



Improved Bactec MGIT 960 Pyrazinamide Test Decreases Detection of False *Mycobacterium tuberculosis* Pyrazinamide Resistance

Alessandro Mustazzolu,^a Angelo Iacobino,^a Federico Giannoni,^a Claudio Piersimoni,^b the Italian Multicentre Study on Resistance to Antituberculosis Drugs (SMIRA) Group, Lanfranco Fattorini^a

Istituto Superiore di Sanità, Department of Infectious Diseases, Rome, Italy^a; Regional Reference Mycobacteria Laboratory, United Hospitals, Ancona, Italy^b

KEYWORDS Bactec MGIT 960, false resistance, *Mycobacterium tuberculosis*, tuberculosis, pyrazinamide

Pyrazinamide (PZA) is a key drug for the treatment of tuberculosis (TB). Resistance to PZA is caused mostly by mutations in the *pncA* gene, encoding pyrazinamidase, which converts the prodrug PZA to the active form, pyrazinoic acid (1, 2). For testing PZA susceptibility, the World Health Organization (WHO) recommends performance of the assay in liquid medium at pH 5.9 in the Bactec MGIT 960 (M960) system (Becton, Dickinson, Sparks, MD, USA) (3). However, false resistance to PZA was reported for this phenotypic assay (4–7) due to a high *Mycobacterium tuberculosis* inoculum, which may impair pyrazinamidase activity by increasing the pH of the medium (8). Indeed, a reduced M960 inoculum decreased detection of false resistance (9, 10) when results were compared with those of the previous reference radiometric method Bactec 460 (Becton, Dickinson) (11) and with *pncA* sequencing, a method providing from 83% to 90% sensitivity (2, 12, 13). Since 2013, the WHO has yearly offered to the global Supranational Reference Laboratory (SRL) network proficiency test panels of *M. tuberculosis* strains for PZA drug susceptibility testing (DST).

At the SRL in Rome, the PZA M960 assay is performed according to the manufacturer's instructions (14), with minor modifications. Briefly, a positive MGIT tube obtained 1 or 2 days after the positivity signal of the M960 instrument (seed tube) is vortexed for 30 s and then allowed to settle for 20 to 30 min. Thereafter, a 1-ml aliquot of the settled seed tube is taken with a 1-ml pipet from the top surface, instead of lower down. Of this aliquot, 0.5 ml is transferred to the PZA test tube containing 100 μg/ml of PZA and 0.5 ml is transferred to a tube containing 4.5 ml of sterile saline. This 1:10 dilution tube is repeatedly mixed with a new pipet, and 0.5 ml is used to inoculate the tube without PZA (growth control tube). After inversion, both the growth control and PZA test tubes are incubated in the M960 instrument. The seed tube is used first for PZA DST and then for other drugs.

In 2013 to 2016, the SRL tested the PZA susceptibility of 106 WHO *M. tuberculosis* strains (41 were PZA resistant and 65 were susceptible) with known *pncA* mutations. Using the modified M960 PZA assay (MMPA), 1 of 106 strains was falsely resistant (0.9%) and no strain was falsely susceptible. In Italy, the SRL coordinates a laboratory network (SMIRA [Italian Multicenter Study on Resistance to Antituberculosis Drugs]) periodically examined by first- and second-line drug proficiency testing exercises (15, 16). In 2016, 17 SMIRA laboratories performed in parallel the standard M960 PZA assay (14) and the MMPA on 10 *M. tuberculosis* strains from the 21st WHO round (4 were resistant and 6

Accepted manuscript posted online 13 September 2017

Citation Mustazzolu A, Iacobino A, Giannoni F, Piersimoni C, the Italian Multicentre Study on Resistance to Antituberculosis Drugs (SMIRA) Group, Fattorini L. 2017. Improved Bactec MGIT 960 pyrazinamide test decreases detection of false *Mycobacterium tuberculosis* pyrazinamide resistance. *J Clin Microbiol* 55:3552–3553. <https://doi.org/10.1128/JCM.01437-17>.

Editor Geoffrey A. Land, Carter BloodCare and Baylor University Medical Center

Copyright © 2017 American Society for Microbiology. All Rights Reserved.

Address correspondence to Lanfranco Fattorini, lanfranco.fattorini@iss.it.

susceptible). Out of a total of 170 strains (68 were resistant and 102 susceptible) examined by each of the two methods, 8/170 showed false resistance by the standard assay (4.7%) and 2/170 showed false resistance by MMPA (1.2%); no strain was falsely susceptible. Overall, these observations suggest that the MMPA performed by withdrawing inoculum from the top surface of the settled MGIT 960 seed tubes may be useful in decreasing the finding of false phenotypic PZA resistance.

ACKNOWLEDGMENTS

The members of the SMIRA laboratory network in Italy involved in this study were Ester Mazzola (Hospital Niguarda, Milan), Paola Cichero (Hospital San Raffaele, Milan), Alessandra Lombardi (Hospital L. Sacco, Milan), Elena Libanori (Hospital of Sondalo), Piero Marone and Vincenzina Monzillo (Hospital San Matteo, Pavia), Marco Arosio and Claudio Farina (Hospital Giovanni XXIII, Bergamo), Marta Peracchi (Hospital of Padua), Claudio Scarparo (Hospital of Udine), Eliana Frizzera (Hospital of Bolzano), Paola Pietrosevoli (Hospital of Modena), Marina Matteucci (AUSL of Romagna-Cesena), Laura Rindi (Hospital of Pisa), Eugenio Luciano and Antonella Mencacci (Hospital of Perugia), Nicoletta Nuzzolese and Eustachio Vitullo (Hospital of Matera), Saveria Dodaro and Cristina Giraldi (Hospital of Cosenza), Salvatore Nisticò and Maria Vinci (Hospital of Lamezia Terme), and Giovanni Salvatore Podda (Hospital Santissima Trinità, Cagliari).

We thank Armand van Deun, Institute of Tropical Medicine, Antwerp, Belgium, Coordinator of the WHO SRL network, for sending the *M. tuberculosis* isolates to L. Fattorini (SRL of Rome) for annual proficiency testing of anti-TB drugs.

This study was supported in part by the CCM Project of the Italian Ministry of Health.

REFERENCES

- Zhang Y, Shi W, Zhang W, Mitchison D. 2014. Mechanisms of pyrazinamide action and resistance. *Microbiol Spectr* 2:MGM2-0023-2013. <https://doi.org/10.1128/microbiolspec.MGM2-0023-2013>.
- Ramirez-Busby SM, Rodwell TC, Fink L, Catanzaro D, Jackson RL, Pettigrove M, Catanzaro A, Valafar F. 2017. A multinational analysis of mutations and heterogeneity in PZase, RpsA, and PanD associated with pyrazinamide resistance in M/XDR *Mycobacterium tuberculosis*. *Sci Rep* 7:3790. <https://doi.org/10.1038/s41598-017-03452-y>.
- World Health Organization. 2014. Companion handbook to the WHO guidelines for the programmatic management of drug-resistant tuberculosis. WHO/HTM/TB/2014.11. World Health Organization, Geneva, Switzerland.
- Pandey S, Newton S, Upton A, Roberts S, Drinković D. 2009. Characterisation of *pncA* mutations in clinical *Mycobacterium tuberculosis* isolates in New Zealand. *Pathology* 41:582–584. <https://doi.org/10.1080/00313020903071587>.
- Chedore P, Bertucci L, Wolfe J, Sharma M, Jamieson F. 2010. Potential for erroneous results indicating resistance when using the Bactec MGIT 960 system for testing susceptibility of *Mycobacterium tuberculosis* to pyrazinamide. *J Clin Microbiol* 48:300–301. <https://doi.org/10.1128/JCM.01775-09>.
- Simons SO, van Ingen J, van der Laan T, Mulder A, Dekhuijzen PN, Boeree MJ, van Soolingen D. 2012. Validation of *pncA* gene sequencing in combination with the mycobacterial growth indicator tube method to test susceptibility of *Mycobacterium tuberculosis* to pyrazinamide. *J Clin Microbiol* 50:428–434. <https://doi.org/10.1128/JCM.05435-11>.
- Hoffner S, Angeby K, Sturegård E, Jönsson B, Johansson A, Sellin M, Werngren J. 2013. Proficiency of drug susceptibility testing of *Mycobacterium tuberculosis* against pyrazinamide: the Swedish experience. *Int J Tuberc Lung Dis* 17:1486–1490. <https://doi.org/10.5588/ijtld.13.0195>.
- Zhang Y, Permar S, Sun Z. 2002. Conditions that may affect the results of susceptibility testing of *Mycobacterium tuberculosis* to pyrazinamide. *J Med Microbiol* 51:42–49. <https://doi.org/10.1099/0022-1317-51-1-42>.
- Piersimoni C, Mustazzolu A, Giannoni F, Bornigia S, Gherardi G, Fattorini L. 2013. Prevention of false resistance results obtained in testing the susceptibility of *Mycobacterium tuberculosis* to pyrazinamide with the Bactec MGIT 960 system using a reduced inoculum. *J Clin Microbiol* 51:291–294. <https://doi.org/10.1128/JCM.01838-12>.
- Piersimoni C, Mustazzolu A, Iacobino A, Giannoni F, Santoro G, Gherardi G, Del Giudice A, Perna R, Fattorini L. 2016. Pyrazinamide susceptibility testing: proposed new standard with the BACTEC™ MGIT™ 960 system. *Int J Tuberc Lung Dis* 20:1677–1680. <https://doi.org/10.5588/ijtld.16.0360>.
- Clinical and Laboratory Standards Institute. 2011. Susceptibility testing of mycobacteria, nocardiae, and other aerobic actinomycetes, 2nd ed. Approved standard. CLSI document M24-A2. CLSI, Wayne, PA.
- Miotto P, Cabibbe AM, Feuerriegel S, Casali N, Drobniewski F, Rodionova Y, Bakonyte D, Stakenas P, Pimkina E, Augustynowicz-Kopeć E, Degano M, Ambrosi A, Hoffner S, Mansjö M, Werngren J, Rüsç-Gerdes S, Niemann S, Cirillo DM. 2014. *Mycobacterium tuberculosis* pyrazinamide resistance determinants: a multicenter study. *mBio* 5(5):e01819-14. <https://doi.org/10.1128/mBio.01819-14>.
- Ramirez-Busby SM, Valafar F. 2015. Systematic review of mutations in pyrazinamidase associated with pyrazinamide resistance in *Mycobacterium tuberculosis* clinical isolates. *Antimicrob Agents Chemother* 59:5267–5277. <https://doi.org/10.1128/AAC.00204-15>.
- Becton, Dickinson and Company. BD BACTEC™ MGIT™ 960 PZA kit for the antimycobacterial susceptibility testing of *Mycobacterium tuberculosis*. Package insert L005686JAA(01) 2014-03. Becton, Dickinson and Company, Sparks, MD.
- Fattorini L, Migliori GB, Cassone A, Mustazzolu A, Piccaro G, Filippini P, Cirillo DM, Borroni E, Italian Multicentre Study on Resistance to Antituberculosis Drugs Group. 2012. Proficiency testing of first- and second-line anti-tuberculosis drugs in Italy. *Eur Respir J* 39:1263–1266. <https://doi.org/10.1183/09031936.00129011>.
- Fattorini L, Mustazzolu A, Borroni E, Piccaro G, Giannoni F, Cirillo DM, Italian Multicentre Study on Resistance to Antituberculosis Drugs Group. 2016. Tuberculosis in migrants from 106 countries, 2008–2014. *Eur Respir J* 47:1273–1276. <https://doi.org/10.1183/13993003.01844-2015>.