



An Interleukin-6 Receptor Antibody Suppresses Atherosclerosis in Atherogenic Mice

Koji Akita, Kikuo Isoda*, Yayoi Sato-Okabayashi, Tomoyasu Kadoguchi, Kenichi Kitamura, Fumie Ohtomo, Kazunori Shimada and Hiroyuki Daida

Department of Cardiovascular Medicine, Juntendo University Graduate School of Medicine, Tokyo, Japan

OPEN ACCESS

Edited by:

Ichiro Manabe,
Chiba University, Japan

Reviewed by:

Willi Jahnen-Dechent,
RWTH Aachen University, Germany
George W. Booz,
University of Mississippi Medical
Center School of Dentistry,
United States

*Correspondence:

Kikuo Isoda
kisoda@juntendo.ac.jp

Specialty section:

This article was submitted
to Atherosclerosis and
Vascular Medicine,
a section of the journal
Frontiers in Cardiovascular Medicine

Received: 30 September 2017

Accepted: 08 December 2017

Published: 22 December 2017

I κ BNS is a nuclear I κ B protein which negatively regulates nuclear factor- κ B activity. We demonstrated that I κ BNS deficiency accelerates atherosclerosis in LDL receptor-deficient (LDLr^{-/-}) mice *via* increased interleukin (IL)-6 production by macrophages. Previous studies showed that the increase in IL-6 might contribute to the development of atherosclerotic lesions. However, whether an anti-mouse IL-6 receptor antibody (MR16-1) can protect atherosclerotic lesions in atherogenic mice remains to be elucidated. We investigated atherosclerotic lesions in LDLr^{-/-} and I κ BNS^{-/-}/LDLr^{-/-} mice after 16 weeks consumption of a high-fat diet. All mice received intraperitoneal injections of MR16-1 or phosphate-buffered saline (PBS) (control) once a week during a high-fat diet consumption. Treatment of MR16-1 yielded no adverse systemic effects, and we detected no significant differences in serum cholesterol levels in either group. The atherosclerotic lesions were significantly increased in I κ BNS^{-/-}/LDLr^{-/-} compared with LDLr^{-/-} mice ($p < 0.01$) under treatment of PBS. However, MR16-1 treatment abolished the significant difference of atherosclerotic lesions between I κ BNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice. Interestingly, MR16-1 also significantly decreased atherosclerotic lesions in LDLr^{-/-} mice compared with PBS treatment ($p < 0.05$). Immunostaining revealed percent phospho-STAT3-positive cell were significantly decreased in the atherosclerotic lesions of MR16-1 treated both I κ BNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice compared with PBS-treated mice, indicating MR16-1 could suppress atherosclerotic lesions *via* the inhibition of IL-6–STAT3 signaling pathway. This study highlights the potential therapeutic benefit of anti-IL-6 therapy in preventing atherogenesis induced by dyslipidemia and/or inflammation.

Keywords: interleukin-6, atherosclerosis, inflammation, I κ BNS, dyslipidemia

Citation:

Akita K, Isoda K, Sato-Okabayashi Y, Kadoguchi T, Kitamura K, Ohtomo F, Shimada K and Daida H (2017) An Interleukin-6 Receptor Antibody Suppresses Atherosclerosis in Atherogenic Mice. *Front. Cardiovasc. Med.* 4:84. doi: 10.3389/fcvm.2017.00084

INTRODUCTION

Nuclear factor- κ B (NF- κ B) plays essential roles in mediating immune systems. Cytoplasmic I κ B family proteins regulate the transcriptional activity of NF- κ B, because excessive activation is detrimental to the host (1). After translocation of NF- κ B from the cytoplasm to the nucleus, nuclear proteins that are structurally similar to cytoplasmic I κ Bs take part in the mediating of NF- κ B transcriptional activity, as activators or inhibitors, by associating with NF- κ B subunits (2). Thus, the regulatory I κ B-like nuclear molecules are described as nuclear I κ B proteins. Although I κ BNS is one of nuclear I κ B

proteins, the role of I κ BNS in the development of atherosclerosis is poorly understood. Our previous study demonstrated that deficiency of I κ BNS induces atherogenesis in LDL receptor-deficient (LDLr^{-/-}) mice fed a high-fat diet and increases in interleukin (IL)-6 production by macrophages, indicating that I κ BNS plays an important role in the suppression of atherosclerotic lesions *in vivo* (3). However, whether or not increased IL-6 production in I κ BNS-deficient LDLr^{-/-} mice accelerates atherogenesis remains undetermined.

Several previous reports indicate that IL-6 could accelerate atherosclerosis. Increased plasma levels of IL-6 were associated with cardiovascular mortality over 5-year independent of other risk factor of atherosclerosis (4, 5). Injection of IL-6 itself accelerated atherosclerosis in apolipoprotein E-null mice and C57Bl/6 mice either (4). The genetic polymorphism in the IL-6 signaling pathway concordantly associates with lifetime lower risks of coronary heart disease (6). However, whether inhibition of IL-6 might be effective for the suppression of atherogenesis remains to be elucidated.

We demonstrated here that treatment of an anti-mouse IL-6 receptor antibody (MR16-1) suppressed atherosclerosis lesion in atherogenic mice *via* the inhibition of IL-6–STAT3 signaling pathway.

MATERIALS AND METHODS

Mice

The generation of LDLr^{-/-} mice that lacked I κ BNS (I κ BNS^{-/-}/LDLr^{-/-}) used in this study has been described previously (3). Details of I κ BNS-deficient mice were described in the previous report (7). We investigated atherosclerotic lesions in LDLr^{-/-} and I κ BNS^{-/-}/LDLr^{-/-} mice after 16 weeks consumption of a high-fat diet (MF diet containing 0.5% cholesterol, Oriental Yeast Co.). This study was performed according to the protocols approved by the Juntendo University Board for Studies in Experimental Animals.

Plasma Lipid Measurement

Plasma total cholesterol, triglyceride, high-density lipoprotein (HDL) cholesterol, and low-density lipoprotein (LDL) cholesterol levels were measured by high-performance liquid chromatography at Skylight Biotech Inc. (Akita, Japan).

Quantification of Atherosclerotic Lesions

After blood collection, the animals were euthanized by pentobarbital injection, and the heart and aorta were flushed with 0.9% NaCl followed by 4% paraformaldehyde. After perfusion procedure, the aorta was harvested and fixed with 10% neutral-buffered formalin for 48 h, embedded in paraffin, and sectioned from just above the aortic valve throughout the aortic sinus (each 6- μ m thickness). We used equally spaced 10 cross sections (100- μ m interval) to qualify arteriosclerotic lesions in the aortic sinus for each mouse. The samples were stained with Elastica van Gieson, and then photographed using a BX53 microscope (OLYMPUS, Tokyo, Japan). The luminal, arteriosclerotic

lesions and medial areas were calculated using NIH Image J 1.42 (National Institutes of Health, public domain software). Quantification of the atherosclerotic lesions was performed by two blinded observers.

The whole aortas were also stained with Sudan IV. The surface atherosclerotic lesions were expressed as the percent of the lesion area extending from the ascending aorta to the iliac bifurcation.

Immunohistochemistry

Activation of STAT-3 was detected by phospho-Stat3 (Tyr705) (pSTAT3) staining (1:50; Cell Signaling Technology, #9145). Activation both of pSTAT3 was evaluated for percentage of positive nuclei to total nuclei in the arteries.

Inhibition of IL-6 by Anti-Mouse IL-6 Receptor Antibody (MR16-1) in Mice

All mice received intraperitoneal injections of phosphate-buffered saline (PBS) or MR16-1 (2 mg) once a week during a high-fat diet consumption. MR16-1 was kindly provided from Chugai Pharmaceutical (Japan).

Statistical Analysis

Results are shown as mean \pm SEM. The two groups were compared using Student's *t*-test. Differences between groups were analyzed using one-way ANOVA test followed by Bonferroni's *t*-test to determine statistical differences after multiple comparisons. *p* < 0.05 was considered statistically significant.

RESULTS

Treatment of MR16-1 Yielded No Adverse in Mice

We investigated atherosclerotic lesions in LDLr^{-/-} and I κ BNS^{-/-}/LDLr^{-/-} mice after 16 weeks consumption of a high-fat diet. Both I κ BNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice received injection of PBS (control) or MR16-1 (2 mg) once a week during a high-fat diet consumption of 16 weeks. MR16-1 treatment yielded no adverse systemic effects. Systolic blood pressure and body weight were similar among four groups (Figures 1A,B). Weight gains of mice after consumption of high-fat diet were similar among the four groups. Moreover, analysis of plasma lipid profiles revealed no statistically significant differences in total cholesterol (Figure 2A), LDL cholesterol (Figure 2B), triglyceride (Figure 2C), and HDL cholesterol (Figure 2D) among these groups.

MR16-1 Inhibited Atherosclerosis in both LDLr^{-/-} and I κ BNS^{-/-}/LDLr^{-/-} Mice

We investigated the effect of MR16-1 treatment on atherogenesis in both LDLr^{-/-} and I κ BNS^{-/-}/LDLr^{-/-} mice after 16 weeks consumption of a high-fat diet. As reported previously (3), the extent of atherosclerosis in the mice aortas (en face) was significantly increased in I κ BNS^{-/-}/LDLr^{-/-} compared with single LDLr^{-/-} mice (*p* < 0.001) under treatment of PBS (Figure 3). However, MR16-1 treatment abolished the significant difference of atherosclerotic

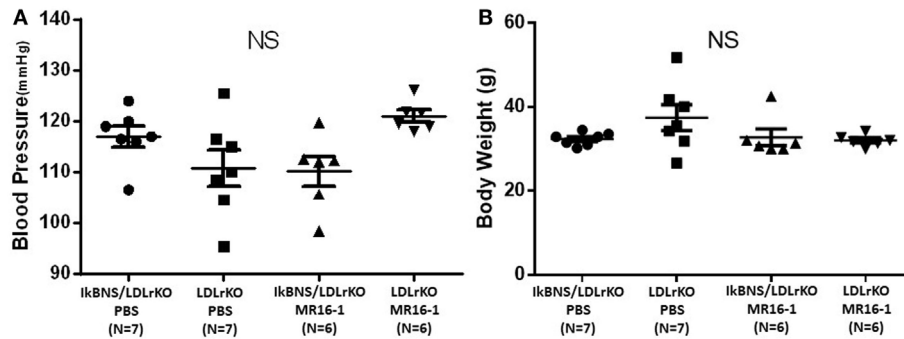


FIGURE 1 | Treatment with MR16-1 did not change blood pressure or body weight in both $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ and $\text{LDLr}^{-/-}$ mice. Both mice were received injection of phosphate-buffered saline (PBS) or MR16-1 (2 mg) once a week during a high-fat diet consumption of 16 weeks. Systolic blood pressure (**A**) and body weight (**B**) were similar among four groups. $\text{I}\kappa\text{BNS}/\text{LDLr}$ KO PBS: PBS-treated $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice, LDLr KO PBS: PBS-treated $\text{LDLr}^{-/-}$ mice. $\text{I}\kappa\text{BNS}/\text{LDLr}$ KO MR16-1: MR16-1-treated $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice, LDLr KO MR16-1: MR16-1-treated $\text{LDLr}^{-/-}$ mice. Data are expressed as mean \pm SEM. NS, not significant.

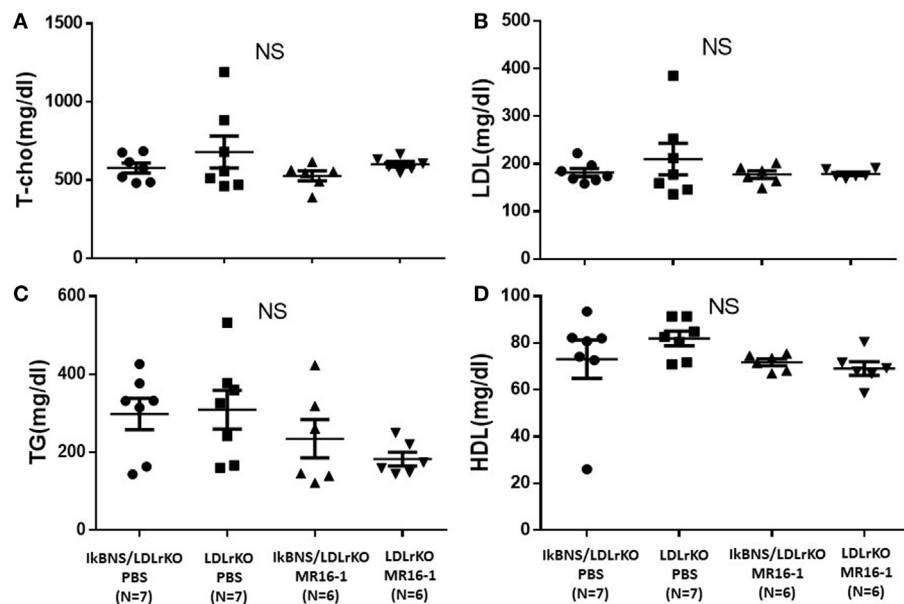


FIGURE 2 | Treatment with MR16-1 did not change cholesterol profiles in both $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ and $\text{LDLr}^{-/-}$ mice. The levels of total cholesterol (T-cho) (**A**), low-density lipoprotein (LDL)-cholesterol (**B**), triglyceride (TG) (**C**), and high-density lipoprotein (HDL)-cholesterol (**D**) after treatment of phosphate-buffered saline (PBS) or MR16-1 (2 mg) once a week during a high-fat diet consumption of 16 weeks. $\text{I}\kappa\text{BNS}/\text{LDLr}$ KO PBS: PBS-treated $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice, LDLr KO PBS: PBS-treated $\text{LDLr}^{-/-}$ mice. $\text{I}\kappa\text{BNS}/\text{LDLr}$ KO MR16-1: MR16-1-treated $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice, LDLr KO MR16-1: MR16-1-treated $\text{LDLr}^{-/-}$ mice. Data are expressed as mean \pm SEM. NS, not significant.

lesions between $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ and $\text{LDLr}^{-/-}$ mice (**Figure 3**). Interestingly, MR16-1 treatment also significantly decreased atherosclerotic lesions in $\text{LDLr}^{-/-}$ mice compared with PBS treatment (**Figure 3**).

Aortic root atherosclerotic lesions of $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice were also significantly larger than those in $\text{LDLr}^{-/-}$ mice ($p < 0.05$) under treatment of PBS (**Figure 4**). MR16-1 treatment significantly reduced atherosclerotic lesions in both $\text{LDLr}^{-/-}$ and $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice compared with PBS treatment (**Figure 4**).

These results show that an anti-mouse IL-6 receptor antibody significantly suppresses $\text{I}\kappa\text{BNS}$ in both $\text{LDLr}^{-/-}$ and $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ mice without major confounding effects on

LDL-cholesterol or HDL-cholesterol, suggesting that inhibition of IL-6 might be significantly effective to prevent atherogenesis.

MR16-1 Suppressed STAT3 Activation in the Atherosclerotic Lesions of both $\text{LDLr}^{-/-}$ and $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$ Mice

To examine STAT activation in the atherosclerotic lesions of both MR16-1 and PBS groups, we performed pSTAT3 staining in the atherosclerotic lesions of four groups after 16 weeks consumption of a high-fat diet. We detected much more pSTAT3-positive cells in the atherosclerotic lesions of PBS-treated $\text{I}\kappa\text{BNS}^{-/-}/\text{LDLr}^{-/-}$

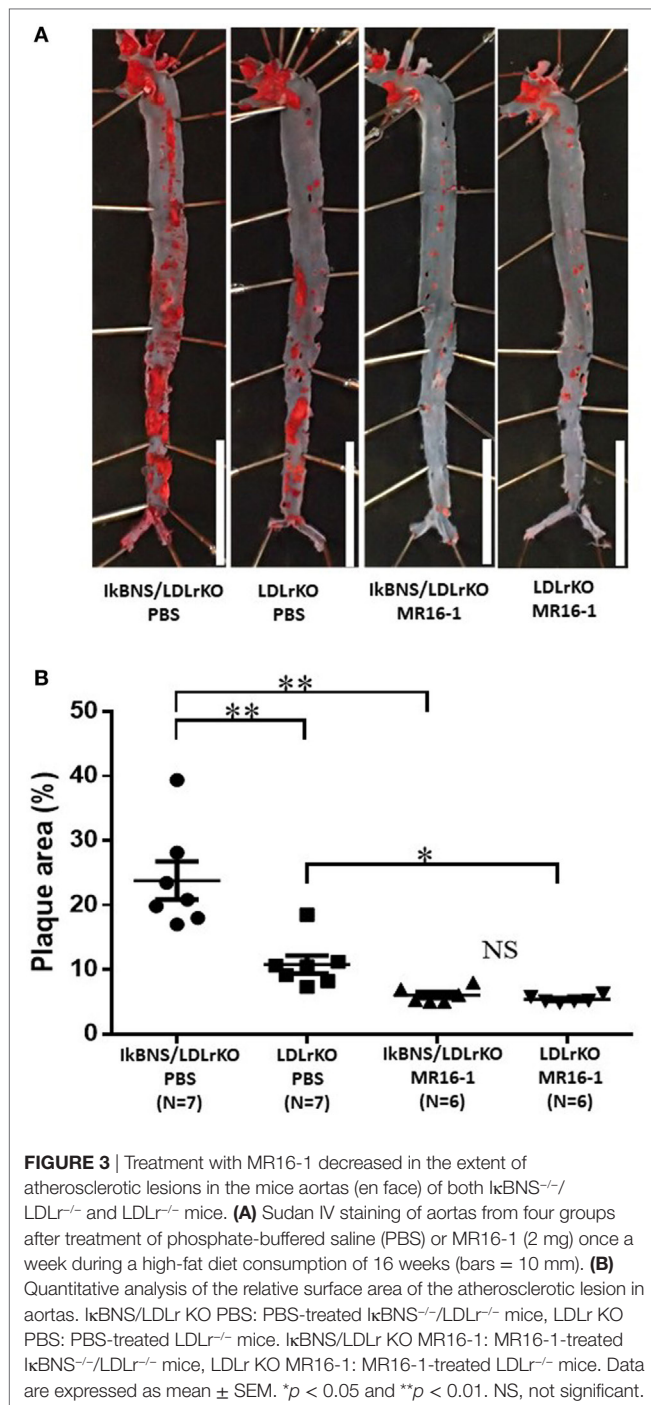


FIGURE 3 | Treatment with MR16-1 decreased in the extent of atherosclerotic lesions in the mice aortas (en face) of both IκBNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice. **(A)** Sudan IV staining of aortas from four groups after treatment of phosphate-buffered saline (PBS) or MR16-1 (2 mg) once a week during a high-fat diet consumption of 16 weeks (bars = 10 mm). **(B)** Quantitative analysis of the relative surface area of the atherosclerotic lesion in aortas. IκBNS/LDLr KO PBS: PBS-treated IκBNS^{-/-}/LDLr^{-/-} mice, LDLr KO PBS: PBS-treated LDLr^{-/-} mice, IκBNS/LDLr KO MR16-1: MR16-1-treated IκBNS^{-/-}/LDLr^{-/-} mice, LDLr KO MR16-1: MR16-1-treated LDLr^{-/-} mice. Data are expressed as mean ± SEM. **p* < 0.05 and ***p* < 0.01. NS, not significant.

mice compared with PBS-treated IκBNS^{-/-}/LDLr^{-/-} mice (Figure 5A). MR16-1 treatment significantly decreased pSTAT3-positive cells in the atherosclerotic lesions of both LDLr^{-/-} and IκBNS^{-/-}/LDLr^{-/-} mice compared with PBS treatment (Figure 5A).

To quantify STAT3 activity, we calculated the percentage of pSTAT3-positive nuclei to total nuclei in the arteries. The percentage of pSTAT3-positive nuclei in the aorta of MR16-1-treated IκBNS^{-/-}/LDLr^{-/-} mice was significantly lower than that in PBS treated group (47.3 ± 3.1 versus 8.4 ± 1.5%; *p* < 0.01; Figure 5B).

Interestingly, MR16-1 treatment also decreased percentage of pSTAT3-positive nuclei in LDLr^{-/-} mice compared with PBS treatment (27.7 ± 1.5 versus 7.2 ± 1.8%; *p* < 0.01; Figure 5B).

DISCUSSION

Persistent IL-6 production has been shown to play a critical role in chronic inflammatory disease (8). IL-6 signaling has also been linked to plaque initiation and destabilization (9) and to adverse outcomes in the setting of acute ischemia (10). IL-6 stimulates growth of promoters of macrophage and vascular smooth muscle cells, which are major components of plaque (11–13), and IL-6 expression is also detected in human atherosclerotic lesions (14). These findings indicate that IL-6 participates in the development of atherosclerosis. Thus, targeting IL-6 is a rational approach for atheroprotection. To block the effect of IL-6 on the development of atherosclerosis, we used anti-mouse IL-6 receptor antibody (MR16-1). MR16-1 has proved to be effective in experimental model of arthritis (15) and humanized anti-IL-6 receptor antibody (tocilizumab) has been approved by the Food and Drug Administration for the treatment in the patients with rheumatoid arthritis.

Our previous study demonstrates that IL-6 expression and STAT3 activation were increased in the foam cell rich-atherosclerotic lesions of IκBNS^{-/-}/LDLr^{-/-} mice *in vivo* and IκBNS^{-/-}/LDLr^{-/-} macrophages produced much higher level of IL-6 than LDLr^{-/-} macrophages *in vitro*. These results indicate deficiency of IκBNS increases in the production of IL-6 in macrophage and an increase of IL-6 contributes to the susceptibility to atherogenesis in IκBNS^{-/-}/LDLr^{-/-} mice (3). Consequently, because we believe that the mice were suitable for evaluating the effect of IL-6-blocking therapy in atherogenesis, we used IκBNS^{-/-}/LDLr^{-/-} mice in this study. As expected, MR16-1 treatment abolished the significant difference of atherosclerotic lesions between IκBNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice. Interestingly, MR16-1 treatment also significantly decreased atherosclerotic lesions in LDLr^{-/-} mice compared with PBS treatment.

Interleukin-6 is an upstream inflammatory cytokine key player, propagating the downstream inflammatory response in atherosclerosis (16). Previous study demonstrated that inflammatory gene, such as IL-6 (17), which is known as an inducer of STAT3 was highly expressed in atherosclerotic plaques. Moreover, activation of STAT3 has been detected in the plaque (18), and its activation is involved in the progression of atherosclerotic lesions (19). In this study, we analyzed pSTAT3-positive nuclei to evaluate the inhibitory effect of MR16-1 in IL-6 signaling pathway. MR16-1 inhibited STAT3 activation and development of atherosclerotic lesions in both IκBNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice, suggesting IL-6 is a main contributor to atherogenesis, and inhibition of IL-6 might be a new method to prevent the development of atherosclerosis.

Our data provide solid evidence for a causal role of the IL-6 pathway in the development of atherosclerosis, raising expectations for ongoing trials testing anti-inflammatory strategies for cardiovascular diseases (20).

Although we could not find any adverse systemic effects including signs of infection in MR16-1-treated mice, further

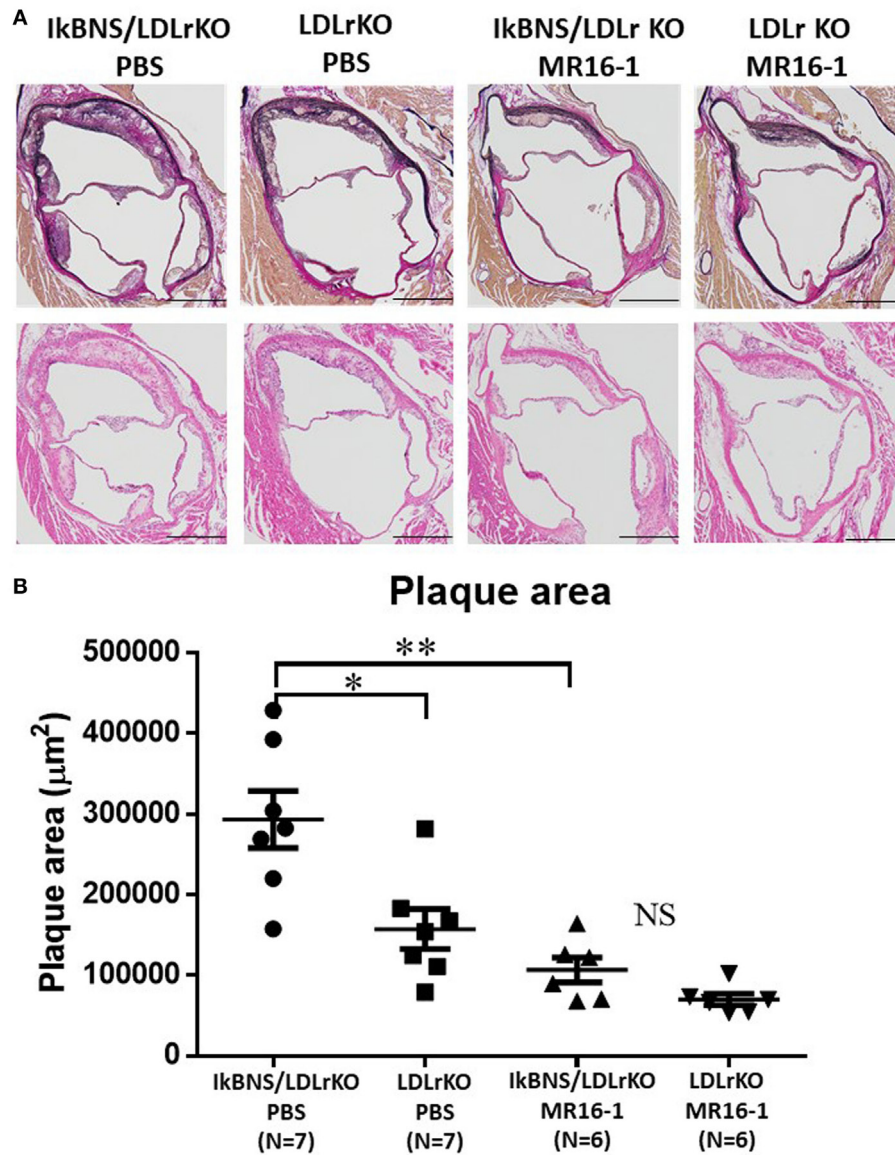


FIGURE 4 | Treatment with MR16-1 decreased in aortic root atherosclerotic lesions of both IκBNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice. **(A)** Representative photomicrographs of sections of aortic sinus plaque from four groups after treatment of phosphate-buffered saline (PBS) or MR16-1 (2 mg) once a week during a high-fat diet consumption of 16 weeks. Adjacent sections were processed for Elastica van Gieson staining (upper panels) and H&E staining (lower panels) (bars = 500 μm). **(B)** Quantitative comparison of the atherosclerotic lesion in the aortic sinus. IκBNS/LDLr KO PBS: PBS-treated IκBNS^{-/-}/LDLr^{-/-} mice, LDLr KO PBS: PBS-treated LDLr^{-/-} mice. IκBNS/LDLr KO MR16-1: MR16-1-treated IκBNS^{-/-}/LDLr^{-/-} mice, LDLr KO MR16-1: MR16-1-treated LDLr^{-/-} mice. Data are expressed as mean ± SEM. **p* < 0.05 and ***p* < 0.01. NS, not significant.

study must be needed for checking any adverse effects of MR16-1 in the immune systems.

In our experiments, levels of IL-6 in plasma and liver were not different between IκBNS^{-/-}/LDLr^{-/-} and LDLr^{-/-} mice after consumption of high-fat diet. On the other hand, Kuwata et al. (7) demonstrated intraperitoneally injection of LPS induced significantly high-serum levels of IL-6 in IκBNS^{-/-} mice compared to wild-type (IκBNS^{+/+}) mice. These findings suggest that feeding a high-fat diet could induce local increase of IL-6 produced by IκBNS^{-/-} macrophages around the vessels,

but not serum levels of IL-6. In our previous study, IκBNS expression was observed in all cell types resident to the plaque microenvironment (i.e., smooth muscle cells, macrophages, and endothelial cells) in LDLr^{-/-} mice, but not in IκBNS^{-/-}/LDLr^{-/-} mice. Because main IκBNS expression cells were macrophages, we previously examined the effects of the IκBNS deficiency in macrophages. We also investigated the effect of IκBNS deficiency in smooth muscle cells using small interference RNA, because they play important roles in atherogenesis. However, IκBNS deficiency in aortic smooth muscle cell did not

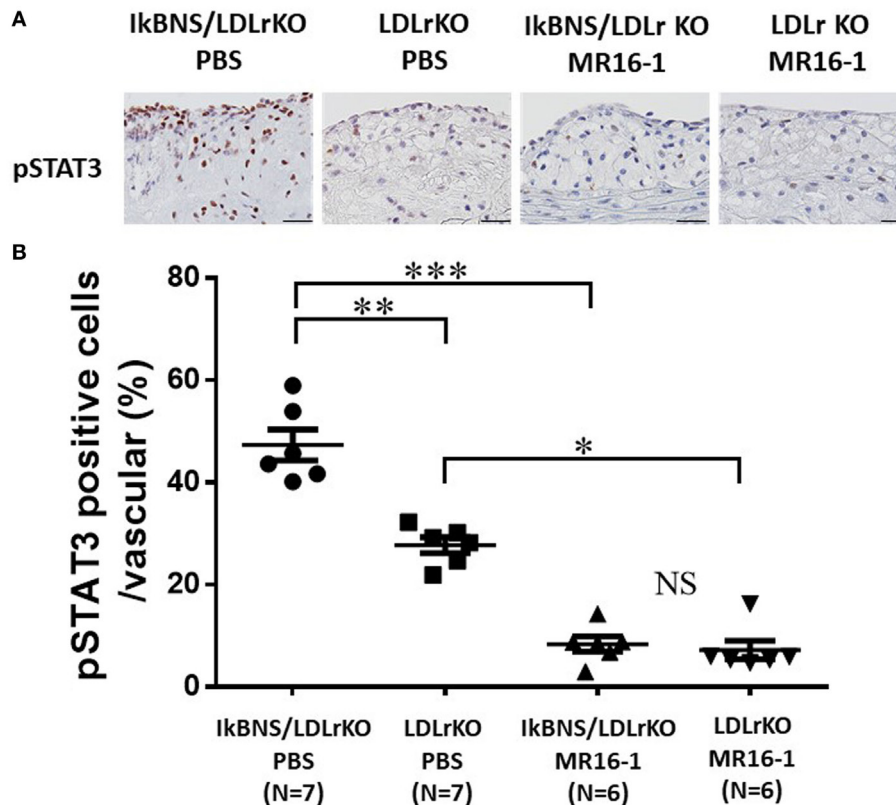


FIGURE 5 | MR16-1 treatment significantly inhibited STAT3 activation in the atherosclerotic lesions of both *IkbNS^{-/-}/LDLr^{-/-}* and *LDLr^{-/-}* mice. **(A)** Representative photomicrographs of sections of aortic sinus plaque from four groups after treatment of phosphate-buffered saline (PBS) or MR16-1 (2 mg) once a week during a high-fat diet consumption of 16 weeks. Sections were processed for pSTAT3 staining (bars = 25 μ m). **(B)** Quantitative analysis of pSTAT3-positive cell in vascular wall in sections from four groups. *IkbNS/LDLr KO PBS*: PBS-treated *IkbNS^{-/-}/LDLr^{-/-}* mice, *LDLr KO PBS*: PBS-treated *LDLr^{-/-}* mice, *IkbNS/LDLr KO MR16-1*: MR16-1-treated *IkbNS^{-/-}/LDLr^{-/-}* mice, *LDLr KO MR16-1*: MR16-1-treated *LDLr^{-/-}* mice. Data are expressed as mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$. NS, not significant.

affect in IL-6 production *in vitro*. Further experiments might be needed to clarify which cells are main players in atherogenesis induced by *IkbNS* deficiency.

In conclusion, this study highlights the potential therapeutic benefit of anti-IL-6 therapy in preventing atherogenesis in the mice with dyslipidemia and/or inflammation.

ETHICS STATEMENT

The studies were performed according to the protocols approved by the Juntendo University Board for Studies in Experimental Animals.

REFERENCES

1. Ghosh S, Karin M. Missing pieces in the NF- κ B puzzle. *Cell* (2002) 109:S81–96. doi:10.1016/S0092-8674(02)00703-1
2. Yamamoto M, Takeda K. Role of nuclear I κ B proteins in the regulation of host immune responses. *J Infect Chemother* (2008) 14:265–9. doi:10.1007/s10156-008-0619-Y
3. Akita K, Isoda K, Okabayashi Y, Shimada K, Daida H. Lack of *IkbNS* accelerates atherosclerosis in LDL receptor-deficient mice via increased interleukin-6 production. *Int J Cardiol* (2016) 211:61–3. doi:10.1016/j.ijcard.2016.02.142

AUTHOR CONTRIBUTIONS

KA, KI, YO-S, TK, and KK performed the research. KI, KS, and HD designed the research study. KA, YO-S, TK, KK, and FO contributed essential reagents or tools. KA, KI, YO-S, TK, KK, FO, KS, and HD analyzed the data. KA, KI, YO-S, TK, KK, and FO, KS, and HD wrote the paper.

FUNDING

This work was supported by Grant-in-Aid for Scientific Research (C) (JSPS KAKENHI Grant Number 16K09522).

4. Huber SA, Sakkinen P, Conze D, Hardin N, Tracy R. Interleukin-6 exacerbates early atherosclerosis in mice. *Arterioscler Thromb Vasc Biol* (1999) 19:2364–7. doi:10.1161/01.ATV.19.10.2364
5. Luc G, Bard JM, Juhan-Vague I, Ferrieres J, Evans A, Amouyel P, et al. C-reactive protein, interleukin-6, and fibrinogen as predictors of coronary heart disease: the PRIME Study. *Arterioscler Thromb Vasc Biol* (2003) 23:1255–61. doi:10.1161/01.ATV.0000079512.66448.1D
6. Ridker PM. From C-reactive protein to interleukin-6 to interleukin-1: moving upstream to identify novel targets for atheroprotection. *Circ Res* (2016) 118:145–56. doi:10.1161/CIRCRESAHA.115.306656

7. Kuwata H, Matsumoto M, Atarashi K, Morishita H, Hirofumi T, Koga R, et al. IkappaBNS inhibits induction of a subset of toll-like receptor-dependent genes and limits inflammation. *Immunity* (2006) 24:41–51. doi:10.1016/j.immuni.2005.11.004
8. Tanaka T, Kishimoto T. Targeting interleukin-6: all the way to treat autoimmune and inflammatory diseases. *Int J Biol Sci* (2012) 8:1227–36. doi:10.7150/ijbs.4666
9. Yudkin JS, Kumari M, Humphries SE, Mohamed-Ali V. Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis* (2000) 148:209–14. doi:10.1016/S0021-9150(99)00463-3
10. Lindmark E, Diderholm E, Wallentin L, Siegbahn A. Relationship between interleukin 6 and mortality patients with unstable coronary artery disease: effects of an early invasive or noninvasive strategy. *JAMA* (2001) 286:2107–13. doi:10.1001/jama.286.17.2107
11. Ikeda U, Ikeda M, Oohara T, Oguchi A, Kamitani T, Tsuruya Y, et al. Interleukin 6 stimulates growth of vascular smooth muscle cells in a PDGF-dependent manner. *Am J Physiol* (1991) 260:H1713–7.
12. Jenkins BJ, Grail D, Inglese M, Quilici C, Bozinovski S, Wong P, et al. Imbalanced gp130-dependent signaling in macrophages alters macrophage colony-stimulating factor responsiveness via regulation of c-fms expression. *Mol Cell Biol* (2004) 24:1453–63. doi:10.1128/MCB.24.4.1453-1463.2004
13. Wang D, Liu Z, Li Q, Karpurapu M, Kundumani-Sridharan V, Cao H, et al. An essential role for gp130 in neointima formation following arterial injury. *Circ Res* (2007) 100:807–16. doi:10.1161/01.RES.0000261350.61711.9e
14. Seino Y, Ikeda U, Ikeda M, Yamamoto K, Misawa Y, Hasegawa T, et al. Interleukin 6 gene transcripts are expressed in human atherosclerotic lesions. *Cytokine* (1994) 6:87–91. doi:10.1016/1043-4666(94)90013-2
15. Takagi N, Mihara M, Moriya Y, Nishimoto N, Yoshizaki K, Kishimoto T, et al. Blockage of interleukin-6 receptor ameliorates joint disease in murine collagen-induced arthritis. *Arthritis Rheum* (1998) 41:2117–21. doi:10.1002/1529-0131(199812)41:12<2117::AID-ART6>3.0.CO;2-P
16. Hartman J, Frishman WH. Inflammation and atherosclerosis: a review of the role of interleukin-6 in the development of atherosclerosis and the potential for targeted drug therapy. *Cardiol Rev* (2014) 22:147–51. doi:10.1097/CRD.0000000000000021
17. Schieffer B, Schieffer E, Hilfiker-Kleiner D, Hilfiker A, Kovanen PT, Kaartinen M, et al. Expression of angiotensin II and interleukin 6 in human coronary atherosclerotic plaques: potential implications for inflammation and plaque instability. *Circulation* (2000) 101:1372–8. doi:10.1161/01.CIR.101.12.1372
18. Recinos A III, Lejeune WS, Sun H, Lee CY, Tieu BC, Lu M, et al. Angiotensin II induces IL-6 expression and the Jak-STAT3 pathway in aortic adventitia of LDL receptor-deficient mice. *Atherosclerosis* (2007) 194:125–33. doi:10.1016/j.atherosclerosis.2006.10.013
19. Grote K, Luchtefeld M, Schieffer B. JANUS under stress-role of JAK/STAT signaling pathway in vascular disease. *Vascul Pharmacol* (2005) 43:357–63. doi:10.1016/j.vph.2005.08.021
20. Ridker PM, Luscher TF. Anti-inflammatory therapies for cardiovascular disease. *Eur Heart J* (2014) 35:1782–91. doi:10.1093/eurheartj/ehu203

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Akita, Isoda, Sato-Okabayashi, Kadoguchi, Kitamura, Ohtomo, Shimada and Daida. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.