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## Biexponential T<sub>2</sub> Relaxation Estimation of Human Knee Cartilage in-vivo at 3T

Azadeh Sharafi, PhD, Gregory Chang, MD, and Ravinder R Regatte, PhD

Bernard and Irene Schwartz Center for Biomedical Imaging, Department of Radiology, New York University School of Medicine, New York, NY, USA

### Abstract

**Purpose**—To evaluate biexponential T<sub>2</sub> relaxation mapping of human knee cartilage in-vivo in clinically feasible scan times.

**Materials and Methods**—T<sub>2</sub> weighted MR images were acquired from eight healthy volunteers using a standard 3T clinical scanner. A 3D Turbo-Flash sequence was modified to enable T<sub>2</sub> weighted imaging with different echo times. Series of T<sub>2</sub>-weighted images were fitted using mono- and biexponential models with two- and four- parametric non-linear approaches, respectively.

**Results**—Biexponential relaxation of T<sub>2</sub> was detected in the knee cartilage in five regions of interest on all eight healthy volunteers. Short/long relaxation components of T<sub>2</sub> were estimated to be  $8.27 \pm 0.68\text{ms}/45.35 \pm 3.79\text{ms}$  with corresponding fractions of  $41.3 \pm 1.1\%/58.6 \pm 4.6\%$  respectively. The monoexponential relaxation of T<sub>2</sub> was measured to be  $26.9 \pm 2.27\text{ms}$ . The experiments showed good repeatability with  $CV_{\text{rms}} < 18\%$  in all regions. The only difference in gender was observed in medial-tibial cartilage where the biexponential T<sub>2</sub> in female volunteers was significantly higher compared to male volunteers ( $P = 0.014$ ). Significant differences were observed in T<sub>2</sub> relaxation between different regions on interest.

**Conclusion**—Biexponential relaxation of T<sub>2</sub> was observed in the human knee cartilage in-vivo. The short and long components are thought to be related to the tightly bound and loosely bound macromolecular water compartments. These preliminary results of biexponential T<sub>2</sub> analysis could potentially be used to increase the specificity for detection of early osteoarthritis by measuring different water compartments and their fractions.

### Keywords

T<sub>2</sub> relaxation; biexponential fitting; articular cartilage

## INTRODUCTION

Osteoarthritis (OA) is a degenerative joint disease which causes changes in biochemical, morphological, functional and structural properties of cartilage and is mainly defined by the

progressive loss of hyaline articular cartilage (1). Several imaging modalities such as radiography, radionuclide imaging, computed tomography (CT), ultrasound, and magnetic resonance imaging (MRI) have been used to diagnose OA (2,3). However, none of these clinical standard techniques are sensitive enough to detect early stage OA (4).

At the early stage of OA, the structure of collagen fibers starts to change. The loss of proteoglycans (PG) leads to an increase in water content in cartilage (5). Different water compartments in articular cartilage lead to different  $T_2$  relaxation components. Additionally,  $T_2$  is sensitive to collagen fibril orientation and anisotropy (6,7). As a result of the change in the biochemistry of cartilage and increase in water content due to OA, elevated  $T_2$  relaxation times is expected (4,5,8–12). The existing  $T_2$  mapping techniques include fast spin-echo (FSE) (9), multiple two-dimensional (2D) spin-echo (SE) acquisitions at different echo times (TEs) (13), and double echo steady state (DESS) acquisition methods (14,15).

Since different components in cartilage such as collagen, PG macromolecules, fragmented PG molecules, water molecules trapped within collagen fibrils, and free water molecules have different  $T_2$  relaxation times a biexponential model may provide more information on different water compartments than a monoexponential model. In most of the previous in-vivo studies (4,5,16),  $T_2$  was described by a monoexponential decay model, which shows the mean of relaxation time from all of the water compartments.

Liu *et al.* (17–19) measured two components of  $T_2$  relaxation in the human knee joint using the multicomponent driven equilibrium single shot observation of  $T_1$  and  $T_2$  (mcDESPOT) technique in which several balanced steady state free-precession (bSSFP) and spoiled gradient-echo (SPGR) scans were taken with different flip angles and radiofrequency phases. Then a two-pool model of longitudinal  $T_1$  and  $T_2$  relaxation were fit to the data (17,20–22). However, the short  $T_2$  component and its fraction suffer from partial volume effects (PVE)(19). Also, the accuracy of the estimation could be affected by the magnetization transfer (MT) effect (19,23).

The goal of this paper is to propose a method for estimating biexponential relaxation times of articular cartilage in the human knee joint using 3T MRI in clinically feasible scan times.

## MATERIALS AND METHODS

### Monte Carlo Simulations

The monoexponential  $T_2$  relaxation can be calculated by fitting the signal intensities of each pixel to:

$$S(TE) = A_m \exp\left(-\frac{TE}{T_{2m}}\right) + s_0 \quad [1]$$

In the same manner, the biexponential relaxation components can be estimated from:

$$S(\text{TE}) = A_s \exp\left(-\frac{\text{TE}}{T_{2s}}\right) + A_l \exp\left(-\frac{\text{TE}}{T_{2l}}\right) + s_0 \quad [2]$$

Where  $T_{2s}$  and  $T_{2l}$  correspond to the short and long relaxation time components respectively. The weightings (fractions) of the short and long components are usually reported in percentage as  $a_s\% = 100 \times A_s / (A_s + A_l)$  and  $a_l\% = 100 \times A_l / (A_s + A_l)$  respectively.  $s_0$  is the average noise level.

Monte Carlo simulations were performed for different  $T_2$  relaxation in the range of possible  $T_2$  values in the cartilage to determine a sufficient number of TEs for estimating four parameters in a biexponential model [2] considering the range. Under given signal to noise ratio (SNR), the smaller set of TEs is desired to minimize the total acquisition time.

To perform the Monte Carlo simulation, a set of signals were generated with a different set of TEs with known  $T_2$  relaxations time and their fractions (assuming  $A_s + A_l = 1$ ). Assuming the SNR of  $1/\sigma$  (24) a random noise with normal distribution  $N(0, \sigma)$  was added to data. The relaxation components were estimated with a biexponential model using the noisy signals. The process was repeated for 500 independent noise trail and the mean absolute percentage error (MAPE) were calculated as:

$$\text{MAPE} = \frac{100}{n} \sum_{m=1}^n \frac{|Y_m - F_m|}{Y_m} \quad [3]$$

Where  $Y_m$  and  $F_m$  are the actual and estimated values respectively.  $n$  is the total number of noise trail ( $n = 500$ ) in Monte Carlo simulation.

### **T<sub>2</sub>-weighted MRI Acquisition**

$T_2$ -weighted MR scans were acquired on a 3T whole-body clinical MRI scanner (Prisma, Siemens Healthcare, Erlangen, Germany) with a 15-channel Tx/Rx knee coil (QED, Cleveland OH). A 3D Cartesian turbo-Flash sequence was modified and  $T_2$  preparation module was added to the sequence for  $T_2$  imaging with variable echo times. The  $T_2$  imaging pulse sequence diagram is shown in FIG. 1.

For  $T_2$  relaxation mapping, a set of 3D scan was acquired with different TEs. The sequence acquisition parameters for all the scans were: TR/TE 1500ms/4ms, flip angle  $8^\circ$ , matrix size  $256 \times 128 \times 64$ , slice thickness = 2mm, field of view (FOV) =  $120\text{mm} \times 120\text{mm}$ , and receiver bandwidth = 515 Hz/pixel. Binomial water excitation pulse and GRAPPA (25) parallel imaging method were used in readout section for fat suppression and decreasing the total acquisition time, respectively.

### **Ex-vivo Bovine Cartilage Study**

Fresh bovine patellae cartilage specimens ( $n=3$ , age= $\sim 6$  months old) were obtained from a slaughterhouse (Max Insel Cohen, Inc., Livingston, NJ) within 24 hours postmortem. The

bovine cartilage specimens were equilibrated in phosphate-buffered saline (PBS) for one hour before the MRI study. After that, the specimen was covered with parafilm to avoid drying up during the scan.  $T_2$  weighted images were taken from three specimens at 15 different TEs including 0.5/2/4/6/7/8/10/12/15/20/25/35/45/55/65ms. The experiment was repeated with different acceleration factor of  $AF = 1-4$  to examine the effect of reduced SNR due to parallel imaging on the estimated relaxation time. The total acquisition time for 3D data set with 15 TEs were 46, 27, 22, and 18 minutes for  $AF = 1$  (fully sampled) 2, 3, and 4 respectively.

### In-vivo knee Study

The study was approved by the institutional review board (IRB) and all eight volunteers provided written informed consent before the scans. Eight healthy volunteers ( $n=4$  females, and  $n=4$  males) were recruited for this study with a mean age of  $30 \pm 4$  years, mean weight of  $63 \pm 15$ kg, and mean height of  $169 \pm 12$ cm. Volunteer exclusion criteria were any knee pain or clinical symptoms, history of osteoarthritis or inflammatory arthritis, previous knee injury, and surgery on either knee

$T_2$  weighted images were taken at 10 different TEs including 2/4/6/8/10/15/25/35/45/55ms. Using GRAPPA with  $AF = 3$  the total acquisition time was decreased from 30 to 15 minutes.

The acquired  $T_2$ -weighted scans were analyzed using a custom-written script in MATLAB (R2016a, The MathWorks Inc., USA). Mono- and biexponential  $T_2$  relaxation times were calculated pixel by pixel over five consecutive slices in five regions of interest (ROI): medial-tibia (MTC), medial-femoral (MFC), lateral-tibia (LTC), lateral-femoral (LFC), and Patellar (PC) cartilages.

In the final biexponential fitting map, the pixels that were not satisfied the following condition were excluded from the map (26).

$$4 \times T_s < T_l \quad [4]$$

The mean values of  $T_2$  were calculated across all volunteers in each ROI. Then, the mixed model with  $P = 0.05$  as the threshold was used to assess the gender difference as well as the significance of the difference in  $T_2$  relaxation time components between different ROIs.

For repeatability investigation, the same scans were acquired from three volunteers, two weeks after their first scan. To evaluate the intra-subject repeatability (i.e. repeating the experiment on the same subject) the coefficient of variation (CV) was calculated for volunteer  $i$  as:

$$CV = \frac{SD_i}{M_i} \quad [5]$$

Where the  $M_i$  and  $SD_i$  are the mean and standard deviation of the estimated relaxation times in two experiments.

The inter-subject repeatability (the repeatability across all volunteers) was reported as root mean sum square of the CVs of individual subject:

$$CV_{\text{rss}} = \sqrt{\frac{\sum_{i=1}^N CV_i^2}{N}} \quad [6]$$

## RESULTS

### Monte Carlo Simulations

The results of Monte Carlo simulation are shown in FIG.2. The estimation error for short and long relaxation components and their fractions decreases by increasing the number of TEs (FIG. 2a) at the cost of longer total acquisition time. As shown in FIG.2a, the estimation error for 10 TE points is less than 10% for all parameters and the improvement with 15 TE points is not significant (less than 2%) considering the fact that the total acquisition time will increase by 50%. Hence, 10 TE points were selected for the rest of the simulations as well as in-vivo studies.

As shown in FIG.2b, higher SNR leads to smaller error. The SNR of an in-vivo knee cartilage scan is expected to be around 60, resulting in an expected error of about 15% for the short component and 10% for the long component in the in-vivo study based on the simulation result. As the ranges of  $T_2$  are different in different tissues and subjects, the accuracy of estimation for different short and long  $T_2$  values are shown in FIG. 2c and FIG. 2d respectively. While the estimation is more accurate for lower values of the short  $T_2$  component, it is more accurate for the higher values of the long  $T_2$  components. The fractions also affect the estimation in a way that the component with the greater fraction has less estimation error than the components with the smaller fraction (FIG. 2e-f)

### Ex-vivo bovine cartilage Study

Mono, short and long  $T_2$  relaxation maps of a bovine patella from a representative slice are shown in FIG. 3a-c respectively. Considering the estimated  $T_2$  values from fully sampled data as a reference, FIG. 3d shows the estimation error for different GRAPPA acceleration factors ( $AF = 2-4$ ). Using acceleration factor of 3 will decrease the total acquisition time from 32 to 15min while the estimated relaxations have less than 5% difference from the estimated values from fully sampled data. Hence;  $AF = 3$  was selected for performing in-vivo experiments.

The differences between the estimation using 15 TEs (as reference) with the estimation using 10 and 6 TEs are shown in FIG. 3e. The result shows less than 3% difference between the relaxation values estimated from 15 TE points and values estimated using 10 TE points, which is in agreement with the simulation study. Hence; 10 TE points was used in in-vivo knee studies as a good tradeoff between time and accuracy.

### In-vivo knee articular cartilage study

FIG. 4 shows the  $T_2$  signal decay with increasing TE in a representative medial slice. An example of  $T_2$  maps in medial, lateral and patellar cartilages are shown in FIG. 5. The binary maps in FIG. 5a1–3 show the distribution of pixels that meet the condition [4].

Approximately 46% of pixels in the ROIs had biexponential relaxation. The  $T_2$  descriptive statistics are summarized in Table 1. The short component has a lower fraction (41.3%) than the long component ( $T_2$ : 58.7%).

FIG. 6 shows the mono- and biexponential fit and their fit residuals in representative voxels. The deviation of data points from the straight line in logarithmic scale shows the existence of more than one exponential term. Moreover, the smaller fit residual in the biexponential model confirms that it better represents the relaxation behavior than the monoexponential model.

The biexponential  $T_2$  components variation was observed in different cartilage zones. As shown in FIG.7 mono- and biexponential  $T_2$  decrease from superficial zone to the subchondral bone.

### Statistical Analysis

FIG. 8 shows the mean mono- and biexponential components of  $T_2$  among all volunteers as well as male and female volunteers. The mixed-model analysis for each component revealed:

- A significantly higher ( $P = 0.014$ ) biexponential  $T_2$  of MTC in female volunteers than in male volunteers.
- Statistically significant difference in  $T_{2m}$  between MTC and PC ( $P = 0.019$ )
- Statistically significant difference in  $T_{2m}$  between MTC and LTC ( $P = 0.025$ ).
- Statistically significant difference in  $T_{2l}$  ( $P = 0.01$ ) and  $T_{2s}$  ( $P = 0.04$ ) between MTC and PC.

### Repeatability

Table 1 shows the intra-subject repeatability data. The CV is less than 10% for mono and biexponential relaxations. Higher CV was detected in the fractions.

As shown in FIG. 9 long  $T_2$  relaxation has less variability (mean: 7% range: [3% – 11%]) than the short  $T_2$  relaxation (mean: 10% range: [4%-17%]), probably because the short component is more sensitive to change and also has larger potential differences between volunteers (27).

The experiments showed good repeatability in the entire region ( $CV < 18\%$ ). No statistically significant difference was observed between CV in different ROIs

## DISCUSSION

A 3T MRI technique for in-vivo, bicomponent  $T_2$  analysis of articular cartilage is presented in this paper.  $T_2$  relaxation time was measured in five ROIs in the articular cartilage of knee joints. The results suggest that a biexponential fit may better represent and differentiate the different relaxation times of different water compartments in the cartilage. It is expected the short component to be related to tightly-bound water in PG and collagen macromolecules while the long component corresponds to the water loosely bound to macromolecules.

Although the  $T_2$  relaxation range was comparable to the other studies (28–32), there were some differences that could be due to the partial volume effects (PVE), the number of echoes (TEs) acquired or the pulse sequence used in our study. The smaller slice thickness (2mm) in our study comparing to 3mm or 4mm of other studies (28–31), decreases the PVE.

Moreover, 10 TE points were acquired in our study to model the  $T_2$  decay behavior which leads to more accurate estimation in comparison with other studies in which only used 4 or 5 data points or only two echoes in case of using double echo steady state (DESS) sequence (14,15,32). Furthermore, using different MR pulse sequences for  $T_2$  relaxation time measurements in articular cartilage can lead to different results. For example, in the Matzat study (33) a  $35\pm 22\%$  difference was observed between the estimated  $T_2$  using 2D-FSE sequence and the estimated  $T_2$  from multi-echo SE (MESE) sequence.

The total acquisition time of our in-vivo study (14:45min) is also close the scan time of other studies (33) using 3D-MAPSS (16:15min) and 3D vfl-FSE (13:56min). However, only 6 echo times were acquired with 3D-MAPSS and 3D vfl-FSE sequences (33) while we acquired 10 TEs resulting in a better  $T_2$  estimation. The acquisition time of 8:54 min using quantitative DESS (qDESS) scans in Matzat study (33) is roughly half of our scan time. However, with only two echoes, this sequence cannot be used for biexponential relaxation estimation. We plan to reduce the total scan time by applying compressed sensing (34) in addition to parallel imaging in the future.

Reiter *et al.* (35) presented a method to measure multiexponential  $T_2$  relaxation in the bovine nasal cartilage of a mature cow. Three components were detected:  $T_{2,1} = 2.3$ ms,  $T_{2,2} = 25.2$  ms, and  $T_{2,3} = 96.3$  ms, with fractions  $a_1 = 6.2\%$ ,  $a_2 = 14.5\%$ , and  $a_3 = 79.3\%$  and the authors confirmed that the shortest component  $T_{2,1}$  is related to the immobile collagen-bound water while  $T_{2,2}$  and  $T_{2,3}$  correspond to the water compartments that are bound and loosely related to PG respectively (35). Comparing to our experiment, using biexponential fitting instead of multiexponential, our short component  $T_{2s} = 7.6$ ms,  $a_s = 41\%$  is related to the water tightly bound to PG and collagen macromolecules while the long component  $T_{2l} = 46.0$ ms,  $a_l = 59\%$  corresponds to water associated with loosely bound to PG and collagen macromolecules. Our estimated fractions follow the same pattern as the one observed in Reiter *et al.* (35), where the largest component has higher fraction than the short component. Using our current protocol, we cannot measure the water related to immobile collagen itself - since the echo time is longer than the ultrashort  $T_2$  of collagen (0.2–1ms). We plan to modify a UTE-based sequence (36) in the future to measure the ultrashort  $T_2$  components.



Liu *et al.* (17–19) measured two components of  $T_2$  relaxation of the human knee joint using multicomponent driven equilibrium single shot observation of  $T_1$  and  $T_2$  (mcDESPOT) technique (17,20,21). First, they showed the feasibility of using the method for in-vivo  $T_2$  measurement in different regions of interest (17) and then compared the relaxation values between asymptomatic volunteers and patients with OA (18). The estimated values of the single, short and long  $T_2$  relaxation time in these studies (17,18) for asymptomatic volunteers (~ 35.5ms, 16ms, 62 ms ) were higher than our measurements (~27ms, 7.5ms, 46ms) while the fraction of the short component is lower (31% vs 41%). In their next study, Liu *et al.*(19) proposed a multicomponent  $T_2$  analysis method with synovial fluid partial volume correction. Using the three-pool model instead of two-pool model they claimed that they corrected the  $T_2$  bias and as a result estimated a shorter long  $T_2$  component (~53ms) which is closer to our estimation (~46ms). However, the short  $T_2$  component and its fraction still suffer from PVE (19). We employed a 2mm slice thickness rather than 3 mm used in these studies so the  $T_2$  estimation would be more accurate. Another drawback of the mcDESPOT method is that the mcDESPOT parameters could be affected by the magnetization transfer (MT) effect (19,23) and as a result less accurate  $T_2$  estimation. Our proposed method does not suffer from MT due to the long delay in the sequence for  $T_1$  recovery. In addition, our method has a shorter acquisition time (15min vs 17min) while acquiring thinner slices. Recently a method called mcRISE has been proposed by Liu *et al.*(37) to correct the MT effect at the cost of increasing the acquisition time from 17min (mcDESPOT) to 25min.

Several studies have reported in-vivo bicomponent  $T_2^*$  measurement in different anatomic locations including articular cartilage (27) and the meniscus (26). The estimated  $T_2^*$  relaxation in (27) is in agreement with our  $T_2$  estimation since all the mono, short and long  $T_2^*$  components are shorter than the estimated  $T_2$  in our studies.

Li X *et al.* (31) investigated the repeatability of  $T_2$  relaxation time measurement and the  $CV_{rms}$  of 4.2%–8.5% has been reported across all the ROIs. Their numbers are in good agreement with our experiments where the  $CV_{rms}$  of 4.4%-11.5% was measured for monoexponential  $T_2$ . Mosher *et al.* (38) investigated the reproducibility of measuring  $T_2$  in the healthy control group and osteoarthritis (OA) volunteers ( $CV_{rms}$ : 8.12%-10.95% in the healthy volunteers). Similar results were also observed in our study. The good agreement was also observed if we compare the  $CV_{rms}$  in different ROIs. Note that the reproducibility was evaluated in Mosher study by acquiring the data from different sites while we investigated the repeatability by acquiring all the data on the same scanner.

Qian *et al.*(27) evaluated the repeatability of two-component  $T_2^*$  estimations in knee articular cartilage. Similar to our study, they observed better repeatability in the long- $T_2$  component ( $CV_{rms}$  ~8%) than in the short- $T_2$  component (~12%).

Our study has some limitations. First, there were a small number of volunteers. Second, the biexponential condition of  $4 \times T_s < T_1$  may produce some bias. We chose this condition based on the suggestion in (26) and confirmed it with our own experiments. Moreover, the magic angle effect can influence  $T_2$  values due to the dipolar interactions of fiber orientation with respect to  $B_0$ . As shown by Henkelman *et al.* (39) in bovine articular cartilage, the



biexponential behavior disappeared when the tissue's orientation to  $B_0$  is about 55(40). Finally, only asymptomatic volunteers were included in this study and we didn't perform validation with cartilage histology/biochemical assay or in knee OA patients with symptoms.

In conclusion, in this study, biexponential  $T_2$  relaxation of human knee articular cartilage is estimated in-vivo on a 3T MRI scanner in clinically feasible scan times. The short component corresponds to the water tightly bound to PG and collagen macromolecules while the long component is expected to be affected by loosely bound water. Our preliminary results show that biexponential  $T_2$  mapping could potentially be used to increase the specificity of early osteoarthritis diagnosis by estimating the relaxation time of different water compartments and their fractions.

## Acknowledgments

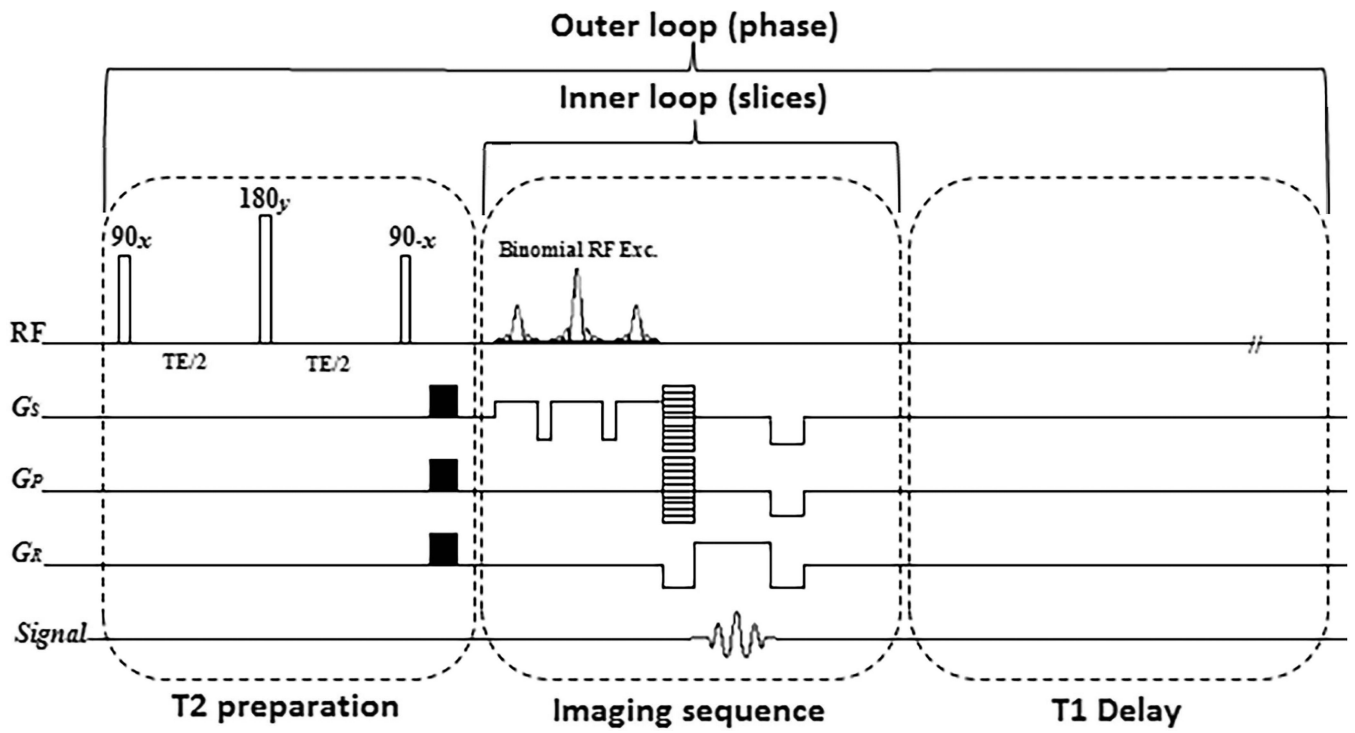
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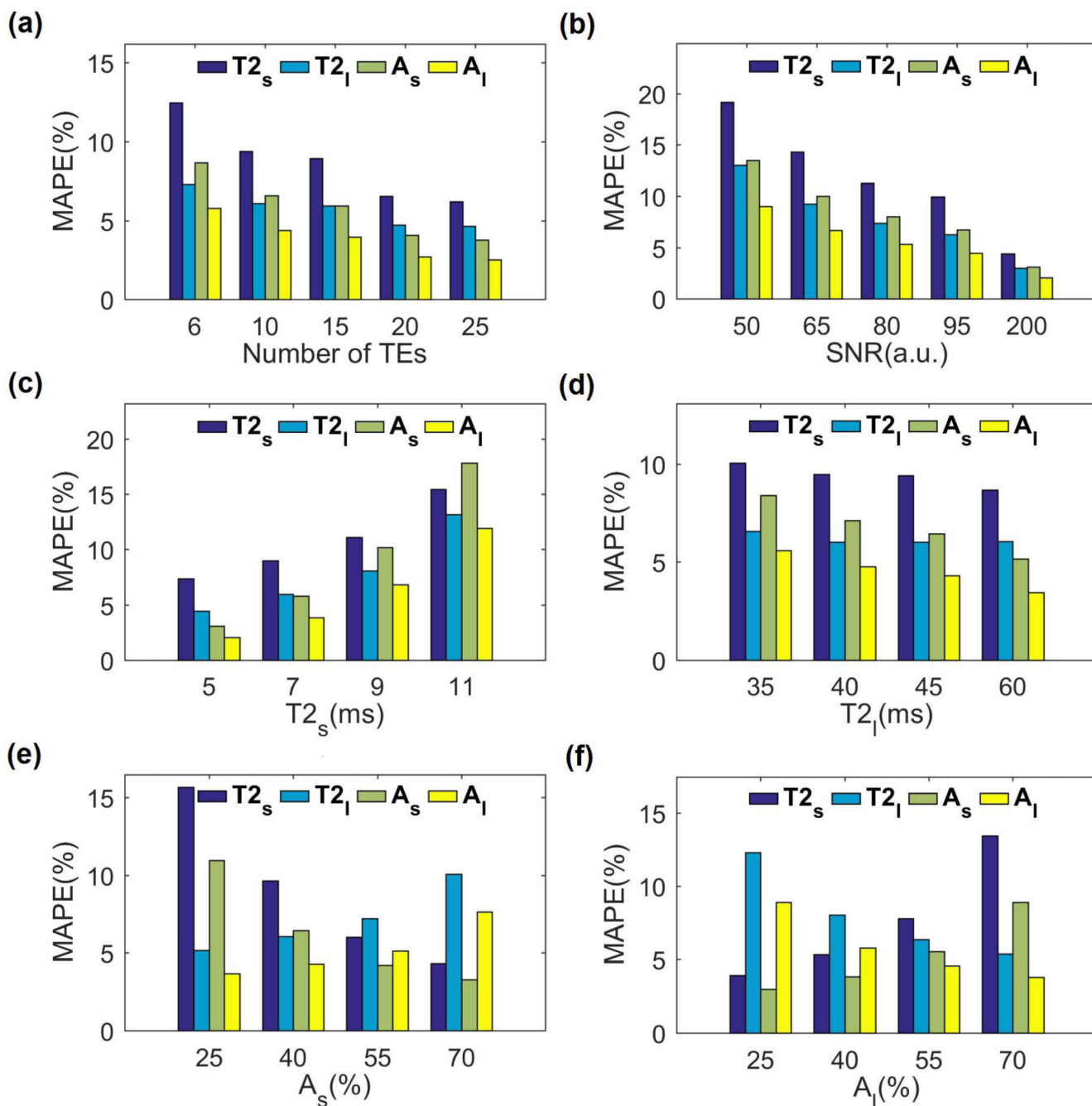
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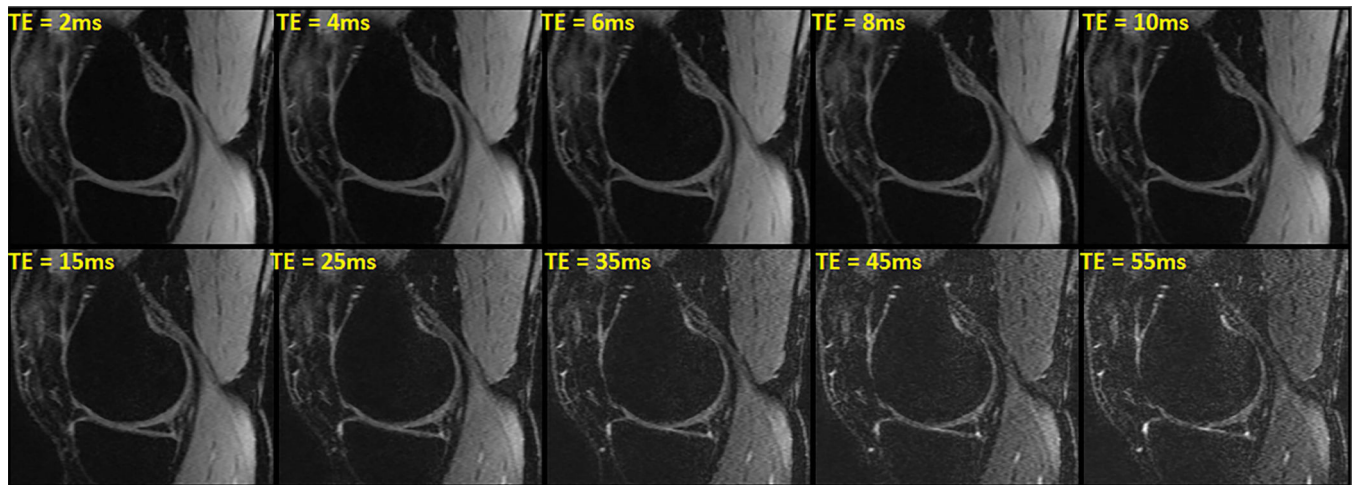
**FIG. 1.**

The T<sub>2</sub> mapping sequence timing diagram with preparation module, 3D turbo-Flash readout, and T<sub>1</sub> recovery delay. One phase line from all slices was acquired after applying the T<sub>2</sub> preparation module (inner loop) followed by a delay for T<sub>1</sub> restoration. Then, the preparation module was applied again to acquire another phase line (outer loop).



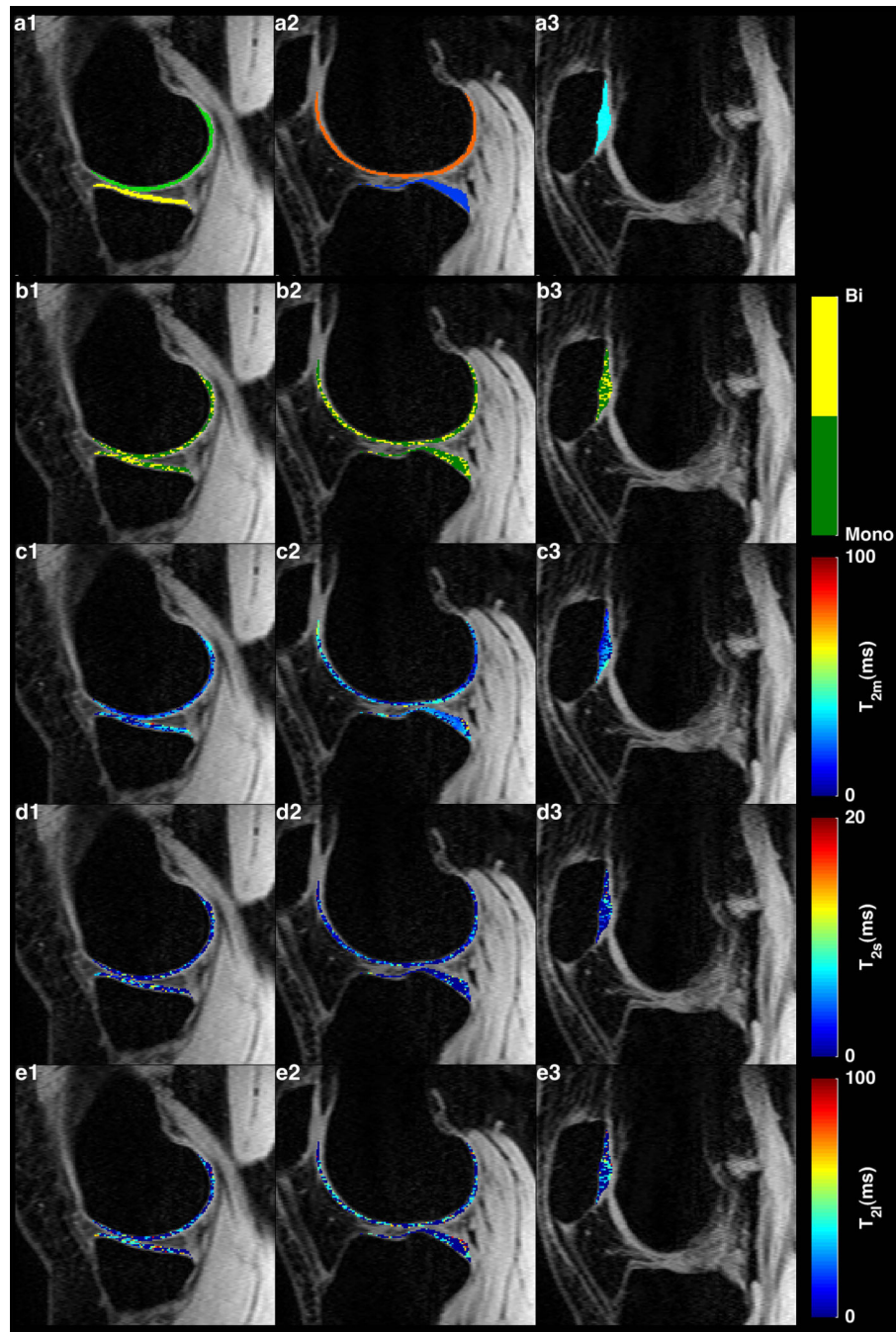
**FIG. 2.** Monte Carlo simulations. (a) The biexponential relaxation estimation error decreases by increasing the number of TE points. (b) Higher SNR results in lower estimation error. (c) The estimation error of short relaxation component is higher for longer short component. (d) The estimation error of long component is higher for shorter long component. (e, f) The component with larger fraction (short in (e) and long in (f)) has been estimated more accurately than the component with smaller fraction.





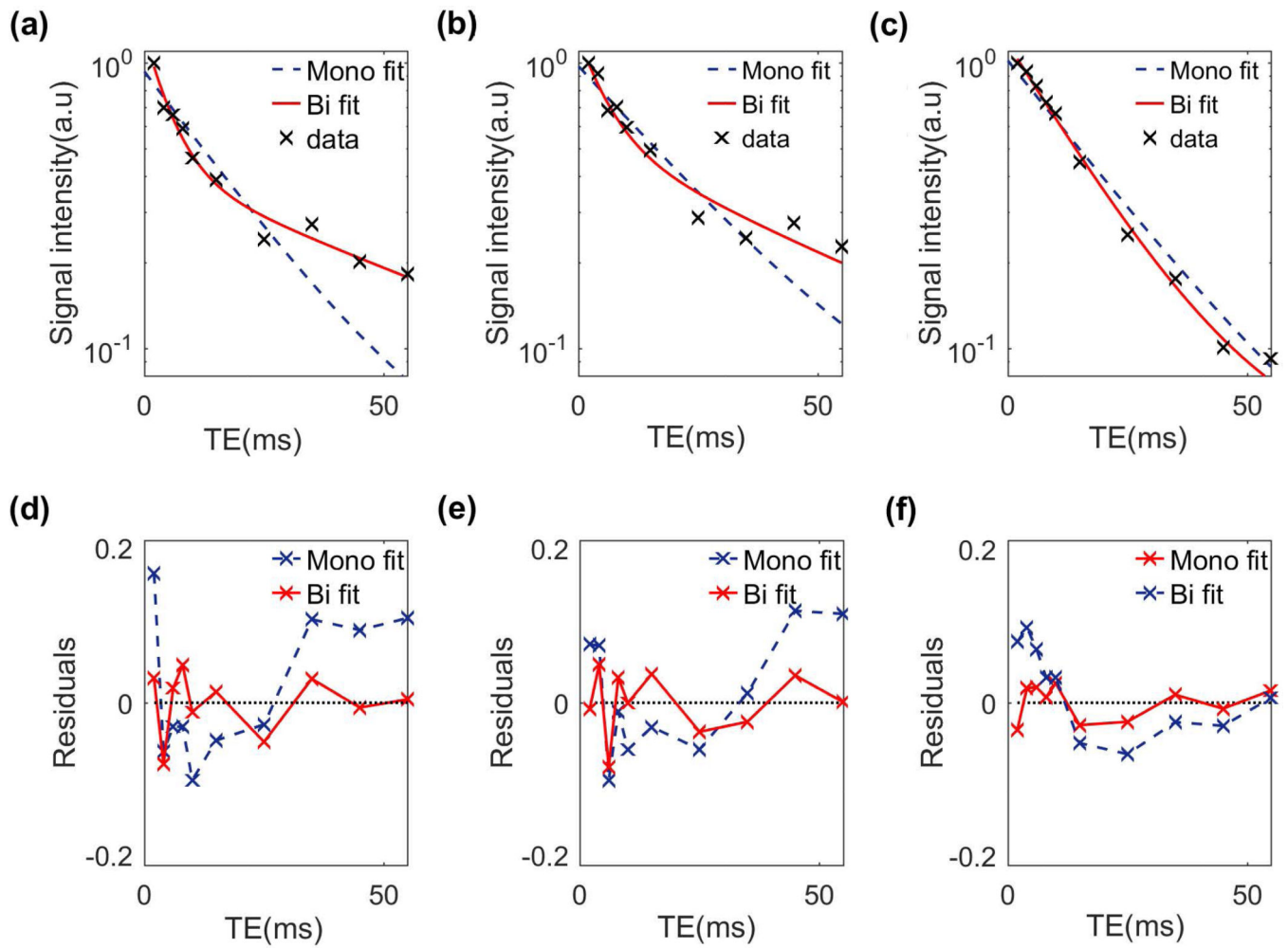
**FIG. 4.** Representative examples of  $T_2$ -weighted images with different TE in a medial slice from one volunteer. The signal intensity decays by increasing TE.



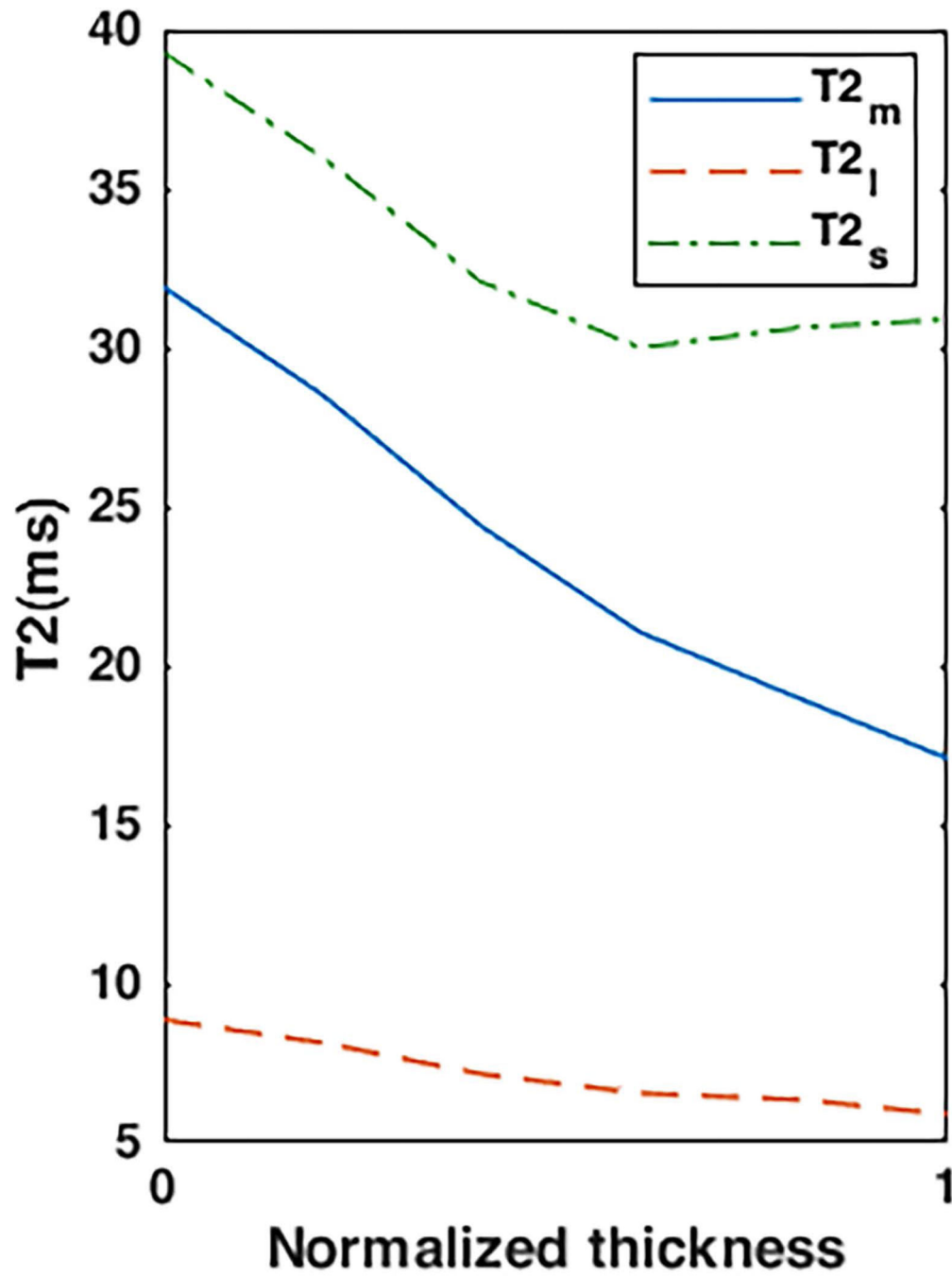


**FIG. 5.**

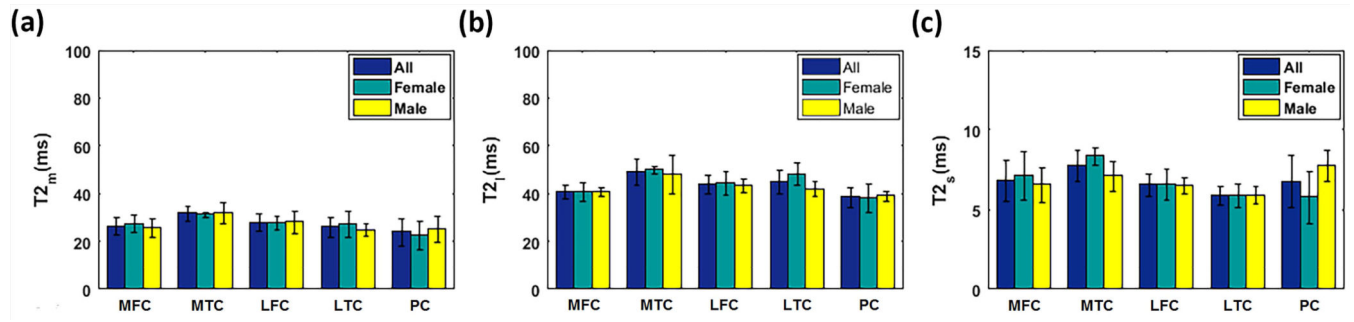
(a1-d1) Five regions of interest for relaxation mapping: medial femoral (a1-green), medial tibial (a1-yellow), lateral femoral (a2-orange), lateral tibial (a2-yellow), and patellar (a3,cyan) cartilages. (b1-b3) binary maps show the distribution of the pixels with mono- and biexponential fitting in each ROI. (c1-c3) Representative examples of monoexponential, (d1-d3) biexponential short and (e1-e3) long  $T_2$  relaxation maps.

**FIG. 6.**

$T_2$  Biexponential versus monoexponential fitting model and residuals in three representative slices from (a, d) lateral, (b, e) medial and (c, f) patellar cartilage.



**FIG. 7.** Representative  $T_2$  relaxation profile in patellar cartilage from the superficial zone (0) to the subchondral bone (1)



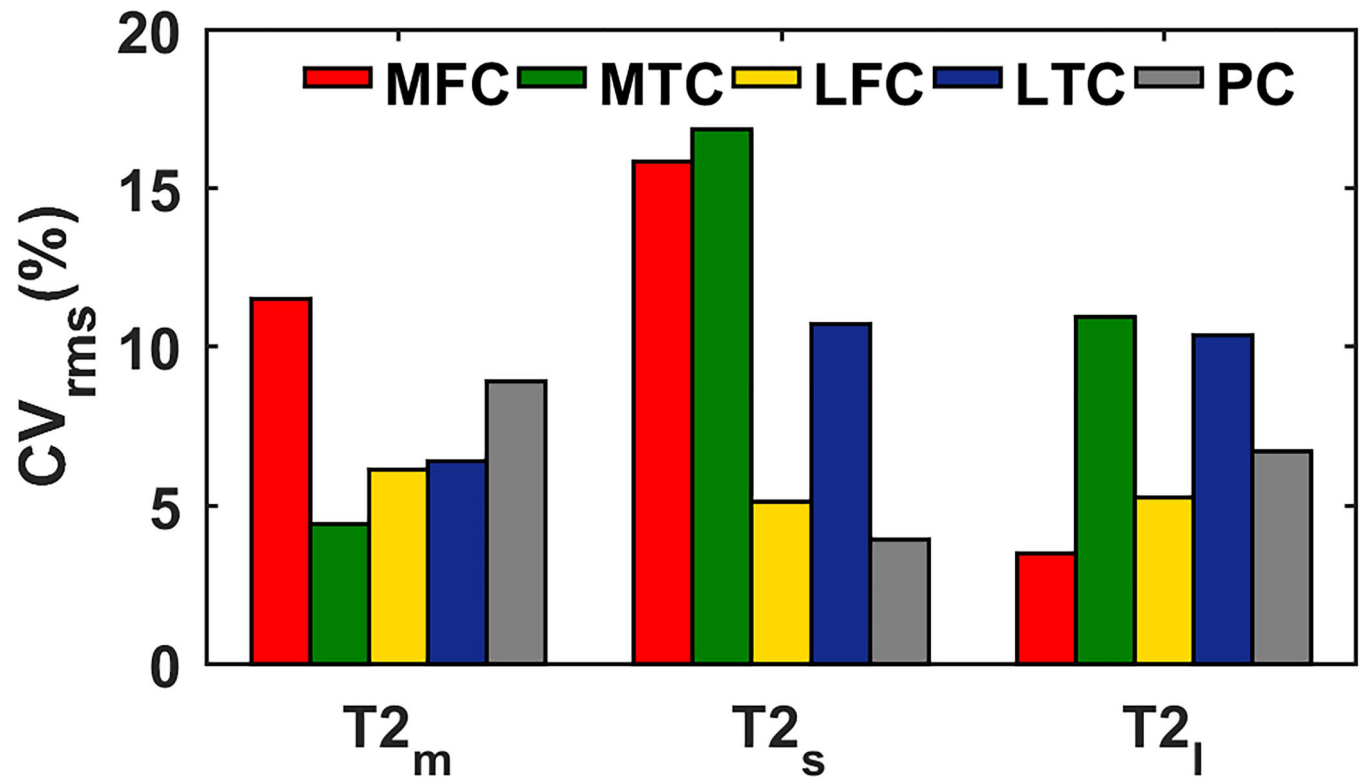
**FIG. 8.** Mean mono and biexponential  $T_2$  estimation of female (N=4), male (N=4), and all volunteers (N=8) in different ROIs.

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**FIG. 9.**  
Inter-subject repeatability

Descriptive statistics of mono- and biexponential relaxation and the ratio of biexponential pixels to total number of pixels calculated in five ROIs.

**Table 1**

ROI		T <sub>mono</sub> (ms)	T <sub>short</sub> (ms)	T <sub>long</sub> (ms)	a <sub>s</sub> (%)	a <sub>l</sub> (%)	Ratio (%)
MTC	Mean	26.98	7.81	44.55	41.64	58.36	46
	SD	2.84	1.23	3.07	1.23	3.07	16
	R <sup>2</sup>	0.99	0.99	0.99	--	--	--
MFC	Mean	30.35	8.69	51.07	41.42	58.58	51
	SD	3.08	0.99	4.09	0.99	4.09	8
	R <sup>2</sup>	0.99	0.99	0.99	--	--	--
LTC	Mean	28.54	7.33	45.82	39.63	60.37	48
	SD	3.09	1.08	4.05	1.08	4.05	11
	R <sup>2</sup>	0.99	0.99	0.99	--	--	--
LFC	Mean	26.23	7.40	47.66	45.17	54.83	40
	SD	4.00	0.74	5.31	0.74	5.31	8
	R <sup>2</sup>	0.98	0.98	0.98	--	--	--
PC	Mean	24.58	6.59	40.71	38.78	61.22	45
	SD	4.68	1.68	6.52	1.68	6.52	19
	R <sup>2</sup>	0.99	0.99	0.99	--	--	--
Global	Mean	27.336	7.564	45.962	41.328	58.672	46
	SD	3.538	1.144	4.608	1.144	4.608	12.4
	R <sup>2</sup>	0.988	0.988	0.988	--	--	--

SD: Standard Deviation

R<sup>2</sup> Adjusted R square

**Table 2**

The intra-subject repeatability.

ROI	T <sub>2m</sub>			T <sub>2s</sub>			T <sub>2l</sub>			a <sub>s</sub>			a <sub>l</sub>		
	Mean (ms)	SD (ms)	CV (%)	Mean (ms)	SD (ms)	CV (%)	Mean (ms)	SD (ms)	CV (%)	Mean (ms)	SD (ms)	CV (%)	Mean (ms)	SD (ms)	CV (%)
MTC	24.5	2.1	8.7	7.8	0.1	0.9	45.5	0.7	1.6	43.5	6.4	14.6	56.5	6.4	11.3
MFC	29.5	0.7	2.4	7.6	0.1	1.9	50.0	1.4	2.8	41.0	1.4	3.4	59.0	1.4	2.4
LTC	25.5	0.7	2.8	6.8	0.6	8.3	41.5	2.1	5.1	38.0	5.7	14.9	62.0	5.7	9.1
LFC	21.5	0.7	3.3	6.5	0.4	5.5	42.0	4.2	10.1	45.0	7.1	15.7	55.0	7.1	12.9
PC	27.0	2.8	10.5	6.4	0.1	1.1	37.5	2.1	5.7	30.0	5.7	18.9	70.0	5.7	8.1