



REVIEW

Recent advances in the biology and therapy of medullary thyroid carcinoma [version 1; referees: 2 approved]

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Abstract

Medullary thyroid cancer (MTC) is a relatively uncommon yet prognostically significant thyroid cancer. Several recent advances in the biology and current or potential treatment of MTC are notable. These include a new understanding of the developmental biology of the thyroid C cell, which heretofore was thought to develop from the neural crest. RET, encoded by the most common driver gene in MTC, has been shown to be a dual function kinase, thus expanding its potential substrate repertoire. Promising new therapeutic developments are occurring; many have recently progressed to clinical development. There are new insights into RET inhibitor therapy for MTC. New strategies are being developed to inhibit the RAS proteins, which are potential therapeutic targets in MTC. Potential emerging immunotherapies for MTC are discussed. However, gaps in our knowledge of the basic biology of the C cell, its transformation to MTC, and the mechanisms of resistance to therapy impede progress; further research in these areas would have a substantial impact on the field.

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Introduction

Medullary thyroid cancer (MTC) is a relatively uncommon (about 1,400 new cases per year in the US) cancer of the thyroid C cells, yet accounts for a substantial fraction of thyroid cancer mortality. Excellent comprehensive reviews of the biology, genetics, and management of MTC have been published¹⁻⁹. Briefly, in 25% of MTC cases, the disease is hereditary, occurring as part of the MEN 2 syndromes (MEN 2A, MEN 2B, and familial medullary thyroid cancer) due to germline activating mutations of the *RET* receptor tyrosine kinase gene. *RET* is also mutated in about 50% of sporadic cases of MTC; in both hereditary and sporadic cases, specific mutations are correlated with phenotype and prognosis^{4,6}. MTC is resistant to cytotoxic chemotherapy. *RET* inhibitors have provided significant clinical benefit; two *RET* inhibitors, vandetanib and cabozantinib, are US Food and Drug Administration (FDA)-approved for the treatment of advanced MTC¹⁰⁻¹². In this short and necessarily selective review, I discuss some recent advances in MTC and in related areas that are likely to affect MTC. I have tried to point out some other areas in which MTC research might be productively focused.

Biology of medullary thyroid cancer

Medullary thyroid cancer is not derived from the neural crest

For almost half a century, thyroid C cells have been thought to be derived from the neural crest. This hypothesis was based largely on chick-quail xenotransplantation studies by Le Douarin *et al.*^{13,14}, which revealed that avian calcitonin-producing ultimobranchial bodies were derived from the neural crest. This model was extrapolated to mammalian species, in which the ultimobranchial bodies are derived from the fourth pharyngeal pouch and invade the thyroid during development, differentiating into C cells. The neural crest derivation of mammalian ultimobranchial bodies and thyroid C cells was challenged by Kameda *et al.*¹⁵, who showed by cell-lineage tracing that mouse ultimobranchial bodies were not derived from neural crest cells. Recently, Johansson *et al.*¹⁶ used an elegant lineage-tracing scheme to confirm that mouse ultimobranchial bodies and thyroid C cells are not derived from neural crest cells but rather from pharyngeal endoderm. Nevertheless, C-cell development is significantly influenced by neural crest-derived cells¹⁷. These findings may have an impact on our understanding of the biology of mixed histology tumors, as discussed by DeLellis and Mangray⁸. While many of these are likely to be “collision tumors”, the endodermal origin of both C cells and follicular cells strengthens the possibility that some of these tumors arise from a single progenitor cell.

Understanding RET

A potential dramatic change in our understanding of *RET* kinase function is underway. Bagheri-Yarmand *et al.*¹⁸ reported that *RET* is not only a membrane-bound receptor tyrosine kinase, but also is localized to the nucleus. There, one of its functions is to inhibit the pro-apoptotic transcription factor ATF4. This antagonism of ATF4 activity provides a potential mechanism for the anti-apoptotic function of *RET*¹⁹. Remarkably, Bagheri-Yarmand *et al.*¹⁸ showed that *RET* appears to accomplish this

inhibition by *threonine* phosphorylation of ATF4, marking it for proteolytic degradation; thus, *RET* is a dual specificity tyrosine-threonine kinase. Plaza-Menacho *et al.*²⁰ recently confirmed and extended this finding of dual specificity, showing that *RET* is autophosphorylated at serine 909 in the activation domain. While they show that this phosphorylation has no effect on *RET* enzymatic activity *in vitro*, they showed that it affects *RET* signaling in an *in vivo* model. In the *Drosophila* Ret2B model of MEN 2B, *dRet* M955T mutation, analogous to human *RET* M918T, results in a rough eye phenotype²¹. When a further *dRet* S946A mutation (analogous to human S909A) was introduced, the rough eye phenotype was rescued. Plaza-Menacho *et al.*²⁰ speculate that *RET* pS909 may function in docking of downstream effector proteins. This expanded specificity has the potential to expand the universe of substrates of *RET*; in any case, important *RET* effector substrate proteins still await identification.

Under-researched area: biology of C-cell transformation

Thus far, relatively little is known regarding the molecular steps of C-cell transformation leading to MTC²². This is due in large part to the rarity of C cells in the normal human and mouse thyroid, limiting their study to *in situ* methods. Cell-sorting methods applied to thyroid glands²³⁻²⁵ suggest that isolation of small numbers of normal or early dysplastic C cells may be feasible; functional and “omics” studies on such cell populations could sharply address our dearth of knowledge in this important area.

Genomics of medullary thyroid cancer

MTC cases typically have few mutations²⁶. As mentioned, about 50% of sporadic cases have an activating point mutation in the *RET* gene. The only other common mutations identified in sporadic MTC are *KRAS* and *HRAS*; *NRAS* mutations are infrequent²⁶⁻³¹. Recent reports have described *ALK* and *RET* gene rearrangements in sporadic MTC^{32,33}. While these rearrangements are infrequent, they indicate the potential for targeted therapy in specific cases; perhaps even more importantly, they suggest the important role that these and other rearrangements may play in MTC. Rearrangements would not likely have been detected in whole exome sequencing studies of MTC²⁶ and can play a dominant driver role in cancer development³⁴⁻³⁶.

Treatment of medullary thyroid cancer

RET inhibitors: fulfilling the promise

Introduction. Vandetanib and cabozantinib have changed the face of MTC systemic therapy, offering an effective targeted treatment. Nevertheless, both drugs are effective in less than half of patients with MTC and have significant adverse effects, and (even in responsive cases) resistance develops, typically within a few years.

Adverse effects. It is not clear which effects are off-target (non-*RET*) and which are on-target (due to *RET* inhibition). Since both vandetanib and cabozantinib are multikinase inhibitors, there is some thought that more specific inhibitors may alleviate these effects. However, it is necessary to consider the possibility that the inhibition of some of the other kinases

targeted by these drugs contributes to their efficacy. The response of *RET* wild-type (wt) tumors may suggest that this occurs; the ability of both vandetanib and cabozantinib to inhibit VEGFR2 suggests it as a potential candidate target, in which case VEGFR2-dependent adverse effects may be inseparable from efficacy.

Resistance. As mentioned above, MTC commonly exhibits intrinsic resistance to RET inhibitors, and even those cases that are initially sensitive to RET inhibitors almost always develop acquired resistance, resulting in disease progression. A critical need in the field is an understanding of the mechanisms of intrinsic and acquired resistance to RET inhibitors in MTC. Numerous mechanisms for resistance to tyrosine kinase inhibitors (TKIs) have been described, including feedback signaling response, secondary mutations, or gene amplification in the TKI, bypass activation of other signaling or survival pathways, and pharmacokinetic/pharmacodynamic (PK/PD) issues; several resistance mechanisms can coexist in a single resistant tumor³⁷. Mathematical modeling of resistance in other cancers has shown that even apparently acquired resistance may exist intrinsically as subclonal cell populations within the tumor, and such resistant cells can be selected by treatment^{38–40}. That an understanding of the mechanisms of resistance can direct subsequent effective therapeutic strategies is not only intuitive; it has been demonstrated dramatically in many instances for TKIs and other targeted therapeutics^{41–46}.

Some information regarding resistance to vandetanib and cabozantinib is available, but much further study is urgently needed. Both inhibitors perform poorly on *RET* mutants with V804M or V804L mutations^{47,48}; these mutations are orthologous to the *BCR-ABL* T315I “gatekeeper” mutation which confers resistance to imatinib in chronic myelogenous leukemia⁴⁹. A recent article identified *RET* I788N as another mutation conferring resistance to cabozantinib and vandetanib⁵⁰; I788 is an ortholog of the V654 residue in *c-KIT*, mutation of which confers resistance to imatinib⁵¹. PK studies of vandetanib and cabozantinib have been extensive. There is limited bioavailability *in vivo* and this is due in part to substantial plasma protein binding (92–94% for vandetanib⁵² and more than 99.7% for cabozantinib^{53,54}). This suggests that resistance may be due in part to limited drug exposure within the tumor. Unfortunately, PD studies in patients treated with these compounds, pivotal for our understanding, have not been reported.

Remarkably, changes in MTC seen on progression after TKI therapy have not yet been reported. Such studies have been uniquely informative in identifying mechanisms of resistance to other therapeutics^{41–46}. In such studies, progression biopsies are compared with pretreatment samples by using next-generation sequencing to look for secondary mutations, amplifications, and other genomic changes, transcriptomic studies such as RNA-seq, or proteomic studies to look for potential gene expression changes and pathway activation that may account for acquired resistance. Given the experience with disease progression on current TKIs and the facile availability of these technologies, one would hope that these elucidating correlative studies soon become standard.

Cabozantinib: who benefits? In the Efficacy of XL184 (Cabozantinib) in Advanced Medullary Thyroid Cancer (EXAM) phase 3 clinical trial of cabozantinib¹², MTC patients receiving the drug had a significantly longer progression-free survival (PFS) than patients receiving the placebo control (median PFS 49 versus 17 weeks; hazard ratio [HR] = 0.28, $p < 0.0001$). Recent correlative subgroup analysis indicated that patients with a *RET* M918T mutation significantly benefited from cabozantinib treatment^{55,56}. All other mutational subgroups (*RET* non-M918T mutation, RAS mutation, RAS wt, *RET* and RAS wt, *RET* unknown status, and *RET* mutation of unknown significance) also exhibited decreased HR in response to cabozantinib treatment; the PFS differences in these subgroups did not reach statistical significance, although in some cases this was likely due to the small size of the subgroups. As the authors note, the effects of cabozantinib in these smaller subgroups will need to be resolved in future prospective studies. Notably, the M918T subgroup was the only group to exhibit a significant increase in overall survival (OS) (44.3 versus 18.9 months; HR = 0.60, $p < 0.03$). Thus, it is clear that cabozantinib is effective for MTC patients with the *RET* M918T mutation, but in no case is it yet possible to exclude patients from cabozantinib treatment on the basis of mutation status, especially given the paucity of effective MTC treatments other than RET multikinase inhibitors.

If, as suggested by the correlative data in the EXAM trial, MTC with *RET* M918T mutations derives more clinical benefit from cabozantinib than do other MTC subgroups, what might be the mechanism? Here, our understanding is impeded by our limited knowledge of the biology of *RET* mutations. A few transcriptomic studies have compared MTC cases with *RET* M918T mutations versus other *RET* mutations^{57–60}, but further work is necessary, since some of the studies were underpowered and consensus among other studies was lacking. One potentially important difference between M918T and other *RET* mutations is in substrate specificity. RET M918T has a relaxed and altered substrate preference⁶¹. This is due to destabilization of the substrate recognition domain, which also leads to increased kinase activity^{62–64}. Speculatively, this altered kinase activity could lead to greater oncogene addiction, which would result in augmented clinical activity of RET inhibitors.

By what mechanism might cabozantinib provide benefit to MTC cases without RET mutations? The potential for other kinase targets of cabozantinib (including VEGFR2, MET, TIE2, RON, and others)⁶⁵ to contribute to angiogenesis⁶⁵, malignant transformation⁶⁶, or other MTC functions has been discussed⁵⁵. The possibility also exists that wt RET is important in the biology of MTC.

New RET inhibitors. Several new RET inhibitors have been reported⁶⁷. Some of these compounds have been designed for increased potency, increased specificity (especially as compared with KDR inhibition), improved PKs, or the ability to inhibit RET gatekeeper mutants. The following four interesting compounds are currently in clinical trials.

Alectinib (Roche) is an ALK inhibitor approved for ALK-rearranged non-small cell lung cancer. It is highly specific, inhibiting little other than ALK and RET (concentration resulting in inhibition of 50% of activity [IC_{50}] for RET = 4.8 nM)⁶⁸. At somewhat higher concentrations, alectinib also effectively targets the RET gatekeeper mutants, V804L (32 nM) and V804M (53 nM). While alectinib effectively reduced phospho-RET levels in MTC cell (TT) xenografts, it had little effect on MTC xenograft growth⁶⁸. This suggests that another target, in addition to RET, may serve to maintain growth in MTC cells. Whether this intrinsic resistance will be seen clinically in MTC is unknown but should be determined shortly; a phase 1 trial for alectinib in MTC and other RET-driven thyroid and lung cancers (NCT03131206) is ongoing.

BLU-667 (Blueprint Medicines) inhibits RET at subnanomolar concentrations⁶⁹. It efficiently inhibits the RET V804M gatekeeper mutant at similar concentrations and has 70-fold specificity versus KDR. It is in a phase 1 clinical trial (NCT03037385) for cancers with activating *RET* gene alterations, including MTC.

LOXO-292 (Loxo Oncology) inhibits RET at nanomolar concentrations⁷⁰. It also inhibits the RET gatekeeper mutant at similar concentrations. Importantly, LOXO-292 has been shown to be very specific for RET in kinase assays. Its efficacy in inhibiting the growth of MTC cell xenografts further confirms that targeting RET alone is a viable therapeutic strategy in MTC. LOXO-292 is in a phase 1 clinical trial (NCT03157128) for cancers with activating *RET* gene alterations, including MTC.

RXDX-105 (Ignyta) was originally developed as a BRAF inhibitor (IC_{50} = 14 nM)⁷¹. It also efficiently inhibits wt RET (IC_{50} = 1.5 nM)⁷¹. This suggests the potential for RXDX-105 to act as a combination treatment, targeting RET and the RAF-MEK-ERK pathway, a combination which may be synergistic in MTC cells (72 and unpublished results). RXDX-105 is reported to have 96–100% bioavailability⁷¹, a substantial improvement over other RET inhibitors. However, while RXDX-105 does not inhibit KDR, it is a broad-specificity inhibitor⁷¹, so it may have significant adverse effects. Moreover, since it does not have activity against CRAF, one can envision “paradoxical activation” of the RAF-MEK-ERK pathway^{73–75}. Nevertheless, in an ongoing phase 1 trial of RXDX-105 for RET-driven solid tumors (NCT01877811), a partial response was noted in a lung adenocarcinoma patient with a RET gene rearrangement⁷⁶.

RAS as a therapeutic target in medullary thyroid cancer

The discovery of RAS mutations in MTC^{26–31} highlighted the possibility of targeting RAS therapeutically in some cases of MTC; since RET signaling functions in part through RAS activation⁷⁷, RAS inhibition also may be a potential therapeutic strategy in MTC with RET mutations. However, RAS has a long history as an intractable target^{78–80}. It was realized early that targeting RAS-guanosine triphosphate (RAS-GTP) interaction would be difficult since GTP has picomolar affinity for RAS while GTP is present at millimolar concentrations in cells⁸¹. A substantial effort to block RAS attachment to the cell membrane

by blocking farnesyltransferase was unsuccessful because of alternative cellular mechanisms (geranylgeranylation) of membrane attachment^{78,79,82–84}. Blocking RAS downstream signaling has been attractive but has commonly suffered from ineffectiveness of blocking one pathway and toxicity of blocking multiple pathways⁸⁵. In recent years, with advances in drug and protein biochemistry, there has been renewed interest and preclinical success in directly inhibiting RAS via a multitude of novel strategies, including small-molecule binding, inhibition of protein interactions, and antisense approaches^{86–111}. A review of this rapidly advancing field, other than the several examples below, is outside the scope of this review, and the reader is directed to the cited references. In addition, it must be noted that some of the promising therapeutic approaches target specific RAS mutations; at least one of these targeted mutations, KRAS G12C^{87,91,92,95,100}, is rarely found in MTC³¹.

PDE6 δ inhibitors. The prenyl-binding chaperone protein PDE6 δ is necessary for RAS membrane localization¹¹¹. Early PDE6 δ inhibitors, including deltarasin and deltatizone, had low nanomolar affinity for PDE6 δ but required micromolar concentrations for RAS inhibitory activity in intact cells^{88,99}. This discrepancy was recently reported to be due to inhibitor release mediated by the release factor Arl2¹⁰⁴. Newly designed PDE6 δ inhibitors with subnanomolar affinity have been shown to be effective in cells and can block KRAS-dependent cell proliferation¹⁰⁴. A third generation of highly specific PDE6 δ inhibitors covalently binds the active site of PDE6 δ , rendering these compounds refractory to Arl2-mediated release¹⁰⁵.

Farnesyltransferase inhibition. As mentioned above, the farnesyltransferase inhibitors (FTIs) failed because of alternative prenylation of RAS proteins. However, *HRAS*, the most commonly mutated RAS gene in MTC, cannot employ this alternative prenylation strategy and is dependent upon farnesyltransferase for activity. Thus, *HRAS*-driven cancers should be sensitive to FTIs. This hypothesis is being tested in a phase 2 study of the FTI tipifarnib for *HRAS* mutant thyroid or head and neck cancers (NCT02383927). Interestingly, a phase 1 clinical trial of a combination of the FTI tipifarnib and the multikinase (including RET) inhibitor sorafenib showed significant activity for MTC with or without RET mutations; *HRAS* mutation status was not evaluated¹¹². While the response rate was greater than that seen in another trial employing sorafenib alone¹¹³, the trials were not powered for statistical comparison. If this combination is effective, it may be due to additional targets of tipifarnib¹¹², or to synergistic pathway inhibition by tipifarnib and sorafenib, similar to the synergy reported in studies of sorafenib and the MEK inhibitor selumetinib in MTC cells *in vitro*⁷².

Farnesyltransferase-mediated delivery of covalent RAS inhibitors. In this recently reported, very novel approach, endogenous farnesyltransferase activity was hijacked to mislocalize KRAS by blocking normal prenylation mediated by both farnesyltransferase and geranylgeranyltransferase¹⁰⁶. Thus, a farnesyltransferase neosubstrate was designed that binds covalently to the RAS CAAX moiety yet does not promote membrane

localization. In cell culture, treatment with this neosubstrate resulted in decreased RAS signaling. The effects were modest, and specificity for transformed cells and efficacy *in vivo* were not addressed. Nevertheless, this report represents the first step in the development of a promising strategy to overcome the ability of KRAS to evade the therapeutic effects of prenylation inhibitors.

Inhibition of RAS-effector interaction. RAS activates its downstream effectors by interaction with their RAS-binding domains (RBDs). Many of the small-molecule RAS direct inhibitors in development appear to work by disrupting these interactions^{97,98,103}. Notably, rigosertib, a kinase inhibitor already in phase 3 clinical development for myelodysplastic syndrome, was recently shown to be a RAS mimetic, binding to the RBDs of RAF, PI3K, and RAL-GEF, preventing interaction with RAS and pathway activation⁹⁴. Rigosertib was shown to inhibit tumorigenesis in several RAS-driven xenograft models.

Antisense oligonucleotides. While therapeutic antisense technology has been hampered by delivery issues, a recent article showed that KRAS expression can be efficiently silenced *in vivo* by systemic treatment with modified 2',4'-constrained ethyl antisense oligonucleotides¹⁰⁷; toxicity was not seen, and xenograft growth inhibition was demonstrated. If successful in further studies, such an approach may also be applicable to KRAS, HRAS, and RET mutations in MTC.

Immunotherapy for medullary thyroid cancer

Exciting recent advances have been made in immunotherapy for a variety of cancers^{114–118}. The current state of immunotherapy for thyroid cancer, including MTC, was reviewed recently¹¹⁹. While effective immunotherapy has not yet reached MTC, there are several points of interest.

Immune checkpoint therapy. It is not clear whether immune checkpoint therapy has promise in MTC. As noted, MTC has a low mutation burden²⁶. MTC also has very low expression of the immune checkpoint ligand PD-L1¹²⁰; both of these correlate with poor response to checkpoint blockade^{118,121,122}. Nevertheless, in early preclinical and clinical studies, MTC cell or calcitonin vaccines elicited a T-cell response, with apparent antitumor activity^{123–126}, suggesting the possibility that checkpoint blockade, perhaps in combination with a vaccine, may be effective. A phase 2 clinical trial (NCT03072160) employing the PD-1 checkpoint-blocking antibody pembrolizumab for MTC will begin to explore this potential therapeutic strategy.

Adoptive cell therapies: tumor-infiltrating lymphocyte, T-cell receptor, and chimeric antigen receptor transfer^{127–130}. The low mutation burden of MTC renders these exciting strategies somewhat challenging. Analysis of the most common RET mutations (M918T, C634 mutations) using NetMHC 3.4 software¹³¹ failed to identify neoepitopes for avid major histocompatibility complex (MHC) binding in the most common serotypes (unpublished data). Wild-type (and mutated) RET have several epitopes predicted to bind avidly to common MHC alleles; while these could be targeted as potential tumor-associated antigens, the

expression of RET in normal cells throughout the body^{132,133} raises significant safety concerns of off-tumor toxicity^{134,135}.

Wang *et al.* reported recently that peptides surrounding the commonly mutated KRAS G12 position are avidly bound and presented by the HLA-A11*01 serotype¹³⁶, facilitating recognition of these KRAS neoepitopes by T-cell receptors. Wang *et al.* demonstrated that KRAS G12V and KRAS G12D could be recognized by T cells. They isolated high-affinity T-cell receptors specific for these neoepitopes and showed that adoptive transfer of peripheral blood lymphocytes transduced with these KRAS-specific T-cell receptors could effect an antitumor immune response against xenografts harboring the cognate KRAS mutation. While this epitope is not presented by most other common MHC serotypes, Wang *et al.* note that HLA-A11*01 is present in 14% of the U.S. Caucasian population and in 40% of residents of southern China. Moreover, HRAS and NRAS are identical with KRAS in this region, so this strategy could be effective for all RAS isoforms. A phase 1 clinical trial has been opened for HLA-A11*01-positive patients with tumors harboring a KRAS G12V mutation (NCT03190941); this trial is open to MTC patients.

Antibody-drug conjugates. MTC and other neuroendocrine tumors commonly express DLL3, a Notch ligand, on their cell surface^{137,138}. A DLL3 antibody linked to a DNA crosslinker warhead (Rovalpituzumab tesirine; Rova-T) was shown to efficiently inhibit the growth of DLL3-expressing xenografts; notably, tumor-initiating cells were also targeted. A phase 1 trial of Rova-T in small cell lung cancer showed significant clinical activity, almost exclusively in DLL3-highly expressing tumors¹³⁹. A phase 2 trial of Rova-T in DLL3-expressing solid tumors, including a cohort for MTC, is now open (NCT02709889).

Intracellular antibodies. In general, antibody targeting of intracellular targets has been challenging. However, a recent report describes the construction and preclinical use of a monoclonal antibody that penetrates the cytoplasm and targets activated RAS¹⁴⁰. The antibody was modified to achieve cellular uptake by endocytosis, followed by endosomal escape into the cytosol. By binding activated RAS, the antibody disrupted downstream signaling pathways and interfered with cell growth. While activity *in vivo* was modest, several strategies were discussed to increase *in vivo* activity. If successful, this promising, albeit nascent, strategy may be applicable not only for RAS but also for targeting other intracellular targets, including RET M918T in MTC.

Peptide receptor radionuclide therapy

Neuroendocrine cancers, including MTC, frequently express somatostatin receptors (SSTRs), and the potential use of somatostatin analogs for imaging, therapy, or therapeutic targeting has been a common focus for many years. In a recently reported phase 3 trial, midgut neuroendocrine tumors were successfully treated by using peptide receptor radionuclide therapy (PRRT)¹⁴¹. Tumors were stratified by ¹¹¹In-DOTATATE scintigraphy for SSTR expression; in SSTR-positive midgut neuroendocrine tumors (NETs), ¹⁷⁷Lu-DOTATATE (Lutathera) treatment resulted

in highly significant prolongation of PFS and OS and modestly increased objective response rate (ORR). These results have led to European approval of Lutathera for the treatment of midgut NETs; a new drug application has been submitted to the FDA. Could this PRRT strategy work in MTC? In five small studies in MTC, using either ⁹⁰Y- or ¹⁷⁷Lu- somatostatin analogs^{142–146} (reviewed in¹⁴⁷), a modest ORR and frequent stable disease were seen. One might envision PRRT as a salvage therapy, stratifying MTC patients with ⁶⁸Ga-SST analog positron emission tomography (PET) (⁶⁸Ga-SST analog PET is more sensitive than ¹¹¹In-DOTATATE scintigraphy and detects lesions in about 70% of MTC patients with advanced or recurrent disease^{147–151}). One concern regarding the potential use of PRRT for MTC is that SSTR expression in MTC has been reported to be focal rather than uniform^{151,152}; this tumor heterogeneity could significantly limit efficacy.

Conclusions

The array of exciting directions for advancing our understanding of MTC, and especially for achieving more effective therapies, has never been more promising. Nevertheless, it must be emphasized that further advances will require careful design of basic, translational, and clinical research. Preclinical resources, including cell lines, animal models, and patient-derived xenografts, are currently very limited. A better understanding of the biology of the C cell and its transformation to MTC is critical. As discussed above, it is imperative to understand the

mechanisms of MTC progression on therapy, and this will require extensive analysis of progression biopsies. The recent and future advances in the field validate the tireless and ingenious effort that so many researchers have devoted.

Abbreviations

EXAM, Efficacy of XL184 (Cabozantinib) in Advanced Medullary Thyroid Cancer; FDA, US Food and Drug Administration; FTL, farnesyltransferase inhibitor; GTP, guanosine triphosphate; HR, hazard ratio; IC₅₀, concentration resulting in inhibition of 50% of activity; MEN, multiple endocrine neoplasia; MHC, major histocompatibility complex; MTC, medullary thyroid cancer; NET, neuroendocrine tumor; ORR, objective response rate; OS, overall survival; PD, pharmacodynamic; PET, positron emission tomography; PFS, progression-free survival; PK, pharmacokinetic; PRRT, peptide receptor radionuclide therapy; RBD, RAS-binding domain; SST, somatostatin; SSTR, somatostatin receptor; TKI, tyrosine kinase inhibitor; wt, wild-type.

Competing interests

The author declares that he has no competing interests.

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References



- Wells SA Jr, Pacini F, Robinson BG, *et al.*: **Multiple endocrine neoplasia type 2 and familial medullary thyroid carcinoma: an update.** *J Clin Endocrinol Metab.* 2013; **98**(8): 3149–64.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Matias-Guiu X, De Lellis R: **Medullary thyroid carcinoma: a 25-year perspective.** *Endocr Pathol.* 2014; **25**(1): 21–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Raue F(Ed.): **Medullary thyroid carcinoma. Biology, management, treatment.** London: Springer. 2015; 249.
[Publisher Full Text](#)
- Wells SA Jr, Asa SL, Dralle H, *et al.*: **Revised American Thyroid Association guidelines for the management of medullary thyroid carcinoma.** *Thyroid.* 2015; **25**(6): 567–610.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Hadoux J, Pacini F, Tuttle RM, *et al.*: **Management of advanced medullary thyroid cancer.** *Lancet Diabetes Endocrinol.* 2016; **4**(1): 64–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Romei C, Ciampi R, Elisei R: **A comprehensive overview of the role of the RET proto-oncogene in thyroid carcinoma.** *Nat Rev Endocrinol.* 2016; **12**(4): 192–202.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Wang TS, Evans DB(Eds.): **Medullary thyroid cancer.** London: Springer. 2016; 197.
[Publisher Full Text](#)
- DeLellis RA, Mangray S: **Medullary thyroid carcinoma: a contemporary perspective.** *AJSP-Reviews and Reports.* 2017; **22**(4): 196–208.
[Reference Source](#)
- Maia AL, Wajner SM, Vargas CV: **Advances and controversies in the management of medullary thyroid carcinoma.** *Curr Opin Oncol.* 2017; **29**(1): 25–32.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Thornton K, Kim G, Maher VE, *et al.*: **Vandetanib for the treatment of symptomatic or progressive medullary thyroid cancer in patients with unresectable locally advanced or metastatic disease: U.S. Food and Drug Administration drug approval summary.** *Clin Cancer Res.* 2012; **18**(14): 3722–30.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Wells SA Jr, Robinson BG, Gagel RF, *et al.*: **Vandetanib in patients with locally advanced or metastatic medullary thyroid cancer: a randomized, double-blind phase III trial.** *J Clin Oncol.* 2012; **30**(2): 134–41.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Elisei R, Schlumberger MJ, Müller SP, *et al.*: **Cabozantinib in progressive medullary thyroid cancer.** *J Clin Oncol.* 2013; **31**(29): 3639–46.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Le Douarin N, Le Lièvre C: **[Demonstration of neural origin of calcitonin cells of ultimobranchial body of chick embryo].** *C R Acad Sci Hebd Seances Acad Sci D.* 1970; **270**(23): 2857–60.
[PubMed Abstract](#)
- Polak JM, Pearse AG, Le Lièvre C, *et al.*: **Immunocytochemical confirmation of the neural crest origin of avian calcitonin-producing cells.** *Histochemistry.* 1974; **40**(3): 209–14.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Kameda Y, Nishimaki T, Miura M, *et al.*: **Mash1 regulates the development of C cells in mouse thyroid glands.** *Dev Dyn.* 2007; **236**(1): 262–70.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Johansson E, Andersson L, Örnros J, *et al.*: **Revising the embryonic origin of thyroid C cells in mice and humans.** *Development.* 2015; **142**(20): 3519–28.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Kameda Y: **Cellular and molecular events on the development of mammalian thyroid C cells.** *Dev Dyn.* 2016; **245**(3): 323–41.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Bagheri-Yarmand R, Sinha KM, Gururaj AE, *et al.*: **A novel dual kinase function of the RET proto-oncogene negatively regulates activating transcription factor 4-mediated apoptosis.** *J Biol Chem.* 2015; **290**(18): 11749–61.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Drosten M, Hilken G, Böckmann M, *et al.*: **Role of MEN2A-derived RET in maintenance and proliferation of medullary thyroid carcinoma.** *J Natl Cancer Inst.* 2004; **96**(16): 1231–9.
[PubMed Abstract](#) | [Publisher Full Text](#)

20. **F** Plaza-Menacho I, Barnouin K, Barry R, *et al.*: **RET functions as a dual-specificity kinase that requires allosteric inputs from juxtamembrane elements.** *Cell Rep.* 2016; **17**(12): 3319–32. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
21. Read RD, Goodfellow PJ, Mardis ER, *et al.*: **A Drosophila model of multiple endocrine neoplasia type 2.** *Genetics.* 2005; **171**(3): 1057–81. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Cote GJ, Grubbs EG, Hofmann MC: **Thyroid C-cell biology and oncogenic transformation.** *Recent Results Cancer Res.* 2015; **204**: 1–39. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
23. Barasch JM, Mackey H, Tamir H, *et al.*: **Induction of a neural phenotype in a serotonergic endocrine cell derived from the neural crest.** *J Neurosci.* 1987; **7**(9): 2874–83. [PubMed Abstract](#)
24. Moerch U, Nielsen HS, Lundsgaard D, *et al.*: **Flow sorting from organ material by intracellular markers.** *Cytometry A.* 2007; **71**(7): 495–500. [PubMed Abstract](#) | [Publisher Full Text](#)
25. Gawade S, Mayer C, Hafen K, *et al.*: **Cell growth dynamics in embryonic and adult mouse thyroid revealed by a novel approach to detect thyroid gland subpopulations.** *Thyroid.* 2016; **26**(4): 591–9. [PubMed Abstract](#) | [Publisher Full Text](#)
26. **F** Agrawal N, Jiao Y, Sausen M, *et al.*: **Exomic sequencing of medullary thyroid cancer reveals dominant and mutually exclusive oncogenic mutations in RET and RAS.** *J Clin Endocrinol Metab.* 2013; **98**(2): E364–9. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
27. **F** Moura MM, Cavaco BM, Pinto AE, *et al.*: **High prevalence of RAS mutations in RET-negative sporadic medullary thyroid carcinomas.** *J Clin Endocrinol Metab.* 2011; **96**(5): E863–8. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
28. Schulten H, Al-Maghrabi J, Al-Ghamdi K, *et al.*: **Mutational screening of RET, HRAS, KRAS, NRAS, BRAF, AKT1, and CTNNB1 in medullary thyroid carcinoma.** *Anticancer Res.* 2011; **31**(12): 4179–83. [PubMed Abstract](#)
29. Boichard A, Croux L, Al Ghuzlan A, *et al.*: **Somatic RAS mutations occur in a large proportion of sporadic RET-negative medullary thyroid carcinomas and extend to a previously unidentified exon.** *J Clin Endocrinol Metab.* 2012; **97**(10): E2031–5. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
30. **F** Ciampi R, Mian C, Fugazzola L, *et al.*: **Evidence of a low prevalence of RAS mutations in a large medullary thyroid cancer series.** *Thyroid.* 2013; **23**(1): 50–7. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
31. Moura MM, Cavaco BM, Leite V: **RAS proto-oncogene in medullary thyroid carcinoma.** *Endocr Relat Cancer.* 2015; **22**(5): R235–52. [PubMed Abstract](#) | [Publisher Full Text](#)
32. **F** Grubbs EG, Ng PK, Bui J, *et al.*: **RET fusion as a novel driver of medullary thyroid carcinoma.** *J Clin Endocrinol Metab.* 2015; **100**(3): 788–93. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
33. **F** Ji JH, Oh YL, Hong M, *et al.*: **Identification of driving ALK fusion genes and genomic landscape of medullary thyroid cancer.** *PLoS Genet.* 2015; **11**(8): e1005467. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
34. **F** Tomlins SA, Rhodes DR, Perner S, *et al.*: **Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer.** *Science.* 2005; **310**(5748): 644–8. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
35. Rubin MA, Maher CA, Chinnaiyan AM: **Common gene rearrangements in prostate cancer.** *J Clin Oncol.* 2011; **29**(27): 3659–68. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
36. Mertens F, Johansson B, Fioretos T, *et al.*: **The emerging complexity of gene fusions in cancer.** *Nat Rev Cancer.* 2015; **15**(6): 371–81. [PubMed Abstract](#) | [Publisher Full Text](#)
37. Gottesman MM, Lavi O, Hall MD, *et al.*: **Toward a better understanding of the complexity of cancer drug resistance.** *Annu Rev Pharmacol Toxicol.* 2016; **56**: 85–102. [PubMed Abstract](#) | [Publisher Full Text](#)
38. **F** Diaz LA Jr, Williams RT, Wu J, *et al.*: **The molecular evolution of acquired resistance to targeted EGFR blockade in colorectal cancers.** *Nature.* 2012; **486**(7404): 537–40. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
39. Bozic I, Gerold JM, Nowak MA: **Quantifying clonal and subclonal passenger mutations in cancer evolution.** *PLoS Comput Biol.* 2016; **12**(2): e1004731. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
40. **F** McGranahan N, Swanton C: **Clonal heterogeneity and tumor evolution: past, present, and the future.** *Cell.* 2017; **168**(4): 613–28. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
41. Kantarjian H, Giles F, Wunderle L, *et al.*: **Nilotinib in imatinib-resistant CML and Philadelphia chromosome-positive ALL.** *N Engl J Med.* 2006; **354**(24): 2542–51. [PubMed Abstract](#) | [Publisher Full Text](#)
42. **F** de Bono JS, Logothetis CJ, Molina A, *et al.*: **Abiraterone and increased survival in metastatic prostate cancer.** *N Engl J Med.* 2011; **364**(21): 1995–2005. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
43. **F** Flaherty KT, Infante JR, Daud A, *et al.*: **Combined BRAF and MEK inhibition in melanoma with BRAF V600 mutations.** *N Engl J Med.* 2012; **367**(18): 1694–703. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
44. **F** Scher HI, Fizazi K, Saad F, *et al.*: **Increased survival with enzalutamide in prostate cancer after chemotherapy.** *N Engl J Med.* 2012; **367**(13): 1187–97. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
45. Zhang C, Spevak W, Zhang Y, *et al.*: **RAF inhibitors that evade paradoxical MAPK pathway activation.** *Nature.* 2015; **526**(7574): 583–6. [PubMed Abstract](#) | [Publisher Full Text](#)
46. Mok TS, Wu YL, Ahn MJ, *et al.*: **Osimertinib or platinum-pemetrexed in EGFR T790M-positive lung cancer.** *N Engl J Med.* 2017; **376**(7): 629–40. [PubMed Abstract](#) | [Publisher Full Text](#)
47. Carlomagno F, Guida T, Anaganti S, *et al.*: **Disease associated mutations at valine 804 in the RET receptor tyrosine kinase confer resistance to selective kinase inhibitors.** *Oncogene.* 2004; **23**(36): 6056–63. [PubMed Abstract](#) | [Publisher Full Text](#)
48. Mologni L, Redaelli S, Morandi A, *et al.*: **Ponatinib is a potent inhibitor of wild-type and drug-resistant gatekeeper mutant RET kinase.** *Mol Cell Endocrinol.* 2013; **377**(1–2): 1–6. [PubMed Abstract](#) | [Publisher Full Text](#)
49. Shah NP, Nicoll JM, Nagar B, *et al.*: **Multiple BCR-ABL kinase domain mutations confer polyclonal resistance to the tyrosine kinase inhibitor imatinib (STI571) in chronic phase and blast crisis chronic myeloid leukemia.** *Cancer Cell.* 2002; **2**(2): 117–25. [PubMed Abstract](#) | [Publisher Full Text](#)
50. Plenker D, Riedel M, Brägelmann J, *et al.*: **Drugging the catalytically inactive state of RET kinase in RET-rearranged tumors.** *Sci Transl Med.* 2017; **9**(394): pii: eaah6144. [PubMed Abstract](#) | [Publisher Full Text](#)
51. Roberts KG, Odell AF, Byrnes EM, *et al.*: **Resistance to c-KIT kinase inhibitors conferred by V654A mutation.** *Mol Cancer Ther.* 2007; **6**(3): 1159–66. [PubMed Abstract](#) | [Publisher Full Text](#)
52. Weil A, Martin P, Smith R, *et al.*: **Pharmacokinetics of vandetanib in subjects with renal or hepatic impairment.** *Clin Pharmacokinet.* 2010; **49**(9): 607–18. [PubMed Abstract](#) | [Publisher Full Text](#)
53. U.S. Food and Drug Administration: **Clinical pharmacology and biometrics review(s).** Silver Spring (MD): U.S. Food and Drug Administration; 2012. [Reference Source](#)
54. Singh H, Brave M, Beaver JA, *et al.*: **U.S. Food and Drug Administration approval: cabozantinib for the treatment of advanced renal cell carcinoma.** *Clin Cancer Res.* 2017; **23**(2): 330–5. [PubMed Abstract](#) | [Publisher Full Text](#)
55. **F** Sherman SI, Clary DO, Elisei R, *et al.*: **Correlative analyses of RET and RAS mutations in a phase 3 trial of cabozantinib in patients with progressive, metastatic medullary thyroid cancer.** *Cancer.* 2016; **122**(24): 3856–64. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
56. Schlumberger M, Elisei R, Müller S, *et al.*: **Overall survival analysis of EXAM, a phase III trial of cabozantinib in patients with radiographically progressive medullary thyroid carcinoma.** *Ann Oncol.* 2017; **28**(11): 2813–2819. [PubMed Abstract](#) | [Publisher Full Text](#)
57. **F** Jain S, Watson MA, DeBenedetti MK, *et al.*: **Expression profiles provide insights into early malignant potential and skeletal abnormalities in multiple endocrine neoplasia type 2B syndrome tumors.** *Cancer Res.* 2004; **64**(11): 3907–13. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
58. Ameur N, Lacroix L, Roucan S, *et al.*: **Aggressive inherited and sporadic medullary thyroid carcinomas display similar oncogenic pathways.** *Endocr Relat Cancer.* 2009; **16**(4): 1261–72. [PubMed Abstract](#) | [Publisher Full Text](#)
59. **F** Maliszewska A, Leandro-Garcia LJ, Castelblanco E, *et al.*: **Differential gene expression of medullary thyroid carcinoma reveals specific markers associated with genetic conditions.** *Am J Pathol.* 2013; **182**(2): 350–62. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
60. Oczko-Wojciechowska M, Swierniak M, Krajewska J, *et al.*: **Differences in the transcriptome of medullary thyroid cancer regarding the status and type of RET gene mutations.** *Sci Rep.* 2017; **7**: 42074. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
61. Songyang Z, Carraway KL 3rd, Eck MJ, *et al.*: **Catalytic specificity of protein-tyrosine kinases is critical for selective signalling.** *Nature.* 1995; **373**(6514): 536–9. [PubMed Abstract](#) | [Publisher Full Text](#)
62. Gujral TS, Singh VK, Jia Z, *et al.*: **Molecular mechanisms of RET receptor-mediated oncogenesis in multiple endocrine neoplasia 2B.** *Cancer Res.* 2006; **66**(22): 10741–9. [PubMed Abstract](#) | [Publisher Full Text](#)
63. Dixit A, Torkamani A, Schork NJ, *et al.*: **Computational modeling of structurally conserved cancer mutations in the RET and MET kinases: the impact on protein structure, dynamics, and stability.** *Biophys J.* 2009; **96**(3): 858–74. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
64. George Priya Doss C, Rajith B, Chakraborty C, *et al.*: **In silico profiling and structural insights of missense mutations in RET protein kinase domain by molecular dynamics and docking approach.** *Mol Biosyst.* 2014; **10**(3): 421–36. [PubMed Abstract](#) | [Publisher Full Text](#)

65. Yakes FM, Chen J, Tan J, *et al.*: **Cabozantinib (XL184), a novel MET and VEGFR2 inhibitor, simultaneously suppresses metastasis, angiogenesis, and tumor growth.** *Mol Cancer Ther.* 2011; **10**(12): 2298–308.
[PubMed Abstract](#) | [Publisher Full Text](#)
66. Fujita-Sato S, Galeas J, Truitt M, *et al.*: **Enhanced MET translation and signaling sustains K-Ras-driven proliferation under anchorage-independent growth conditions.** *Cancer Res.* 2015; **75**(14): 2851–62.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
67. Mologni L, Gambacorti-Passerini C, Goekjian P, *et al.*: **RET kinase inhibitors: a review of recent patents (2012–2015).** *Expert Opin Ther Pat.* 2017; **27**(1): 91–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
68. Kodama T, Tsukaguchi T, Satoh Y, *et al.*: **Alectinib shows potent antitumor activity against RET-rearranged non-small cell lung cancer.** *Mol Cancer Ther.* 2014; **13**(12): 2910–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
69. Rahal R, Evans EK, Hu W, *et al.*: **Abstract 2641: the development of potent, selective RET inhibitors that target both wild-type RET and prospectively identified resistance mutations to multi-kinase inhibitors.** *Cancer Res.* 2016; **76**(14): 2641.
[Publisher Full Text](#)
70. Brandhuber B, Haas J, Tuch B, *et al.*: **The development of a potent, KDR/VEGFR2-sparing RET kinase inhibitor for treating patients with RET-dependent cancers.** *Eur J Cancer.* 2016; **69**(Supplement 1): S144.
[Publisher Full Text](#)
71. James J, Ruggeri B, Armstrong RC, *et al.*: **CEP-32496: a novel orally active BRAF^{V600E} inhibitor with selective cellular and in vivo antitumor activity.** *Mol Cancer Ther.* 2012; **11**(4): 930–41.
[PubMed Abstract](#) | [Publisher Full Text](#)
72. Koh YW, Shah MH, Agarwal K, *et al.*: **Sorafenib and Mek inhibition is synergistic in medullary thyroid carcinoma in vitro.** *Endocr Relat Cancer.* 2012; **19**(1): 29–38.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
73. **F** Hatzivassiliou G, Song K, Yen I, *et al.*: **RAF inhibitors prime wild-type RAF to activate the MAPK pathway and enhance growth.** *Nature.* 2010; **464**(7287): 431–5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
74. **F** Heidorn SJ, Milagre C, Whittaker S, *et al.*: **Kinase-dead BRAF and oncogenic RAS cooperate to drive tumor progression through CRAF.** *Cell.* 2010; **140**(2): 209–21.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
75. **F** Poulidakos PI, Zhang C, Bollag G, *et al.*: **RAF inhibitors transactivate RAF dimers and ERK signalling in cells with wild-type BRAF.** *Nature.* 2010; **464**(7287): 427–30.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
76. Li GG, Somwar R, Joseph J, *et al.*: **Antitumor activity of RXD-105 in multiple cancer types with RET rearrangements or mutations.** *Clin Cancer Res.* 2017; **23**(12): 2981–90.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
77. Ohiwa M, Murakami H, Iwashita T, *et al.*: **Characterization of Ret-Shc-Grb2 complex induced by GDNF, MEN 2A, and MEN 2B mutations.** *Biochem Biophys Res Commun.* 1997; **237**(3): 747–51.
[PubMed Abstract](#) | [Publisher Full Text](#)
78. Cox AD, Fesik SW, Kimmelman AC, *et al.*: **Drugging the undruggable RAS: mission possible?** *Nat Rev Drug Discov.* 2014; **13**(11): 828–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
79. Stephen AG, Esposito D, Bagni RK, *et al.*: **Dragging ras back in the ring.** *Cancer Cell.* 2014; **25**(3): 272–81.
[PubMed Abstract](#) | [Publisher Full Text](#)
80. Papke B, Der CJ: **Drugging RAS: know the enemy.** *Science.* 2017; **355**(6330): 1158–63.
[PubMed Abstract](#) | [Publisher Full Text](#)
81. Goody RS, Frech M, Wittinghofer A: **Affinity of guanine nucleotide binding proteins for their ligands: facts and artefacts.** *Trends Biochem Sci.* 1991; **16**(9): 327–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
82. James GL, Goldstein JL, Brown MS: **Polylysine and CVIM sequences of K-RasB dictate specificity of prenylation and confer resistance to benzodiazepine peptidomimetic in vitro.** *J Biol Chem.* 1995; **270**(11): 6221–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
83. Rowell CA, Kowalczyk JJ, Lewis MD, *et al.*: **Direct demonstration of geranylgeranylation and farnesylation of Ki-Ras in vivo.** *J Biol Chem.* 1997; **272**(22): 14093–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
84. Whyte DB, Kirschmeier P, Hockenberry TN, *et al.*: **K- and N-Ras are geranylgeranylated in cells treated with farnesyl protein transferase inhibitors.** *J Biol Chem.* 1997; **272**(22): 14459–64.
[PubMed Abstract](#) | [Publisher Full Text](#)
85. Grilley-Olson JE, Bedard PL, Fasolo A, *et al.*: **A phase Ib dose-escalation study of the MEK inhibitor trametinib in combination with the PI3K/mTOR inhibitor GSK2126458 in patients with advanced solid tumors.** *Invest New Drugs.* 2016; **34**(6): 740–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
86. **F** Maurer T, Garrenton LS, Oh A, *et al.*: **Small-molecule ligands bind to a distinct pocket in Ras and inhibit SOS-mediated nucleotide exchange activity.** *Proc Natl Acad Sci U S A.* 2012; **109**(14): 5299–304.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
87. **F** Ostrem JM, Peters U, Sos ML, *et al.*: **K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interactions.** *Nature.* 2013; **503**(7477): 548–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
88. **F** Zimmermann G, Papke B, Ismail S, *et al.*: **Small molecule inhibition of the KRAS-PDEδ interaction impairs oncogenic KRAS signalling.** *Nature.* 2013; **497**(7451): 638–42.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
89. Zorde Khvalevsky E, Gabai R, Rachmut IH, *et al.*: **Mutant KRAS is a druggable target for pancreatic cancer.** *Proc Natl Acad Sci U S A.* 2013; **110**(51): 20723–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
90. Burns MC, Sun Q, Daniels RN, *et al.*: **Approach for targeting Ras with small molecules that activate SOS-mediated nucleotide exchange.** *Proc Natl Acad Sci U S A.* 2014; **111**(9): 3401–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
91. Hunter JC, Gurbani D, Ficarro SB, *et al.*: **In situ selectivity profiling and crystal structure of SML-8-73-1, an active site inhibitor of oncogenic K-Ras G12C.** *Proc Natl Acad Sci U S A.* 2014; **111**(24): 8895–900.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
92. **F** Lim SM, Westover KD, Ficarro SB, *et al.*: **Therapeutic targeting of oncogenic K-Ras by a covalent catalytic site inhibitor.** *Angew Chem Int Ed Engl.* 2014; **53**(1): 199–204.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
93. **F** Upadhyaya P, Qian Z, Selner NG, *et al.*: **Inhibition of Ras signaling by blocking Ras-effector interactions with cyclic peptides.** *Angew Chem Int Ed Engl.* 2015; **54**(26): 7602–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
94. **F** Athuluri-Divakar SK, Vasquez-Del Carprio R, Dutta K, *et al.*: **A small molecule RAS-mimetic disrupts RAS association with effector proteins to block signaling.** *Cell.* 2016; **165**(3): 643–55.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
95. **F** Lito P, Solomon M, Li LS, *et al.*: **Allele-specific inhibitors inactivate mutant KRAS G12C by a trapping mechanism.** *Science.* 2016; **351**(6273): 604–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
96. Lu S, Jang H, Gu S, *et al.*: **Drugging Ras GTPase: a comprehensive mechanistic and signaling structural view.** *Chem Soc Rev.* 2016; **45**(18): 4929–52.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
97. McCormick F: **K-Ras protein as a drug target.** *J Mol Med (Berl).* 2016; **94**(3): 253–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
98. **F** Ostrem JM, Shokat KM: **Direct small-molecule inhibitors of KRAS: from structural insights to mechanism-based design.** *Nat Rev Drug Discov.* 2016; **15**(11): 771–85.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
99. Papke B, Murarka S, Vogel HA, *et al.*: **Identification of pyrazolopyridazinones as PDEδ inhibitors.** *Nat Commun.* 2016; **7**: 11360.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
100. **F** Patricelli MP, Janes MR, Li LS, *et al.*: **Selective inhibition of oncogenic KRAS output with small molecules targeting the inactive state.** *Cancer Discov.* 2016; **6**(3): 316–29.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
101. Cetin M, Evenson WE, Gross GG, *et al.*: **RasIns: genetically encoded intrabodies of activated Ras proteins.** *J Mol Biol.* 2017; **429**(4): 562–73.
[PubMed Abstract](#) | [Publisher Full Text](#)
102. Kauke MJ, Traxlmayr MW, Parker JA, *et al.*: **An engineered protein antagonist of K-Ras/B-Raf interaction.** *Sci Rep.* 2017; **7**(1): 5831.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
103. Keeton AB, Salter EA, Piazza GA: **The RAS-effector interaction as a drug target.** *Cancer Res.* 2017; **77**(2): 221–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
104. **F** Martín-Gago P, Fansa EK, Klein CH, *et al.*: **A PDEδ-KRAS inhibitor chemotype with up to seven H-Bonds and picomolar affinity that prevents efficient inhibitor release by Arl2.** *Angew Chem Int Ed Engl.* 2017; **56**(9): 2423–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
105. Martín-Gago P, Fansa EK, Winzker M, *et al.*: **Covalent protein labeling at glutamic acids.** *Cell Chem Biol.* 2017; **24**(5): 589–597.e5.
[PubMed Abstract](#) | [Publisher Full Text](#)
106. **F** Novotny CJ, Hamilton GL, McCormick F, *et al.*: **Farnesyltransferase-mediated delivery of a covalent inhibitor overcomes alternative prenylation to mislocalize K-Ras.** *ACS Chem Biol.* 2017; **12**(7): 1956–62.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
107. **F** Ross SJ, Revenko AS, Hanson LL, *et al.*: **Targeting KRAS-dependent tumors with AZD4785, a high-affinity therapeutic antisense oligonucleotide inhibitor of KRAS.** *Sci Transl Med.* 2017; **9**(394): pii: eaa15253.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
108. Sakamoto K, Kamada Y, Sameshima T, *et al.*: **K-Ras(G12D)-selective inhibitory peptides generated by random peptide T7 phage display technology.** *Biochem Biophys Res Commun.* 2017; **484**(3): 605–11.
[PubMed Abstract](#) | [Publisher Full Text](#)
109. **F** Spencer-Smith R, Koide A, Zhou Y, *et al.*: **Inhibition of RAS function through targeting an allosteric regulatory site.** *Nat Chem Biol.* 2017; **13**(1): 62–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)

110. **F** Welsch ME, Kaplan A, Chambers JM, *et al.*: **Multivalent small-molecule pan-RAS inhibitors.** *Cell.* 2017; **168**(5): 878–889.e29.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
111. **F** Chandra A, Grecco HE, Pisupati V, *et al.*: **The GDI-like solubilizing factor PDE δ sustains the spatial organization and signalling of Ras family proteins.** *Nat Cell Biol.* 2011; **14**(2): 148–58.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
112. Hong DS, Cabanillas ME, Wheler J, *et al.*: **Inhibition of the Ras/Raf/MEK/ERK and RET kinase pathways with the combination of the multitikinase inhibitor sorafenib and the farnesyltransferase inhibitor tipifarnib in medullary and differentiated thyroid malignancies.** *J Clin Endocrinol Metab.* 2011; **96**(4): 997–1005.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
113. Lam ET, Ringel MD, Kloos RT, *et al.*: **Phase II clinical trial of sorafenib in metastatic medullary thyroid cancer.** *J Clin Oncol.* 2010; **28**(14): 2323–30.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
114. **F** Brahmer JR, Tykodi SS, Chow LQ, *et al.*: **Safety and activity of anti-PD-L1 antibody in patients with advanced cancer.** *N Engl J Med.* 2012; **366**(26): 2455–65.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
115. **F** Topalian SL, Hodi FS, Brahmer JR, *et al.*: **Safety, activity, and immune correlates of anti-PD-1 antibody in cancer.** *N Engl J Med.* 2012; **366**(26): 2443–54.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
116. Couzin-Frankel J: **Breakthrough of the year 2013. Cancer immunotherapy.** *Science.* 2013; **342**(6165): 1432–3.
[PubMed Abstract](#) | [Publisher Full Text](#)
117. **F** Sharma P, Allison JP: **The future of immune checkpoint therapy.** *Science.* 2015; **348**(6230): 56–61.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
118. **F** Chen DS, Mellman I: **Elements of cancer immunity and the cancer-immune set point.** *Nature.* 2017; **541**(7637): 321–30.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
119. French JD, Bible K, Spitzweg C, *et al.*: **Leveraging the immune system to treat advanced thyroid cancers.** *Lancet Diabetes Endocrinol.* 2017; **5**(6): 469–81.
[PubMed Abstract](#) | [Publisher Full Text](#)
120. Bongiovanni M, Rebecchini C, Saglietti C, *et al.*: **Very low expression of PD-L1 in medullary thyroid carcinoma.** *Endocr Relat Cancer.* 2017; **24**(6): L35–L38.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
121. **F** Herbst RS, Soria JC, Kowanzet M, *et al.*: **Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients.** *Nature.* 2014; **515**(7528): 563–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
122. **F** Rizvi NA, Hellmann MD, Snyder A, *et al.*: **Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer.** *Science.* 2015; **348**(6230): 124–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
123. Schott M, Feldkamp J, Klucken M, *et al.*: **Calcitonin-specific antitumor immunity in medullary thyroid carcinoma following dendritic cell vaccination.** *Cancer Immunol Immunother.* 2002; **51**(11–12): 663–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
124. Papewalis C, Wuttke M, Seissler J, *et al.*: **Dendritic cell vaccination with xenogenic polypeptide hormone induces tumor rejection in neuroendocrine cancer.** *Clin Cancer Res.* 2008; **14**(13): 4298–305.
[PubMed Abstract](#) | [Publisher Full Text](#)
125. Wuttke M, Papewalis C, Meyer Y, *et al.*: **Amino acid-modified calcitonin immunization induces tumor epitope-specific immunity in a transgenic mouse model for medullary thyroid carcinoma.** *Endocrinology.* 2008; **149**(11): 5627–34.
[PubMed Abstract](#) | [Publisher Full Text](#)
126. Bachleitner-Hofmann T, Friedl J, Hassler M, *et al.*: **Pilot trial of autologous dendritic cells loaded with tumor lysate(s) from allogeneic tumor cell lines in patients with metastatic medullary thyroid carcinoma.** *Oncol Rep.* 2009; **21**(6): 1585–92.
[PubMed Abstract](#) | [Publisher Full Text](#)
127. **F** Kalos M, Levine BL, Porter DL, *et al.*: **T cells with chimeric antigen receptors have potent antitumor effects and can establish memory in patients with advanced leukemia.** *Sci Transl Med.* 2011; **3**(95): 95ra73.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
128. Rosenberg SA, Restifo NP: **Adoptive cell transfer as personalized immunotherapy for human cancer.** *Science.* 2015; **348**(6230): 62–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
129. Porter DL, Hwang W, Frey NV, *et al.*: **Chimeric antigen receptor T cells persist and induce sustained remissions in relapsed refractory chronic lymphocytic leukemia.** *Sci Transl Med.* 2015; **7**(303): 303ra139.
[PubMed Abstract](#) | [Publisher Full Text](#)
130. Johnson LA, June CH: **Driving gene-engineered T cell immunotherapy of cancer.** *Cell Res.* 2017; **27**(1): 38–58.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
131. **NetMHC 3.4.**
[Reference Source](#)
132. Plaza-Menacho I, Burzynski GM, de Groot JW, *et al.*: **Current concepts in RET-related genetics, signaling and therapeutics.** *Trends Genet.* 2006; **22**(11): 627–36.
[PubMed Abstract](#) | [Publisher Full Text](#)
133. Runeberg-Roos P, Saarma M: **Neurotrophic factor receptor RET: structure, cell biology, and inherited diseases.** *Ann Med.* 2007; **39**(8): 572–80.
[PubMed Abstract](#) | [Publisher Full Text](#)
134. Linette GP, Stadtmayer EA, Maus MV, *et al.*: **Cardiovascular toxicity and titin cross-reactivity of affinity-enhanced T cells in myeloma and melanoma.** *Blood.* 2013; **122**(6): 863–71.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
135. Morgan RA, Chinnsamy N, Abate-Daga D, *et al.*: **Cancer regression and neurological toxicity following anti-MAGE-A3 TCR gene therapy.** *J Immunother.* 2013; **36**(2): 133–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
136. **F** Wang QJ, Yu Z, Griffith K, *et al.*: **Identification of T-cell receptors targeting KRAS-mutated human tumors.** *Cancer Immunol Res.* 2016; **4**(3): 204–14.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
137. Saunders LR, Bankovich AJ, Anderson WC, *et al.*: **A DLL3-targeted antibody-drug conjugate eradicates high-grade pulmonary neuroendocrine tumor-initiating cells in vivo.** *Sci Transl Med.* 2015; **7**(302): 302ra136.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
138. Saunders LR, Williams SA, Bheddah S, *et al.*: **Abstract 3093: expression of DLL3 in metastatic melanoma, glioblastoma and high-grade extrapulmonary neuroendocrine carcinomas as potential indications for rovalpituzumab tesirine (Rova-T; SC16LD6.5), a delta-like protein 3 (DLL3)-targeted antibody drug conjugate (ADC).** *Proc Amer Assoc for Cancer Res.* 2017; **7**(13 Suppl): Abstract 3093.
[PubMed Abstract](#) | [Publisher Full Text](#)
139. **F** Rudin CM, Pietanza MC, Bauer TM, *et al.*: **Rovalpituzumab tesirine, a DLL3-targeted antibody-drug conjugate, in recurrent small-cell lung cancer: a first-in-human, first-in-class, open-label, phase 1 study.** *Lancet Oncol.* 2017; **18**(1): 42–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
140. **F** Shin SM, Choi DK, Jung K, *et al.*: **Antibody targeting intracellular oncogenic Ras mutants exerts anti-tumour effects after systemic administration.** *Nat Commun.* 2017; **8**: 15090.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
141. **F** Strosberg J, El-Haddad G, Wolin E, *et al.*: **Phase 3 Trial of ¹⁷⁷Lu-dotatate for midgut neuroendocrine tumors.** *N Engl J Med.* 2017; **376**(2): 125–35.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
142. Waldherr C, Schumacher T, Pless M, *et al.*: **Radiopeptide transmitted internal irradiation of non-iodophil thyroid cancer and conventionally untreatable medullary thyroid cancer using.** *Nucl Med Commun.* 2001; **22**(6): 673–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
143. Bodei L, Handkiewicz-Junak D, Grana C, *et al.*: **Receptor radionuclide therapy with ⁹⁰Y-DOTATOC in patients with medullary thyroid carcinomas.** *Cancer Biother Radiopharm.* 2004; **19**(1): 65–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
144. Makis W, McCann K, McEwan AJ: **Medullary thyroid carcinoma (MTC) treated with ¹⁷⁷Lu-DOTATATE PRRT: a report of two cases.** *Clin Nucl Med.* 2015; **40**(5): 408–12.
[PubMed Abstract](#) | [Publisher Full Text](#)
145. Vaisman F, Rosado de Castro PH, Lopes FP, *et al.*: **Is there a role for peptide receptor radionuclide therapy in medullary thyroid cancer?** *Clin Nucl Med.* 2015; **40**(2): 123–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
146. Salavati A, Puranik A, Kulkarni HR, *et al.*: **Peptide receptor radionuclide therapy (PRRT) of medullary and nonmedullary thyroid cancer using radiolabeled somatostatin analogues.** *Semin Nucl Med.* 2016; **46**(3): 215–24.
[PubMed Abstract](#) | [Publisher Full Text](#)
147. Conry BG, Papanthasiou ND, Prakash V, *et al.*: **Comparison of ⁶⁸Ga-DOTATATE and ¹⁸F-fluorodeoxyglucose PET/CT in the detection of recurrent medullary thyroid carcinoma.** *Eur J Nucl Med Mol Imaging.* 2010; **37**(1): 49–57.
[PubMed Abstract](#) | [Publisher Full Text](#)
148. Treglia G, Castaldi P, Villani MF, *et al.*: **Comparison of ¹⁸F-DOPA, ¹⁸F-FDG and ⁶⁸Ga-somatostatin analogue PET/CT in patients with recurrent medullary thyroid carcinoma.** *Eur J Nucl Med Mol Imaging.* 2012; **39**(4): 569–80.
[PubMed Abstract](#) | [Publisher Full Text](#)
149. Ozkan ZG, Kuyumcu S, Uzum AK, *et al.*: **Comparison of ⁶⁸Ga-DOTATATE PET-CT, ¹⁸F-FDG PET-CT and ^{99m}Tc-(V)DMSA scintigraphy in the detection of recurrent or metastatic medullary thyroid carcinoma.** *Nucl Med Commun.* 2015; **36**(3): 242–50.
[PubMed Abstract](#) | [Publisher Full Text](#)
150. Tran K, Khan S, Taghizadehasl M, *et al.*: **Gallium-68 Dotatate PET/CT is superior to other imaging modalities in the detection of medullary carcinoma of the thyroid in the presence of high serum calcitonin.** *Hell J Nucl Med.* 2015; **18**(1): 19–24.
[PubMed Abstract](#)
151. Herac M, Niederle B, Raderer M, *et al.*: **Expression of somatostatin receptor 2A in medullary thyroid carcinoma is associated with lymph node metastasis.** *APMIS.* 2016; **124**(10): 839–45.
[PubMed Abstract](#) | [Publisher Full Text](#)
152. Papotti M, Kumar U, Volante M, *et al.*: **Immunohistochemical detection of somatostatin receptor types 1–5 in medullary carcinoma of the thyroid.** *Clin Endocrinol (Oxf).* 2001; **54**(5): 641–9.
[PubMed Abstract](#) | [Publisher Full Text](#)

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