies. *European journal of nuclear medicine and molecular imaging*. 2017; 44:517–32. DOI: 10.1007/s00259-016-3560-9 [PubMed: 27844106]
192. Jauw YW, Menke-van der Houven van Oordt CW, Hoekstra OS, Hendrikse NH, Vugts DJ, Zijlstra JM, et al. Immuno-Positron Emission Tomography with Zirconium-89-Labeled Monoclonal Antibodies in Oncology: What Can We Learn from Initial Clinical Trials? *Frontiers in pharmacology*. 2016; 7:131. doi: 10.3389/fphar.2016.00131 [PubMed: 27252651]
193. Halpern SE, Dillman RO, Witztum KF, Shega JF, Hagan PL, Burrows WM, et al. Radioimmunodetection of melanoma utilizing In-111 96.5 monoclonal antibody: a preliminary report. *Radiology*. 1985; 155:493–9. DOI: 10.1148/radiology.155.2.3983401 [PubMed: 3983401]
194. Rosenblum MG, Murray JL, Haynie TP, Glenn HJ, Jahns MF, Benjamin RS, et al. Pharmacokinetics of ¹¹¹In-labeled anti-p97 monoclonal antibody in patients with metastatic malignant melanoma. *Cancer research*. 1985; 45:2382–6. [PubMed: 3986780]
195. Dadachova E, Moadel T, Schweitzer AD, Bryan RA, Zhang T, Mints L, et al. Radiolabeled melanin-binding peptides are safe and effective in treatment of human pigmented melanoma in a mouse model of disease. *Cancer biotherapy & radiopharmaceuticals*. 2006; 21:117–29. DOI: 10.1089/cbr.2006.21.117 [PubMed: 16706632]
196. Dadachova E, Revskaya E, Sesay MA, Damania H, Boucher R, Sellers RS, et al. Pre-clinical evaluation and efficacy studies of a melanin-binding IgM antibody labeled with ¹⁸⁸Re against experimental human metastatic melanoma in nude mice. *Cancer biology & therapy*. 2008; 7:1116–27. [PubMed: 18535406]

197. Dadachova E, Nosanchuk JD, Shi L, Schweitzer AD, Frenkel A, Nosanchuk JS, et al. Dead cells in melanoma tumors provide abundant antigen for targeted delivery of ionizing radiation by a mAb to melanin. *Proceedings of the National Academy of Sciences of the United States of America*. 2004; 101:14865–70. DOI: 10.1073/pnas.0406180101 [PubMed: 15469923]
198. Revskaya E, Jongco AM, Sellers RS, Howell RC, Koba W, Guimaraes AJ, et al. Radioimmunotherapy of experimental human metastatic melanoma with melanin-binding antibodies and in combination with dacarbazine. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2009; 15:2373–9. DOI: 10.1158/1078-0432.CCR-08-2376 [PubMed: 19293257]
199. Jandt T, Revskaya E, Jiang Z, Bryan RA, Casadevall A, Dadachova E. Complement-dependent cytotoxicity of an antibody to melanin in radioimmunotherapy of metastatic melanoma. *Immunotherapy*. 2013; 5:357–64. DOI: 10.2217/imt.13.16 [PubMed: 23557419]
200. Suzuki M, Cheung NK. Disialoganglioside GD2 as a therapeutic target for human diseases. *Expert Opin Ther Targets*. 2015; 19:349–62. DOI: 10.1517/14728222.2014.986459 [PubMed: 25604432]
201. Voss SD, Smith SV, DiBartolo N, McIntosh LJ, Cyr EM, Bonab AA, et al. Positron emission tomography (PET) imaging of neuroblastoma and melanoma with ⁶⁴Cu-SarAr immunoconjugates. *Proceedings of the National Academy of Sciences of the United States of America*. 2007; 104:17489–93. DOI: 10.1073/pnas.0708436104 [PubMed: 17954911]
202. Dearling JL, Voss SD, Dunning P, Snay E, Fahey F, Smith SV, et al. Imaging cancer using PET--the effect of the bifunctional chelator on the biodistribution of a (⁶⁴Cu)-labeled antibody. *Nuclear medicine and biology*. 2011; 38:29–38. DOI: 10.1016/j.nucmedbio.2010.07.003 [PubMed: 21220127]
203. Vavere AL, Butch ER, Dearling JL, Packard AB, Navid F, Shulkin BL, et al. ⁶⁴Cu-p-NH₂-Bn-DOTA-hu14.18K322A, a PET radiotracer targeting neuroblastoma and melanoma. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2012; 53:1772–8. DOI: 10.2967/jnumed.112.104208
204. Herhaus P, Habringer S, Vag T, Steiger K, Slotta-Huspenina J, Gerngross C, et al. Response assessment with the CXCR4-directed positron emission tomography tracer [⁶⁸Ga]Pentixafor in a patient with extranodal marginal zone lymphoma of the orbital cavities. *EJNMMI research*. 2017; 7:51.doi: 10.1186/s13550-017-0294-z [PubMed: 28577295]
205. Werner RA, Weich A, Higuchi T, Schmid JS, Schirbel A, Lassmann M, et al. Imaging of Chemokine Receptor 4 Expression in Neuroendocrine Tumors - a Triple Tracer Comparative Approach. *Theranostics*. 2017; 7:1489–98. DOI: 10.7150/thno.18754 [PubMed: 28529632]
206. Lapa C, Schreder M, Schirbel A, Samnick S, Kortum KM, Herrmann K, et al. [⁶⁸Ga]Pentixafor-PET/CT for imaging of chemokine receptor CXCR4 expression in multiple myeloma - Comparison to [¹⁸F]FDG and laboratory values. *Theranostics*. 2017; 7:205–12. DOI: 10.7150/thno.16576 [PubMed: 28042328]
207. Lapa C, Luckerath K, Kleinlein I, Monoranu CM, Linsenmann T, Kessler AF, et al. (⁶⁸Ga)-Pentixafor-PET/CT for Imaging of Chemokine Receptor 4 Expression in Glioblastoma. *Theranostics*. 2016; 6:428–34. DOI: 10.7150/thno.13986 [PubMed: 26909116]
208. Derlin T, Jonigk D, Bauersachs J, Bengel FM. Molecular Imaging of Chemokine Receptor CXCR4 in Non-Small Cell Lung Cancer Using ⁶⁸Ga-Pentixafor PET/CT: Comparison With ¹⁸F-FDG. *Clinical nuclear medicine*. 2016; 41:e204–5. DOI: 10.1097/RLU.0000000000001092 [PubMed: 26756098]
209. Liang Z, Zhan W, Zhu A, Yoon Y, Lin S, Sasaki M, et al. Development of a unique small molecule modulator of CXCR4. *PloS one*. 2012; 7:e34038.doi: 10.1371/journal.pone.0034038 [PubMed: 22485156]
210. Vag T, Gerngross C, Herhaus P, Eiber M, Philipp-Abbrederis K, Graner FP, et al. First Experience with Chemokine Receptor CXCR4-Targeted PET Imaging of Patients with Solid Cancers. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2016; 57:741–6. DOI: 10.2967/jnumed.115.161034
211. Norain A, Dadachova E. Targeted Radionuclide Therapy of Melanoma. *Seminars in nuclear medicine*. 2016; 46:250–9. DOI: 10.1053/j.semnuclmed.2015.12.005 [PubMed: 27067506]

212. Kam BL, Teunissen JJ, Krenning EP, de Herder WW, Khan S, van Vliet EI, et al. Lutetium-labelled peptides for therapy of neuroendocrine tumours. *European journal of nuclear medicine and molecular imaging*. 2012; 39(Suppl 1):S103–12. DOI: 10.1007/s00259-011-2039-y [PubMed: 22388631]
213. Strosberg J, El-Haddad G, Wolin E, Hendifar A, Yao J, Chasen B, et al. Phase 3 Trial of 177Lu-Dotatate for Midgut Neuroendocrine Tumors. *The New England journal of medicine*. 2017; 376:125–35. DOI: 10.1056/NEJMoa1607427 [PubMed: 28076709]
214. Horsch D, Ezziddin S, Haug A, Gratz KF, Dunkelmann S, Miederer M, et al. Effectiveness and side-effects of peptide receptor radionuclide therapy for neuroendocrine neoplasms in Germany: A multi-institutional registry study with prospective follow-up. *Eur J Cancer*. 2016; 58:41–51. DOI: 10.1016/j.ejca.2016.01.009 [PubMed: 26943056]
215. Shibuya M. Vascular endothelial growth factor and its receptor system: physiological functions in angiogenesis and pathological roles in various diseases. *Journal of biochemistry*. 2013; 153:13–9. DOI: 10.1093/jb/mvs136 [PubMed: 23172303]
216. Ferrara N, Adamis AP. Ten years of anti-vascular endothelial growth factor therapy. *Nature reviews Drug discovery*. 2016; 15:385–403. DOI: 10.1038/nrd.2015.17
217. Camacho X, Calzada V, Fernandez M, Alonso O, Chammas R, Riva E, et al. 177Lu-DOTA-Bevacizumab: Radioimmunotherapy agent for melanoma. *Current radiopharmaceuticals*. 2016
218. Guo H, Miao Y. Melanoma targeting property of a Lu-177-labeled lactam bridge-cyclized alpha-MSH peptide. *Bioorganic & medicinal chemistry letters*. 2013; 23:2319–23. DOI: 10.1016/j.bmcl.2013.02.069 [PubMed: 23473679]
219. Joyal JL, Barrett JA, Marquis JC, Chen J, Hillier SM, Maresca KP, et al. Preclinical evaluation of an 131I-labeled benzamide for targeted radiotherapy of metastatic melanoma. *Cancer research*. 2010; 70:4045–53. DOI: 10.1158/0008-5472.CAN-09-4414 [PubMed: 20442292]
220. Bonnet M, Mishellany F, Papon J, Cayre A, Penault-Llorca F, Madelmont JC, et al. Anti-melanoma efficacy of internal radionuclide therapy in relation to melanin target distribution. *Pigment cell & melanoma research*. 2010; 23:e1–11. DOI: 10.1111/j.1755-148X.2010.00716.x
221. Viallard C, Perrot Y, Boudhraa Z, Jouberton E, Miot-Noirault E, Bonnet M, et al. [(1)(2)(3)I]ICF01012 melanoma imaging and [(1)(3)(1)I]ICF01012 dosimetry allow adapted internal targeted radiotherapy in preclinical melanoma models. *European journal of dermatology : EJD*. 2015; 25:29–35. DOI: 10.1684/ejd.2014.2481 [PubMed: 25548082]
222. Viallard C, Chezal JM, Mishellany F, Ranchon-Cole I, Pereira B, Herbette A, et al. Targeting DNA repair by coDbait enhances melanoma targeted radionuclide therapy. *Oncotarget*. 2016; 7:12927–36. DOI: 10.18632/oncotarget.7340 [PubMed: 26887045]
223. Mier W, Kratochwil C, Hassel JC, Giesel FL, Beijer B, Babich JW, et al. Radiopharmaceutical therapy of patients with metastasized melanoma with the melanin-binding benzamide 131I-BA52. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine*. 2014; 55:9–14. DOI: 10.2967/jnumed.112.112789
224. Maisonia A, Kuhnast B, Papon J, Boisgard R, Bayle M, Vidal A, et al. Single photon emission computed tomography/positron emission tomography imaging and targeted radionuclide therapy of melanoma: new multimodal fluorinated and iodinated radiotracers. *Journal of medicinal chemistry*. 2011; 54:2745–66. DOI: 10.1021/jm101574q [PubMed: 21417462]
225. Maisonia A, Billaud EM, Besse S, Rbah-Vidal L, Papon J, Audin L, et al. Synthesis, radioiodination and in vivo screening of novel potent iodinated and fluorinated radiotracers as melanoma imaging and therapeutic probes. *European journal of medicinal chemistry*. 2013; 63:840–53. DOI: 10.1016/j.ejmech.2012.11.047 [PubMed: 23603044]
226. Rbah-Vidal L, Vidal A, Billaud EM, Besse S, Ranchon-Cole I, Mishellany F, et al. Theranostic Approach for Metastatic Pigmented Melanoma Using ICF15002, a Multimodal Radiotracer for Both PET Imaging and Targeted Radionuclide Therapy. *Neoplasia*. 2017; 19:17–27. DOI: 10.1016/j.neo.2016.11.001 [PubMed: 27987437]
227. Morton JJ, Bird G, Refaeli Y, Jimeno A. Humanized Mouse Xenograft Models: Narrowing the Tumor-Microenvironment Gap. *Cancer research*. 2016; 76:6153–8. DOI: 10.1158/0008-5472.CAN-16-1260 [PubMed: 27587540]

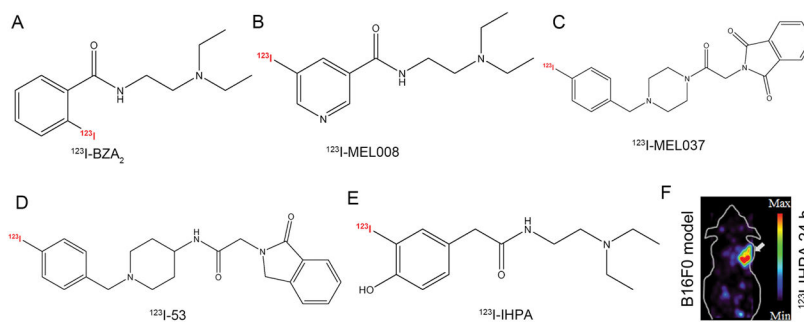


Figure 1. Representative melanin-targeting probes. Chemical structures of (A) ^{123}I -BZA₂, (B) ^{123}I -MEL008, (C) ^{123}I -MEL037, (D) ^{123}I -53, and (E) ^{123}I -IHPA. (F) MicroSPECT image of C57BL/6 mice bearing B16/F0 melanotic melanoma at 24 h postinjection of approximately 11.1 MBq of ^{123}I -IHPA, the tumor is indicated by the white arrow. Adapted and modified with permission from references [34, 36, 41, 42, 44].

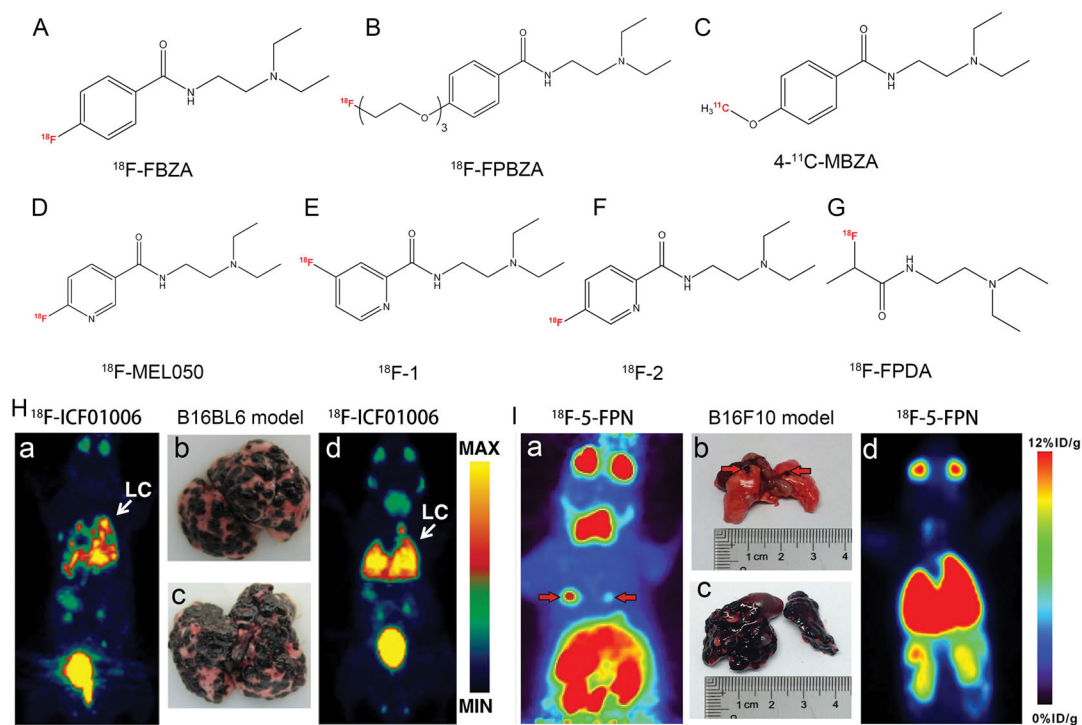


Figure 2.

Representative melanin-targeting probes and *in vivo* study images. Chemical structures of (A) ^{18}F -FBZA, (B) ^{18}F -FPBZA, (C) $4\text{-}^{11}\text{C}$ -MBZA, (D) ^{18}F -MEL050, (E) ^{18}F -1, (F) ^{18}F -2, and (G) ^{18}F -FPDA. (H) Whole-body maximum intensity projection (MIP) images of ^{18}F -ICF01006 and corresponding lung photographs of B16/BL6 melanoma-bearing mice at the early stage (a, b) and late stage (c, d) of tumor development. (I) ^{18}F -5-FPN PET images of two mice with lung metastases from melanoma. Note that this probe was able to detect both micrometastases (a, b) and wide spread lung metastases (c, d) from melanoma. Tumors are indicated by red arrows. Adapted and modified with permission from references [45–47, 49, 51, 52, 66, 67].

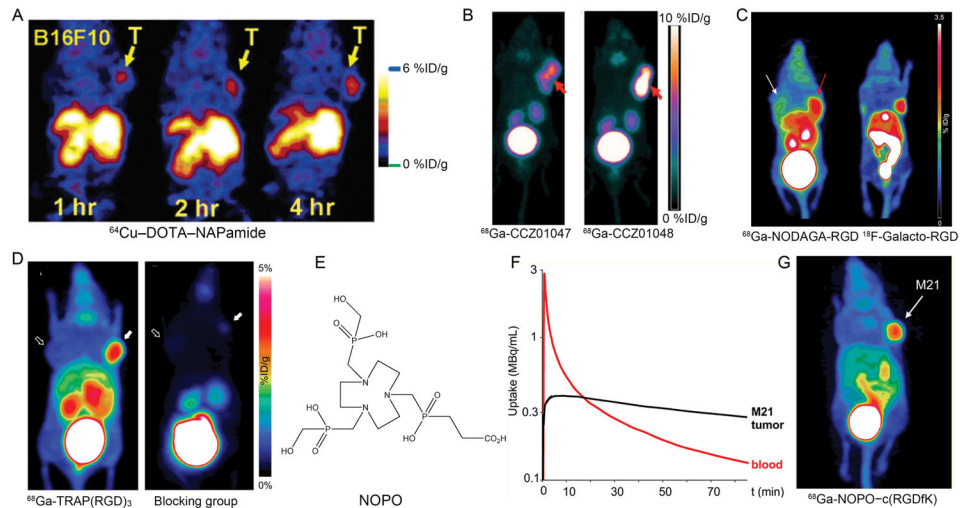


Figure 3.

Examples of peptide-based imaging of melanoma. (A) Coronal microPET images of mice bearing B16/F10 tumors at different time points after tail vein injection of ^{64}Cu -DOTA-NAPamide. (B) PET images of ^{68}Ga -CCZ01047 and ^{68}Ga -CCZ01048 at 1 h postinjection of the corresponding tracer in mice bearing B16/F10 tumors. (C) MIP PET images of mice bearing M21 and M21L tumor xenografts on right and left shoulder, respectively. ^{68}Ga -NODAGA-RGD (left) showed more intense tumor uptake than ^{18}F -Galacto-RGD (right) in $\alpha_v\beta_3$ positive M21 melanoma models. (D) MIP images of microPET scans of M21 (solid arrows) and M21L (outline arrows) human melanoma models. ^{68}Ga -TRAP(RGD)₃ showed high-contrast visualization of the M21 tumor. (E) Chemical structure of bifunctional chelator NOPO. (F) The relative uptake of ^{68}Ga -NOPO-c(RGDfK) in M21 tumor tissue and blood over time. (G) MIP of ^{68}Ga -NOPO-c(RGDfK) PET imaging in M21/M21L xenografted mice. Tumor site was marked with white arrow. Adapted and modified with permission from references [96, 105, 120, 130–132].

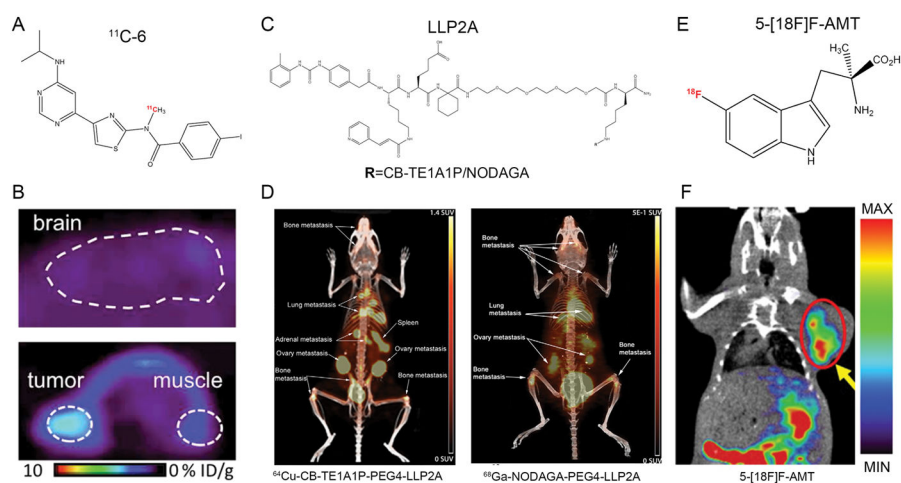


Figure 4.

Other PET probes for melanoma imaging. (A) Chemical structure of ^{11}C -6 for imaging of mGlu1 receptor. (B) Representative PET images of mice bearing B16/F10 after injection of ^{11}C -6. Upper: sagittal image of the brain. Lower: Axial image of tumor and muscle. (C) Chemical structures of NODAGA-PEG4-LLP2A and CB-TE1A1P-PEG4-LLP2A. (D) Small animal PET/CT imaging at 2 h after injection of the radiotracers (7.4 MBq). Both ^{64}Cu -CB-TE1A1P-PEG4-LLP2A and ^{68}Ga -NODAGA-PEG4-LLP2A were able to image melanoma lung metastases with high contrast and minimal lung background. (E) Chemical structure of 5-[^{18}F]F-AMT. (F) Coronal PET/CT image of B16/F10 melanoma 30 min after injection of 5-[^{18}F]F-AMT. Adapted and modified with permission from references [166, 172, 180].

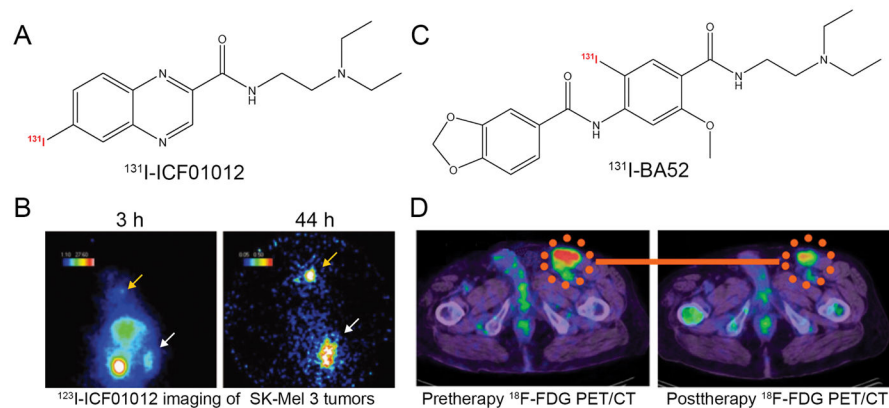


Figure 5. Representative radionuclide-labeled therapeutic agents for melanoma. (A) Chemical structure of ^{131}I -ICF01012. (B) Gamma-camera imaging of SK-Mel 3 melanoma-bearing mice after injection of 3.7 MBq ^{123}I -ICF01012. A clear concentration of ^{123}I -ICF01012 occurred 3 hours after radiotracer administration and a rapid elimination of ^{123}I -ICF01012 from non-specific organs was observed at 44 h postinjection. Tumor and thyroid were indicated by white and orange arrows, respectively. (C) Chemical structure of ^{131}I -BA52. (D) ^{18}F -FDG PET/CT examinations in a melanoma patient before and after ^{131}I -BA52 treatment. After treatment using ^{131}I -BA52, post-therapeutic ^{18}F -FDG PET/CT examination demonstrated that SUV of the inguinal lymph node metastasis decreased from 9.02 to 5.81. Adapted and modified with permission from references [221, 223].

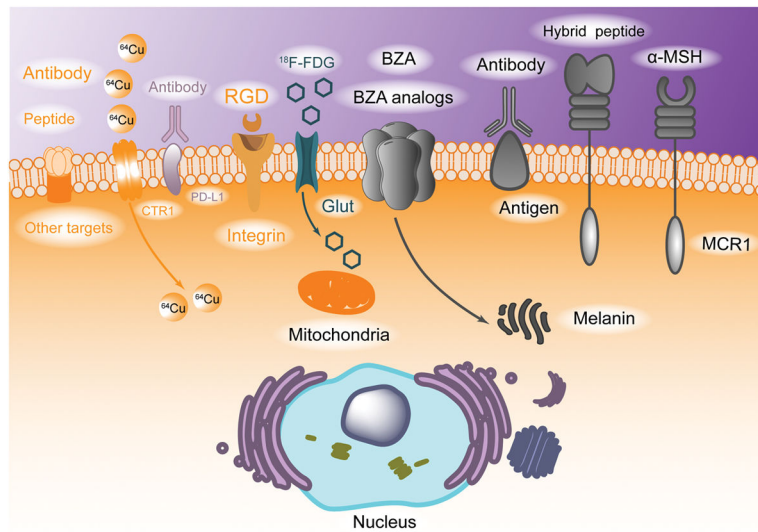


Figure 6. Pictorial abstract showing PET and SPECT imaging probes for malignant melanoma. Molecular targets discussed above can be divided into two groups, melanoma specific targets which are indicated by grey/black models and melanoma nonspecific targets which are indicated by colorful models.