


Article

Diversity and Phylogenetic Analyses of Bacterial Symbionts in Three Whitefly Species from Southeast Europe

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Abstract: *Bemisia tabaci* (Gennadius), *Trialeurodes vaporariorum* (Westwood), and *Siphoninus phillyreae* (Haliday) are whitefly species that harm agricultural crops in many regions of the world. These insects live in close association with bacterial symbionts that affect host fitness and adaptation to the environment. In the current study, we surveyed the infection of whitefly populations in Southeast Europe by various bacterial symbionts and performed phylogenetic analyses on the different symbionts detected. *Arsenophonus* and *Hamiltonella* were the most prevalent symbionts in all three whitefly species. *Rickettsia* was found to infect mainly *B. tabaci*, while *Wolbachia* mainly infected both *B. tabaci* and *S. phillyreae*. Furthermore, *Cardinium* was rarely found in the investigated whitefly populations, while *Fritschea* was never found in any of the whitefly species tested. Phylogenetic analyses revealed a diversity of several symbionts (e.g., *Hamiltonella*, *Arsenophonus*, *Rickettsia*), which appeared in several clades. Reproductively isolated *B. tabaci* and *T. vaporariorum* shared the same (or highly similar) *Hamiltonella* and *Arsenophonus*, while these symbionts were distinctive in *S. phillyreae*. Interestingly, *Arsenophonus* from *S. phillyreae* did not cluster with any of the reported sequences, which could indicate the presence of *Arsenophonus*, not previously associated with whiteflies. In this study, symbionts (*Wolbachia*, *Rickettsia*, and *Cardinium*) known to infect a wide range of insects each clustered in the same clades independently of the whitefly species. These results indicate horizontal transmission of bacterial symbionts between reproductively isolated whitefly species, a mechanism that can establish new infections that did not previously exist in whiteflies.

Keywords: bacterial symbionts; phylogenetic analyses; mtCOI; diversity; whiteflies

1. Introduction

Insects share a long-term and intimate association with bacterial symbionts, which has helped their hosts to become well adapted to numerous challenges present in their environment [1–4]. Primary bacterial symbionts are essential for insects to live on nutritionally poor diets, while secondary

bacterial symbionts can additionally affect traits such as development, reproduction, thermal tolerance, protection from natural enemies, and immunity [2,5–9].

Whitefly species such as *Bemisia tabaci* (Gennadius), *Trialeurodes vaporariorum* (Westwood), and *Siphoninus phillyreae* (Haliday) are phloem-feeding pests that damage numerous agricultural crops [10–13]. In particular, *B. tabaci* and *T. vaporariorum* are globally distributed, extremely polyphagous, and cause indirect damage by transmitting important plant viruses [14–16]. All three whitefly species are invasive and have a long history of resistance to insecticides from different chemical classes [13,17,18]. *B. tabaci* presents a species complex, composed of at least 40 morphologically indistinguishable species [10,15,19–27]. *B. tabaci* species Middle East-Asia Minor 1 (MEAM1) and Mediterranean (MED) are globally distributed and the most challenging to control [15,28–30]. The high diversity in mitochondrial cytochrome oxidase I (mtCOI) gene sequences revealed differentiation within *B. tabaci* (MED) into four groups: Q1 (western Mediterranean populations), Q2 (Middle Eastern populations), Q3 (Burkina Faso populations), and Africa Silver Leafing (ASL) (Ivory Coast, Burkina Faso, and Cameroon) [31–33].

All whitefly species harbor a primary symbiont, *Portiera aleyrodidarum*, which supplements the insects' diet with essential amino acids and carotenoids [34]. In *B. tabaci*, several bacterial symbionts have been described, including *Hamiltonella*, *Rickettsia*, *Wolbachia*, *Cardinium*, *Arsenophonus*, and *Fritschea* [35–38]. Several studies have attempted to link infection with the secondary symbiotic community with *B. tabaci* speciation and performance [33,38]. However, due to horizontal transmission, this is difficult to prove [33,39,40].

Diverse secondary symbionts have been found in *T. vaporariorum* and *S. phillyreae* populations as well [39,41]. *T. vaporariorum* was infected with *Hamiltonella*, *Rickettsia*, *Wolbachia*, *Cardinium*, and *Arsenophonus*, while *S. phillyreae* contained the same symbiotic composition as *T. vaporariorum* with no detected presence of *Rickettsia*. *Wolbachia*, *Rickettsia*, *Cardinium*, and *Arsenophonus* are known to manipulate host reproduction in many insect species [2,42–44]. In *B. tabaci*, *Rickettsia* induces the expression of genes required for thermotolerance [45], provides the host with fitness benefits [46], increases its susceptibility to insecticides [47], and is involved in hosts' response to parasitism [48]. *Arsenophonus* was additionally found to act as an obligatory mutualist in lice [49,50]. The GroEL proteins produced by *Arsenophonus* in *B. tabaci* (Asia II species) and *Hamiltonella* in Israeli populations of *B. tabaci* (MEAM1) were found to interact with the coat protein of begomovirus and therefore facilitate virus transmission [51,52]. The GroEL produced by other symbionts of *B. tabaci* (MEAM1 and MED) did not interact with the virus and therefore were not involved in virus transmission [52]. Furthermore, in the pea aphid, *Hamiltonella* confers resistance to parasitoids and increases thermotolerance [53,54]. Recently, the genome of *Hamiltonella* from *B. tabaci* (MED) was sequenced, and this dataset provided insights into nutritional benefits this symbiont may provide to the whitefly host [55,56]. *Fritschea* has been reported only in the New World *B. tabaci* species, and at present, nothing is known concerning its function [57,58].

The focus of this and our previous studies were whiteflies from Southeast Europe (Croatia, Montenegro, Bosnia and Herzegovina, Macedonia, and Serbia) and their associated bacterial symbionts. In this region, *B. tabaci*, *T. vaporariorum*, and *S. phillyreae* have been identified and possibly predominate [39,41]. *T. vaporariorum* has been reported as the predominant whitefly species in the surveyed region, while *B. tabaci* and *S. phillyreae* have never been reported from Serbia [41] (J. Bocanski, pers. comm.). Based on molecular markers, two invasive species of *B. tabaci* were detected; in Montenegro, both the MEAM1 and MED species were detected, whereas in Croatia and Bosnia and Herzegovina, only the MED species was confirmed [39,41]. Q1 and Q2 groups of *B. tabaci* (MED) were reported from Croatia and Montenegro [19,41]. All three whitefly species had unique co-infections with bacterial symbionts compared to previously reported studies [28,37,58]. Our aim was to add further details concerning the global phylogenetic diversity of bacterial symbionts between the three whitefly species. This may give further insights about the functions these symbionts may have in whitefly biology.

2. Materials and Methods

2.1. Whitefly Populations

Populations of the three whitefly species were collected in the summers from 2009 to 2013 in regions with intense agricultural activity in Croatia, Montenegro, Macedonia, and Serbia (Figure 1, Table S1). Whitefly specimens were collected from open fields or greenhouses when high population densities were available. Sampling locations, whitefly species, and host plants are shown in Table S1 and Table 3. Whitefly populations from Macedonia and Serbia were newly examined; two laboratory populations of *B. tabaci* from Israel were used for comparison, whereas other whitefly populations from Croatia and Montenegro were previously screened in Skaljac et al. (2010, 2013) [39,41] and used in this study as well. Adults of the three whitefly species were collected using a Pasteur pipette attached to a hand-held aspirator and then moved into absolute ethanol and kept at $-20\text{ }^{\circ}\text{C}$ until processing.

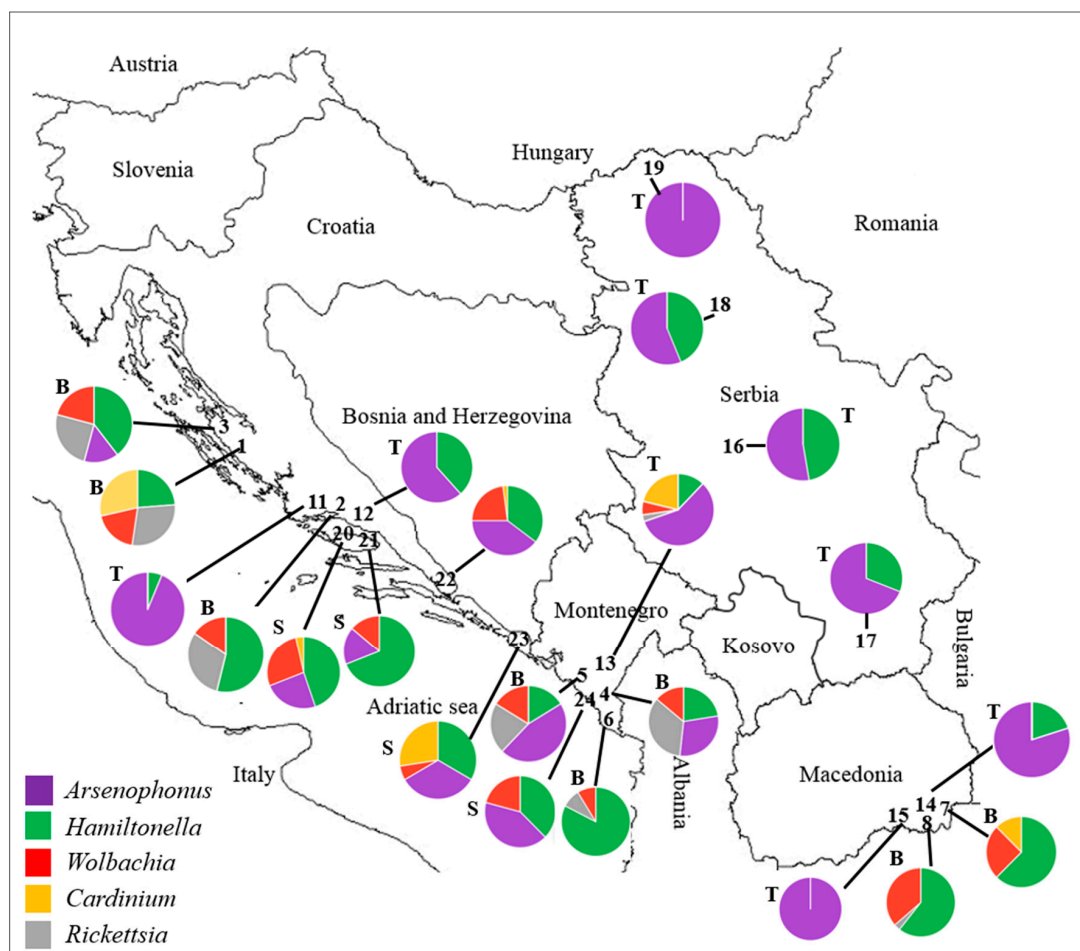


Figure 1. Locations of collected whitefly populations (according to population numbers in Table S1) with overview of overall infection frequencies of secondary bacterial symbionts (according to Table 2) in the three whitefly species in Southeast Europe. Each chart is marked with a code (B, T, and S) in order to distinguish between the whitefly species. B, *Bemisia tabaci*; T, *Trialeurodes vaporariorum*; S, *Siphoninus phillyrae*.

2.2. Screening and Sequencing of Secondary Bacterial Symbionts and Molecular Identification of *B. tabaci* Species

Adults ($n = 10\text{--}20$ per population) from the whitefly populations (Table S1) were tested for the presence of secondary symbionts and *B. tabaci* species (MED or MEAM1) identification. Genomic DNA of each individual was extracted in lysis buffer as previously described in Chiel et al. (2007) [37] and

used for further screening. *Bemisia tabaci* MED or MEAM1 groups were identified using microsatellite markers by polymerase chain reaction (PCR) amplification using Bem 23 primers and fragment sizes. The product size obtained from MEAM1 was 200 bp, and from the MED, it was 400 bp. In addition, we amplified mtCOI gene fragment for phylogenetic analyses of *B. tabaci* species (Table 1). Secondary bacterial symbionts were tested by PCR using genus-specific primers amplifying 16S or 23S rDNA (Table 1). We did not detect *Fritschea* in our previous studies, and due to the lack of a positive control, *Fritschea* was not considered in this study.

Table 1. The primers used in this study.

Primer Name	Sequence (5'→3')	Annealing (°C)/Size (bp)	Gene	Reference
Bem 23 F Bem 23 R	CGGAGCTTGGCGCTTAGTC CGGCTTTATCATAGCTCTCGT	55/MEAM1 = 200; MED = 400	Microsatellite	[59]
C1-J-2195 L2-N-3014	TTGATTTT TGGTCATCCAGAAGT TCCAATGCACTAATCTGCCATATTA	51/850	Cytochrome oxidase I (mtCOI)	[60]
Por-F Por-R	TGCAAGTCGAGCGGCATCAT AAAGTTCGCCCTTATGCGT	59/1000	<i>Portiera</i> 16S rDNA	[35]
Rb F Rb R	GCTCAGAACGAACGCTATC GAAGGAAAGCATCTCTGC	59/900	<i>Rickettsia</i> 16S rDNA	[36]
92 F Hb R	TGAGTAAAGTCTGGGAATCTGG AGTTCAAGACCGCCAACCTC	62/700	<i>Hamiltonella</i> 16S rDNA	[35]
Ars23S-1 Ars23S-2	CGTTTGATGAATTCATAGTCAAAA GGTCCTCCAGTTAGTGTACCCAAC	59/600	<i>Arsenophonus</i> 23S rDNA	[61]
Wol16S F Wol16S R	CGG GGGAAAAATTATTGCT AGCTGTAATACAGAAAGTAAA	55/650	<i>Wolbachia</i> 16S rDNA	[38]
CFB F CFB R	GCGGTGTAATAATGAGCGTG ACCTMTTCTTAAGCTCAAGCCT	59/500	<i>Cardinium</i> 16S rDNA	[62]

PCR was carried out in 25 µL volumes containing 4 µL DNA lysate, 10 pmol of each primer, 10 mM dNTP mix, 10× PCR Rxn Buffer, 2 mM MgCl₂, and 5 units/µL of Platinum Taq DNA polymerase (Invitrogen). PCR products were visualized on a 1.5% agarose gel containing ethidium bromide. The PCR products were eluted, and DNA was ligated into a pGEM-T Easy Vector (Promega, Madison, WI, USA) following the manufacturer's instructions. After transfection into RapidTrans TAM1 competent *E. coli* (Active Motif, Carlsbad, CA, USA), cells were grown on LB agar (Invitrogen) at 37 °C overnight. Clones were then screened for PCR inserts using the primers listed in Table 1. Colonies were then grown overnight in 5 mL LB medium containing ampicillin (100 µg/mL) in a shaking incubator at 200 rpm and 37 °C. Plasmid DNA from independent colonies was purified using a QIAprep Spin Miniprep kit (Qiagen, Hilden, Germany) and sent for sequencing (3730xl DNA analyzer, Macrogen Europe, Amsterdam, The Netherlands). The sequences were obtained from 10–20 individual specimens per primer set and compared with those in the databases using the BLAST algorithm at the NCBI [63].

2.3. Phylogenetic Analyses

Mitochondrial COI and secondary symbiont sequences for all *B. tabaci*, *T. vaporarium*, and *S. phillyreae* obtained during this study, together with reference sequences from GenBank, were aligned using the program MAFFT 7 [64] followed by visual inspection and manual adjustment. The appropriate model of evolution was estimated using the Bayesian Information Criterion (BIC) with Jmodeltest 2.1.6 on CIPRES Science Gateway [65], and selected models were the same for ML inferences. Analyses were initiated from random starting trees. The models selected were GTR+G for mtCOI, HKY for *Hamiltonella*, GTR+G for *Arsenophonus* and *Wolbachia*, and HKY+I for *Cardinium* and *Rickettsia*. Using these models, phylogenetic reconstruction was carried out using Bayesian inference (BI). For BI analysis, two independent runs of Markov Chain Monte Carlo (MCMC) were done for 10 million generations using eight chains and a sampling frequency of 1000 generations by MrBayes ver. 3.2 [66]. The first 25% of the samples were discarded by setting the burn-in fraction to 0.25. This burn-in setting

was shared across the SUMT command to discard 25% of sampled trees. These phylogenetic trees were viewed by FigTree V. 1.4.2, available at <http://tree.bio.ed.ac.uk/software/figtree/>. Pairwise genetic distances for whitefly mtCOI sequences were analyzed using the p-distance model of evolution in MEGA 6.0 [67]. The whitefly sequences characterized in this study were assigned to different species on the basis of the rule of >3.5% pairwise sequence divergence [19].

2.4. Nucleotide Sequence Accession Numbers

The nucleotide sequences of the genes identified in this study have been deposited in GenBank under the accession numbers KY623456 to KY623475 for *Arsenophonus*, from KY621483 to KY621519 for *Hamiltonella*, from KY994554 to KY994569 for *Rickettsia*, from KY994545 to KY994553 for *Wolbachia*, from KY994541 to KY994544 for *Cardinium*, and from KY628431 to KY628448 for mtCOI from *B. tabaci*.

3. Results

3.1. Whitefly Infection with Secondary Bacterial Symbionts

Ten to 20 individuals from each population of the three whitefly species were tested for the presence of the secondary bacterial symbionts. Infection frequencies with multiple secondary symbionts present in 24 whitefly populations used in this and our previous studies are given in Tables 2 and 3, and Figure S1. All the tested individuals were positive for the primary symbiont of whiteflies, *P. aleyrodidarum*, and this was considered to control the quality of extracted DNA.

We could not find a correlation between infection with bacterial symbionts and host plants that whiteflies infested, as was suggested in our previous studies (Table 2) [39,41].

3.1.1. *Bemisia tabaci* Infection with Secondary Bacterial Symbionts

B. tabaci populations were collected in Croatia, Montenegro, and Macedonia (Figure 1; Table S1). All populations used in this study were identified as MED species and they belonged to groups Q1 and Q2 (Tables 2 and 3). These populations had high prevalence of mixed infections with *Hamiltonella*, *Arsenophonus*, *Rickettsia*, *Wolbachia*, and *Cardinium* (Table 2). *Hamiltonella* was highly prevalent or fixed in *B. tabaci* MED (Q1) populations from Macedonia (Doiran, Gevgelija) and Montenegro (Darza). *Arsenophonus* was prevalent in populations from Montenegro (Bar—MED Q2, Lastva Grbaljska—MED Q1), whereas *Rickettsia* was fixed in population collected in Bar (MED Q2). Individuals of both Q1 and Q2 groups of *B. tabaci* MED were mainly infected with one (5–75%) or two symbionts (15–35%), whereas infections with three and four symbionts were only common in Q2 *B. tabaci* MED populations from Zadar (Croatia) and Bar (Montenegro) (Table 3). The two populations of *B. tabaci* MED from Croatia (Turanj-Q2, Split-Q1) had 35% of individuals that did not contain any of the tested secondary symbionts. *Rickettsia* infected around 70–75% of individuals of both Israeli *B. tabaci* MED (Q2) and MEAM1 populations. Additionally, *Arsenophonus* was fixed in Israeli MED (Q2), whereas *Hamiltonella* was fixed in MEAM1 population. Beside *Rickettsia* and *Arsenophonus*, around 20% of individuals of the Israeli MED (Q2) population were infected with *Wolbachia*. All individuals of both Israeli MED (Q2) and MEAM1 populations contained at least one secondary symbiont. In Israeli MED (Q2), 25% of individuals were infected with one, 55% with two symbionts, and 20% of individuals with three symbionts. Israeli MEAM1 population contained individuals infected with one (30%) and two (70%) secondary symbionts.

Table 2. Overview of overall infection frequencies of secondary bacterial symbionts in the three whitefly species in Southeast Europe with associated host plants.

Population Number, Location and Species			Host Plant	n	Infection of Bacterial Symbiont (%)					Reference
					H	A	R	W	C	
1	Croatia/Turanj	MED (Q2)	<i>Cucumis sativus</i>	20	25	-	30	20	30	[39]
2	Croatia/Split	MED (Q1)	<i>Euphorbia pulcherrima</i>	20	35	-	20	10	-	This study
3	Croatia/Zadar ¶	MED (Q1 and Q2)	<i>Hibiscus</i> sp.	20	95	35	60	50	-	[41]
4	Montenegro/Bar	MED (Q2)	<i>Dipladenia sanderi</i>	20	65	85	100	40	-	
5	Montenegro/Lastva Grbaljska	MED (Q1)	<i>Cucumis sativus</i>	20	30	85	40	30	-	This study
6	Montenegro/Darza	MED (Q1)	<i>Cucumis melo</i>	20	95	-	10	10	-	
7	Macedonia/Doiran	MED (Q1)	<i>Lycopersicon esculentum</i>	20	100	-	-	40	20	
8	Macedonia/Gevgelija	MED (Q1)	<i>Cucumis melo</i>	20	100	-	5	60	-	
9	Israel	MED (Q2)	<i>Gossypium hirsutum</i>	20	-	100	75	20	-	[39]
10	Israel	MEAM1	<i>Gossypium hirsutum</i>	20	100	-	70	-	-	
11	Croatia/Split	<i>T. vaporariorum</i>	<i>Sonchus oleraceus</i>	20	5	75	-	-	-	[41]; This study
12	Croatia/Split§	<i>T. vaporariorum</i>	<i>Euphorbia pulcherrima</i>	10	50	80	-	-	-	
13	Montenegro/Podgorica	<i>T. vaporariorum</i>	<i>Sonchus oleraceus</i>	20	20	95	5	10	35	
14	Macedonia/Doiran	<i>T. vaporariorum</i>	<i>Lycopersicon esculentum</i>	20	25	100	-	-	-	This study
15	Macedonia/Grciste	<i>T. vaporariorum</i>	<i>Lycopersicon esculentum</i>	20	-	100	-	-	-	
16	Serbia/Trbusani	<i>T. vaporariorum</i>	<i>Cucurbita pepo</i>	20	90	100	-	-	-	
17	Serbia/Vranjska Banja	<i>T. vaporariorum</i>	<i>Gerbera</i> sp.	20	45	100	-	-	-	
18	Serbia/Opovo	<i>T. vaporariorum</i>	<i>Lycopersicon esculentum</i>	20	70	90	-	-	-	[41]
19	Serbia/Zorka Subotica	<i>T. vaporariorum</i>	<i>Chrysanthemum</i> sp.	20	-	95	-	-	-	
20	Croatia/Brac-Supetar	<i>S. phillyreae</i>	<i>Punica granatum</i>	20	65	35	-	40	5	
21	Croatia/Brac-Pucisca	<i>S. phillyreae</i>	<i>Punica granatum</i>	20	100	25	-	20	-	
22	Croatia/Opuzen	<i>S. phillyreae</i>	<i>Punica granatum</i>	20	85	95	-	55	5	
23	Croatia/Ljuta	<i>S. phillyreae</i>	<i>Punica granatum</i>	20	85	85	-	15	70	
24	Montenegro/Bar	<i>S. phillyreae</i>	<i>Punica granatum</i>	20	90	100	-	50	-	

n, total number of individual tested per population; H, *Hamiltonella*; A, *Arsenophonus*; R, *Rickettsia*; W, *Wolbachia*; C, *Cardinium*; -, zero. ¶ Population imported by trade from Italy (Rome, region Lazio). § Population imported by trade from Slovenia (Catez ob Savi). For additional information on imported whitefly populations, please refer to Skaljic et al. (2013) [41].

Table 3. Infection frequencies of different secondary bacterial symbiont combinations in the three whitefly species in this study.

Population Number, Location, and Species		Infection Frequencies of Bacterial Symbiont Combination (%)																									
		R	H	A	W	C	RH	RA	HA	HW	HC	AW	AC	WC	RHA	RHW	RAW	RWC	HAW	HAC	HWC	AWC	RHWC	RHAW	HAWC	No Infection	
1	Croatia/Turanj MED (Q2)	10	10	-	5	15	15	-	-	-	-	-	-	5	-	-	-	5	-	-	-	-	5	-	-	-	35
2	Croatia/Split MED (Q1)	20	35	-	10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	35
3	Croatia/Zadar [¶] MED (Q1 and Q2)	-	20	-	-	-	15	-	15	5	-	-	-	-	-	25	5	-	-	-	-	-	-	15	-	-	
4	Montenegro/Bar MED (Q2)	5	-	-	-	-	5	20	-	-	-	-	-	-	30	5	10	-	-	-	-	-	-	25	-	-	
5	Montenegro/L. Grbaljska MED (Q1)	-	-	35	5	-	-	15	-	-	-	5	-	-	10	-	-	-	5	-	-	-	-	15	-	10	
6	Montenegro/Darza MED (Q1)	-	75	-	-	-	10	-	-	10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	5	
7	Macedonia/Doiran MED (Q1)	-	55	-	-	-	-	-	-	25	5	-	-	-	-	-	-	-	-	-	15	-	-	-	-	-	
8	Macedonia/Gevgelija MED (Q1)	-	40	-	-	-	-	-	-	55	-	-	-	-	-	5	-	-	-	-	-	-	-	-	-	-	
9	Israel MED (Q2)	-	-	25	-	-	-	55	-	-	-	-	-	-	-	-	20	-	-	-	-	-	-	-	-	-	
10	Israel MEAMI	-	30	-	-	-	70	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
11	Croatia/Split [§] <i>T. vaporariorum</i>	-	-	70	-	-	-	-	5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	25	
12	Croatia [§] /Split <i>T. vaporariorum</i>	-	-	30	-	-	-	-	50	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	20	
13	Montenegro/Podgorica <i>T. vaporariorum</i>	-	-	35	5	-	-	-	20	-	-	5	35	-	-	-	-	-	-	-	-	-	-	-	-	-	
14	Macedonia/Doiran <i>T. vaporariorum</i>	-	-	75	-	-	-	-	25	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
15	Macedonia/Grciste <i>T. vaporariorum</i>	-	-	100	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
16	Serbia/Trbusani <i>T. vaporariorum</i>	-	-	10	-	-	-	-	90	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
17	Serbia/Vranjska Banja <i>T. vaporariorum</i>	-	-	55	-	-	-	-	45	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
18	Serbia/Opovo <i>T. vaporariorum</i>	-	5	25	-	-	-	-	65	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	5	
19	Serbia/Z. Subotica <i>T. vaporariorum</i>	-	-	95	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	5	
20	Croatia/Brac-Supetar <i>S. phillyreae</i>	-	25	-	10	-	-	-	20	10	-	5	-	5	-	-	-	-	10	-	-	-	-	-	-	15	
21	Croatia/Brac-Pucisca <i>S. phillyreae</i>	-	60	-	-	-	-	-	20	15	-	-	-	-	-	-	-	-	5	-	-	-	-	-	-	-	
22	Croatia/Opuzen <i>S. phillyreae</i>	-	5	5	-	-	-	-	35	-	-	10	-	-	-	-	-	-	40	-	-	-	-	-	5	-	
23	Croatia/Ljuta <i>S. phillyreae</i>	-	10	5	-	-	-	-	15	-	-	-	5	-	-	-	-	-	-	50	-	5	-	-	10	-	
24	Montenegro/Bar <i>S. phillyreae</i>	-	-	5	-	-	-	-	45	-	-	5	-	-	-	-	-	-	45	-	-	-	-	-	-	-	

H, *Hamiltonella*; A, *Arsenophonus*; R, *Rickettsia*; W, *Wolbachia*; C, *Cardinium*; -, zero. [¶] Population imported by trade from Italy (Rome, region Lazio). [§] Population imported by trade from Slovenia (Catez ob Savi). For additional information on imported whitefly populations, please refer to Skaljac et al. (2013) [41].

3.1.2. *Trialeurodes vaporariorum* Infection with Secondary Bacterial Symbionts

T. vaporariorum populations were collected in Croatia, Montenegro, Macedonia, and Serbia (Figure 1 and Table S1). All populations used in this study were predominantly infected with *Hamiltonella* and *Arsenophonus* (Table 2). The only exception was *T. vaporariorum* population collected in Montenegro (Podgorica), which was additionally infected with *Rickettsia*, *Wolbachia*, and *Cardinium*. *Arsenophonus* was fixed or close to fixation in the populations collected in Montenegro, Macedonia, and Serbia, while the infection rate for *Hamiltonella* varied between 5% and 90% independently of the location (Table 2). Individuals of *T. vaporariorum* were mainly infected with one (10–100%) or two symbionts (5–90%), whereas there were no infections detected with three or four symbionts. The populations of *T. vaporariorum* from Croatia (Split) and Serbia (Opovo, Zorka Subotica) had 5–25% of individuals that did not contain any of the tested secondary symbionts.

3.1.3. *Siphoninus phillyreae* Infection with Secondary Bacterial Symbionts

S. phillyreae populations were collected in Croatia and Montenegro (Figure 1 and Table S1). All populations used in this study had high prevalence of mixed infections with *Hamiltonella*, *Arsenophonus*, *Wolbachia*, and *Cardinium* (Table 2). Interestingly, *Rickettsia* was not found in any of the tested populations. Both *Arsenophonus* and *Hamiltonella* were highly prevalent in all the tested *S. phillyreae* populations (25–100% and 65–100%, respectively). All the tested *S. phillyreae* populations were infected with *Wolbachia* (15–55%), while *Cardinium* was detected only in populations from Croatia (Supetar, Pucisca, Ljuta). Individuals of *S. phillyreae* were commonly infected with one (5–60%), two (20–50%), and three symbionts (5–55%) (Table 3). Infections with four symbionts were rare and appeared only in individuals from Croatian *S. phillyreae* populations from Opuzen and Ljuta in 5% and 10%, respectively. Overall, all *S. phillyreae* individuals contained at least one secondary symbiont.

3.2. Phylogenetic Relationships of *B. tabaci* Sequences

A total of 18 *B. tabaci* sequences were characterized from Croatia, Montenegro, and Macedonia. In this study, *B. tabaci* sequences from Israeli populations served as a reference. A phylogenetic tree was constructed from the mtCOI sequences of *B. tabaci* and divergence analyses were compared with 40 putative species previously reported [10,15,19–27] (Figure 2). Our results showed that *B. tabaci* populations from Croatia, Montenegro, Macedonia, and Israel were identified as MED (Q1 and Q2) species. Populations of *B. tabaci* MED from Macedonia (Doiran, Gevgelija), Montenegro (Darza, Lastva Grbaljska), and Croatia (Split) belonged to the Q1 group, whereas populations from Croatia (Turanj) and Montenegro (Bar) belonged to the Q2 group (Figure 2). The exception was a *B. tabaci* MED population from Croatia (Zadar), which contained individuals from both Q1 and Q2 groups (Figure 2). In addition, the *B. tabaci* MEAM1 species was identified from the Israeli population only.

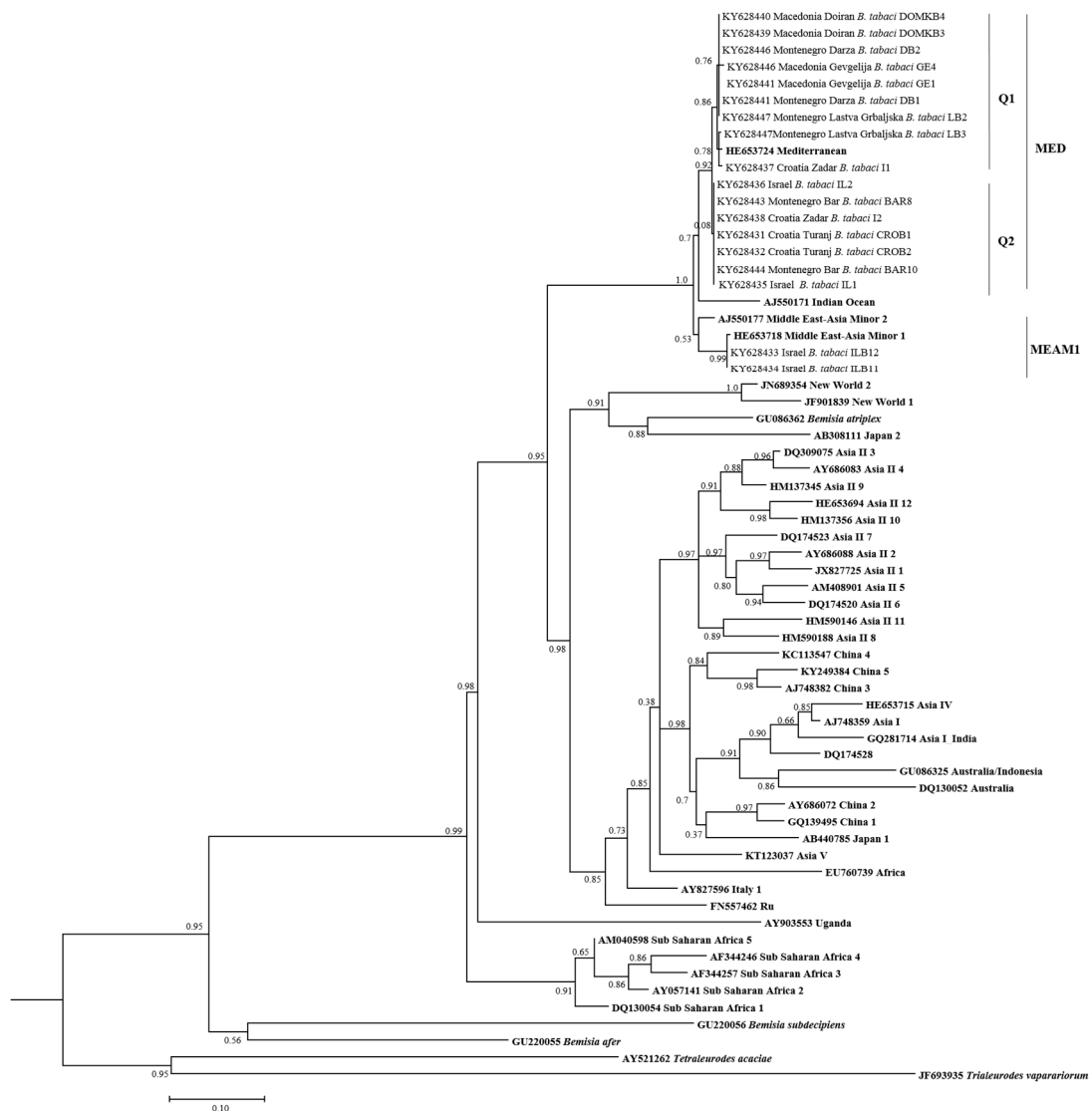


Figure 2. Phylogenetic tree based on the Bayesian analysis of 657 bp of mtCOI gene sequences of *Bemisia tabaci* populations. Sequences from GenBank are indicated in bold, while those obtained in this study are rearranged based on the geographical origin of *B. tabaci* individuals. Posterior probabilities are indicated at nodes.

3.3. Phylogenetic Analysis of Secondary Symbionts in Whiteflies

All secondary symbionts sequenced in this study were compared with bacterial reference sequences from GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>). Sequence analysis revealed that, in addition to species diversity, several bacterial subgroups/strains may exist within the different bacterial genera tested. For *Hamiltonella*, the sequences from *B. tabaci*, *T. vaporarium*, and *S. phillyreae* formed two groups (H1 and H2) (Figure 3). Clade H1 contains *Hamiltonella* from *B. tabaci* and *T. vaporarium* from Croatia, Montenegro, Serbia, and Macedonia. Group H2 contains *Hamiltonella* from *S. phillyreae* and *T. vaporarium* from Croatia and Montenegro, respectively. 23rDNA sequences of *Arsenophonus* cluster into two major groups A1 and A2 (Figure 4). The A1 group contains strains from *B. tabaci* and *T. vaporarium*. Group A2 contains a distinct *Arsenophonus* strain from *S. phillyreae* from Croatia and *T. vaporarium* from Montenegro, which differs from the A1 group. *Rickettsia* was grouped into two major clusters, R1 and R2 (Figure 5). The top clade (R1) consists of *Rickettsia* from both *B. tabaci* and *T. vaporarium* from Croatia and Montenegro. Clade R2 consists of *Rickettsia* from only *T. vaporarium*

from Montenegro. Interestingly, none of the *S. phillyreae* individuals tested positive for *Rickettsia*. In the case of *Wolbachia*, to date, 16 supergroups successively named A–Q infecting a wide range of arthropods and filarial nematodes have been reported [68]. In our analysis, the B supergroup was observed in all three species tested from Croatia, Montenegro, and Macedonia (Figure 6). 16S rDNA sequences of *Cardinium* from *B. tabaci* and *S. phillyreae* are grouped under a single clade (Figure 7).

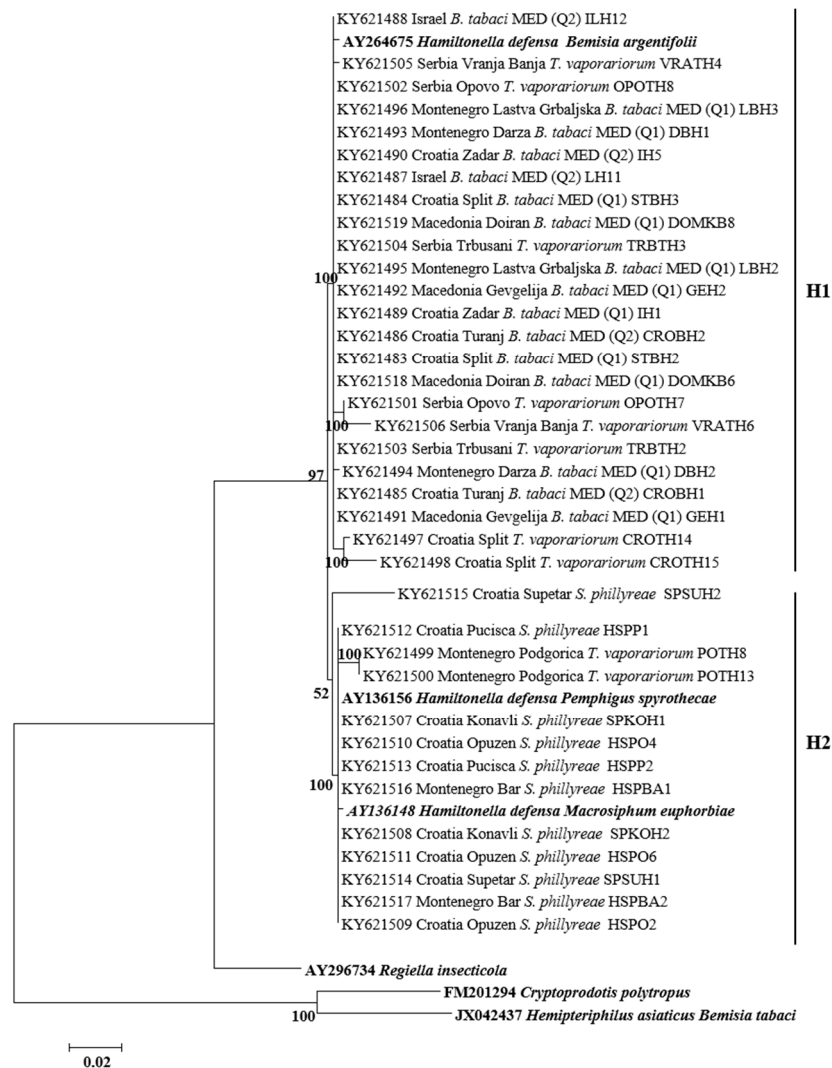


Figure 3. Molecular phylogenetic placements of *Hamiltonella* from whitefly species. The tree was constructed via Bayesian inference (BI) using an HKY substitution model. Sequences from GenBank are indicated in bold, while those obtained in this study are rearranged based on geographical origin and associated whitefly hosts, *B. tabaci*, *T. vaporariorum*, and *S. phillyreae*.

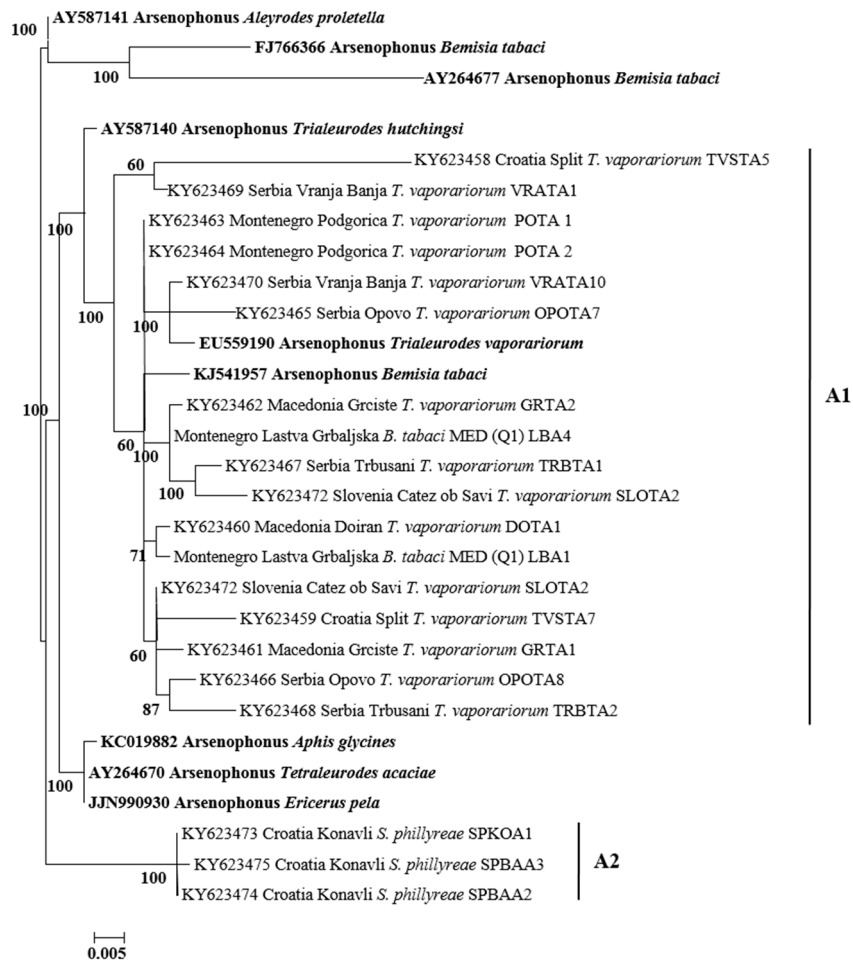


Figure 4. Molecular phylogenetic placements of *Arsenophonus* from whitefly species. The tree was constructed via Bayesian inference (BI) using a TPM1 μ f substitution model. Sequences from GenBank are indicated in bold, while those obtained in this study are rearranged based on geographical origin and associated whitefly hosts, *B. tabaci*, *T. vaporariorum*, and *S. phillyreae*.

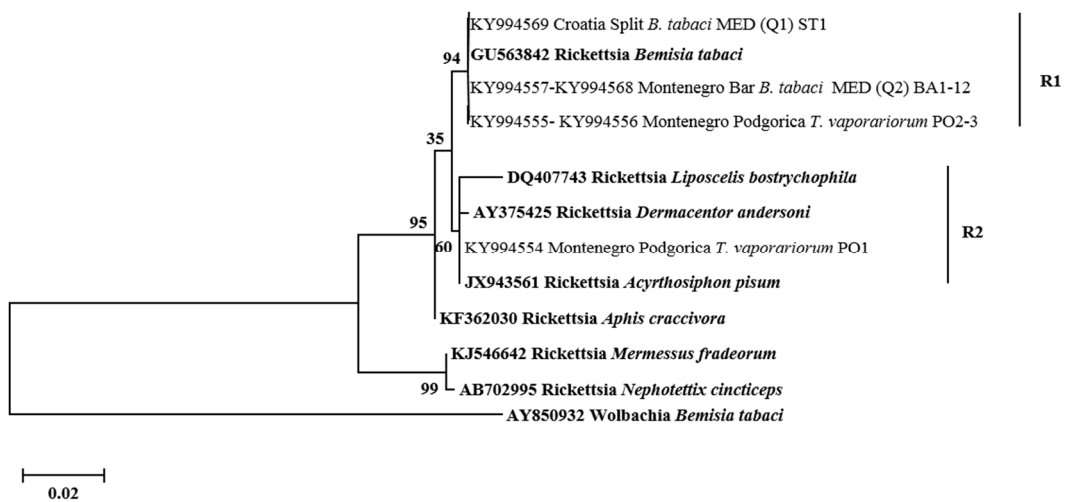


Figure 5. Molecular phylogenetic placements of *Rickettsia* from whitefly species. The tree was constructed via Bayesian inference (BI) using a GTR+G substitution model. Sequences from GenBank are indicated in bold, while those obtained in this study are rearranged based on geographical origin and associated whitefly hosts, *B. tabaci* and *T. vaporariorum*.

or parasitoids [38,73,74]. These events are necessary for the symbiont to spread to novel hosts [75]. Such acquisition of bacterial symbionts can directly impact host biology [46]. For example, the spread of *Rickettsia* in *B. tabaci* from the United States resulted in large fitness benefits and female bias in the tested whiteflies [46].

Intensive plant trade occurs between Mediterranean countries and this provides opportunities for introducing new whitefly species and symbionts in regions where they have not existed before [76]. Several studies reported the invasion of *B. tabaci* MED (Q2), previously associated only with the Middle East countries of the Mediterranean basin, into Spain, France, and Italy, where it coexisted with the native *B. tabaci* MED (Q1) [76,77]. Our study revealed the presence of both Q1 and Q2 groups of *B. tabaci* MED in the investigated region (Table 2, Figure 2). Two populations identified as Q2 were collected in Croatia (Turanj) and Montenegro (Bar). Interestingly, a *B. tabaci* MED population (Croatia-Zadar) collected on plants imported from Italy consisted of both Q1 and Q2 groups and they carried *Arsenophonus*, which was not previously detected in existing *B. tabaci* MED (Q1) populations (Table 2, Figure 2) [39].

Variations in the diversity and infection frequencies of bacterial symbionts between the three whitefly species in this study were substantial (Tables 2 and 3, Figure S1).

Our results revealed that *Arsenophonus* and *Hamiltonella* from both *B. tabaci* and *T. vaporariorum* were phylogenetically grouped in the same clades (with several exceptions; e.g., *Hamiltonella* from *T. vaporariorum* Podgorica), compared with those symbionts from *S. phillyreae*, which were placed in more distant clades (Figures 3 and 4). *B. tabaci* and *T. vaporariorum* frequently share host plants, therefore the possibility of horizontal transmission of bacterial symbionts between these two whitefly species is high [39]. *S. phillyreae* was only found on pomegranate plants, which was not a host for the other two whitefly species [40,42]. Placement of *Hamiltonella* in different clades was also reflected on its localization pattern in the whitefly hosts. *Hamiltonella* found in *B. tabaci* and *T. vaporariorum* was localized exclusively in bacteriosomes, while in *S. phillyreae* it was found to be scattered throughout the entire bodies of nymphs and adults [39,41].

Hamiltonella was present at high frequencies in the investigated species of *B. tabaci* MED (Q1) and MEAM1 (Table 2), suggesting it had a major impact on the host biology. The infection frequencies of *Hamiltonella* were in accordance with the global screening reports of bacterial symbionts in *B. tabaci* MED (Q1) and MEAM1 [33,38,40,78,79]. However, these studies showed absence of *Hamiltonella* in *B. tabaci* species such as MED (Q2, ASL), Sub-Saharan Africa 1–5, Asia II, and China 1 [33,78,80]. This was contrary to our results, where *Hamiltonella* infected 25% and 65% of individuals from MED (Q2) populations from Croatia (Turanj) and Montenegro (Bar), respectively (Table 2). Infection of *Hamiltonella* varied in *T. vaporariorum* populations (from absence of infection to high prevalence) in this study, whereas infection with this symbiont was highly prevalent in *S. phillyreae*. Surprisingly, a large study by Kapantaidaki et al. (2014) [81] did not report infection of *Hamiltonella* in *T. vaporariorum* populations from different global locations.

In general, the infection frequencies of *Arsenophonus* in *B. tabaci* varied greatly and sometimes depended on the genetic group. *B. tabaci* MEAM1 had low infection rates with *Arsenophonus* [33,38–40]. This was also similar to several *B. tabaci* (MED) populations from Southeast Europe (Q1) and China, but some MED genetic groups from Montenegro (Q1), Israel (Q2), and Burkina Faso (Q3) were highly infected with *Arsenophonus* [33,39,41,78]. This variable occurrence of *Arsenophonus* in *B. tabaci* is in agreement with our results. For example, *Arsenophonus* was fixed in *B. tabaci* MED (Q2) from Israel and prevalent in MED (Q1 and Q2) from Montenegro (Lastva Grbaljska, Bar) (85% infection rate). However, this symbiont was absent in other MED (Q1 and Q2) populations (Table 2). *Arsenophonus* was constantly present at high frequencies in *T. vaporariorum* and *S. phillyreae* in our and other studies [39–41,81].

In this study, *Rickettsia* was present in nearly all populations of *B. tabaci* MED. However, infection rates were much lower independently of Q1 and Q2 groups compared to Israeli MED and MEAM1 populations (Table 2). This suggests recent introduction of *Rickettsia* in *B. tabaci* populations in the investigated region. Other studies have shown that *Rickettsia* was present at high infection

rates in the MEAM1, MED (Q2), and China 1 groups of *B. tabaci*, while lower infection rates were observed in Asia II and the MED groups of *B. tabaci* from Burkina Faso (Q3) and China [33,38,40,78,79]. This symbiont was rarely detected in *T. vaporariorum*, and no infection was found in *S. phillyreae* populations (Table 2) [41,81].

Wolbachia is one of the most commonly present symbionts in insects [82]. In this study, nearly all *B. tabaci* and *S. phillyreae* populations were infected with *Wolbachia* (10–60% and 15–50%, respectively), while it was rarely found in *T. vaporariorum* (Table 2). Several studies reported highly diverse infection rates of *Wolbachia* in *B. tabaci* MED (Q1, Q2, ASL), while it was not observed in other genetic groups [33,40]. Bing et al. (2013) [78] found that native genetic groups of *B. tabaci* (Asia II, China 1) were highly infected with *Wolbachia*, whereas this symbiont was not detected in invasive MED and MEAM1 groups. *Cardinium*, another known reproductive manipulator, was rarely detected among populations of the three whitefly species in this study (Table 2) [39,41,43]. Other studies have revealed large differences in rates of *Cardinium* infection in populations from the *B. tabaci* species complex [33,38,40,78,79]. Phylogenetically, *Wolbachia* and *Cardinium* present in the three whitefly species from this study appeared in the same clades, together with other reported sequences of the same symbionts (Figures 6 and 7).

Mixed infections and heterogeneity among populations in our study could be a result of competition for space and resources among the symbionts or the tolerance of the host to maintain diverse bacterial community [39,83]. This can contribute to multiple events of horizontal transmissions of secondary symbionts within the same whitefly species and reproductively isolated *B. tabaci* and *T. vaporariorum* (Tables 2 and 3). In our study, horizontal transmission seems likely for *Hamiltonella*, *Arsenophonus*, *Wolbachia*, and *Cardinium* between *B. tabaci* and *T. vaporariorum*. *Rickettsia* was mostly associated with *B. tabaci*, and there were rare individuals from *T. vaporariorum* that were positive for this symbiont, which could reflect isolated events of horizontal transmission [41].

Chiel et al. (2009) [75] demonstrated horizontal transmission of *Rickettsia* in *B. tabaci* using parasitoid wasps, while transmission of *Hamiltonella* failed in this system. In addition, *Rickettsia* was successfully shown in *B. tabaci* to be transmitted via host plants [73]. There are evidences for horizontal transmission of bacterial symbionts such as *Arsenophonus*, *Cardinium*, and *Wolbachia* either among whiteflies, between whiteflies and arthropods present in the environment, or, as previously mentioned, via host plants [62,84–86]. Phylogenies of secondary symbionts are largely incongruent with those of their hosts, which is not the case with the phylogeny of primary symbionts [86]. This suggests an ancient infection of a host with primary symbiont followed by vertical transmission and consequent cospeciation with the insect host. In contrast, incongruent phylogenies suggest multiple infections and horizontal transfer between different hosts [86–88].

In the investigated bacterial community of the three whiteflies, we have found more than 20 different symbiotic combinations in a range from no infected individuals to those that were infected with four symbionts (Table 3). In populations of *B. tabaci* and *T. vaporariorum* single infections dominated (43% and 56%, respectively), while in *S. phillyreae* single (25%), double (38%), and triple (31%) infections were present in similar range. Double infections were present in 25% and 38% of individuals of *B. tabaci* and *T. vaporariorum*, respectively. Interestingly, *T. vaporariorum* in this study had no infections with three or four symbionts, while those infections existed in *B. tabaci* in 14% and 7.5%, respectively. In *B. tabaci*, *T. vaporariorum*, and *S. phillyreae*, individuals with no infections were present in 11%, 5%, and 3%, respectively. Our results were slightly different from study of Gueguen et al. (2010) [33], where infections of global populations of *B. tabaci* with two or more symbionts were common and double infections dominated (59%). Furthermore, several studies on sub-Saharan populations of *B. tabaci* report increasing number of secondary symbiont-free whiteflies [88,89]. It is likely to conclude that heterogeneity of symbiotic community of the whiteflies depends on location, population, and time [88].

5. Conclusions

The present study found substantial diversity of bacterial symbionts in the three whitefly species, especially between those that shared habitat (*B. tabaci* and *T. vaporariorum*) and in *S. phillyreae*, which was exclusively found on pomegranate plants. Dynamic processes of horizontal transmission could bring phylogenetically distant symbionts together in previously uninfected whiteflies (e.g., via parasitoids). This finding can contribute to the discovery of new functions that symbionts such as *Hamiltonella* or *Arsenophonus* may fulfill in their whitefly hosts.

Supplementary Materials: The following are available online at www.mdpi.com/2075-4450/8/4/113/s1, Figure S1: Individual and multiple infections by secondary bacterial symbionts in five *B. tabaci* and six *T. vaporariorum* populations from Croatia, Montenegro, Macedonia, and Serbia. Each square represents one population and each column represents one type of symbiont; the 20 rows per table represent the 20 individuals tested per population. Gray fields indicate positive infection for the tested symbiont. Population number, country and geographical location, species, and number of tested individuals are indicated at the top of each table. Symbionts: R—*Rickettsia*, H—*Hamiltonella*, A—*Arsenophonus*, W—*Wolbachia*, C—*Cardinium*; Table S1. The whitefly populations used in this study.

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Conflicts of Interest: The authors declare no conflict of interest.

References

1. Moran, N.A.; McCutcheon, J.P.; Nakabachi, A. Genomics and evolution of heritable bacterial symbionts. *Annu. Rev. Genet.* **2008**, *42*, 165–190. [[CrossRef](#)] [[PubMed](#)]
2. Dale, C.; Moran, N.A. Molecular interactions between bacterial symbionts and their hosts. *Cell* **2006**, *126*, 453–465. [[CrossRef](#)] [[PubMed](#)]
3. Brownlie, J.C.; Johnson, K.N. Symbiont-mediated protection in insect hosts. *Trends Microbiol.* **2009**, *17*, 348–354. [[CrossRef](#)] [[PubMed](#)]
4. Sanchez-Contreras, M.; Vlisidou, I. The diversity of insect-bacteria interactions and its applications for disease control. *Biotechnol. Genet. Eng. Rev.* **2008**, *25*, 203–244. [[CrossRef](#)]
5. Kikuchi, Y.; Hosokawa, T.; Nikoh, N.; Meng, X.Y.; Kamagata, Y.; Fukatsu, T. Host-symbiont co-speciation and reductive genome evolution in gut symbiotic bacteria of acanthosomatid stinkbugs. *BMC Biol.* **2009**, *7*, 2. [[CrossRef](#)] [[PubMed](#)]
6. Feldhaar, H. Bacterial symbionts as mediators of ecologically important traits of insect hosts. *Ecol. Entomol.* **2011**, *36*, 533–543. [[CrossRef](#)]
7. Dunbar, H.E.; Wilson, A.C.C.; Ferguson, N.R.; Moran, N.A. Aphid thermal tolerance is governed by a point mutation in bacterial symbionts. *PLoS Biol.* **2007**, *5*. [[CrossRef](#)] [[PubMed](#)]
8. Tsuchida, T.; Koga, R.; Matsumoto, S.; Fukatsu, T. Interspecific symbiont transfection confers a novel ecological trait to the recipient insect. *Biol. Lett.* **2011**, *7*, 245–248. [[CrossRef](#)] [[PubMed](#)]
9. Ferrari, J.; Vavre, F. Bacterial symbionts in insects or the story of communities affecting communities. *Phil. Trans. R. Soc. B Biol. Sci.* **2011**, *366*, 1389–1400. [[CrossRef](#)] [[PubMed](#)]
10. De Barro, P.J.; Liu, S.S.; Boykin, L.M.; Dinsdale, A.B. Bemisia tabaci: A statement of species status. *Annu. Rev. Entomol.* **2011**, *56*, 1–19. [[CrossRef](#)] [[PubMed](#)]
11. Ash Whitefly, Siphoninus Phillyreae (Haliday) (Insecta: Aleyrodidae: Aleyrodinae). Available online: <http://www.doacs.state.fl.us/pi/enpp/ento/entcirc/ent337.pdf> (accessed on 18 May 2012).

12. Brown, J.K. The bemisia tabaci complex: Genetic and phenotypic variation and relevance to tylocv-vector interactions. In *Tomato Yellow Leaf Curl Virus Disease (Management, Molecular Biology, Breeding for Resistance)*; Czosnek, H., Ed.; Springer: Houten, The Netherlands, 2007; pp. 25–57.
13. CABI. Improving Lives by Solving Problems in Agriculture and the Environment. Available online: <http://www.cabi.org> (accessed on 1 September 2017).
14. Jones, D.R. Plant viruses transmitted by whiteflies. *Eur. J. Plant Pathol.* **2003**, *109*, 195–219. [[CrossRef](#)]
15. Hu, J.; De Barro, P.; Zhao, H.; Wang, J.; Nardi, F.; Liu, S.-S. An extensive field survey combined with a phylogenetic analysis reveals rapid and widespread invasion of two alien whiteflies in china. *PLoS ONE* **2011**, *6*. [[CrossRef](#)] [[PubMed](#)]
16. Gao, R.R.; Zhang, W.P.; Wu, H.T.; Zhang, R.M.; Zhou, H.X.; Pan, H.P.; Zhang, Y.J.; Brown, J.K.; Chu, D. Population structure of the greenhouse whitefly, *trialeurodes vaporariorum* (westwood), an invasive species from the americas, 60 years after invading china. *Int. J. Mol. Sci.* **2014**, *15*, 13514–13528. [[CrossRef](#)] [[PubMed](#)]
17. Sparks, T.C.; Nauen, R. Irac: Mode of action classification and insecticide resistance management. *Pestic. Biochem. Physiol.* **2015**, *121*, 122–128. [[CrossRef](#)] [[PubMed](#)]
18. Kapantaidaki, D.E.; Sadikoglou, E.; Tsakireli, D.; Kampanis, V.; Stavrakaki, M.; Schorn, C.; Ilias, A.; Riga, M.; Tsiamis, G.; Nauen, R.; et al. Insecticide resistance in *trialeurodes vaporariorum* populations and novel diagnostics for *kdr* mutations. *Pest. Manag. Sci.* **2017**. [[CrossRef](#)] [[PubMed](#)]
19. Dinsdale, A.; Cook, L.; Riginos, C.; Buckley, Y.; Barro, P.D. Refined global analysis of *bemisia tabaci* (hemiptera: Sternorrhyncha: Aleyrodoidea: Aleyrodidae) mitochondrial cytochrome oxidase 1 to identify species level genetic boundaries. *Ann. Entomol. Soc. Am.* **2010**, *103*, 196–208. [[CrossRef](#)]
20. Firdaus, S.; Vosman, B.; Hidayati, N.; Jaya Supena, E.D.; Visser, R.G.F.; van Heusden, A.W. The *bemisia tabaci* species complex: Additions from different parts of the world. *Insect Sci.* **2013**, *20*, 723–733. [[CrossRef](#)] [[PubMed](#)]
21. Chowda-Reddy, R.; Kirankumar, M.; Seal, S.E.; Muniyappa, V.; Valand, G.B.; Govindappa, M.; Colvin, J. *Bemisia tabaci* phylogenetic groups in india and the relative transmission efficacy of tomato leaf curl bangalore virus by an indigenous and an exotic population. *J. Integr. Agric.* **2012**, *11*, 235–248. [[CrossRef](#)]
22. Hu, J.; Jiang, Z.L.; Nardi, F.; Liu, Y.Y.; Luo, X.R.; Li, H.X.; Zhang, Z.K. Members of *bemisia tabaci* (hemiptera: Aleyrodidae) cryptic species and the status of two invasive alien species in the Yunnan province (China). *J. Insect Sci. Online* **2014**, *14*. [[CrossRef](#)] [[PubMed](#)]
23. Alemandri, V.; De Barro, P.; Bejerman, N.; Arguello Caro, E.B.; Dumon, A.D.; Mattio, M.F.; Rodriguez, S.M.; Truoli, G. Species within the *bemisia tabaci* (hemiptera: Aleyrodidae) complex in soybean and bean crops in argentina. *J. Econ. Entomol.* **2012**, *105*, 48–53. [[CrossRef](#)] [[PubMed](#)]
24. Ueda, S.; Kitamura, T.; Kijima, K.; Honda, K.I.; Kanmiya, K. Distribution and molecular characterization of distinct asian populations of *bemisia tabaci* (hemiptera: Aleyrodidae) in Japan. *J. Appl. Entomol.* **2009**, *133*, 355–366. [[CrossRef](#)]
25. Parrella, G.; Scassillo, L.; Giorgini, M. Evidence for a new genetic variant in the *bemisia tabaci* species complex and the prevalence of the biotype q in Southern Italy. *J. Pest Sci.* **2012**, *85*, 227–238. [[CrossRef](#)]
26. Esterhuizen, L.L.; Mabasa, K.G.; van Heerden, S.W.; Czosnek, H.; Brown, J.K.; van Heerden, H.; Rey, M.E.C. Genetic identification of members of the *bemisia tabaci* cryptic species complex from south africa reveals native and introduced haplotypes. *J. Appl. Entomol.* **2013**, *137*, 122–135. [[CrossRef](#)]
27. Hu, J.; Zhang, X.; Jiang, Z.; Zhang, F.; Liu, Y.; Li, Z.; Zhang, Z. New putative cryptic species detection and genetic network analysis of *bemisia tabaci* (hemiptera: Aleyrodidae) in China based on mitochondrial *coi* sequences. *Mitochondrial DNA Part A DNA Mapp. Seq. Anal.* **2017**, 1–11. [[CrossRef](#)] [[PubMed](#)]
28. Tsagkarakou, A.; Mouton, L.; Kristoffersen, J.B.; Dokianakis, E.; Grispou, M.; Bourtzis, K. Population genetic structure and secondary endosymbionts of q *bemisia tabaci* (hemiptera: Aleyrodidae) from greece. *Bull. Entomol. Res.* **2012**, *102*, 353–365. [[CrossRef](#)] [[PubMed](#)]
29. Gauthier, N.; Clouet, C.; Perrakis, A.; Kapantaidaki, D.; Peterschmitt, M.; Tsagkarakou, A. Genetic structure of *bemisia tabaci* med populations from home-range countries, inferred by nuclear and cytoplasmic markers: Impact on the distribution of the insecticide resistance genes. *Pest Manag. Sci.* **2014**, *70*, 1477–1491. [[CrossRef](#)] [[PubMed](#)]
30. Chen, W.; Hasegawa, D.K.; Kaur, N.; Klot, A.; Pinheiro, P.V.; Luan, J.; Stensmyr, M.C.; Zheng, Y.; Liu, W.; Sun, H.; et al. The draft genome of whitefly *bemisia tabaci* meam1, a global crop pest, provides novel insights into virus transmission, host adaptation, and insecticide resistance. *BMC Biol.* **2016**, *14*, 110. [[CrossRef](#)] [[PubMed](#)]

31. Boykin, L.M.; Shatters, R.G., Jr.; Rosell, R.C.; McKenzie, C.L.; Bagnall, R.A.; De Barro, P.; Frohlich, D.R. Global relationships of *bemisia tabaci* (hemiptera: Aleyrodidae) revealed using bayesian analysis of mitochondrial coi DNA sequences. *Mol. Phylogenet. Evol.* **2007**, *44*, 1306–1319. [[CrossRef](#)] [[PubMed](#)]
32. Chu, D.; Wan, F.-H.; Tao, Y.-L.; Liu, G.-X.; Fan, Z.-X.; Bi, Y.-P. Genetic differentiation of *bemisia tabaci* (gennadius) (hemiptera: Aleyrodidae) biotype q based on mitochondrial DNA markers. *Insect Sci.* **2008**, *15*, 115–123. [[CrossRef](#)]
33. Gueguen, G.; Vavre, F.; Gnankine, O.; Peterschmitt, M.; Charif, D.; Chiel, E.; Gottlieb, Y.; Ghanim, M.; Zchori-Fein, E.; Fleury, F. Endosymbiont metacommunities, mtdna diversity and the evolution of the *bemisia tabaci* (hemiptera: Aleyrodidae) species complex. *Mol. Ecol.* **2010**, *19*, 4365–4376. [[CrossRef](#)] [[PubMed](#)]
34. Sloan, D.B.; Moran, N.A. Endosymbiotic bacteria as a source of carotenoids in whiteflies. *Biol. Lett.* **2012**, *8*, 986–989. [[CrossRef](#)] [[PubMed](#)]
35. Zchori-Fein, E.; Brown, J.K. Diversity of prokaryotes associated with *bemisia tabaci* (gennadius) (hemiptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* **2002**, *95*, 711–718. [[CrossRef](#)]
36. Gottlieb, Y.; Ghanim, M.; Chiel, E.; Gerling, D.; Portnoy, V.; Steinberg, S.; Tzuri, G.; Horowitz, A.R.; Belausov, E.; Mozes-Daube, N.; et al. Identification and localization of a rickettsia sp. in *bemisia tabaci* (homoptera: Aleyrodidae). *Appl. Environ. Microbiol.* **2006**, *72*, 3646–3652. [[CrossRef](#)] [[PubMed](#)]
37. Chiel, E.; Gottlieb, Y.; Zchori-Fein, E.; Mozes Daube, N.; Katzir, N.; Inbar, M. Biotype-dependent secondary symbiont communities in sympatric populations of *bemisia tabaci*. *Bull. Entomol. Res.* **2007**, *97*, 407. [[CrossRef](#)] [[PubMed](#)]
38. Gottlieb, Y.; Ghanim, M.; Gueguen, G.; Kontsedalov, S.; Vavre, F.; Fleury, F.; Zchori-Fein, E. Inherited intracellular ecosystem: Symbiotic bacteria share bacteriocytes in whiteflies. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **2008**, *22*, 2591–2599. [[CrossRef](#)] [[PubMed](#)]
39. Skaljic, M.; Zanic, K.; Ban, S.G.; Kontsedalov, S.; Ghanim, M. Co-infection and localization of secondary symbionts in two whitefly species. *BMC Microbiol.* **2010**, *10*, 142. [[CrossRef](#)] [[PubMed](#)]
40. Marubayashi, J.M.; Kliot, A.; Yuki, V.A.; Rezende, J.A.M.; Krause-Sakate, R.; Pavan, M.A.; Ghanim, M. Diversity and localization of bacterial endosymbionts from whitefly species collected in brazil. *PLoS ONE* **2014**, *9*, e108363. [[CrossRef](#)] [[PubMed](#)]
41. Skaljic, M.; Zanic, K.; Hrnčić, S.; Radonjic, S.; Perovic, T.; Ghanim, M. Diversity and localization of bacterial symbionts in three whitefly species (hemiptera: Aleyrodidae) from the east coast of the adriatic sea. *Bull. Entomol. Res.* **2013**, *103*, 48–59. [[CrossRef](#)] [[PubMed](#)]
42. Gherna, R.L.; Werren, J.H.; Weisburg, W.; Cote, R.; Woese, C.R.; Mandelco, L.; Brenner, D.J. *Arsenophonus nasoniae* gen. Nov., sp. Nov., the causative agent of the son-killer trait in the parasitic wasp *nasonia vitripennis*. *Int. J. Syst. Evol. Microbiol.* **1991**, *41*, 563–565. [[CrossRef](#)]
43. Zchori-Fein, E.; Perlman, S.J. Distribution of the bacterial symbiont *cardinium* in arthropods. *Mol. Ecol.* **2004**, *13*, 2009–2016. [[CrossRef](#)] [[PubMed](#)]
44. Werren, J.H.; Baldo, L.; Clark, M.E. *Wolbachia*: Master manipulators of invertebrate biology. *Nat. Rev. Microbiol.* **2008**, *6*, 741–751. [[CrossRef](#)] [[PubMed](#)]
45. Brumin, M.; Kontsedalov, S.; Ghanim, M. *Rickettsia* influences thermotolerance in the whitefly *bemisia tabaci* b biotype. *Insect Sci.* **2011**, *18*, 57–66. [[CrossRef](#)]
46. Himler, A.G.; Adachi-Hagimori, T.; Bergen, J.E.; Kozuch, A.; Kelly, S.E.; Tabashnik, B.E. Rapid spread of a bacterial symbiont in an invasive whitefly is driven by fitness benefits and female bias. *Science* **2011**, *332*, 254–256. [[CrossRef](#)] [[PubMed](#)]
47. Kontsedalov, S.; Zchori-Fein, E.; Chiel, E.; Gottlieb, Y.; Inbar, M.; Ghanim, M. The presence of *rickettsia* is associated with increased susceptibility of *bemisia tabaci* (homoptera: Aleyrodidae) to insecticides. *Pest. Manag. Sci.* **2008**, *64*, 789–792. [[CrossRef](#)] [[PubMed](#)]
48. Mahadav, A.; Gerling, D.; Gottlieb, Y.; Czosnek, H.; Ghanim, M. Parasitization by the wasp *eretmocerus mundus* induces transcription of genes related to immune response and symbiotic bacteria proliferation in the whitefly *bemisia tabaci*. *BMC Genom.* **2008**, *9*, 342. [[CrossRef](#)] [[PubMed](#)]
49. Allen, J.M.; Reed, D.L.; Perotti, M.A.; Braig, H.R. Evolutionary relationships of “*candidatus riesia* spp.,” endosymbiotic enterobacteriaceae living within hematophagous primate lice. *Appl. Environ. Microbiol.* **2007**, *73*, 1659–1664. [[CrossRef](#)] [[PubMed](#)]

50. Perotti, M.A.; Allen, J.M.; Reed, D.L.; Braig, H.R. Host-symbiont interactions of the primary endosymbiont of human head and body lice. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **2007**, *21*, 1058–1066. [[CrossRef](#)] [[PubMed](#)]
51. Rana, V.S.; Singh, S.T.; Priya, N.G.; Kumar, J.; Rajagopal, R. Arsenophonus groel interacts with clcuv and is localized in midgut and salivary gland of whitefly b. Tabaci. *PLoS ONE* **2012**, *7*, e42168. [[CrossRef](#)] [[PubMed](#)]
52. Gottlieb, Y.; Zchori-Fein, E.; Mozes-Daube, N.; Kontsedalov, S.; Skaljic, M.; Brumin, M.; Sobol, I.; Czosnek, H.; Vavre, F.; Fleury, F.; et al. The transmission efficiency of tomato yellow leaf curl virus by the whitefly bemisia tabaci is correlated with the presence of a specific symbiotic bacterium species. *J. Virol.* **2010**, *84*, 9310–9317. [[CrossRef](#)] [[PubMed](#)]
53. Oliver, K.M.; Russell, J.A.; Moran, N.A.; Hunter, M.S. Facultative bacterial symbionts in aphids confer resistance to parasitic wasps. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 1803–1807. [[CrossRef](#)] [[PubMed](#)]
54. Russell, J.A.; Moran, N.A. Horizontal transfer of bacterial symbionts: Heritability and fitness effects in a novel aphid host. *Appl. Environ. Microbiol.* **2005**, *71*, 7987–7994. [[CrossRef](#)] [[PubMed](#)]
55. Rao, Q.; Wang, S.; Su, Y.-L.; Bing, X.-L.; Liu, S.-S.; Wang, X.-W. Draft genome sequence of “candidatus hamiltonella defensa,” an endosymbiont of the whitefly bemisia tabaci. *J. Bacteriol.* **2012**, *194*. [[CrossRef](#)] [[PubMed](#)]
56. Su, Q.; Xie, W.; Wang, S.; Wu, Q.; Liu, B.; Fang, Y.; Xu, B.; Zhang, Y. The endosymbiont hamiltonella increases the growth rate of its host bemisia tabaci during periods of nutritional stress. *PLoS ONE* **2014**, *9*. [[CrossRef](#)] [[PubMed](#)]
57. Thao, M.L.; Baumann, L.; Hess, J.M.; Falk, B.W.; Ng, J.C.; Gullan, P.J.; Baumann, P. Phylogenetic evidence for two new insect-associated chlamydia of the family simkaniaceae. *Curr. Microbiol.* **2003**, *47*, 46–50. [[CrossRef](#)] [[PubMed](#)]
58. Everett, K.D.E.; Thao, M.; Horn, M.; Dyszynski, G.E.; Baumann, P. Novel chlamydiae in whiteflies and scale insects: Endosymbionts “candidatus fritschea bemisiae” strain falk and “candidatus fritschea eriococci” strain elm. *Int. J. Syst. Evol. Microbiol.* **2005**, *55*, 1581–1587. [[CrossRef](#)] [[PubMed](#)]
59. De Barro, P.J.; Scott, K.D.; Graham, G.C.; Lange, C.L.; Schutze, M.K. Isolation and characterization of microsatellite loci in bemisia tabaci. *Mol. Ecol. Notes* **2003**, *3*, 40–43. [[CrossRef](#)]
60. Frohlich, D.R.; Torres-Jerez, I.I.; Bedford, I.D.; Markham, P.G.; Brown, J.K. A phylogeographical analysis of the bemisia tabaci species complex based on mitochondrial DNA markers. *Mol. Ecol.* **1999**, *8*, 1683–1691. [[CrossRef](#)] [[PubMed](#)]
61. Thao, M.L.; Baumann, P. Evidence for multiple acquisition of arsenophonus by whitefly species (sternorrhyncha: Aleyrodidae). *Curr. Microbiol.* **2004**, *48*, 140–144. [[CrossRef](#)] [[PubMed](#)]
62. Weeks, A.; Breeuwer, J. A new bacterium from the cytophaga-flavobacterium—bacteroides phylum that causes sex-ratio distortion. In *Insect Symbiosis*; CRC Press: Boca Raton, FL, USA, 2003; pp. 165–176.
63. Altschul, S.F.; Gish, W.; Miller, W.; Myers, E.W.; Lipman, D.J. Basic local alignment search tool. *J. Mol. Biol.* **1990**, *215*, 403–410. [[CrossRef](#)]
64. Katoh, K.; Standley, D.M. Mafft multiple sequence alignment software version 7: Improvements in performance and usability. *Mol. Biol. Evol.* **2013**, *30*, 772–780. [[CrossRef](#)] [[PubMed](#)]
65. Miller, M.A.; Pfeiffer, W.; Schwartz, T. Creating the cipes science gateway for inference of large phylogenetic trees. In *Gateway Computing Environments Workshop (GCE), 2010*; IEEE: New Orleans, LA, USA, 2010; pp. 1–8.
66. Ronquist, F.; Huelsenbeck, J.P. Mrbayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **2003**, *19*, 1572–1574. [[CrossRef](#)] [[PubMed](#)]
67. Tamura, K.; Stecher, G.; Peterson, D.; Filipski, A.; Kumar, S. Mega6: Molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.* **2013**, *30*, 2725–2729. [[CrossRef](#)] [[PubMed](#)]
68. Glowska, E.; Dragun-Damian, A.; Dabert, M.; Gerth, M. New wolbachia supergroups detected in quill mites (acar: Symbiophila). *Infect. Genet. Evol.* **2015**, *30*, 140–146. [[CrossRef](#)] [[PubMed](#)]
69. Ghanim, M.; Kontsedalov, S. Susceptibility to insecticides in the q biotype of bemisia tabaci is correlated with bacterial symbiont densities. *Pest. Manag. Sci.* **2009**, *65*, 939–942. [[CrossRef](#)] [[PubMed](#)]
70. Degnan, P.H.; Yu, Y.; Sisneros, N.; Wing, R.A.; Moran, N.A. Hamiltonella defensa, genome evolution of protective bacterial endosymbiont from pathogenic ancestors. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 9063–9068. [[CrossRef](#)] [[PubMed](#)]
71. Singh, S.T.; Priya, N.G.; Kumar, J.; Rana, V.S.; Ellango, R.; Joshi, A. Diversity and phylogenetic analysis of endosymbiotic bacteria from field caught bemisia tabaci from different locations of north india based on 16s rdna library screening. *Infect. Genet. Evol.* **2012**, *12*, 411–419. [[CrossRef](#)] [[PubMed](#)]

72. Xie, W.; Meng, Q.S.; Wu, Q.J.; Wang, S.L.; Yang, X.; Yang, N.N.; Li, R.M.; Jiao, X.G.; Pan, H.P.; Liu, B.M.; et al. Pyrosequencing the bemisia tabaci transcriptome reveals a highly diverse bacterial community and a robust system for insecticide resistance. *PLoS ONE* **2012**, *7*. [[CrossRef](#)] [[PubMed](#)]
73. Caspi-Fluger, A.; Inbar, M.; Mozes-Daube, N.; Katzir, N.; Portnoy, V.; Belausov, E.; Hunter, M.S.; Zchori-Fein, E. Horizontal transmission of the insect symbiont rickettsia is plant-mediated. *Proc. R. Soc. B Biol. Sci.* **2012**, *279*, 1791–1796. [[CrossRef](#)] [[PubMed](#)]
74. Gonella, E.; Pajoro, M.; Marzorati, M.; Crotti, E.; Mandrioli, M.; Pontini, M.; Bulgari, D.; Negri, I.; Sacchi, L.; Chouaia, B.; et al. Plant-mediated interspecific horizontal transmission of an intracellular symbiont in insects. *Sci. Rep.* **2015**, *5*. [[CrossRef](#)] [[PubMed](#)]
75. Chiel, E.; Zchori-Fein, E.; Inbar, M.; Gottlieb, Y.; Adachi-Hagimori, T.; Kelly, S.E.; Aspken, M.K.; Hunter, M.S. Almost there: Transmission routes of bacterial symbionts between trophic levels. *PLoS ONE* **2009**, *4*. [[CrossRef](#)] [[PubMed](#)]
76. Terraz, G.; Gueguen, G.; Arno, J.; Fleury, F.; Mouton, L. Nuclear and cytoplasmic differentiation among mediterranean populations of bemisia tabaci: Testing the biological relevance of cytotypes. *Pest. Manag. Sci.* **2014**, *70*, 1503–1513. [[CrossRef](#)] [[PubMed](#)]
77. Parrella, G.; Nappo, A.G.; Manco, E.; Greco, B.; Giorgini, M. Invasion of the q2 mitochondrial variant of mediterranean bemisia tabaci in southern italy: Possible role of bacterial endosymbionts. *Pest. Manag. Sci.* **2014**, *70*, 1514–1523. [[CrossRef](#)] [[PubMed](#)]
78. Bing, X.; Ruan, Y.; Rao, Q.; Wang, X.; Liu, S. Diversity of secondary endosymbionts among different putative species of the whitefly bemisia tabaci. *Insect Sci.* **2013**, *20*, 194–206. [[CrossRef](#)] [[PubMed](#)]
79. Chu, D.; Gao, C.S.; De Barro, P.; Zhang, Y.J.; Wan, F.H.; Khan, I.A. Further insights into the strange role of bacterial endosymbionts in whitefly, bemisia tabaci: Comparison of secondary symbionts from biotypes b and q in china. *Bull. Entomol. Res.* **2011**, *101*, 477–486. [[CrossRef](#)] [[PubMed](#)]
80. Ghosh, S.; Bouvaine, S.; Maruthi, M. Prevalence and genetic diversity of endosymbiotic bacteria infecting cassava whiteflies in africa. *BMC Microbiol.* **2015**, *15*. [[CrossRef](#)] [[PubMed](#)]
81. Kapantaidaki, D.E.; Ovcarenko, I.; Fytrou, N.; Knott, K.E.; Bourtzis, K.; Tsagkarakou, A. Low levels of mitochondrial DNA and symbiont diversity in the worldwide agricultural pest, the greenhouse whitefly trialeurodes vaporariorum (hemiptera: Aleyrodidae). *J. Hered.* **2014**, *106*, 80–92. [[CrossRef](#)] [[PubMed](#)]
82. Correa, C.C.; Ballard, J. Wolbachia associations with insects: Winning or losing against a master manipulator. *Front. Ecol. Evol.* **2016**, *3*. [[CrossRef](#)]
83. Vautrin, E.; Vavre, F. Interactions between vertically transmitted symbionts: Cooperation or conflict? *Trends Microbiol.* **2009**, *17*, 95–99. [[CrossRef](#)] [[PubMed](#)]
84. Sintupachee, S.; Milne, J.R.; Poonchaisri, S.; Baimai, V.; Kittayapong, P. Closely related wolbachia strains within the pumpkin arthropod community and the potential for horizontal transmission via the plant. *Microb. Ecol.* **2006**, *51*, 294–301. [[CrossRef](#)] [[PubMed](#)]
85. Mouton, L.; Thierry, M.; Henri, H.; Baudin, R.; Gnankine, O.; Reynaud, B.; Zchori-Fein, E.; Becker, N.; Fleury, F.; Delatte, H. Evidence of diversity and recombination in arsenophonus symbionts of the bemisia tabaci species complex. *BMC Microbiol.* **2012**, *12* (Suppl. 1). [[CrossRef](#)] [[PubMed](#)]
86. Vavre, F.; Fleury, F.; Lepetit, D.; Fouillet, P.; Bouletreau, M. Phylogenetic evidence for horizontal transmission of wolbachia in host-parasitoid associations. *Mol. Biol. Evol.* **1999**, *16*, 1711–1723. [[CrossRef](#)] [[PubMed](#)]
87. Ahmed, M.Z.; De Barro, P.J.; Ren, S.X.; Greeff, J.M.; Qiu, B.L. Evidence for horizontal transmission of secondary endosymbionts in the bemisia tabaci cryptic species complex. *PLoS ONE* **2013**, *8*. [[CrossRef](#)] [[PubMed](#)]
88. Tajebe, L.S.; Guastella, D.; Cavalieri, V.; Kelly, S.E.; Hunter, M.S.; Lund, O.S.; Legg, J.P.; Rapisarda, C. Diversity of symbiotic bacteria associated with bemisia tabaci (homoptera: Aleyrodidae) in cassava mosaic disease pandemic areas of tanzania. *Ann. Appl. Biol.* **2015**, *166*, 297–310. [[CrossRef](#)]
89. Ghosh, S.; Bouvaine, S.; Richardson, S.C.W.; Ghanim, M.; Maruthi, M.N. Fitness costs associated with infections of secondary endosymbionts in the cassava whitefly species bemisia tabaci. *J. Pest Sci.* **2017**. [[CrossRef](#)]

