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LETTER TO THE EDITOR

Aged T cells and cardiovascular disease

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emory T cells undergo replicative senescence via repetitive proliferation. As in other cells, replicative senescence of T cells is characterized by shortened telomeres.1 Immunophenotypes are also altered in replicative senescent T cells compared to young T cells. In humans, replicative senescent T cells typically lose expression of CD28, a costimulatory receptor, and acquire the expression of CD57 antigen, a terminally sulfated glycan carbohydrateepitope,1 though the mechanism underlying this regulation has not yet been clearly elucidated. Therefore, CD28⁻ or CD57⁺ T cells are considered replicative senescent T cells in humans, and the frequency of CD28⁻ or CD57⁺ T cells in the peripheral blood increases with aging.^{2,3}

Replicative senescent CD28⁻ or CD57⁺ T cells gain the expression of NK cell-related receptors, such as KLRG-1 and CD85j, and chemokine receptors for homing to peripheral inflammatory sites, such as CX3CR1 and CCR5 (Figure 1).^{1,4} Moreover, CD28⁻ or CD57⁺ T cells express higher levels of cytotoxic proteins, such as perforin and granzyme B, and produce

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higher amounts of IFN-y and tumor necrosis factor upon T cell receptor (TCR) stimulation compared to nonsenescent CD28+ or CD57- T cells (Figure 1).⁵ In terms of cell proliferation, CD28⁻ or CD57⁺ T cells are less responsive to TCR or IL-7 stimulation than their CD28+ or CD57- counterparts (Figure 1).^{1,4} In particular, CD28⁻ or CD57⁺ T cells express reduced levels of IL-7 receptor α -chain (CD127). However, IL-15-induced proliferation is intact in CD28⁻ or CD57⁺ T cells. Furthermore, senescent T cells are different from non-senescent T cells in regards to intracellular signaling. In senescent human T cells, the mitogen-activated protein kinase (MAPK) p38 pathway is activated by intracellular changes, leading to inhibition of autophagy, proliferation and telomerase activity (Figure 1).6-8 An immune-inhibitory role of the sestrin-MAPK activation complex in senescent T cells was also reported recently.9

In principle, replicative senescence of T cells can be induced by any antigen or microbial pathogen if it is encountered repetitively. Among various human pathogens, cytomegalovirus (CMV) is the major virus responsible for the accumulation of replicative senescent CD28⁻ or CD57⁺ T cells in humans. ^{10,11} CMV is a human herpesvirus that latently infects a majority of the population. CMV usually infects individuals asymptomatically during early childhood and establishes a lifelong latent infection, ^{10,11} but it is occasionally reactivated even in non-immunocompromised healthy hosts depending on the immune

status of the host over their lifetime. Repetitive cycles of CMV reactivation and resolution result in repetitive antigenic stimulation of CMV-specific memory T cells and consequent accumulation of replicative senescent CMV-specific T cells over a person's lifetime. 10–12 A sustained increase and maintenance of CMV-specific memory T cells over a lifetime is known as 'memory inflation'. 10,12

Possible roles of replicative senescent T cells in human cardiovascular disease were first reported in patients with unstable angina.¹³ The number of CD4 +CD28⁻ T cells in the peripheral blood was significantly increased in patients with unstable angina compared to patients with chronic stable angina, 13 and a higher frequency of CD4+CD28-T cells was significantly associated with the recurrence of acute coronary events in patients with unstable angina.14 In addition, accelerated telomere shortening was demonstrated in T cells from patients with atherosclerosis. 15 Senescent T cells were also reported to be important in human hypertension. The relative frequency of CD57+ or CD28- cells in the peripheral CD8+ T-cell population was increased in patients with essential hypertension compared to age- and sexmatched control subjects.¹⁶ Interestingly, increased expression of CXCL11 and T-cell infiltration were observed in renal tissues from patients with hypertension. Senescent CD8+ T cells are also associated with the prognosis of acute myocardial infarction (MI). The relative frequency of CD57+ cells in the

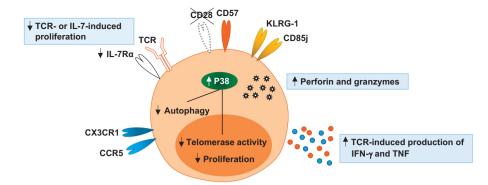


Figure 1 Characteristics of replicative senescent T cells. Replicative senescent T cells lose the expression of CD28 and acquire the expression of CD57, NK cell-related receptors, and chemokine receptors for peripheral homing. They have high levels of perforin and granzymes, and produce high amounts of IFN-γ and TNF upon TCR stimulation. However, these cells are less proliferative to TCR or IL-7 stimulation compared to young T cells. They exhibit constitutive p38 mitogen-activated protein kinase (MAPK) activation, leading to inhibition of autophagy, proliferation, and telomerase activity. TCR; T cell receptor, TNF; tumor necrosis factor.

peripheral CD8⁺ T-cell population has been shown to significantly correlate with cardiovascular mortality 6 months after acute MI.¹⁷

Recently, an association between replicative senescence of T cells and arterial stiffness has been demonstrated in healthy individuals.¹⁸ Arterial stiffness is increased in the presence of cardiovascular risk factors, including aging, and known to be associated with inflammation. In this study, the relative frequency of CD57+cells in the peripheral CD8+ T-cell population independently correlated with arterial stiffness after adjusting for traditional cardiovascular risk factors.¹⁸ More importantly, the relative frequency of CMV pp65-specific CD8+ T cells in the peripheral CD8⁺ T-cell population, which was evaluated by the production of IFN-v or tumor necrosis factor and CD107a exposure upon CMV pp65 stimulation, independently correlated with arterial stiffness. 18 This study first demonstrated that arterial stiffness, representing vascular aging, is associated with senescent CD8+CD57+ T cells and CMV pp65-specific CD8⁺T cells.

As described above, there is a growing body of evidence indicating that replicative senescent CD28⁻ or CD57⁺ T cells are involved in the development and progression of cardiovascular diseases, such as atherosclerosis, acute MI and hypertension, in humans. However, how replicative senescent T cells are regulated and contribute to the pathogenesis of cardiovascular diseases remains to be elucidated. For example, it is not yet clear whether senescent CD8⁺ and CD4⁺ T cells

exert distinct functions in cardiovascular disease. In addition, it is important to determine how T-cell effector functions are altered during replicative senescence, which effector function of senescent T cells has a major role in the pathogenesis of cardiovascular diseases, how their effector functions are stimulated in the presence or absence of cognate antigens, and which tissue is the primary site for senescent T-cell infiltration and action. By answering these questions, we will be able to develop better strategies for the prevention and treatment of cardiovascular diseases.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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