FOCUSSED RESEARCH REVIEW



# **High immunosuppressive burden in cancer patients: a major hurdle for cancer immunotherapy**

**Suresh Gopi Kalathil1 · Yasmin Thanavala1**

Received: 16 September 2015 / Accepted: 8 February 2016 / Published online: 24 February 2016 © Springer-Verlag Berlin Heidelberg 2016

**Abstract** A bottleneck for immunotherapy of cancer is the immunosuppressive microenvironment in which the tumor cells are located. Regardless of the fact that large numbers of tumor-specific T cells can be generated in patients by active immunization or adoptive transfer, these T cells do not readily translate to tumor cell killing in vivo. The immune regulatory mechanism that prevents autoimmunity may be harnessed by tumor cells for the evasion of immune destruction. Regulatory T cells, myeloid-derived suppressor cells, inhibitory cytokines and immune checkpoint receptors are the major components of the immune system acting in concert with causing the subversion of anti-tumor immunity in the tumor microenvironment. This redundant immunosuppressive network may pose an impediment to efficacious immunotherapy, thus facilitating tumor progression. Cancer progression clearly documents the failure of immune control over relentless growth of tumor cells. Detailed knowledge of each of these factors responsible for creating an immunosuppressive shield to protect tumor cells from immune destruction is essential for the development of novel immune-based therapeutic interventions of cancer. Multipronged targeted depletion of these suppressor cells may restore production of granzyme

This paper is a Focussed Research Review based on a presentation given at the *Fourth International Conference on Cancer Immunotherapy and Immunomonitoring* (CITIM 2015), held in Ljubljana, Slovenia, 27th–30th April 2015. It is part of a series of Focussed Research Reviews and meeting report in *Cancer Immunology, Immunotherapy*.

 $\boxtimes$  Yasmin Thanavala yasmin.thanavala@roswellpark.org

B by  $CD8^+$  T cells and increase the number of IFN- $\gamma$ producing CD4<sup>+</sup> T cells.

**Keywords** Regulatory T cells · Myeloid-derived suppressor cells  $\cdot$  PD-1<sup>+</sup> T cells  $\cdot$  Immunosuppression  $\cdot$ Hepatocellular cancer · CITIM 2015

#### **Abbreviations**



# **Introduction**

Emerging clinical data suggest that cancer immunotherapy is likely to become a key part of the clinical management of cancer. The major advantage of immunotherapy is that it specifically targets the tumor cells, while sparing normal healthy cells, thus preventing the debilitating side effects

 $1$  Department of Immunology, Roswell Park Cancer Institute, Elm and Carlton Streets, Buffalo, NY 14263, USA

that are inevitable with chemo- or radiotherapy. Timely administration of immunotherapeutic strategies may eliminate minimal residual disease, thus preventing recurrence and metastasis.

A major hurdle for immunotherapy of cancer is that in most of the cases the tumor cells are located in an immunosuppressive microenvironment. Regardless of the fact that large numbers of tumor-specific T cells can be generated in patients by active immunization or adoptive transfer, these T cells cannot fully achieve their tumoricidal potential in vivo. Thus, large numbers of tumor antigen-specific T cells present in the periphery do not readily translate to tumor cell killing. Tumor cells employ various suppressive mechanisms to accomplish immune suppression, which include induction of regulatory T cells (Tregs), myeloidderived suppressor cells [\[1](#page-4-0)] and expression of programmed death ligand (PD-L1).

#### **Immunosuppression by Tregs**

Regulatory T cells (Tregs) are potent inhibitors of the immune system, crucially important for maintaining peripheral tolerance as well as regulating immune responses, preventing excessive immune activation and autoimmunity. Tregs suppress effector T cells, NK cells and dendritic cells either through cell–cell contact or through secretion of immunosuppressive cytokines and indoleamine 2,3 dioxygenase (IDO). Studies have demonstrated that many cancers can induce the proliferation of Tregs and/or promote their generation from naive T cells, resulting in the accumulation of these cells in the tumor beds and in the periphery. Importantly, the elimination and/or functional inactivation of tumor-induced Tregs can promote anti-tumor immunity and enhance the efficacy of immunotherapy.

Chronic inflammation characterized by continued expression of pro-inflammatory cytokines and recruitment of immune cells to the liver contributes to hepatic carcinogenesis [[2\]](#page-4-1). The inhibition of tumor-specific immune surveillance in this chronic inflammatory milieu is mediated by increased frequency of Tregs, MDSCs, changes in the expression pattern of inhibitory immune checkpoints, alterations in the function of dendritic cells (DC) and release of immunosuppressive cytokines. Decline in  $CD4<sup>+</sup>$  T effector cell function with progressive tumor growth in HCC is related to increased Treg-mediated suppression [\[3](#page-4-2), [4](#page-4-3)]. Even though the suppressive role of Tregs during the inflammation is beneficial for maintenance of normal immune homeostasis, increase in the number of Tregs during tumor progression suppresses  $CD8<sup>+</sup> T$  effector cell-mediated anti-tumor immunity, hence creating a barrier to successful immunotherapy of cancer  $[5-7]$  $[5-7]$ . On the contrary, a

paradox has been reported in colorectal cancer; while some studies concluded that high density of Tregs in the tumor microenvironment is associated with better prognosis, other studies reported their elevated numbers are associated with advanced stage tumors and poor overall survival [[8\]](#page-4-6). However, the benefit or hazard of Tregs in colorectal cancer (CRC) is highly debated. It seems that Tregs have differential influences at different stages of CRC development. Excessive activation of Tregs also contributes to persistent HBV infection and progression of HCC  $[9]$  $[9]$ , and HBV<sup>+</sup> HCC patients have Tregs with greater suppressive potential than non-HBV<sup>+</sup> HCC patients  $[10]$  $[10]$ . Negative role of Tregs through dampening favorable  $CD8<sup>+</sup>$  T cell responses to tumor-associated antigens has been reported in breast cancer patients, and patients with low  $CD8<sup>+</sup>$  T cells/Tregs ratio had a better survival benefit [\[11](#page-5-2)]. Tregs also play an important role in the progression and metastasis of lung cancer, the most lethal disease globally which has no effective therapies and expression of tumor necrosis receptor factor type II (TNRF2) by circulating Tregs in these patients correlated with poor clinical outcome [\[12](#page-5-3), [13](#page-5-4)]. The inhibitory effect of Tregs on  $CD4^+$  and  $CD8^+$  T cells can be abrogated by treatment with anti-chemokine receptor type-4 (CCR4) and anti-CD56 antibody in vitro which represent a novel strategy for the elimination of Tregs in the therapeutic settings of lung cancer patients [[14\]](#page-5-5).

# **Role of MDSCs in suppression of anti‑tumor immunity**

MDSCs are a heterogeneous population of immature myeloid cells that originate from the bone marrow, and normally differentiate into dendritic cells (DCs) or macrophages. However, their maturation is halted in malignancy, and they infiltrate into tumor microenvironment. MDSCs induce a state of chronic tissue inflammation and immune suppression that is characterized by the production of reactive oxygen species (ROS), nitric oxide (NO), arginase-1 (Arg-1) and cytokines IL-1, IL-6 and TNF-α. They can induce Tregs and attract suppressive tumor-associated macrophages (TAM) [[15\]](#page-5-6). The expansion of both granulocytic and monocytic MDSCs in a broad spectrum of human cancers has been extensively reviewed by Stromnes et al. [\[16](#page-5-7)]. Abrogation of MDSC activity by targeted depletion has been shown to enhance endogenous CTL-mediated tumor cell killing, which suggests that failure of immune surveillance to cancer may be in part due to MDSCs [\[17](#page-5-8)]. Several strategies for therapeutic targeting of MDSCs in malignancy have been detailed, and such strategies include depletion of MDSCs, interfering with suppressive activity of MDSCs by using cyclooxygenase-2 (COX2) inhibitors and phosphodiesterase-5 (PDE-5) inhibitors [\[16](#page-5-7)[–19](#page-5-9)].

MDSCs are recruited by IDO expression in the tumor microenvironment. In addition to attracting MDSCs, IDO is instrumental in the deprivation of tryptophan in the tumor microenvironment, thus inhibiting T cell activation [[16\]](#page-5-7). IDO-mediated tryptophan depletion inhibits the kinases mammalian target of rapamycin (mTOR) and protein kinase C (PKC). The novel experimental inhibitor of IDO, 1-methyl p-tryptophan, relieves the inhibition of mTOR and PKC by acting as a tryptophan mimetic. Thus, IDO represents an attractive potential target for cancer therapy [\[20](#page-5-10)].

Increased levels of HLA-DR<sup>low/neg</sup> CD14<sup>+</sup> MDSCs detected in the circulation of patients with melanoma [\[21](#page-5-11)], HCC [\[9](#page-5-0), [22\]](#page-5-12), rectal cancer [\[23](#page-5-13)] and prostate cancer [[24\]](#page-5-14) correlated with poor prognosis; furthermore, a reciprocal relationship between Tregs and MDSCs has been demonstrated in HCC and prostate cancer patients [\[9](#page-5-0), [24](#page-5-14), [25\]](#page-5-15). In contrast, an inverse correlation between the frequency of MDSCs and the presence of functional antigen-specific T cells found in melanoma patients and breast cancer patients highlights the potent immunosuppressive role of MDSCs in cancer [[11,](#page-5-2) [21\]](#page-5-11). Thus, several studies conducted in cancer patients and preclinical models suggest the pro-tumorigenic nature of MDSCs [[1\]](#page-4-0) and therapeutic targeting of MDSCs to maximize the efficacy of immune-based therapies are being considered in clinical trial settings [[16,](#page-5-7) [26–](#page-5-16)[28\]](#page-5-17).

### **Inhibitory checkpoint receptors and "T cell exhaustion"**

PD-1 is a checkpoint receptor expressed on activated T cells that down-modulates effector T cell function and prevents the formation of immune memory. Interaction of activated PD-1<sup>+</sup> T cells with PD-L1-expressing tumor cells, immune cells such as DCs, macrophages, NK cells, monocytes and B cells results in suppression of T cells. Upregulation of PD-L1 by neoplastic cells allows tumors to escape the anti-tumor effector T cell responses. Studies conducted in different types of human cancers have shown that PD-L1 expression on tumor cells reflects an immune reactive microenvironment, and responsiveness to anti-PD-1 therapy has strong correlation with tumor PD-L1 expression [\[29](#page-5-18)]. Impressive clinical responses have been achieved by therapeutic targeting of T cell inhibitory pathways using monoclonal antibodies directed against immune checkpoints PD-1, PD-L1 and CTLA-4. While therapeutic efficacy of PD-1/PD-L1 blockade has been shown to be promising with accomplishment of better survival and durable remission reported in different types of solid malignancies [\[30](#page-5-19)[–32](#page-5-20)], this represents a potential strategy to target immune suppression in a variety of hematological malignancies [\[33](#page-5-21)]. Soluble CD80-mediated intervention of PD-1/PD-L1 axis can restore the activation of both CD4<sup>+</sup> and  $CD8^+$  T cells with IFN- $\gamma$  production [\[34](#page-5-22), [35](#page-5-23)]. Virotherapy using oncolytic viruses has been reported to reduce the systemic resistance to PD-1 immunotherapy in an experimental model of liver cancer, thus providing the rationale for combining oncolytic viruses and immune checkpoint blockade in clinical trial [[36\]](#page-5-24). Immune checkpoint receptors are recognized as important players of immune suppression in HCC [\[37](#page-5-25), [38](#page-5-26)]. We have reported the increased frequency of exhausted T cells expressing PD-1 in HCC patients with advanced stage of the disease, which contributes to incompetent T effector cell function; selective in vitro depletion of the immunosuppressive cells resulted in partial restoration of T effector cell function in HCC  $[25,$  $[25,$ [39](#page-5-27)].

CTLA-4 is another inhibitory receptor pathway hindering T cell function in cancer; it competes with CD28 for binding to the co-stimulatory receptors CD80 and CD86 on antigen-presenting cells, thus impairing initial stages of T cell activation and proliferation. While PD-1 plays a central role in downregulating activated T cells in the periphery, CTLA-4 predominantly regulates early stages of T cell activation. Ipilimumab, an anti-CTLA-4 monoclonal antibody, augmented anti-CD3-driven T cell proliferation in vitro and enhanced bispecific antibody-directed cytolysis of tumor cells by activated T cells; this is implicated to attenuation of Treg-mediated suppression by ipilimumab [[40\]](#page-5-28). Ipilimumab has been administered for the treatment of melanoma in combination with bevacizumab, and improvement in overall survival was recorded in melanoma patients [\[41](#page-5-29)]. In a recent study, combination therapy using ipilimumab and anti-PD-1 antibody, nivolumab, has been shown to produce significant improvement in the progression-free survival of metastatic melanoma patients as compared to ipilimumab alone. This combination therapy was found to be more effective in PD-L1-negative tumors [\[42](#page-5-30)].

## **Immune suppression in HCC**

HCC is an inflammation-associated malignancy, and majority of the patients suffer from cirrhosis of viral etiology or chronic alcohol consumption. Immunogenicity of HCC renders it a suitable candidate for immunotherapeutic approaches. Alfa-fetoprotein (AFP), glypican-3 (GPC-3), New York esophageal squamous cell cancer-1 (NY-ESO-1) and melanoma antigen-A (MAGE-A) are the prominent and most studied antigens in HCC. Spontaneous cellular or humoral responses against NY-ESO1 antigen have been reported in 50 % of the patients [\[43](#page-5-31)]. Infiltration of tumorassociated antigen-specific  $CD8<sup>+</sup>$  T cells was reported in 50 % of HCC patients and antigen-specific  $CD8<sup>+</sup>$  T cell responses correlated with progression-free survival [\[44](#page-6-0)]. Nevertheless, HCC exploits multiple immunosuppressive pathways to effectively evade the host's anti-tumor immune responses. The immune regulatory mechanism that prevents autoimmunity may be harnessed by tumor cells to accomplish immune escape or the evasion of immune destruction. Regulatory T cells (Tregs), myeloid-derived suppressor cells  $[1]$  $[1]$  and PD-1<sup>+</sup> T cells are major components of the immune system acting in concert with causing the subversion of anti-tumor immunity in the tumor microenvironment [[25,](#page-5-15) [39,](#page-5-27) [44](#page-6-0), [45](#page-6-1)]. This redundant immunosuppressive network in HCC poses an important obstacle to success of immunotherapy. Detailed knowledge of these factors responsible for creating an immunosuppressive milieu is essential for the development of novel immunebased therapeutic modalities.

Several studies have reported that dysregulated immune function poses an impediment to immunotherapeutic approaches in HCC. While many of these studies have focused on investigating one single aspect of dysfunctional immunity in HCC the impact of either Tregs [\[4](#page-4-3)] or MDSCs [\[22](#page-5-12)] on abrogation of anti-tumor immunity, we were the first to perform a global analysis of immune dysfunction in this patient population [\[25](#page-5-15)]. HCC exploits multiple immunosuppressive mechanisms to evade active immune surveillance of the host; this includes induction of Tregs, recruitment of MDSCs, accumulation of exhausted T effector cells, defective antigen presentation by dendritic cells (DCs) and overproduction of inhibitory cytokines such as IL-10 and TGF-β1 [\[25](#page-5-15), [37](#page-5-25), [38](#page-5-26), [44](#page-6-0)].

Since CD25 is expressed on both activated T effector cells and Tregs, this marker alone is incapable of accurately discriminating Tregs from activated effector T cells. Expression of Foxp3 alone is also not a reliable marker, as this protein is also expressed on non-immunosuppressive Tregs. Therefore, we have used multiple markers (CD3+CD4+Foxp3+CD127−) for the identification and characterization of Tregs and detected significantly elevated frequency and absolute number of this phenotype in HCC patients [[25\]](#page-5-15). Following a recent workshop study, a consensus has been reached over the controversy regarding the markers that should be used for the identification of Tregs, and the same markers used in our study have been validated and proposed as essential marker set for the identification of Tregs from PBMC [\[46](#page-6-2)]. Furthermore, intracellular localization of Foxp3 protein precludes its use for therapeutic targeting and depletion of suppressive Tregs using specific antibodies. In order to identify highly immunosuppressive Tregs bearing markers that can be therapeutically targeted, we evaluated the expression of CTLA-4 and glycoprotein A repetition predominant (GARP) on the surface of  $F\alpha p3$ <sup>+</sup> Tregs. High prevalence of this phenotype in patients with advanced HCC highlighted the severe immune dysfunction in these patients and provided the rationale for therapeutic depletion of immunosuppressive Tregs.

The central role of MDSCs in dampening anti-tumor immunity in HCC has not been appreciated before, despite its high importance as a potent immunosuppressive cell subset in other cancers. Recognizing the potential relationship between MDSCs and induction of Tregs during the progression of malignancy, we therefore evaluated the frequency of CD14−HLA-DR−CD11b+CD33+ MDSCs. In conjunction with elevated levels of Tregs, the frequency and absolute number of MDSCs were significantly high in HCC patients. Additionally, the frequency of MDSCs showed excellent correlation with frequency of Tregs, indicating that the accumulation of MDSC is in concurrence with Treg accumulation and the interplay between these immunosuppressive cell subsets is very critical in the establishment of tolerogenic tumor microenvironment in HCC (Fig. [1a](#page-4-7), Copyright permission of [[39\]](#page-5-27)). Thus, depletion of one of these subsets is unlikely to have a beneficial impact on anti-tumor immune responses in HCC, as one of these cell types can induce the generation of other by creating a feedback loop. This provided the rationale for the combined depletion of Tregs and MDSCs, which resulted in restoration of T effector cell functions, enhancement in T cell proliferation and granzyme B production (Fig. [1b](#page-4-7)).

Heightened immunosuppression in HCC is additionally reflected by elevated levels of Treg-derived cytokines IL-10 and TGF-β1 with concomitant downregulation of IFN-γ, a potent anti-tumor cytokine whose diminished levels could be attributed to high prevalence of suppressive Tregs in HCC. Serum interferon-γ levels may reflect host's anti-tumor immunity and may be potential marker of HCC recurrence after curative therapy in patients [[47\]](#page-6-3).

Since HCC has strong association with chronic viral infections (HBV/HCV), appearance of exhausted T cells expressing PD-1 may be considered as a hallmark of HCC patients with viral etiology. Thus, abundance of  $PD-1$ <sup>+</sup> T cells detected in HCC patients is an indication of yet another level of immune dysregulation that would need to be circumvented, in order to accomplish efficacious anti-tumor immune responses. Functional defects in both  $CD4^+$  and  $CD8^+$  T cells reflected by decreased proliferation and diminished granzyme B or IFN-γ secretion in response to anti-CD3/CD28/mitogenic stimulation is not surprising; this could be due to the cumulative effect of underlying immunosuppressive burden in these patients (Fig. [1](#page-4-7)a). Multi-pronged depletion of suppressive cell subsets restored the proliferation of both  $CD4<sup>+</sup>$  T helper cells and  $CD8<sup>+</sup>$  CTL, but not to the levels equivalent to T cells from healthy controls. Nonetheless, combined depletion of suppressive cells fully restored granzyme B production



<span id="page-4-7"></span>Fig. 1 Targeted depletion of Tregs, exhausted PD-1<sup>+</sup> T helper cells and myeloid-derived suppressor cells may restore effective anti-tumor T cell function. **a** Tumor cell-mediated induction of exhausted effector T helper cells and immunosuppressive GARP+CTLA-4+ Tregs and myeloid-derived suppressor cells diminish the anti-tumor capac-

by CD8+ T cells to the levels equivalent to those observed with T cells from healthy donors (Fig. [1b](#page-4-7)). IFN- $\gamma$  production by T cells also improved but to a lesser extent. Thus, elevated levels of immunosuppressive cells in HCC patients compromise CTL function, partly by inhibiting granzyme B production, resulting in attenuated tumor cell killing.

#### **Conclusions**

In conclusion, even though there is abundant in vitro evidence for human T cell reactivity against tumors, such responses are often ineffective in vivo as tumor cells exploit multiple mechanisms to avoid immune cell recognition and anti-tumor effector cell function, thereby limiting the clinical benefits of immunotherapeutic strategies. Mitigation of immunosuppressive cells to augment Th1 and CTL response is critical for improving the efficacy of immunotherapy of cancer, which may impact the survival of patients.

**Acknowledgments** This research in the Thanavala Lab was supported in part through discretionary funds available to Dr. Thanavala. We gratefully acknowledge Dr. Paul Wallace and Earl Timm for their help in designing flow cytometry experiments. The patient samples were obtained through the Roswell Park Cancer Institute Data Bank and Biorepository which is a Cancer Center Support Grant (CCSG) Shared Resource supported by National Institute of Health P30 CA016056-27.

ity of Th1 and CTL by inhibition of IFN-γ and granzyme B production. **b** The restoration of effector T cell proliferation and granzyme B production (but not IFNγ secretion) by the combined removal of all these immunosuppressive cells may restore anti-tumor Th1 and CTL responses (adapted from [\[39\]](#page-5-27))

#### **Compliance with ethical standards**

**Conflict of interest** None of the authors have any conflict of interest.

#### **References**

- <span id="page-4-0"></span>1. Schrader J (2013) The role of MDSCs in hepatocellular carcinoma—in vivo veritas? J Hepatol 59:921–923
- <span id="page-4-1"></span>2. Capece D, Fischietti M, Verzella D, Gaggiano A, Cicciarelli G, Tessitore A et al (2013) The inflammatory microenvironment in hepatocellular carcinoma: a pivotal role for tumorassociated macrophages. Biomed Res Int 2013:187204. doi[:10.1155/2013/187204](http://dx.doi.org/10.1155/2013/187204)
- <span id="page-4-2"></span>3. Cabrera R, Szabo G (2013) Another armed CD4(+) T cell ready to battle hepatocellular carcinoma. Hepatology 58:1–3
- <span id="page-4-3"></span>4. Fu J, Zhang Z, Zhou L, Qi Z, Xing S, Lv J et al (2013) Impairment of CD4<sup>+</sup> cytotoxic T cells predicts poor survival and high recurrence rates in patients with hepatocellular carcinoma. Hepatology 58:139–149
- <span id="page-4-4"></span>5. Pastille E, Bardini K, Fleissner D, Adamczyk A, Frede A, Wadwa M et al (2014) Transient ablation of regulatory T cells improves antitumor immunity in colitis-associated colon cancer. Cancer Res 74:4258–4269
- 6. Makarova-Rusher OV, Medina-Echeverz J, Duffy AG, Greten TF (2015) The yin and yang of evasion and immune activation in HCC. J Hepatol 62:1420–1429
- <span id="page-4-5"></span>7. Huang Y, Wang F, Wang Y, Zhu Z, Gao Y, Ma Z et al (2014) Intrahepatic interleukin-17<sup>+</sup> T cells and  $FoxP3$ <sup>+</sup> regulatory T cells cooperate to promote development and affect the prognosis of hepatocellular carcinoma. J Gastroenterol Hepatol 29:851–859
- <span id="page-4-6"></span>8. Zhang X, Kelaria S, Kerstetter J, Wang J (2015) The functional and prognostic implications of regulatory T cells in colorectal carcinoma. J Gastrointest Oncol 6:307–313
- <span id="page-5-0"></span>9. Kondo Y, Shimosegawa T (2015) Significant roles of regulatory T cells and myeloid derived suppressor cells in hepatitis B virus persistent infection and hepatitis B virus-related HCCs. Int J Mol Sci 16:3307–3322
- <span id="page-5-1"></span>10. Sharma S, Khosla R, David P, Rastogi A, Vyas A, Singh D et al (2015) CD4+CD25+CD127 (low) regulatory T cells play predominant antitumor suppressive role in hepatitis B virus-associated hepatocellular carcinoma. Front Immunol 6:49. doi[:10.3389/fimmu.2015.00049](http://dx.doi.org/10.3389/fimmu.2015.00049)
- <span id="page-5-2"></span>11. Bailur JK, Gueckel B, Derhovanessian E, Pawelec G (2015) Presence of circulating Her2-reactive CD8<sup>+</sup> T-cells is associated with lower frequencies of myeloid-derived suppressor cells and regulatory T cells, and better survival in older breast cancer patients. Breast Cancer Res 17:34. doi[:10.1186/](http://dx.doi.org/10.1186/s13058-015-0541-z) [s13058-015-0541-z](http://dx.doi.org/10.1186/s13058-015-0541-z)
- <span id="page-5-3"></span>12. Zhang D, Chen Z, Wang DC, Wang X (2015) Regulatory T cells and potential inmmunotherapeutic targets in lung cancer. Cancer Metastasis Rev 34:277–290
- <span id="page-5-4"></span>13. Yan F, Du R, Wei F, Zhao H, Yu J, Wang C et al (2015) Expression of TNFR2 by regulatory T cells in peripheral blood is correlated with clinical pathology of lung cancer patients. Cancer Immunol Immunother 64:1475–1485
- <span id="page-5-5"></span>14. Kurose K, Ohue Y, Sato E, Yamauchi A, Eikawa S, Isobe M et al (2015) Increase in activated Treg in TIL in lung cancer and in vitro depletion of Treg by ADCC using an antihuman CCR4 mAb (KM2760). J Thorac Oncol 10:74–83
- <span id="page-5-6"></span>15. Talmadge JE, Gabrilovich DI (2013) History of myeloid-derived suppressor cells. Nat Rev Cancer 13:739–752
- <span id="page-5-7"></span>Stromnes IM, Greenberg PD, Hingorani SR (2014) Molecular pathways: myeloid complicity in cancer. Clin Cancer Res 20:5157–5170
- <span id="page-5-8"></span>17. Stromnes IM, Brockenbrough JS, Izeradjene K, Carlson MA, Cuevas C, Simmons RM et al (2014) Targeted depletion of an MDSC subset unmasks pancreatic ductal adenocarcinoma to adaptive immunity. Gut 63:1769–1781
- 18. Mao Y, Poschke I, Wennerberg E, de Pico CY, Egyhazi BS, Schultz I et al  $(2013)$  Melanoma-educated CD14<sup>+</sup> cells acquire a myeloid-derived suppressor cell phenotype through COX-2-dependent mechanisms. Cancer Res 73:3877–3887
- <span id="page-5-9"></span>19. Califano JA, Khan Z, Noonan KA, Rudraraju L, Zhang Z, Wang H et al (2015) Tadalafil augments tumor specific immunity in patients with head and neck squamous cell carcinoma. Clin Cancer Res 21:30–38
- <span id="page-5-10"></span>20. Metz R, Rust S, Duhadaway JB, Mautino MR, Munn DH, Vahanian NN et al (2012) IDO inhibits a tryptophan sufficiency signal that stimulates mTOR: a novel IDO effector pathway targeted by D-1-methyl-tryptophan. Oncoimmunology 1:1460–1468
- <span id="page-5-11"></span>21. Weide B, Martens A, Zelba H, Stutz C, Derhovanessian E, Di Giacomo AM et al (2014) Myeloid-derived suppressor cells predict survival of patients with advanced melanoma: comparison with regulatory T cells and NY-ESO-1- or melan-A-specific T cells. Clin Cancer Res 20:1601–1609
- <span id="page-5-12"></span>22. Arihara F, Mizukoshi E, Kitahara M, Takata Y, Arai K, Yamashita T et al  $(2013)$  Increase in CD14<sup>+</sup> HLA-DR -/low myeloidderived suppressor cells in hepatocellular carcinoma patients and its impact on prognosis. Cancer Immunol Immunother 62:1421–1430
- <span id="page-5-13"></span>23. Napolitano M, D'Alterio C, Cardone E, Trotta AM, Pecori B, Rega D et al (2015) Peripheral myeloid-derived suppressor and T regulatory PD-1 positive cells predict response to neoadjuvant short-course radiotherapy in rectal cancer patients. Oncotarget 6:8261–8270
- <span id="page-5-14"></span>24. Idorn M, Kollgaard T, Kongsted P, Sengelov L, Thor SP (2014) Correlation between frequencies of blood monocytic myeloidderived suppressor cells, regulatory T cells and negative prognostic markers in patients with castration-resistant metastatic prostate cancer. Cancer Immunol Immunother 63:1177–1187
- <span id="page-5-15"></span>25. Kalathil S, Lugade AA, Miller A, Iyer R, Thanavala Y (2013) Higher frequencies of  $GARP(+)CTLA-4(+)Foxp3(+)T regular$ tory cells and myeloid-derived suppressor cells in hepatocellular carcinoma patients are associated with impaired T-cell functionality. Cancer Res 73:2435–2444
- <span id="page-5-16"></span>26. Medina-Echeverz J, Eggert T, Han M, Greten TF (2015) Hepatic myeloid-derived suppressor cells in cancer. Cancer Immunol Immunother 64:931–940
- 27. Diaz-Montero CM, Finke J, Montero AJ (2014) Myeloid-derived suppressor cells in cancer: therapeutic, predictive, and prognostic implications. Semin Oncol 41:174–184
- <span id="page-5-17"></span>28. Zhu Y, Hawkins WG, DeNardo DG (2015) Regramming myeloid responses to improve cancer immunotherapy. Oncoimmunology 4:e974399. doi:[10.4161/2162402X.2014.974399](http://dx.doi.org/10.4161/2162402X.2014.974399)
- <span id="page-5-18"></span>29. Taube JM, Klein A, Brahmer JR, Xu H, Pan X, Kim JH et al (2014) Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. Clin Cancer Res 20:5064–5074
- <span id="page-5-19"></span>30. Topalian SL, Sznol M, McDermott DF, Kluger HM, Carvajal RD, Sharfman WH et al (2014) Survival, durable tumor remission, and long-term safety in patients with advanced melanoma receiving nivolumab. J Clin Oncol 32:1020–1030
- 31. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R et al (2013) Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. N Engl J Med 369:134–144
- <span id="page-5-20"></span>32. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF et al (2012) Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med 366:2443–2454
- <span id="page-5-21"></span>33. Andersen MH (2014) The targeting of immunosuppressive mechanisms in hematological malignancies. Leukemia 28:1784–1792
- <span id="page-5-22"></span>34. Haile ST, Dalal SP, Clements V, Tamada K, Ostrand-Rosenberg S (2013) Soluble CD80 restores T cell activation and overcomes tumor cell programmed death ligand 1-mediated immune suppression. J Immunol 191:2829–2836
- <span id="page-5-23"></span>35. Ostrand-Rosenberg S, Horn LA, Haile ST (2014) The programmed death-1 immune-suppressive pathway: barrier to antitumor immunity. J Immunol 193:3835–3841
- <span id="page-5-24"></span>36. Woller N, Gurlevik E, Fleischmann-Mundt B, Schumacher A, Knocke S, Kloos AM et al (2015) Viral infection of tumors overcomes resistance to PD-1-immunotherapy by broadening neoantigenome-directed T-cell responses. Mol Ther 23:1630–1640
- <span id="page-5-25"></span>37. Schmidt N, Flecken T, Thimme R (2014) Tumor-associated antigen specific CD8 T cells in hepatocellular carcinoma—a promising target for immunotherapy. Oncoimmunology 3:e954919. doi: [10.4161/21624011.2014.954919](http://dx.doi.org/10.4161/21624011.2014.954919)
- <span id="page-5-26"></span>38. Hato T, Goyal L, Greten TF, Duda DG, Zhu AX (2014) Immune checkpoint blockade in hepatocellular carcinoma: current progress and future directions. Hepatology 60:1776–1782
- <span id="page-5-27"></span>39. Lugade AA, Kalathil S, Miller A, Iyer R, Thanavala Y (2013) High immunosuppressive burden in advanced hepatocellular carcinoma patients: Can effector functions be restored? Oncoimmunology 2:e24679. doi:[10.4161/onci.24679](http://dx.doi.org/10.4161/onci.24679)
- <span id="page-5-28"></span>40. Yano H, Thakur A, Tomaszewski EN, Choi M, Deol A, Lum LG (2014) Ipilimumab augments antitumor activity of bispecific antibody-armed T cells. J Transl Med 12:191. doi[:10.1186/1479-5876-12-191](http://dx.doi.org/10.1186/1479-5876-12-191)
- <span id="page-5-29"></span>41. Hodi FS, Lawrence D, Lezcano C, Wu X, Zhou J, Sasada T et al (2014) Bevacizumab plus ipilimumab in patients with metastatic melanoma. Cancer Immunol Res 2:632–642
- <span id="page-5-30"></span>42. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD et al (2015) Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. N Engl J Med 373:23–34
- <span id="page-5-31"></span>43. Korangy F, Ormandy LA, Bleck JS, Klempnauer J, Wilkens L, Manns MP et al (2004) Spontaneous tumor-specific humoral and cellular immune responses to NY-ESO-1 in hepatocellular carcinoma. Clin Cancer Res 10:4332–4341
- <span id="page-6-0"></span>44. Flecken T, Schmidt N, Hild S, Gostick E, Drognitz O, Zeiser R et al (2014) Immunodominance and functional alterations of tumor-associated antigen-specific CD8+ T-cell responses in hepatocellular carcinoma. Hepatology 59:1415–1426
- <span id="page-6-1"></span>45. Zhao HQ, Li WM, Lu ZQ, Yao YM (2014) Roles of Tregs in development of hepatocellular carcinoma: a meta-analysis. World J Gastroenterol 20:7971–7978
- <span id="page-6-2"></span>46. Santegoets SJ, Dijkgraaf EM, Battaglia A, Beckhove P, Britten CM, Gallimore A et al (2015) Monitoring regulatory T cells in

clinical samples: consensus on an essential marker set and gating strategy for regulatory T cell analysis by flow cytometry. Cancer Immunol Immunother 64:1271–1286

<span id="page-6-3"></span>47. Lee IC, Huang YH, Chau GY, Huo TI, Su CW, Wu JC et al (2013) Serum interferon gamma level predicts recurrence in hepatocellular carcinoma patients after curative treatments. Int J Cancer 133:2895–2902