

# Toxicological Evaluation of the Aqueous Extract of *Caralluma europaea* and Its Immunomodulatory and Inflammatory Activities

Zineb Issiki, Chaimaa Moundir, Farida Marnissi<sup>1</sup>, Nadia Seddik, Naima Benjelloun, Younes Zaid<sup>2</sup>, Mounia Oudghiri

Department of Biology, Immunology and Biodiversity Laboratory, Faculty of Sciences, Hassan II University of Casablanca, B.P 5366 Maarif, <sup>1</sup>Department of Anatomopathology, University Hospital Center Ibn Rochd, 19, rue Tarik Bnou Ziad, Mers Sultan, <sup>2</sup>National Research Laboratory, University Mohammed VI of Health Sciences, Rue Ali Bnou Abi Taleb, Quartier Parc de la Ligue Arabe, Casablanca, Morocco

## ABSTRACT

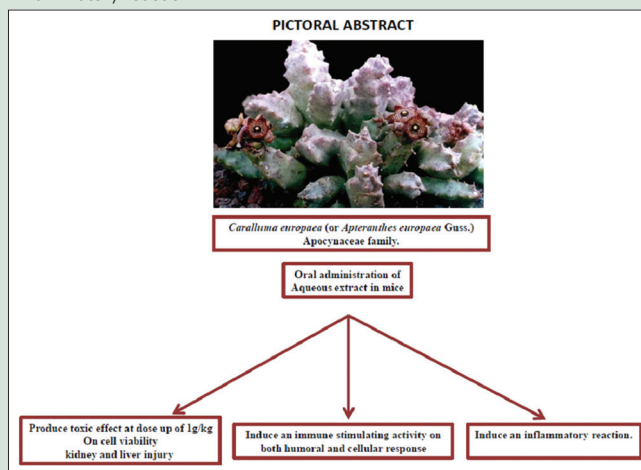
**Background:** *Caralluma europaea* (CE) has been studied for its chemical constituents, and no information is available on its toxicity or its pharmacological activities. **Objective:** To determine the toxicity of an aqueous extract of CE stems *in vitro* and *in vivo* after acute and subchronic oral gavages in Swiss albino's mice and its immunomodulatory and inflammatory activities. **Materials and Methods:** The extract was administrated in single oral dose at 5 g/kg body weight for the acute toxicity test and by gavages daily at doses of 1, 2.5, or 5 g/kg for 30 consecutive days for the subchronic toxicity test. The immunomodulatory activities and inflammatory activities were tested by the evaluation of hemagglutination antibodies (HAs) titers and delayed-type hypersensitivity (DTH) response. **Results:** For the dose of 1 g/kg, no visible toxic effects were observed. However, for the higher doses, clinical observations of toxicity were noted after 1 week of treatment. This was confirmed by the biochemical parameters values and the histology analyses of the spleen, liver, and kidney tissues. The high cellular mortality rate *in vitro* when treated with CE extract confirmed their toxicity potential. There was also increase of "HA titer" and "DTH" response in mice treated with nontoxic dose of CE (1 g/kg) compared to control group. This immune activity was confirmed by the high number of lymphocytes infiltrates noted in the different organs. **Conclusion:** We conclude that CE at the dose up of 1 g/kg produced toxic effect in mice that induced an immune inflammatory reaction.

**Key words:** *Caralluma europaea*, immunomodulatory reaction, inflammatory reaction, toxicity *in vivo* and *in vitro*

## SUMMARY

- Caralluma europaea (CE) has been studied for its chemical constituents, and no information is available on its toxicity or its pharmacological activities. The objective is to determine the toxicity of an aqueous extract of CE stems *in vitro* and *in vivo* after acute and subchronic oral gavages in Swiss albino's mice and its immunomodulatory and inflammatory activities. For the dose of 1 g/kg, no visible toxic effects were observed. However, for the higher doses, clinical observations of toxicity were noted after 1 week of treatment. This was confirmed by the biochemical parameters values and the histology analyses of the spleen, liver, and kidney tissues. The high cellular mortality rate *in vitro* confirmed their toxicity potential. There was also increase of "hemagglutination antibody titer" and "delayed-type hypersensitivity" response in mice treated with nontoxic dose of CE (1 g/kg) compared to control group. This immune activity was confirmed by the high number of

lymphocytes infiltrates noted in the different organs. We conclude that CE at the dose up of 1 g/kg produced toxic effect in mice that induced an immune inflammatory reaction.



**Abbreviations Used:** CE: *Caralluma europaea*, ALT: Alanine aminotransferase, AST: Aspartate aminotransferase, RRBCs: Rat red blood cells, DTH: Delayed-type hypersensitivity response, PBS: Phosphate buffer solution.

## Correspondence:

Prof. Mounia Oudghiri,  
Immunology and Biodiversity Laboratory,  
Faculty of Sciences, Hassan II University of  
Casablanca,  
B.P 5366 Maarif, Casablanca, Morocco.  
E-mail: mouniaoudghiri@gmail.com,  
mounia.oudghiri@univh2c.ma  
DOI: 10.4103/pr.pr\_24\_17

## Access this article online

Website: [www.phcogres.com](http://www.phcogres.com)

## Quick Response Code:



## INTRODUCTION

*Caralluma europaea* (CE) (or *Apteranthes europaea* Guss.) a member of *Apocynaceae* family<sup>[1]</sup> is distributed in Egypt, Spain, Italy, Libya, Tunisia, Algeria, and Morocco.<sup>[2]</sup> Several members of genus *Caralluma* have found various medicinal uses: antidiabetic, antihyperglycemic, antiparasitic, antitrypanosomal, antiulcer, neuroprotective, antipyretic, anti-inflammatory, antinociceptive, antioxidant, antiobesogenic, and antiatherosclerotic properties.<sup>[3-12]</sup> This genus has been extensively explored for a variety of pharmacological activities as compared with other species, but not all species were tested for their biological activity.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

**For reprints contact:** [reprints@medknow.com](mailto:reprints@medknow.com)

**Cite this article as:** Issiki Z, Moundir C, Marnissi F, Seddik N, Benjelloun N, Zaid Y, et al. Toxicological evaluation of the aqueous extract of *Caralluma europaea* and its immunomodulatory and inflammatory activities. *Phcog Res* 2017;9:390-5.

Locally known as “Daghmous,” “Zakkum,” or “Tikiwt,” CE is a plant species communally used in Moroccan traditional medicine for their presumed anticancer activity.<sup>[13,14]</sup> The only few studies on CE cited in the literature were on their chemical constituents (monoterpenoids, terpinolene [23.3%],  $\alpha$ -terpinene [19.1%] and linalool [18.4%], flavonoids) and their possible role in the biology of pollination.<sup>[15,16]</sup> Zito *et al.* have reported that aromatic compounds found in stems and fruits of CE are semi-chemicals for many insects. Some constituents show antimicrobial activity against *Candida albicans*, *Clostridium welchii*, and *Staphylococcus aureus*<sup>[17]</sup> or antifungal activities on the plant pathogenic fungi such as *Rhizoctonia solani*, *Pythium ultimum*, *Pyrenophora avenae*, and *Crinipellis pernicioso*.<sup>[18]</sup> However, no sufficient information is available on the toxicity of CE or its derived effects *in vivo*.

The toxic effect of plant *in vivo* or *in vitro* and its immunomodulatory and anti-inflammatory potential have never been investigated before. The present study was carried out to determine the toxicity of an aqueous extract of CE after acute and subchronic oral gavages in mice at different doses and its immunomodulatory and inflammatory activities.

## MATERIALS AND METHODS

### Plant material

The aerial part of CE was collected in Beni-Mellal city, Morocco, and was authenticated by Professor Najat Khiyati, a Plant Taxonomist, at the Department of Biology, Faculty of Sciences, University Hassan II of Casablanca. A voucher specimen of the plant sample was deposited in the National Scientific Institute, Rabat, for future reference.

### Animals

Young adult male mice Swiss (20–30 g) were purchased from the animal house of the Department of Biology, Faculty of Sciences, Mohammed V University, Rabat, Morocco. The animals were kept in plastic cages in environmental conditions (22°C–24°C, 12-h:12-h dark/light cycle) allowed to drink water *ad-libitum* and standard pellet diet. Mice were deprived of food but with access to water 16–18 h prior the experiments. An adaptation period of 2 weeks was allowed before each experiment.

### Preparation of the aqueous extract of *Caralluma europaea*

The aerial part of CE has been air-dried and then pulverized. The aqueous extract was prepared by adding 1.5 L of distilled water to 150 g of CE powder, and the mixture has been heated under reflux at 60°C for 1 h in a round-bottom flask; then, the boiled decoction was centrifuged, filtered, and then concentrated in a rotary vacuum evaporator at 40°C. The extracted material was stored at –20°C until used. For oral administration (gavages), the crude extract was dissolved in water at a desired concentration which was prepared on the day of the experimental studies.

### Toxic activity of aqueous extract of *Caralluma europaea* *in vivo*

#### Acute toxicity test

The assessment of acute toxicity was performed according to the World Health Organization (WHO) guideline (WHO 2000) and the Organisation for Economic Cooperation and Development (OECD) guideline for testing of chemicals 420 (OECD 2001).<sup>[19]</sup> To conduct these studies, we receive the ethics committee approval of the university.

The aqueous extract was administrated in single oral dose at 5 g/kg body weight in 200  $\mu$ L while the control group received distilled water (5 mice/group). Signs of toxicity and mortality have been recorded at the 1<sup>st</sup>, 2<sup>nd</sup>, 4<sup>th</sup>, and 6<sup>th</sup> h after oral administration and then once daily for 14 days.<sup>[20,21]</sup>

#### Subacute toxicity test

Mice were divided into four groups containing five animals per group and housed in separate cages during the study. The aqueous extract was given orally by gavages to different groups daily at a dose of 1, 2.5, or 5 g/kg body weight for 30 consecutive days, while the control group received the vehicle only. The chosen doses were based on the dose of no-observed-adverse-effect-level (NOAEL) that was obtained from the acute toxicity study, 1 g/kg. The animals were closely monitored daily for general behavior and toxicity signs throughout the experimental period.<sup>[22]</sup> At the end of the treatment period (30 days), the mice were sacrificed and the blood samples and organs have been collected. Blood was collected in tubes with the anticoagulant, ethylenediaminetetraacetate. The blood was allowed to clot before centrifugation (3000 rpm at 4°C for 10 min) to obtain serum, which was analyzed for creatinine, urea, and the activity of liver enzymes: alanine aminotransferase (ALT) and aspartate aminotransferase (AST). Freshly dissected mice's liver, kidneys, and spleen were cut carefully in small slices and fixed in buffered formaldehyde solution (10%), dehydrated in ascending series of ethanol solutions, and embedded in paraffin. Then, 4–5- $\mu$ m thick sections of each tissue was prepared and stained with hematoxylin-eosin and examined under a light microscope; photomicrographs of the samples were recorded and interpreted by a pathologist.

### Toxic activity of aqueous extract of *Caralluma europaea* *in vitro*

Spleen cells from mice were cultured in RPMI (Invitrogen, Merelbeke, Belgium), media supplemented with 10% heat inactivated fetal calf serum, 4 mM glutamine, 100  $\mu$ g/mL gentamicin, and penicillin–streptomycin (200 U/mL and 200  $\mu$ g/mL) (Invitrogen) and 100  $\mu$ L of the plant extract at 0.5 or 2 mg/ml. The cells were maintained in continuous culture in a humid atmosphere at 37°C for 24 h. *In vitro* cytotoxicity assay was carried every 2 h, using Trypan blue (dye exclusion) method for mortality rate determination.<sup>[23]</sup>

### Immunomodulatory activities of assay

#### Antigen

Fresh rat blood was obtained by cardiac puncture. The rat red blood cells (RRBCs) were washed three times in a large volume of phosphate buffer solution (PBS) and centrifuged at 3000  $\times$  g for 10 min before use.

#### Assessment of humoral immune function

The method of Bin Hafeez *et al.* was used to determine the effect of the extracts on the antibody level resulting from sensitization with RRBC.<sup>[24]</sup> Briefly, mice were immunized by intraperitoneal (i.p) injection of 200  $\mu$ L of RRBCs suspension (30% v/v in PBS) on day 0. Mice of the study groups (5 mice) were treated with aqueous extract at 1 g/kg body weight administered orally 3 days before immunization and continued once daily for 7 days. The control group (5 mice) received only the vehicle. The mice were sacrificed by decapitation and blood samples were collected on day 7 for serum preparation. The blood was incubated for 1 h at 37°C, centrifuged, and supernatants pooled. The sera were incubated for 30 min at 56°C to inactivate complement and stored at –20°C until use. The primary antibody titer was determined by hemagglutination technique.<sup>[25]</sup>

#### Hemagglutination antibody titer

A micro technique employing 96-well microplates was used. Each well of the plate received 25  $\mu$ L of serial two-fold dilutions of sera in PBS. The dilution sera were challenged with 25  $\mu$ L of 1% (v/v) RRBCs in the plate. After incubating the mixtures for 2 h at room temperature, the hemagglutination capacity of the sera was read visually. Titers of sera

were determined as the reciprocal of the maximal dilution presenting positive hemagglutination. Each assay in this experiment was repeated three times.

#### Delayed-type hypersensitivity response

The effect of the plant extract on the antigen-specific cellular immune response in experimental animals was measured by determining the degree of DTH response using the footpad swelling test.<sup>[26]</sup> For sensitization, seven animals per group (control and treated) were immunized on day 0 by i.p injection with 200  $\mu$ L of a RRBC suspension (30% v/v in PBS). Seven days later (day + 7), these animals were injected subcutaneous with 50  $\mu$ L of same RRBC suspension in PBS into the right hind footpad for elicitation of the DTH reaction. Then, a footpad swelling was measured 24 h after the injection. The difference between the means of right and left hind footpad thickness gave a degree of footpad swelling which was used for group comparisons. To establish the effect of the extract on this immune response, a daily dose of 700  $\mu$ L of plant extract at 1 g/kg body weight in PBS was administered orally at different stages of the reaction: 2 days before the sensitization (day 0) and for 7 days after the induction. Simultaneously, another group of animals (controls) was inoculated in the same conditions with 700  $\mu$ L of PBS.

#### Statistical analysis

All studies mentioned above were done in triplicate. All values were expressed as mean  $\pm$  standard error of the mean and were analyzed by one-way analysis of variance, followed by Scheffe *post-hoc* test, and statistically significant findings were considered those in which  $P < 0.05$ .

## RESULTS

### Chronic toxicity studies of aqueous extract of *Caralluma europaea* in mice: Clinical observations of intoxication

Asthenia, hypoactivity, and urination were noticed immediately after gavages (15 min) and were more pronounced and persisted until the end of experimentation. The main behavioral signs of toxicity observed from the 3<sup>rd</sup> day of daily oral administration of aqueous extract of CE at 5 g/kg were atypical locomotion, anorexia, asthenia, ataxia, diarrhea, and urination. No mortality was observed after 14 days of treatment [Table 1]. Daily clinical observations are of major importance as well as the final observations.<sup>[27,28]</sup> The doses to be evaluated in chronic toxicity must be larger than that suggested for use in humans. This dose selection is critical for the study.<sup>[28]</sup> Finalizing, the studies carried out suggest that in 1 g/kg dose, the product seems to be safe. However, in 2 and 5 g/kg doses, some adverse effects were observed despite the fact of this dose being

much higher than that usually utilized in human beings (7–10 times the maximum indicated therapeutic dose in folk medicine).<sup>[13,14]</sup>

### Subchronic toxicity studies of aqueous extract of *Caralluma europaea* in mice

#### Clinical observations of intoxication

Daily oral administration of aqueous extract of CE at 1 g/kg, 2.5 g/kg, and 5 g/kg of body weight has induced an hyperactivity during the 1<sup>st</sup> week of the experimentation. At the end of the 2<sup>nd</sup> week, asthenia, hypoactivity, and urination with the loss of hair and the weight (20%) were observed with the doses of 2.5 and 5 g/kg. For the dose of 1 g/kg, no visible toxic effects were observed. The NOAEL was considered to be 1 g/kg/day. No death was observed for all doses.

#### Effects of aqueous extract of *Caralluma europaea* on biochemical parameters

Serum levels of ALT and AST and concentration of creatinine and urea were markedly and significantly ( $P < 0.05$ ) increased in mice treated by 2.5 and 5.0 g/kg of aqueous extract of the plant [Table 2]. This increase was dose dependent. In the groups that received 1 g/kg of CE, no differences were observed. The activities of AST and ALT are indicators of liver functions and the level of creatinine is an indicator for kidney activity. Therefore, CE induced serious kidney and liver injury for the higher doses 2.5 and 5 g/kg.

#### Effects of aqueous extract of *Caralluma europaea* on the spleen, kidney, and liver

Histopathological examination of the spleen, kidneys, and liver [Figure 1] at the end of the study showed that for all the doses used, there were histopathological changes. For the spleen, some hemosiderin deposits with foci of hemorrhage for the three doses tested. In the liver, lobular hepatocellular necrosis, centrilobular, macrovesicular steatosis, and centrilobular lymphocytic inflammatory infiltrations were observed for mice group treated with 1 g/kg. For the dose of 2 g/kg, splitting of nuclei and infiltration around the centrilobular veins were observed. This was more pronounced for 5 g/kg in space carries in intralobular and in centralobular cells. In the kidney, the changes were observed for the doses of 2.5 and 5 g/kg. An interstitial inflammation with nodular system composed of lymphocytes and plasmocytes indicating interstitial nephritis.

### Toxicity activity of aqueous extract of *Caralluma europaea* *in vitro*

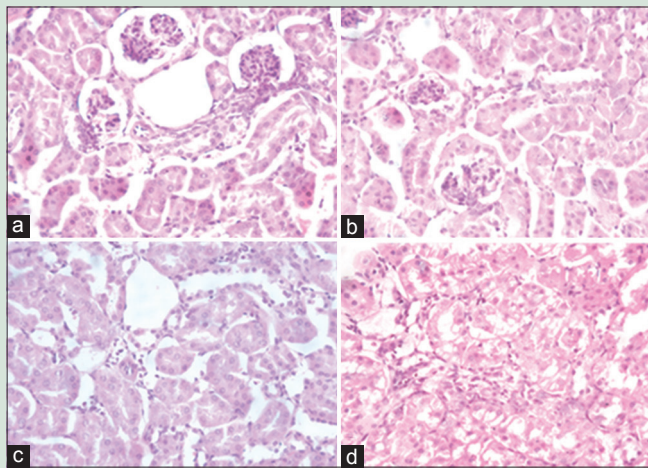
The cytotoxicity activity of the aqueous extract of CE *in vitro* was tested on spleen cell suspension for 24 h with two doses: 0.5 and 2 g/ml and cell viability determined by the Trypan blue exclusion test. The

**Table 1:** Signs observed in chronic toxicity after oral administration of aqueous extract of *Caralluma europaea* in mice

Period of signs observation	Number of mice treated/number of dead (n=5)	Signs of toxicity observed
15 min-1 h	5/0	Asthenia, hypoactivity, urination (+)
1 h-2 h	5/0	Asthenia, hypoactivity, urination (+)
4 h-6 h	5/0	Asthenia, hypoactivity, urination (++)
24 h	5/0	Asthenia, hypoactivity, urination (+++)
48 h	5/0	Asthenia, hypoactivity, urination (+++)
3 days	5/0	Asthenia, hypoactivity, urination (+++)
		Atypical locomotion (back limbs falling) (+)
		Anorexia, asthenia, ataxia and diarrhea (+)
14 days	5/0	Asthenia, hypoactivity, urination (+++)
		Atypical locomotion, anorexia, asthenia, ataxia, and diarrhea (+++)

Period of observation after administration of 5 g/kg of CE: 14 days (observations=15 min, 1, 2, 4, 6 h and in every 24 h to the 14<sup>th</sup> day).

Grade of signs observed: (+) Present or slightly increased; (++) Moderately increased and (+++) Intensively increased



**Figure 1:** Histopathological observations of spleen, liver, and kidney of mice after subchronic treatment with aqueous extract of *Caralluma europaea* (H and E stain,  $\times 400$ ). (a) Control mice. (b) Mice treated with 1 g/kg. (c) Mice treated with 2.5 g/kg. (d) Mice treated with 5 g/kg. The aqueous extract of the plant was given daily by the oral route to groups of mice ( $n = 5$  per group) at the following doses: 0 g/kg, 1.0 g/kg, 2.5 g/kg, and 5 g/kg for 30 days

**Table 2:** Blood chemistry values of mice in sub chronic toxicity study in control and groups treated with different doses of aqueous extract of *Caralluma europaea* on biochemical parameters

Treatments*	AST U/l	ALT U/l	Creatinine mg/L	Urea g/L
Control	111 $\pm$ 0.02	93 $\pm$ 0.01	5.11 $\pm$ 0.04	0.20 $\pm$ 0.03
1.0 g/kg	122 $\pm$ 8.0	93.7 $\pm$ 3.0	4.85 $\pm$ 0.11	0.25 $\pm$ 0.03
2.5 g/kg	130 $\pm$ 7.0	94.7 $\pm$ 9.5	5.25 $\pm$ 0.5	0.30 $\pm$ 0.035
5.0 g/kg	156.7 $\pm$ 4.0	124.3 $\pm$ 6.0	6.10 $\pm$ 0.15	0.35 $\pm$ 0.01

\*The aqueous extract of the plant was given daily by the oral route to groups of mice ( $n=5$  per group) at the following doses: 0 g/kg, 1.0 g/kg, 2.5 g/kg, and 5 g/kg for 30 days. Data are expressed as mean $\pm$ SD ( $n=5$ )  $P < 0.05$  versus the control group. ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; SD: Standard deviation

results [Figure 2] have shown a mortality rate of 80% and 90% of cells after 24 h of incubation, respectively, for 0.5 and 2 g/ml doses compared at 60% for the control. The mortality rate was dose and time dependent. This confirms the toxicity effect *in vivo*.

## Effect of *Caralluma europaea* extract on immune functions in mice

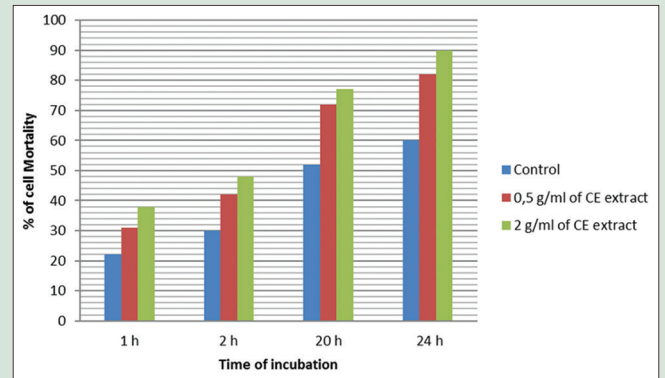
### Effect on humoral immunity

The effect of the aqueous extract treatment on the production of hemagglutination antibodies (HAs) in mice was tested. A significant ( $P < 0.05$ ) increase (16 times) in primary titer values of antibodies (32,768) at limit dose of 1 g/kg was observed as compared to control (2048). The toxic effect of the plant extract on the different tissues has induced lymphocyte B-cell stimulation and antibodies production in high titers compared to controls.

### Effect on the delayed-type hypersensitivity reaction

The plant extract at dose of 1 g/kg elicited a significant increase in DTH response (46%) in comparison to control animals (21%) ( $P < 0.05$ ). This indicates an inflammatory reaction with attraction of inflammatory cells in the site of injection.

The CE extract has shown a significant ( $P < 0.05$ ) immunostimulating effect on both humoral and cellular immune response.



**Figure 2:** Effect of toxicity activity of aqueous extract of *Caralluma europaea* *in vitro* spleen cells from mice were cultured in RPMI supplemented with 10% heat inactivated fetal calf serum, 4 mM glutamine, 100  $\mu$ g/mL gentamicin, and penicillin–streptomycin (200U/mL and 200  $\mu$ g/mL) and 100  $\mu$ l of the plant extract at 0.5 or 2 mg/ml. The cells were maintained in continuous culture in a humid atmosphere at 37°C for 24 h. *In vitro* cytotoxicity assay was carried using Trypan blue (dye exclusion) method for mortality rate determination

## DISCUSSION

There are no publications dealing with CE in its pharmacological activity or toxicity *in vivo*. The aqueous extract of CE was tested *in vivo* in mice and *in vitro* on spleen cell suspensions for its cytotoxic activity. Our results have shown that the dose of 1 g/kg bw was not toxic compared to control. For higher doses, when given orally, the aqueous extract of CE produced toxic effects in mice. The multiple dose study with natural products is necessary to determine the safety of drugs and plant products for human use. The doses selected for chronic and subchronic toxicity studies should be at and above the suggested human dose. In the present study, the doses used were higher than that used in folk medicine in Morocco, which is about 50–100 ml of the liquid preparation containing about 10 g of plant material per liter of water.

In the chronic toxicity study, mice administered an oral dose of 5 g/kg of the CE extract exhibited adverse effects. In the subchronic toxicity study, there was a significant decrease (20%) in bodyweight gain in mice receiving the CE extract orally at doses of 2 and 5 g/kg as compared to control group of mice. A decrease in body weight has been used as an indicator of adverse effects of drugs and chemicals. This is as results of decreased appetite. Other adverse effects have been observed; asthenia, hypoactivity, and urination with the loss of hair.

Among the biochemical parameters evaluated, AST and ALT are considered liver function markers. The increased values of these serum enzymes suggest changes in cell permeability in the hepatocytes, and this has been confirmed by histopathological examinations of the liver that indicate cellular lesions.

Kidney toxicity has also been reported after use of phytotherapeutic products<sup>[29,30]</sup> what makes essential its evaluation. In that case, creatinine and urea determinations are critical as these substances are markers of kidney function. In the present study, significant differences in the parameters were detected between treated mice and controls. This was also confirmed through histopathological examination of the kidney where an interstitial inflammation with nodular system composed of lymphocytes and plasmocytes, indicating interstitial nephritis.

A hyperreactivity in spleen white pulp with lymphoid follicles presenting increased germinal centers was observed for the doses 2 and 5 g/kg. This

change suggests a reaction of the immune system to the effects of the plant extract.

Most common uses of this genus have been recorded as food without any reported adverse effects till date.<sup>[31,32]</sup> *Caralluma* genus seems to be safe for most people when 500 mg of the extract is taken twice a day for up to 60 days; however, the long-term safety is not known. *Caralluma* might cause some mild side effects such as stomach upset, gastric problem, and constipation. These side effects usually go away after a week of use.

Only six *Caralluma* species have reported toxic activity among the 2500 species of the genus.<sup>[33]</sup> Methanolic, ethanolic, and ethyl acetate extracts of *Caralluma tuberculata* show significant toxicity while aqueous extract was found to be nontoxic.<sup>[34]</sup> *Caralluma dalzielii*, *Caralluma retrospiciens*, *Caralluma quadrangula*, *Caralluma negevensis*, and *C. tuberculata* have also shown cytotoxic activity *in vitro*. *Caralluma fimbriata* has shown genotoxicity activity.<sup>[7]</sup> In an animal-conducted trail on Wistar rats, acute toxicity of the ethanolic extract of *C. dalzielii* was found at 2.154 mg/kg orally.<sup>[35]</sup>

We report for the first time the acute and subchronic toxicity of the aqueous extract of the CE *in vivo* at dose up to 1 g/kg bw. This toxicity has been confirmed by the variations of the biochemical parameters and the histopathology changes in the liver, spleen, and kidney and the high rate of cellular mortality *in vitro* compared to the control. The lymphocytic infiltrates were very pronounced in all tissues examined suggesting an immunostimulating effect on both humoral and cellular immune functions in mice. The plant showed a significant enhancement of antibody responsiveness to RRBC in mice as a result of both pre- and post-plant treatment which indicates the enhanced responsiveness of B-lymphocytes involved in antibody synthesis. The mechanism behind this elevated DTH response indicates a stimulatory effect of the plant extract, which has occurred on the lymphocytes and accessory cell types required for the expression of this reaction. Increase in both, antibodies titer, and DTH response indicated that CE extract potentiates humoral as well as the cellular immunity. This plant is a rich source of terpenoids and flavonoids which may act as immunomodulatory, which could justify the high number of lymphocytes infiltrates found in all tissues examined.<sup>[1]</sup>

Since the toxicity studies in experimental animals cannot always be totally extrapolated to humans, and a reasonable estimate of the self-administered dose is difficult to make, and in view of the widespread traditional use of this plant,<sup>[13,14]</sup> recommendations are necessary to protect the population from possible toxic effects of the plant, especially in patients treated for cancer who are already taking cytotoxic treatments.<sup>[36]</sup>

## CONCLUSION

At the dose consumed empirically in traditional Moroccan medicine, CE appears to be relatively toxic. It can cause liver, spleen, and kidney toxicity. Considering these data, we could state that CE stems possess immunomodulatory properties and suggest their involvement of immune responses in the toxic lesions. *Caralluma* species may be tested against different tumor cell lines to explore further its anticancer activity or their inhibiting action on the cancer cell proliferation. Indeed, *Caralluma* species screened for phytochemical constituent seemed to have the potential to act as a source of useful drugs and also to improve the health status.

## Financial support and sponsorship

Hassan II University of Casablanca, Morocco.

## Conflicts of interest

There are no conflicts of interest.

## REFERENCES

1. Meve U, Liede S. Subtribal division of Ceropegieae (*Apocynaceae-Asclepiadoideae*). *Taxon* 2004;53:61-72.
2. Meve U, Heneidak S. A morphological, karyological and chemical study of the *Apteranthes* (*Caralluma*) *Europaea* complex. *Biol J Linn Soc* 2005;149:419-32.
3. Zakaria MN, Islam MW, Radhakrishnan R, Chen HB, Kamil M, Al-Gifri AN, *et al.* Anti-nociceptive and anti-inflammatory properties of *Caralluma arabica*. *J Ethnopharmacol* 2001;76:155-8.
4. Venkatesh S, Reddy GD, Reddy BM, Ramesh M, Rao AV. Antihyperglycemic activity of *Caralluma attenuata*. *Fitoterapia* 2003;74:274-9.
5. Habibuddin M, Daghri HA, Humaira T, Al Qahtani MS, Hefzi AA. Antidiabetic effect of alcoholic extract of *Caralluma sinaica* L. on streptozotocin-induced diabetic rabbits. *J Ethnopharmacol* 2008;117:215-20.
6. Abdel-Sattar E, Harraz FM, Al-ansari SM, El-Mekawy S, Ichino C, Kiyohara H, *et al.* Acylated pregnane glycosides from *Caralluma tuberculata* and their antiparasitic activity. *Phytochemistry* 2008;69:2180-6.
7. Kamalakkannan S, Rajendran R, Venkatesh RV, Clayton P, Akbarsha MA. Antitumorogenic and antiatherosclerotic properties of *Caralluma fimbriata* extract. *J Nutr Metab* 2010;2010:285301.
8. Adnan M, Jan S, Mussarat S, Tariq A, Begum S, Afroz A, *et al.* A review on ethnobotany, phytochemistry and pharmacology of plant genus *Caralluma* R. Br. *J Pharm Pharmacol* 2014;66:1351-68.
9. Gujjala S, Putakala M, Gangarapu V, Nukala S, Bellamkonda R, Ramaswamy R, *et al.* Protective effect of *Caralluma fimbriata* against high-fat diet induced testicular oxidative stress in rats. *Biomed Pharmacother* 2016;83:167-76.
10. Khan MZ, Atlas N, Nawaz W. Neuroprotective effects of *Caralluma tuberculata* on ameliorating cognitive impairment in a d-galactose-induced mouse model. *Biomed Pharmacother* 2016;84:387-94.
11. Garg S, Srivastava S, Singh K, Sharma A, Garg K. Ulcer healing potential of ethanolic extract of *Caralluma attenuata* on experimental diabetic rats. *Anc Sci Life* 2016;35:222-6.
12. Wen S, Chen Y, Lu Y, Wang Y, Ding L, Jiang M, *et al.* Cardenolides from the apocynaceae family and their anticancer activity. *Fitoterapia* 2016;112:74-84.
13. Bellakhdar J, Claisse R, Fleurentin J, Younos C. Repertory of standard herbal drugs in the Moroccan pharmacopoea. *J Ethnopharmacol* 1991;35:123-43.
14. Bellakhdar J. The Traditional Moroccan Pharmacopoeia. Paris: Ibis Press; 1997. p. 764.
15. Formisano C, Senatore F, Della Porta G, Scognamiglio M, Bruno M, Maggio A, *et al.* Headspace volatile composition of the flowers of *Caralluma europaea* N.E.Br. (*Apocynaceae*). *Molecules* 2009;14:4597-613.
16. Zito P, Sajeve M, Bruno M, Maggio A, Rosselli S, Formisano C, *et al.* Essential oil composition of stems and fruits of *Caralluma europaea* N.E.Br. (*Apocynaceae*). *Molecules* 2010;15:627-38.
17. Bodoprost J, Rosemeyer H. Analysis of phenacyl ester derivatives of fatty acids from human skin surface sebum by reversed-phase HPLC: Chromatographic mobility as a function of physico chemical properties. *Int J Mol Sci* 2007;8:1111-24.
18. Walters D, Raynor L, Mitchell A, Walker R, Walker K. Antifungal activities of four fatty acids against plant pathogenic fungi. *Mycopathologia* 2004;157:87-90.
19. Organization of Economic Co-Operation and Development (OECD). Guideline for Testing of Chemicals, TG420 (OECD, The OECD Guideline for Testing of Chemical: 407 Repeated Dose Oral Toxicity Rodent: 28-day or 14-Day Study. Paris, France: OECD; 2001.
20. Ha H, Lee JK, Lee HY, Seo CS, Kim JH, Lee MY, *et al.* Evaluation of safety of the herbal formula Ojeok-san: Acute and sub-chronic toxicity studies in rats. *J Ethnopharmacol* 2010;131:410-6.
21. United States Environmental Protection Agency; 2002. Health Effects Test Guidelines: Acute oral toxicity OPPTS 870.1100; 2002.
22. United States Environmental Protection Agency; 2000. Health Effects Test Guidelines: Repeated dose 28 day oral toxicity study in rodents. OPPTS 870.3050; 2000.
23. Forabosco A, Zaffe D, Tosato L. On the dye exclusion of test cell vitality. I. Evaluation of the optimal concentration. *Boll Soc Ital Biol Sper* 1972;48:33-6.
24. Bin-Hafeez B, Ahmad I, Haque R, Raisuddin S. Protective effect of *Cassia occidentalis* L. On cyclophosphamide-induced suppression of humoral immunity in mice. *J Ethnopharmacol* 2001;75:13-8.
25. Sallander S, Shanwell A, Aqvist M. Evaluation of a solid-phase test for erythrocyte antibody screening of pregnant women, patients and blood donors. *Vox Sang* 1996;71:221-5.
26. Benencia F, Courrèges MC, Coulombié FC. *In vivo* and *in vitro* immunomodulatory

- activities of *Trichilia glabra* aqueous leaf extracts. J Ethnopharmacol 2000;69:199-205.
27. Stevens KR, Mylecraine L. Issues in chronic toxicology. In: Hayes AW, editor. Principles and Methods of Toxicology. 3<sup>rd</sup> ed. New York: Raven Press; 1994. p. 673.
  28. Eaton DL, Klaassen CD. Principles of toxicology. In: Klaassen CD, editor. Casarett and Doull's Toxicology: The Basic Science of Poisons. 5<sup>th</sup> ed. New York: McGraw-Hill; 1996. p. 13.
  29. Corns CM. Herbal remedies and clinical biochemistry. Ann Clin Biochem 2003;40(Pt 5):489-507.
  30. Isnard Bagnis C, Deray G, Baumelou A, Le Quintrec M, Vanherweghem JL. Herbs and the kidney. Am J Kidney Dis 2004;44:1-11.
  31. Naik RM, Venugopalan V, Kumaravelayutham P, Krishnamurthy YL. Nutritive value and mineral composition of edible *Caralluma* and *Boucerosia* species from the arid areas of Karnataka. Int J Agric Environ Biotechnol 2012;5:117-25.
  32. Gilbert MG. A review of *Caralluma* R. Br. and its segregates. Bradleya 1990;8:1-32.
  33. Khan MZ, Khan RA, Ahmed M, Muhammad N, Khan MR, Khan HU, *et al.* Biological screening of methanolic crude extracts of *Caralluma tuberculata*. Int J Indig Med Plants 2013;46:2051-4263.
  34. Rizwani GH. Phytochemical and Biological Studies on Medicinal Herbs, *C. Tuberculata* and *C. Edulis*. A Thesis Submitted to the University of Karachi for the Degree of Doctor of Philosophy, Department of Pharmacognosy, Faculty of Pharmacy, and University of Karachi; 1991.
  35. Tanko Y, Sada NH, Mohammed K, Jimoh A, Yerima M. Effect of ethanolic extract of *Caralluma diazielli* on serum lipid profiles on fructose induced diabetes in Wistar rats. Ann Biol Res 2013;4:157-61.
  36. Chebat A, Skalli S, Errihani H, Boulaâmane L, Mokrim M, Mahfoud T, *et al.* Prevalence study of undesirable effects related to the use of medicinal plants by patients of National Institute of Oncology, Rabat 2014;12:25-32.