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NFAT3/c4-mediated excitotoxicity in hippocampal apoptosis during radiation-induced brain injury

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ABSTRACT

Whole brain irradiation (WBI) has become an indispensible tool in the treatment of head and neck cancer, and it has greatly improved patient survival rate and total survival time. In addition, prophylactic cranial irradiation (PCI) has dramatically decreased the incidence of brain metastatic carcinoma. However, WBI may induce temporary functional deficits or even progressive, irreversible cognitive dysfunction that compromises the quality of life for survivors. Unfortunately, the exact molecular mechanisms for cognitive damage remain elusive, and no treatment or preventative measures are available for use in the clinic. In the present study, the nuclear factor of activated T cells isoform 4 (NFAT3/c4) was found to play a vital role in excitotoxic hippocampus cell apoptosis induced by radiation. Sprague-Dawley (SD) rats received 20 Gy WBI, after which we detected NFAT3/c4-mediated excitotoxicity. We found that radiation caused hippocampus excitotoxicity, resulting from overactivation of the N-methyl-D-aspartate receptor (NMDAR) and always accompanied by subsequent elevation of the intracellular calcium level and activation of calcineurin (CaN). P-NFAT3/c4 was the principal downstream target of CaN, including regulation of its nuclear translocation as well as transcriptional activities. Radiation recruited NMDAR/NFAT3/c4 activation and subsequent Bax induction in hippocampus cells. Once treated with the NFAT3/c4 inhibitor 11R-VIVIT peptide pre-irradiation, hippocampal proliferation and neuron survival (dentate gyrus cells in particular) were protected from radiation-induced injury, resulting in inhibition of the apoptosis marker Bax. Our principal aim was to illuminate the role of NFAT3/c4-mediated excitotoxicity in hippocampal apoptosis during radiation-induced brain injury. This study is the first time that radiation-induced activation of NFAT3/c4 has been recorded, and our results suggest that NFAT3/c4 may be a novel target for prevention and treatment of radiation-induced brain injury.

KEYWORDS: NMDAR, calcineurin, NFAT3/c4, apoptosis, Bax, radiation

INTRODUCTION

Radiation therapy (RT), especially whole brain irradiation (WBI), has become an essential treatment method for primary and metastatic brain tumors. With the rapid development of modern irradiation techniques, many patients receive WBI each year, and the survival of patients has increased. However, the normal brain tissue damage that may inevitably occur is often accompanied by neurological complications, such as temporary brain functional deficits, or

even progressive, irreversible cognitive dysfunction [1]. These dysfunctions seriously affect the quality of life for survivors [2]. The exact mechanisms of radiation-induced cognitive decline have not been completely characterized. It was hypothesized that injury to hippocampal proliferation and neuronal survival may be central to the pathogenesis of radiation-induced cognitive decline [3, 4].

The N-methyl-D-aspartate receptor (NMDAR) is widespread in the CNS and is associated with multiple physiological and pathological processes [5-7], especially the occurrence of excitotoxic neuronal cell death and receptor overactivation [8,9]. The main downstream events of NMDAR overactivation are calcium influx and calcineurin (CaN) activation [10, 11]. CaN subsequently mediates NFAT3/c4 nuclear translocation and its transcriptional activities.

The transcription factor NFAT3/c4 has been shown to be active in neurogenic brain regions such as the hippocampus [12]. Previous studies have shown that the NFATc4/3-mediated pathway plays an important role in long-term changes in neuronal function [13]. Cognitive decline in Alzheimer's disease and traumatic brain injury have been associated with selective changes in CaN/NFAT signaling [14, 15]. Upon stimulation of CaN, NFAT3/c4 is dephosphorylated and then translocated from the cytosol into the nucleus, whereas it is highly phosphorylated in the cytoplasm of resting cells. The re-phosphorylation and subsequent export of NFAT3/c4 from the nucleus is mediated by several kinases, such as GSK- 3β .

In this study, our data show that the NMDAR/CaN/NFAT3/c4 pathway initiates induction of the neuronal apoptosis marker Bax and hippocampal cell apoptosis. Such results are validated by dentate gyrus (DG) cell injuries, which improved after treatment with NFAT3/c4 inhibitor 11R-VIVIT peptide pre-irradiation.

MATERIALS AND METHODS Animals

A total of 120 male Sprague–Dawley rats (21 days old) weighing ~50–60 g were purchased from the Experimental Animal Center of Soochow University (Suzhou, China). The experimental animals were raised in temperature- and humidity-controlled conditions with a 12 h/12 h light/dark cycle (lights on at 7 AM) at 22 \pm 2°C and free access to food and water ad libitum prior to the experiment. All the rules and regulations involving the care and protection of animals complied with the Soochow University Animal Care and Ethics Guidelines, which agree with national laboratory animal care standards.

The animals were divided into four groups: (i) the sham group (Sham), (ii) the sham with 11R-VIVIT peptide injection group (Sham +11R-VIVIT peptide), (iii) the irradiation group (IR), and (iv) the irradiation and 11R-VIVIT peptide injection group (IR +11R-VIVIT peptide) (n = 30 per group).

Irradiation

The rats were anesthetized with 3.6% chloral hydrate $(360\,\text{mg/kg})$ via intraperitoneal injection. WBI treatments were administered with a single dose of 0 (control) or 20 Gy using a 4 MeV electron beam and a linear accelerator (Philips SL-18, UK) at room temperature.

Brain and body weight reduction

Six rats were randomly selected at 2 months post irradiation to be weighed and sacrificed. Their brains were collected and weighed. Brain and body weight reductions were calculated.

Drug treatment

The 11R-VIVIT peptide (RRRRR-GGG-MAGPHPVIVITGPHEE) was purchased from Sigma Genosys (Woodlands, TX). The dosage of the 11R-VIVIT peptide used for each rat was $100\,\mu\text{g}/\text{kg}$. Both the second group and the fourth group were injected intraperitoneally with

11R-VIVIT peptide for 3 days before radiation. Then, the first group of animals was sacrificed and hippocampus tissues were collected for western blot assays. The second group of animals were treated with 11R-VIVIT peptide for 3 days post irradiation and then sacrificed for immunofluorescence staining of hippocampus-proliferating cells. The third group of animals were treated with 11R-VIVIT peptide for 7 days post irradiation and were sacrificed for immunofluorescence staining of mature hippocampal neurons eight weeks after irradiation (n=3 per group per time point).

5-Bromodeoxyuridine (BrdU) labeling and immunofluorescence staining

BrdU (Sigma, St Louis, MO, USA) at 50 mg/kg/day was administered intraperitoneally (i.p.) for 7 consecutive days (twice a day) to animals 24 h before they were sacrificed at 3 days post irradiation. The rats were perfused with cold PBS and 4% paraformaldehyde, followed by incubation in the paraformaldehyde for 24 h at 4°C. The tissue was dehydrated in 15% and 30% sucrose for 48 h. Frozen serial sections of the brains were cut (30 μm thick) through the entire hippocampus using a microtome.

The tissue slices were treated with 2 M HCl at 45°C for 30 min to denature the DNA, and then the reaction was neutralized in 0.1 M borate buffer (pH 8.5) for 10 min. The sections were incubated in buffer (10% bovine serum albumin and 0.5% Triton X-100 in PBS) to block non-specific binding for 2 h at room temperature. The slices were incubated overnight at 4°C with a primary antibody. The tissues were washed in PBS and incubated with a secondary antibody for 1.5 h, and we used an Olympus microscope equipped with a digital camera (Olympus) to analyze the sections. The total number of BrdU+ or NeuN+ cells was counted on every ninth section throughout the hippocampus (seven sections per animal). The number of positives per DG of the hippocampus was obtained by multiplying the value by 9. In addition, we had three people perform the counts independently and calculated the mean value as the ultimate result. The antibodies were purchased and diluted as follows: rat anti-body BrdU (1:500, Biolegend), NeuN (1:500, Abcam), Alexa Fluor 488 donkey anti-goat, Alexa Fluor 555 goat anti-rat, and Alexa Fluor 405 mouse anti-rabbit (1:500, Jackson ImmunoResearch, USA).

Western blot

Four groups of rat hippocampus tissues were used to extract the total protein, and NFAT3/c4 protein was extracted using a Nuclear and Cytoplasmic Extraction Kit (Cwbio, China). All the experiments were conducted on ice. Protein concentration measurements were collected using the BCA (Biomiga, China) method. The samples were incubated at 99°C for 10 min. Then, 30 μ g of total protein were was separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE). The separated proteins were transferred to PVDF membranes (Millipore, Billerica, MA, USA). The membranes were blocked with 5% non-fat milk in TBST for 2 h at room temperature and were incubated overnight at 4°C with the proper primary antibodies. The antibodies were purchased and diluted as follows: rabbit anti-calcineurin (1:10 000, Abcam), rabbit anti-NMDAR (1:1000, Abcam), rabbit anti-NFAT3/c4 (1:500, Sigma), rabbit anti-GSK-3 β (1:1000, Cell Signal), rabbit anti-Bax (1:1000, Cell Signal), rabbit anti-Bcl-2 (1:1000, Cell Signal), rabbit

anti-GAPDH (1:1000, Goodhere), rabbit anti-Histon3 (1:500, Sigma). The membranes were then incubated with an HRP-conjugated secondary antibody (Beyotime, Nantong, China) for 1.5 h at room temperature. The membranes were immunolabeled. After washing the samples with TBST buffer, the membranes were subsequently assayed using the Thermo Scientific Pierce Femto ECL (Rockford, IL, USA).

Statistical analysis

Data were expressed as the mean \pm SD and analyzed with a two-tiered Student's t test or one-way ANOVA in GraphPad Prism (version 5.0; GraphPad, La Jolla, USA). A value of P < 0.05 was considered to be statistically significant.

RESULTS

Body weight and brain weight measurements

Obvious depilation was observed in the irradiated rats at 2-3 weeks post irradiation, which gradually disappeared and was absent 2 months later (Fig. 1A). Compared with the average body weight of the sham group, that of the irradiation group declined (P < 0.05, Fig. 1B), and the same trend was observed for average brain weight (P < 0.05, Fig. 1C). The 11R-VIVIT peptide had no impact on either body weight or brain weight (P > 0.05, Fig. 1B–C).

Cognitive deficits were produced after whole-brain irradiation

Our previous study demonstrated that 20 Gy of WBI resulted in cognitive damage in the surviving rats at 2 months post irradiation [16].

Whole-brain radiation activated the NMDAR/NFAT3/ c4/Bax pathway and induced apoptosis

WBI significantly elevated NMDAR subunit expression (NR2A, not NR1 or NR2B) in the hippocampus at 6 h post irradiation compared with that of the sham group (P < 0.05, Fig. 2A).

The expression level of CaN and GSK-3ß experienced only mild enhancement (P > 0.05, Fig. 2D), but these molecules were significantly activated, as demonstrated by the translocation of NFAT3/c4 between the nuclei and cytoplasm. With radiation stress, NFAT3/c4 rapidly accumulated in the nuclei after dephosphorylation of p-NFAT3/c4 (P < 0.05, Fig. 2B). The transcriptional activities of NFAT3/c4 promoted expression of the apoptosis marker Bax (P < 0.05, Fig. 2C).

The 11R-VIVIT peptide rescued the activation of NFAT3/c4-dependent apoptosis after radiation

Treatment with the 11R-VIVIT peptide did not have any impact on the expression of the NMDAR subunits (NR1, NR2A or NR2B) (P > 0.05, Fig. 3A). However, the 11R-VIVIT peptide prevented nuclear translocation of NFAT3/c4 and abolished its nuclear accumulation (P < 0.05, Fig. 3B). Subsequently, the expression of the apoptosis marker Bax was inhibited. (P < 0.05, Fig. 3C).

The 11R-VIVIT peptide alleviated the decline in the DG proliferating cells and mature neurons

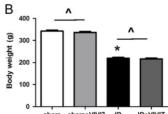
The protective effects of the 11R-VIVIT peptide on radiationinduced brain injury were assessed by analysis of BrdU proliferation cell assay and NeuN immunofluorescence staining. Our data showed that the IR+11R-VIVIT peptide promoted an increase in BrdU+ cells in the DG at 3 days post irradiation compared with the levels in the irradiated group (P < 0.05, Fig. 4A). Similarly, we found that the total number of NeuN+ mature neurons in the IR+ 11R-VIVIT peptide group increased compared with the number in the IR group at 8 weeks post irradiation (P < 0.05, Fig. 4B).

DISCUSSION

In the present study, we investigated hippocampus cell apoptosis in a model of radiation-induced brain injury that was characterized by cognitive dysfunction. We also evaluated the potential mechanisms of NFAT3/c4-mediated excitotoxicity. A single-dose exposure of 20 Gy WBI was chosen to establish the model of radiation-induced brain injury in rats. We showed that: (i) WBI significantly induced hippocampus cell apoptosis and reduction of DG proliferating cells as well as mature neurons; (ii) WBI activated the NMDAR/ NFAT3/c4/Bax pathway; (iii) 11R-VIVIT peptide, an NFAT3/c4 inhibitor, abolished the nuclear translocation of NFAT3/c4 and the induction of Bax; and (iv) the 11R-VIVIT peptide reversed the decline in the number of DG proliferating cells and mature neurons by protecting hippocampal proliferation and neuron survival.

An animal model for radiation-induced brain injury has not yet been clearly defined. Wong-Goodrich et al. employed a single dose





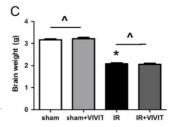


Fig. 1. The observation of the rats' skin reaction and the measurement of body weight and brain weight after WBI (20 Gy). We observed the representative infield depilation at 2-3 weeks post irradiation (A). Ironizing radiation induced a significant decrease in the body weight of rats, and 11R-VIVIT had no remarkable effect (B). Similarly, we observed a conspicuous decrease in the brain weight of irradiated rats, and 11R-VIVIT caused no remarkable changes with respect to this (C). Data are expressed as mean \pm SD, *P < 0.05 vs sham group, #P < 0.05 vs irradiation group.

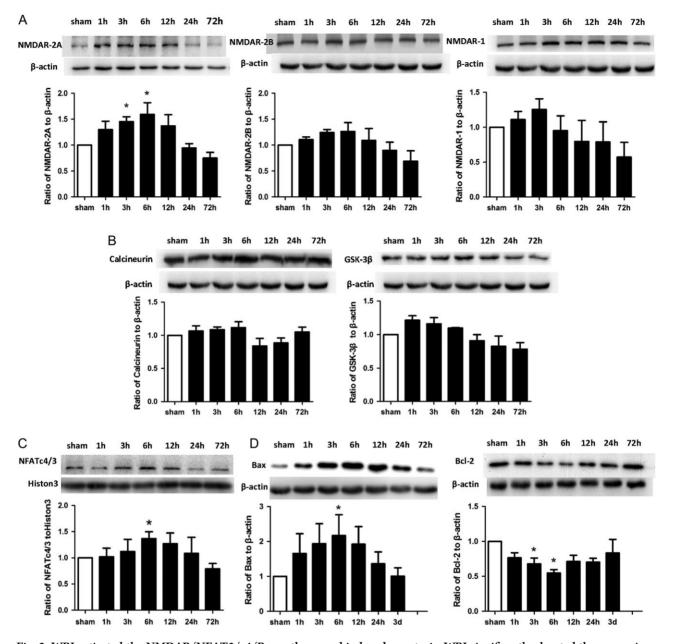


Fig. 2. WBI activated the NMDAR/NFAT3/c4/Bax pathway and induced apoptosis. WBI significantly elevated the expression of: NMDAR subunits (NR2A, not NR1 or NR2B) (A), NFAT3/c4 in the nucleus (C), the apoptosis marker Bax (D) compared with in the sham group. In contrast, the expression of CaN and GSK-3 β showed only mild enhancement compared with in the sham group (B). Data are expressed as mean \pm SD, *P < 0.05 vs sham group, #P < 0.05 vs irradiation group.

of 5 Gy to produce cognitive dysfunction in C57BL/J6 male mice [17], while Conner *et al.* selected a total dose of 40 Gy/8 fractions to establish a model of radiation-induced brain injury with Fischer 344 rats [18]. A large single-dose exposure was more damaging than fractionated irradiation because the former lead to strong reaction in the tissue. Our preliminary work indicated that it was appropriate to apply a single-dose exposure of 20 Gy to establish the model [16], which is what we employed in this study to develop our animal model. Adolescent rats (21 days old) were selected in

our study, mainly because adolescent rats are more sensitive to ionizing radiation and suffer greater damage than adult rats, so it is easier to establish a model of radiation-induced brain injury. We selected time-points of 3 days and 2 months after exposure to detect the reduction of DG proliferating cells and mature neurons, respectively. Both time-points demonstrated radiation-induced brain injury.

Previous studies have suggested that apoptosis of hippocampus cells, especially DG proliferating cells and immature neurons, plays a key role in radiation-induced cognitive damage [19, 20]. DG

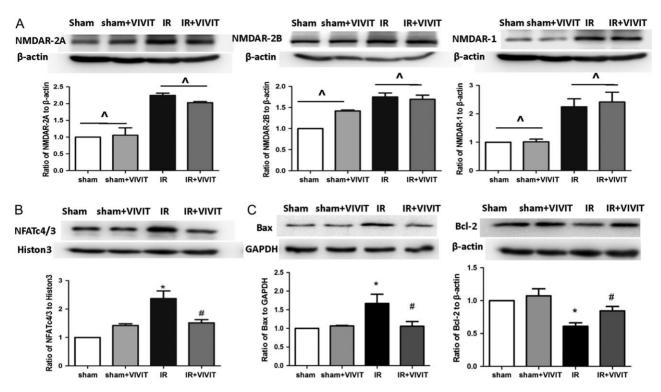


Fig. 3. 11R-VIVIT peptide relieved apoptosis induced by the activation of NFAT3/c4 after irradiation. Treatment with 11R-VIVIT peptide didn't have any impact on the expression of NMDAR subunits (NR1, NR2A or NR2B) (A). 11R-VIVIT peptide prevented nuclear translocation of NFAT3/c4 and appearance of Bax (B-C). Data are expressed as mean ± SD, *P < 0.05 vs sham group, $^{*}P < 0.05$ vs irradiation group, $^{^{*}}P > 0.05$.

proliferating cells were capable of generating new neurons, astrocytes and oligodendrocytes, which are associated with brain selfrecovery function [21-23]. A radiation-induced decrease in proliferating cells was accompanied by a reduction in new neurons [24, 25]. The significant decrease in mature neurons induced by radiation has been associated with hippocampal-dependent spatial learning and memory dysfunction [26, 27]. Single whole-brain irradiation (2-10 Gy) caused a sharp elevation in apoptosis in the hippocampal DG [28, 29]. Apoptosis was accompanied by a subsequent decrease in both proliferating cells and neurons [27, 30, 31]. In the present study, we found a significantly elevated expression of the apoptosis marker Bax accompanied by a steep decrease in hippocampus cells. In addition, the 11R-VIVIT peptide, an NFAT3/c4 inhibitor, inhibited the appearance of apoptosis and reversed the decline in the number of DG proliferating cells and mature neurons.

DNA damage was the most common effect of radiation injury, and DNA transcription and protein synthesis were simultaneously disrupted. Numerous potential mechanisms of radiation-induced apoptosis were present [32-34]. The radiation-induced perturbations of extracellular-signal-regulated kinase (ERK1/ERK2) and downstream signaling pathways severely affected neuronal survival [35]. The secretion of TRAIL by irradiated neural stem cells (NSCs) induced a death-signaling cascade in non-targeted NSCs [32]. Regarding these potential mechanisms, a recent NMDAR-mediated non-conventional apoptotic response to radiation has been reported, so we investigated the excitotoxic effects of overactivated NMDAR [36]. We observed that overexpression of NMDAR caused nuclear translocation and activation of the transcriptional factor NFAT3/c4, which regulates the expression of numerous proteins. Its transcriptional activities promoted the elevation of Bax. Together, our results suggested a hitherto unknown vital role of NFAT3/c4 in the study of radiationinduced brain injury.

Some limitations of this study need mentioning. First, radiationinduced apoptosis requires more extensive study to clarify the mechanisms underlying it and how it develops, including TUNEL assays and Bax immunofluorescence assays. Second, we would now like to use an NFAT3/c4 knock-out rat model to further demonstrate the important role of NFAT3/c4 in radiation-induced brain injury. Third, NFAT3/c4 acts as a powerful transcriptional factor, and the rest of its apoptotic targets need further investigation. Finally, we need to explore further the pathological changes in brain morphology and structure post irradiation with H&E staining. Exploration of the changes in synapse morphology and structure with Golgi-Cox staining is our next goal.

In summary, our present study showed for the first time that NFAT3/c4 mediated excitotoxicity in apoptosis of hippocampus cells in radiation-induced brain injury. Moreover, the results from the inhibitor 11R-VIVIT peptide supported a regulatory role for NFAT3/c4. In addition, we clarified the potential mechanisms of an

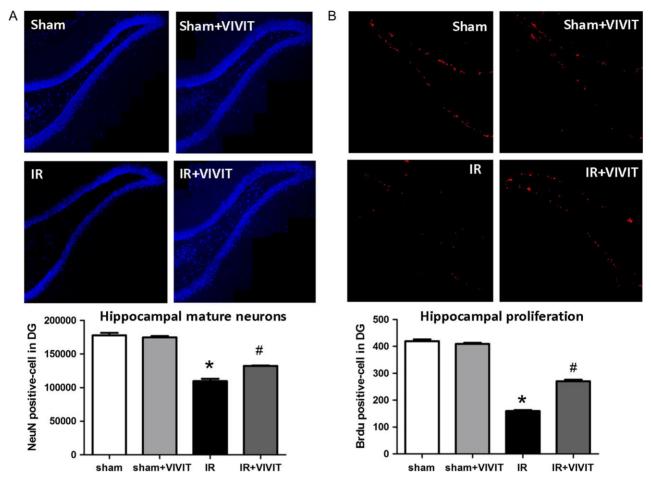


Fig. 4. 11R-VIVIT peptide alleviated the decline in the DG proliferating cells and mature neurons at 3 days and 2 months after WBI (20 Gy). Representative images indicated that 11R-VIVIT peptide significantly increased the total number of DG proliferating cells (BrdU+, red)at 3 days post irradiation (A). Similarly, 11R-VIVIT peptide increased the number of DG mature neurons (NeuN+, blue) at 2 months post irradiation (B). The original objective magnification was $\times 200$. Data are expressed as mean \pm SD, *P < 0.05 vs sham group, #P < 0.05 vs irradiation group.

NMDAR-mediated non-conventional apoptotic response to radiation. Finally, our results highlighted the importance of the NMDAR/NFAT3/c4/Bax pathway and provided evidence of its use in a potential therapeutic intervention.

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