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Memory-less response and OPENviolation of the fuctuationdissipation theorem in colloids suspended in an active bath

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We investigate experimentally and numerically the stochastic dynamics and the time-dependent response of colloids subject to a small external perturbation in a dense bath of motile *E***.** *coli* **bacteria. The external feld is a magnetic feld acting on a superparamagnetic microbead suspended in an active medium. The measured linear response reveals an instantaneous friction kernel despite the complexity of the bacterial bath. By comparing the mean squared displacement and the response function we detect a clear violation of the fuctuation dissipation theorem.**

The dynamics and statistical mechanics of self-propelled particles is attracting a considerable attention both from the fundamental point of view and for its potential applications^{1-[4](#page-5-1)}. These active particles are thermodynamically out of equilibrium as they constantly consume fuel dissipating energy in the surrounding fuid. From a dynamical point of view, however, the trajectories of non-interacting active particles show no sign of non-equilibrium as time-reversal symmetry is preserved and no entropy is produced⁵. Differently, strong deviations from equilibrium can be observed when particle trajectories result from the combination of self-propulsion and interaction forces due to other particles or external felds. Under the assumption that the external forces do not alter the "internal" propulsion mechanism, schematic models of active particles simply assume that the particle veloc-ity is instantaneously determined by the superposition of the external forces and the "propulsion force"^{4[,6](#page-5-3)}. In this framework the reaction of the active particle's velocity to the external feld is instantaneous as dictated by low-Reynolds number hydrodynamics of Newtonian fuids. Diferently the random propulsion force relaxes on fnite time-scale which depends on the specifc propulsion mechanism, for example the propulsion velocity in swimming wild-type *E*. *coli* changes abruptly during "tumbles["7](#page-5-4) , while in chemically propelled Janus particles the swimming direction changes gradually because of rotational diffusion⁸. This combination of an instantaneous response and "colored" noise leads, by construction, to a non-equilibrium dynamics which violates the second Kubo fluctuation dissipation theorem^{[9](#page-5-6)} (FDT) already at the level of individual active particles. This can be seen as the origin of many non-equilibrium phenomena observed in active particles such as the strong deviations from the Boltzmann distribution^{[4,](#page-5-1)[6](#page-5-3)[,10](#page-5-7),[11](#page-5-8)}, the emergence of novel non-equilibrium phase transitions¹², and the striking rectifcation efects induced by asymmetric boundaries and objects. For example it has been shown that swimming bacteria and Janus particles can autonomously assemble with asymmetric microstructures and form self-propelled micromachines $^{13-17}$.

Interestingly also passive colloids, interacting with active particles, inherit the non-equilibrium properties of the active bath^{18–24} and show peculiar off-equilibrium phenomena. In fact it has been shown that these "activated" colloids" violate the equilibrium equipartition theorem²⁰ and are subject to an effective attraction even in pres-ence of purely repulsive forces, deviating strongly from the Boltzmann law^{[21](#page-5-15)}. Moreover an active bath can induce a directed transport of colloids over micro-fabricated asymmetric barriers produced by laser litography^{22,23}.

All this fundamental and applied research motivates the efort in modeling accurately the dynamics of pas-sive colloids suspended in active baths^{14,[18](#page-5-12)[,20](#page-5-14)[,25](#page-6-0)[,26](#page-6-1)}. The dynamics of these tracers is determined by the dynamics of the active particles and their interactions (steric, hydrodynamic, etc.) with the passive colloids. Adopting a

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Figure 1. Scheme of the experimental set-up (**a**) Side view, a current-carrying copper wire (gray) is placed parallel to a glass tube containing superparamagnetic beads and bacteria. (**b**) Top view, the arrow indicates the current fow. (**c**) 3-dimensional view, the paramagnetic bead can be pulled and released by the magnetic feld produced by the current.

simplifying approach *á la Langevin* we focus only on the degrees of freedom of the colloid and treat the active bath as a source of "noise". In addition to this the colloid is also subject to the interactions with the surrounding fuid that introduces thermal fuctuations. We thus can model the dynamics of the colloid in the active bath with the generalized Langevin equation (GLE):

$$
\int_{-\infty}^{t} dt' \Gamma(t - t') \dot{\mathbf{r}}(t') = \mathbf{f} + \boldsymbol{\eta} + \boldsymbol{\xi}
$$
\n(1)

where Γ is the friction memory kernel, $\mathbf{r}(t) = (x(t), y(t), z(t))$ is the position of the colloidal particle in 3-dimensions, *ξ* is the "active noise" term, *η* is the standard Langevin thermal noise and **f** is the external force. Since the colloid is coupled to the non-equilibrium active bath we expect that the FDT is violated resulting in a Γ(*t*−*t*′) having characteristic time-scale which is diferent from the one of the correlation of the noise: 〈*ξ*(*t*′)*ξ*(*t*)〉. Previous experimental and numerical result²⁰ suggest that ξ can be modeled by an exponentially correlated (colored) noise: $\langle \xi_a(t)\xi_\beta(t')\rangle = \gamma^2 D_A \delta_{\alpha\beta}e^{-|t-t'|/\tau}/\tau$ where α and β represent the individual Cartesian components, γ is the drag coefficient of the colloid, D_A is the "active" diffusivity, and τ is the relaxation time of the noise. Differently the thermal noise is assumed to be delta-correlated with $\langle \eta_\alpha(t) \eta_\beta(t') \rangle = 2\gamma^2 D_T \delta_{\alpha\beta} \delta(t-t')$ where D_T is thermal diffusion coefficient: $D_T = k_B T / \gamma$ (*T* being the temperature).

It is clear that to fully characterize the dynamics described by the GLE (1) it is necessary to design experiments aimed at measuring directly the friction kernel Γ. By combining optical tweezers-based passive and active microrheology it was shown that a bacterial bath of swimming *E*. *coli* displays a viscosity without any signifcant frequency dependence¹⁹. This indicates that the friction kernel is instantaneous: $\Gamma = \gamma \delta_{\alpha\beta} \delta(t - t')$ so that Eq. [\(1\)](#page-1-0) reduces to:

$$
\gamma \dot{\mathbf{r}} = \xi + \eta + \mathbf{f} \tag{2}
$$

However this study was limited to very low bacterial densities (about 3×10^{-3} volume fraction) and it is not clear to what extent the optical traps can afect the dynamics of swimming bacteria.

In this work we experimentally measure the time-dependent displacement of a superparamegnetic bead in a dense bacterial bath (10¹⁰ cells/ml corresponding to a volume fraction of about 2×10^{-2}). The external force is applied by using a controlled magnetic field which acts uniquely on the colloidal particle. The linear response reveals an instantaneous friction kernel validating the model of Eq. ([2](#page-1-1)). Diferently the mean squared displacement of the particle shows the typical combination of ballistic and diffusive behavior. This results in clear violation of the FDT which has been investigated intensively in active matter both theoretically $5,27-30$ $5,27-30$ $5,27-30$ and numerically $31,32$ $31,32$. We also show that the same qualitative results are obtained by numerical simulations of "run and tumble" bacteria interacting with a passive colloid. Finally our numerical results suggest that even by further increasing the bacterial density the friction memory kernel remains instantaneous while the efective drag *γ* increases substantially.

Results

Experiment. We prepare motile *E*. *coli* cells following the protocol described in ref.[20](#page-5-14) (see also Methods). We use superparamagnetic microbeads of radius $a = 3.4 \mu$ m (COMPEL, Bangs). These are first diluted in deionized water and then mixed with bacteria directly on a glass slide. The final bacteria density is estimated to be $\sim 10^{10}$ cells/ml. Te bacteria-colloids solution is loaded in a microcapillary glass tube (Vitrocom) of internal radius $R = 25 \mu m$ by capillarity. The sample is left open for few minutes and then sealed with index matching oil. The flled capillary tube is then placed parallel to a copper wire (radius 200*μ*m) at an approximate distance of 100*μ*m (see Fig. [1](#page-1-2)). The copper wire is connected to a computer-controlled current generator. This set-up allows the combined measurement of the response of the microbeads subjected to a controlled magnetic feld and the measurement of its spontaneous active dynamics in absence of external perturbations.

Fluctuations. When the current is zero no external feld acts on the super paramagnetic particles. In this situation the colloids sediment at the bottom of the capillary and fuctuate because of the collisions with swimming bacteria and because of thermal agitation. We characterize the active dynamics of one single colloid one at a time in absence of external filed by collecting bright field images using a $20\times$ microscope objective (NA=0.25). We

Figure 3. (Lef) Same experimental data as in Fig. [2\(a\)](#page-2-0) shown in double log-scale. (Right) Same simulation data as in Fig. $2(c)$ shown in double log-scale.

obtain particle trajectories by center of mass tracking directly while streaming images at a rate of 100 fps. To characterize the dynamics of the microbead in absence of external perturbation we measure the mean squared displacement (MSD) along the *y* axis $\langle \Delta y^2(t) \rangle$ where no force is present (differently along *x* the curvature of the capillary and gravity result in an elastic force as discussed below). These measurements are obtained by averaging over about 600 trajectories (lasting for 1 s) of the same bead. The MSD for one single bead is reported in Fig. [2\(a\)](#page-2-0) (full symbols, see also in Fig. [3\)](#page-2-1) and show the typical transition from a ballistic behaviour at short times followed by a diffusive behaviour at longer timescales (see Fig. [3](#page-2-1)) These data can be fitted very well by the theoretical MSD obtained from Eq. [\(2\)](#page-1-1):

$$
\langle \Delta y^2(t) \rangle = 2D_T t + 2D_A[t - \tau(1 - e^{-t/\tau})]
$$
\n(3)

as shown by the full line in Fig. $2(a)$. Fitting by Eq. ([3](#page-2-2)) allows also to extract the parameters of interest, i.e. the thermal diffusivity $D_T = (2.43 \pm 0.02) \times 10^{-2} \mu m^2/s$, the active diffusivity $D_A = (3.55 \pm 0.03) \times 10^{-2} \mu m^2/s$ and the relaxation time τ = 0.22 \pm 0.03 s. The thermal diffusion constant can be used to extract the mobility of the particle as $\mu = D/(k_B T) = 5.92 \pm 0.06 \ \mu m/(s \text{ pN})$ which is the inverse of the drag coefficient $\mu = \gamma^{-1}$. The measured μ is about a factor two smaller than the bulk mobility value and this is consistent with an increase of the drag caused by the presence of the capillary wall^{20,33}.

It is also interesting to consider the statistics of displacements in absence of external forces. According to Eq. [\(2\)](#page-1-1), (see also ref.[34\)](#page-6-7) the displacement of the particle results from the combination of the active noise *ξ* and the thermal noise *η*. Since *η* is Gaussian distributed any deviation from the Gaussian in the displacement distribution has to be attributed to *ξ*. In Fig. [4\(a\)](#page-3-0) we show the probability distribution of displacements along the *y*-axis at diferent time lags *t* indicated by $P_t(\Delta y)$. It is evident that as *t* grows the deviation from the Gaussian fit (dashed lines in Fig. $4(a)$) become more evident showing that $P_t(\Delta y)$ develops "fat tails". To quantify this effect we compute the non-Gaussian parameter as³⁵: $\alpha_2(t) = \frac{\Delta y^4(t)}{5\Delta y^2(t)} - 3/5$. This is shown in Fig. [4\(b\)](#page-3-0) and it is found to increase and then decrease as *t* grows. The parameter α_2 reaches a maximum at the characteristic relaxation time of the active force $t \approx \tau$ indicating that ξ is the responsible for the non-Gaussian behavior of $P_t(\Delta y)$. Our results are in qualitative agreement with the results of ref.³⁶ where robust exponential tails where found in the distribution of displacements of tracer particles suspended in an active bath of swimming algae.

Finally, for the the discussion that follows, it is also important to look at the (static) position distribution along *x*. Being in a cylindrical capillary the bead is subjected to a nearly elastic force *fe* directed along *x* due to the com-bination of the confinement and gravity as discussed in ref.^{[20](#page-5-14)}. This force is given by by $f_e = -kx$ where the elastic constant *k*=*mg*/(*R*−*a*) is determined by the buoyant mass of the particle *m* and the acceleration due to gravity *g*.

Figure 4. (Experimental data) (**a**) Probability distribution of the displacements along *y* (full lines) at diferent times (see legend). The dashed lines are fits with Gaussian functions. (**b**) Non-Gaussian parameter as a function of the time lag *t* reaching a maximum at *t*≈*τ*. (**c**) Probability distribution of the bead's position along *x* (shaded area). The full line is a fit with a Gaussian.

Figure 5. (Experimental data) (**a**) Trajectories of the same particle obtained by repeatedly switching on the feld at time *t*=0 s (all trajectories are rescaled so that the position of the particle coincides with the origin at $t=0$). An evident drift of the particles along the magnetic field gradient (along *x*) is observed. (**b**) Probability distribution of the final displacement $(t=1 s)$ after switching the field on at $t=0 s$. The probability peaks at Δx > 0 while the peak is symmetric with respect to *y* = 0.

The effect of this force is to confine the particle's motion along x. This is seen in the probability distribution of the colloid's *x*-coordinate $P(x)$ reported in Fig. [4\(c\)](#page-3-0). This figure shows that $P(x)$ is nearly Gaussian-distributed (as also found in ref.^{[20](#page-5-14)}) with a variance $\langle x^2 \rangle = 1.01 \pm 0.06 \,\mu m^2$. Following the theory of ref.²⁰ we can compute the variance from the formula $\langle x^2 \rangle = (D_T + D_A)/(\mu k) = 0.9 \pm 0.2 \mu m^2$ that is compatible with the measured value (the large uncertainty is due to the uncertainty on the particle's density). The value of *k* could be tuned by changing the particle size and/or density or by using capillary tubes with smaller radii.

Response. When the electric current flows a magnetic filed is generated and this allows to measure the time-dependent response of the superparamagnetic particle. The magnetic field acts on the particle inducing a magnetic dipole moment and that interacts with the magnetic feld itself. In this way the particle is attracted toward regions where the magnetic field is more intense. The force on the superparamgnetic bead subject to the magnetic field generated by a current-carrying wire takes the form^{[37,](#page-6-10)38}:

$$
f_m = (\mu_0 a^3 \chi) \frac{I^2}{3\pi r^3}
$$
 (4)

where *I* is the current, μ_0 is the magnetic permeability of vacuum, χ is the (dimensionless) magnetic susceptibility of the bead and *r* is the distance from the center of the wire. Tis force acts on the *x*-axis and pulls the particle toward the wire. By knowing the susceptibility³⁷ χ = 0.170 \pm 0.07 and the current *I* = 0.5 A we can estimate the force acting on a bead placed in the center of the capillary tube $f_m = 0.029 \pm 0.01$ pN. For small displacements≤0.5*μ*m near the center of the capillary, as those observed in this experiment, this force is *fe*≈4×10[−]3pN and therefore can be neglected respect to the magnetic force *fm*. However afer having induced a displacement we switch off the magnetic field and this elastic force brings back the particle to the initial position making the capillary a convenient geometry for taking repeated measurement on the same bead.

To perform the measurements the current is switched on at time *t*=0 and kept constant for 1 s during which we measure the displacement of the particle along x. After 1 s the current is switched off and the particle is released. This procedure is repeated 600 times and the resulting displacements are averaged to obtain the average displacement 〈Δ*x*(*t*)〉 induced by the external force. Some of the trajectories obtained by the tracking of one single bead are displayed in Fig. $5(a)$ which shows how the external field induces a net drift of the particle from their

Figure 6. (Simulation data) (**a**) Symbols represent the MSD of a particle in a bacterial bath at three diferent densities (0.012, 0.038 and 0.062 μ m⁻² from bottom to top), the lines are fits with Eq. [\(3\)](#page-2-2). (**b**) Time-dependent displacement (symbols) of the particle induced by same the external feld (turned on at time *t*=0s) in numerical simulation at three different densities (same as in (a)). The full lines are a linear fits passing through zero. (c) Resulting mobility (circles) from the drif velocity found in (**a**) compared with the mobility from the MSD data (squares) shown in (b) (error bars not show). Te mobility smoothly decreases from the bare mobility (dashed line) upon increasing density as suggested by the ftting (full line).

initial position $(x=0)$ towards the right $(x>0)$. This is even more clear when we compute the probability of the displacement $P(\Delta x, \Delta y)$ after a time $t=1$ s (see Fig. [5\(b\)\)](#page-3-1) which is clearly peaked around positive Δx while being fairly symmetric with respect to the *y* axis.

The results obtained by averaging these measurements are shown in Fig. $2(a)$ (open symbols) for the same bead whose MSD is shown in the Figure. The measured $\langle \Delta x(t) \rangle$ can be fitted very well with a straight line $\langle \Delta x(t) \rangle = vt$ (constant speed drift) with $v = 0.201 \pm 0.005 \mu$ m/s. This confirms that the memory friction kernel of colloids in active baths is indeed instantaneous. Note that to plot 〈Δ*x*(*t*)〉 together with the MSD in Fig. [2\(a\)](#page-2-0) we have normalized it by the fitted speed *v* defining $\langle \Delta \tilde{x}(t) \rangle = \langle \Delta x(t) \rangle / v$ and we have multiplied this by the thermal diffusivity D_T . In this way the MSD and the displacement follow each other at small-times and separate over longer timescales. Moreover having estimated the force we can extract the mobility as $\mu = v/f_m = 6.8 \pm 0.5 \mu$ m/(s pN) which is in fairly good agreement with the measurement of *μ* obtained from the MSD given above.

Having measured the MSD and the displacement following the perturbation we can detect a clear diference between the noise kernel and the friction kernel. To show this we build the fluctuation-dissipation plot (FD-plot)³⁹ in which we report the normalized response $2D_r\langle\Delta \tilde{x}(t)\rangle$ versus the MSD where time is a parameter as shown in Fig. [2\(b\)](#page-2-0). If the system were in equilibrium the data would follow the dashed line in Fig., i.e. $2D_T\langle\Delta\tilde{x}(t)\rangle = \langle\Delta x^2(t)\rangle$. Differently the non-equilibrium dynamics generated by the active bath leads to a quasi-equilibrium regime at timescales *t*≤0.1 shorter than *τ*, where the FDT is valid, followed by a clear violation of the theorem at longer timescales where $2(D_T + D_A)\langle\Delta\tilde{x}(t)\rangle = \langle\Delta x^2(t)\rangle$ characterized by an enhanced diffusivity. This can be translated in an effective temperature that, over long timescales, is higher than the bath temperature: $k_B T_{\text{eff}} = (D_T + D_A)/\mu = (2.5 \pm 0.1) \times k_B T$ for the data in Fig. [2\(b\)](#page-2-0).

Simulations. The numerical simulations are performed by considering one single spherical colloidal particle of radius *a* immersed in a bath of bacteria modeled as self-propelling dumbbells following a "run and tumble" dynamics. Both particles and bacteria move in a 2-dimensional box with periodic boundary conditions. All interactions between bacteria and between bacteria and particles are modeled by steric repulsive forces. In addition we apply a constant force to the colloidal particle when we want to measure the response function. We include Brownian motion only for particles dynamics and neglect hydrodynamic interactions^{[14](#page-5-18),[21](#page-5-15)}. A detailed description of the simulation can be found in refs^{[13,](#page-5-10)[21](#page-5-15)}. We first focus on the case where the bacterial density is low $(\rho = 1.2 \times 10^{-2} \,\mu m^{-2})$ which yelds results more similar to experimental case.

The axial MSD of the particle is shown in Fig. $2(c)$ (full symbols) and is qualitatively very similar to the one found in experiments. These numerical data can also be fitted very well with Eq. [3](#page-2-2) as shown by the full line in Fig. [2\(c\)](#page-2-0) and in Fig. [3.](#page-2-1) The resulting fitting parameters are $D_A = 0.260 \pm 0.004 \mu m^2/s$, $\tau = 0.071 \pm 0.003$ s and $D_T\!=\!0.056\pm0.002$ μ m²/s. We note that the fitting parameters from the simulation data are of the same order of magnitude of the experimental ones although the active diffusivity D_A is considerably higher in simulations than in experiments as found also in ref.^{[20](#page-5-14)} Also for simulations we extract the mobility as $\mu = D_T/k_BT = 13.8 \pm 0.4 \,\mu$ m/ (s pN). As expected at such low density this is very close to the bare mobility used in the simulation $\mu = 13.6 \,\mu$ m/ (s pN). For measuring the externally induced displacement we apply a constant force *f*=0.2 pN directed along the *x*-axis at $t = 0$. This is kept constant for 1 s, during which we measure the time dependent displacement, and the procedure is repeated for 10 times. The normalized displacement $\langle \Delta \tilde{x}(t) \rangle$ is reported in Fig. [2\(c\)](#page-2-0) (open symbols) and reveals again a constant speed drift corresponding to an instantaneous friction kernel. By fitting the displacement with a constant-speed motion $\langle \Delta x(t) \rangle = vt$ we get $v = 2.68 \pm 0.05 \,\mu \text{m/s}$ from which we obtain the mobility $\mu = v/f = 13.7 \pm 0.3 \ \mu m/(s \text{ pN})$. From the combination of the numerical MSD and displacement we can build the FD-plot shown in Fig. $2(d)$. This shows a form that is qualitatively similar to the one found in experiments with the FDT being violated at long timescales. However D_A is higher in simulations than in experiments and consequently we get a higher effective temperature $k_B T_{\text{eff}} = (D_T + D_A)/\mu = (5.6 \pm 0.2) \times k_B T$.

Upon increasing the density of the active bacteria we observe that the MSD increases considerably in ampli-tude (see Fig. [6\(a\)](#page-4-0)). These data can be very well fitted with Eq. ([3](#page-2-2)) allowing to estimate the parameters D_A , D_T

and τ . We find that τ and D_A both increase respectively from 0.10 to 0.18 s and from 0.26 to 3.3 μ m²/s upon increasing *ρ* from 0.012 to 0.062 μ m⁻². Differently D_T systematically decreases from 5.6 \times 10⁻² to 4.5 \times 10⁻² μ m²/s in the same density range. This indicates that the effective mobility $\mu = D_T/k_B T$ of the particle in the bacterial bath,probed by the thermal fluctuations, decreases upon increasing the density. The response measured upon changing density remains instantaneous resulting in a linear drif of the particle following the activation of the external field (see Fig. $6(b)$). While this generalizes the validity of Eq. ([2\)](#page-1-1) to the moderately high-density regime we also observe that the efective mobility of the particle found in simulations decreases systematically confrming the trend observed in the thermal difusivity. By ftting the data in Fig. [6\(b\)](#page-4-0) with straight lines we obtain the drift speed of the particles from which we can estimate again the mobility. This compares very well with the one extracted from D_T as shown in Fig. [6\(c\)](#page-4-0). These data can be fitted with a simple polynomial equation $\mu = \mu_0$ / $(1+\alpha\rho+\beta\rho^2)$ with μ_0 being the bare mobility of the particle and with $\alpha>0$, $\beta>0$. This suggests that, at very low densities, the mobility approaches the bare mobility of the particle set by the fuid and that at high *ρ* the packing of the bacteria becomes important lowering significantly the mobility μ .

Discussion

We have studied numerically and experimentally the time-dependent response and fuctuations of a particle immersed in a bacterial bath. We have shown that this response is instantaneous which, in combination with the persistent behaviour of fuctuations, leads to a dramatic breakdown of the fuctuation-dissipation theorem. Our results further confrm the validity of the model that is currently used to describe the efect of an active bath on a passive particle: the (thermal) Langevin dynamics is simply modifed by adding active (persistent) forces without changing the response function. Moreover our results show some intriguing analogy with recent simulations on active red-blood cells membrane in which it has been shown that the response function of the membrane in presence of active fuctuations is the same that in complete absence of activity while the fuctuations change substantially in the two cases⁴⁰. This is consistent with our results in which, by adding the activity of the bacterial bath, the response of the probe particle does not change. Further experimental, theoretical and numerical studies would be welcomed to frmly asses how general is the scenario in which the active bath leads to non-equilibrium fuctuations but to an equilibrium-like response.

Methods

Bacteria preparation. *E*. *coli* cells (MG1655) are grown overnight at 33 °C in tryptone broth (TB, Difco) containing 1% tryptone and 0.5% NaCl. The saturated culture is then diluted 1:100 (50 μ l in 5 ml) into fresh medium and grown at 33 °C until OD600 = 0.4 (optical density at 600 nm wavelength) is reached. This optical density corresponds to to the half of the extent of the exponential growth phase (i.e. the so-called "middle-log" phase^{[41](#page-6-14)}). Bacterial cells are then harvested from culture media by centrifugation at 2200 rpm for 10 minutes at room temperature. The pellet is resuspended by gently mixing in a pre-warmed motility buffer composed by 10 mM potassium phosphate, 0.1 mM Na-EDTA (pH 7.0), 76 mM NaCl and 0.002% Tween-20⁴². This motility bufer does not sustain cell replication at room temperature, so the population remains constant in the experiment that follows.

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Author Contributions

C.M. performed the experiments and data analysis. M.P. performed computer simulations. R.D.L. and C.M. designed the experiment. L.A. and M.P. designed the numerical simulations. All authors contributed to the writing of the manuscript.

Additional Information

Competing Interests: The authors declare that they have no competing interests.

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