

Review

The Role of Hypoxia in Glioblastoma Invasion

Ana Rita Monteiro ¹, Richard Hill ² , Geoffrey J. Pilkington ² and Patrícia A. Madureira ^{1,2,*} 

¹ Centre for Biomedical Research (CBMR), University of Algarve, Campus of Gambelas, Building 8, Room 3.4, 8005-139 Faro, Portugal; a50039@ualg.pt

² Brain Tumour Research Centre of Excellence, Institute of Biomedical and Biomolecular Sciences, University of Portsmouth, Portsmouth PO1 2DT, UK; richard.hill@port.ac.uk (R.H.); geoff.pilkington@port.ac.uk (G.J.P.)

* Correspondence: pamadureira@ualg.pt; Tel.: +351-289-244-443

Received: 31 October 2017; Accepted: 21 November 2017; Published: 22 November 2017

Abstract: Glioblastoma multiforme (GBM), a grade IV astrocytoma, is the most common and deadly type of primary malignant brain tumor, with a patient's median survival rate ranging from 15 to 17 months. The current treatment for GBM involves tumor resection surgery based on MRI image analysis, followed by radiotherapy and treatment with temozolomide. However, the gradual development of tumor resistance to temozolomide is frequent in GBM patients leading to subsequent tumor regrowth/relapse. For this reason, the development of more effective therapeutic approaches for GBM is of critical importance. Low tumor oxygenation, also known as hypoxia, constitutes a major concern for GBM patients, since it promotes cancer cell spreading (invasion) into the healthy brain tissue in order to evade this adverse microenvironment. Tumor invasion not only constitutes a major obstacle to surgery, radiotherapy, and chemotherapy, but it is also the main cause of death in GBM patients. Understanding how hypoxia triggers the GBM cells to become invasive is paramount to developing novel and more effective therapies against this devastating disease. In this review, we will present a comprehensive examination of the available literature focused on investigating how GBM hypoxia triggers an invasive cancer cell phenotype and the role of these invasive proteins in GBM progression.

Keywords: GBM; hypoxia; HIF; invasion; chemotherapy

1. Introduction

Glioblastoma multiforme (GBM) is the most common type of primary malignant brain tumor, classified by the World Health Organization (WHO) as a grade IV astrocytoma. Patients' median survival is low, ranging from 15 to 17 months [1–3]. Under the Stupp protocol, the current treatment for GBM commences with tumor resection surgery based on MRI image analysis (if possible depending on the location and size of the tumor), followed by radiotherapy and treatment with temozolomide [4]. Radiotherapy alone can significantly increase median survival, although the most common radiological response is to stabilize the disease but ultimately tumor progression follows. Subgroup analysis of GBM patients demonstrated that a discernible clinical response to temozolomide was principally limited to those tumors containing a specific epigenetic alteration, namely promoter methylation of the O6-methylguanine DNA methyltransferase (*MGMT*) gene [5]. Critically, while these patients show an improvement (compared to surgical resection and radiotherapy alone), almost all demonstrate eventual progressive disease in the absence of *MGMT* promoter methylation [6]. The development of novel and more effective therapeutic approaches for GBM is paramount, taking into account the low median survival rate for GBM patients even when undergoing the current treatment protocols.

Genomic studies have identified four subtypes of GBM based on gene expression patterns, namely the classical, proneural, neural, and mesenchymal subtypes [7,8]. The classical subtype was shown to

present amplification or mutation of the epidermal growth factor receptor (EGFR) and high expression of the neural precursor and stem cell marker NES, as well as of the Notch (NOTCH3, JAG1, and LFNG) and Sonic hedgehog (SMO, GAS1, and GLI2) signaling pathway proteins [8]. The proneural subtype shows amplification of platelet-derived growth factor receptor A (PDGFRA) and deletion of the *TP53* tumor suppressor gene [8]. The proneural signature further contains high expression of several proneural developmental genes, such as *SOX*, *DCX*, *DLL3*, *ASCL1*, and *TCF4* [9]. Interestingly, within this subtype isocitrate dehydrogenase (*IDH*) gene mutations associated with improved patient outcome are also observed [8]. The neural subtype is characterized by the expression of neuron markers such as *NEFL*, *GABRA1*, *SYT1*, and *SLC12A5* [8]. Finally, the mesenchymal subtype as the name indicates expresses mesenchymal markers such as *CHI3L1* and *MET* [9] and has been associated with mutations or deletion of the *neurofibromin 1 (NF1)* gene [8,10,11]. Neurofibromin protein stimulates RAS intrinsic GTPase activity, thereby promoting the conversion of active RAS-GTP into its inactive RAS-GDP state. Hence, the loss of *NF1* gene results in sustained intracellular levels of active RAS-GTP, subsequently promoting oncogenesis [12].

More recently, the WHO updated the classification of the tumors of the Central Nervous System (CNS) using molecular parameters in addition to histology [13]. According to the new WHO classification, CNS tumor diagnoses should consist of the histopathological name followed by the genetic features. This new classification subdivides GBM into GBM, *IDH*-wildtype (approximately 90% of all cases) and GBM, *IDH*-mutant. The *IDH* enzymes are responsible for the oxidative carboxylation of isocitrate to α -ketoglutarate producing nicotinamide adenine dinucleotide phosphate (NADPH). Mutations in the *IDH* genes lead to an approximately 50% decrease of the normal *IDH* activity. Taking into account the impaired function of the mitochondria in GBM cells, the production of bioenergy and intermediates is decreased in *IDH* mutant GBM, hence the tumor growth is impaired when compared to *IDH*-wildtype GBM, which translates into a better overall patient prognosis [14].

The majority of GBMs develop *de novo* and as such are classified as primary GBMs. These tumors tend to grow rapidly and usually do not have *IDH* mutations. However, a minority of GBMs can slowly develop from low-grade astrocytomas, known as secondary GBMs. Mutations in *IDH* are more frequently observed in this class of GBMs [15]. Histologically, both primary and secondary GBMs are identical. GBM distinctive pathological features include necrotic foci with surrounding cellular pseudopalisades and microvascular hyperplasia (Figure 1), which are believed to play a main role in the accelerated growth and invasion of GBM [16]. Research on the origin of pseudopalisades suggests that this morphological structure is created by tumor cells migrating away from a central hypoxic (poorly oxygenated) region and forming an invasive front [17,18]. Microvascular hyperplasia is an exacerbated form of angiogenesis that occurs in response to the secretion of proangiogenic factors (i.e., vascular endothelial growth factors (VEGFs), interleukin-8 (IL-8)) by the cells that form the pseudopalisades [19]. Microvascular hyperplasia is characterized by a large number of enlarged and rapidly dividing endothelial cells, pericytes, and smooth muscle cells that form tuft micro-aggregates at the leading edge of sprouting blood vessels [20]. It can take the shape of 'glomeruloid bodies' that constitute a characteristic feature of GBM. The excessive VEGF production observed in GBM favors the hyper-proliferation and recruitment of endothelial cells in detriment to pericytes that provide coverage and support to the blood vessels [21]. This results in the formation of defective and permeable blood vessels that can easily collapse yielding hypoxic foci within the GBM. Low tumor oxygenation, also known as hypoxia, constitutes a major problem for GBM patients, since it promotes tumor cell spreading (invasion) into the healthy brain tissue in order to evade this adverse environment [22,23]. Tumor invasion not only constitutes a major obstacle to therapy, but it is also the main cause of death in GBM patients. For this reason, an increasing amount of research has been focused on characterizing the molecular and cellular mechanisms that regulate GBM cell invasiveness. Since the hypoxic environment is a major inducer of the GBM cell invasive phenotype, understanding how hypoxia triggers the GBM cells to invade is paramount for the development of novel and more effective therapies against this devastating disease. In this review, we will present a comprehensive

examination of the available literature regarding how GBM hypoxia triggers an invasive cancer cell phenotype, the role of these invasive proteins in GBM progression, and whether any of these are therapeutically targetable.

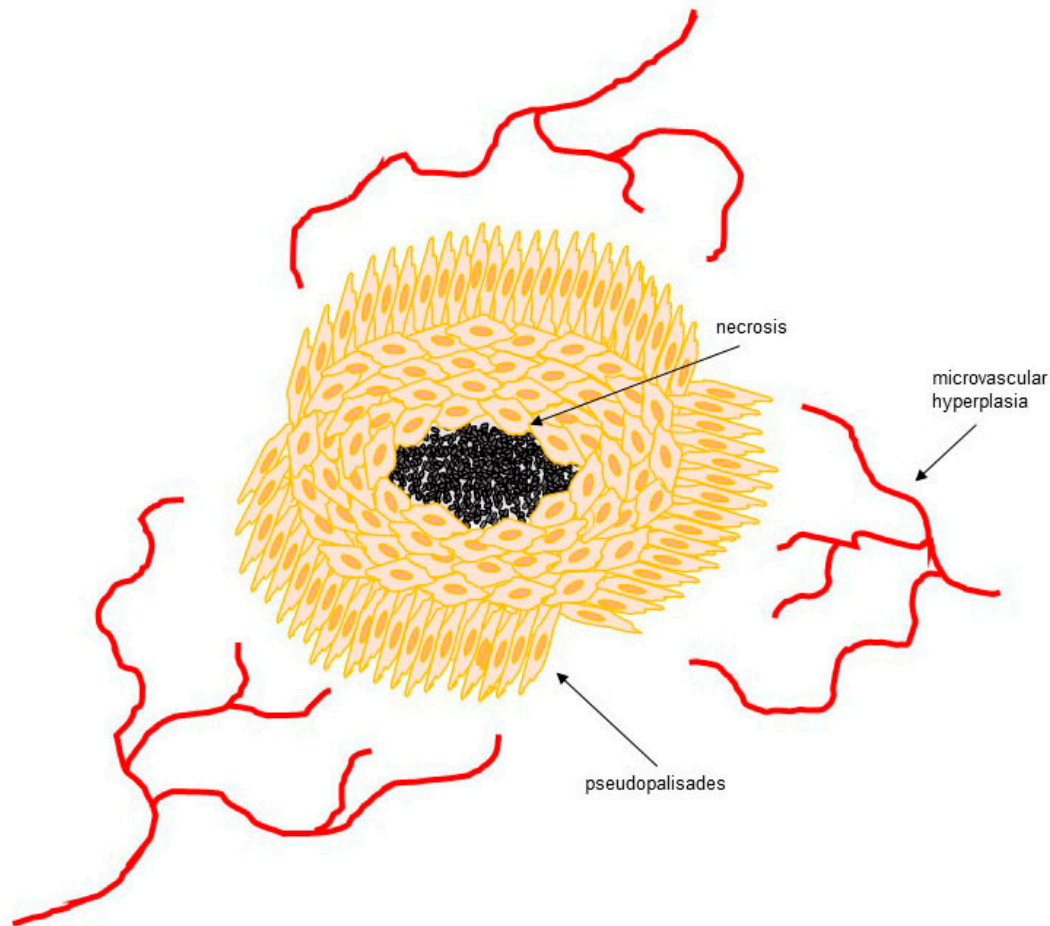


Figure 1. GBM distinctive pathological features. GBM is characterized by necrotic foci with surrounding cellular pseudopalisades and microvascular hyperplasia. Pseudopalisades are created by tumor cells migrating away from a central hypoxic (low oxygenated) region and forming an invasive front. Microvascular hyperplasia is an exacerbated form of angiogenesis that occurs in response to the secretion of proangiogenic factors (e.g., vascular endothelial growth factors (VEGFs), interleukin-8 (IL-8)) by the cells that form the pseudopalisades.

2. Hypoxia in GBM

Hypoxia is a condition in which an organism or a cell is deprived of adequate oxygenation (e.g., when oxygen demand exceeds oxygen supply, O_2 partial pressure (pO_2) less than 10 mmHg). Hypoxia is frequent in solid tumors, being the natural consequence of the increased oxygen diffusion distance due to tumor expansion. The clinical-pathological effects of hypoxia in GBM can be observed by magnetic resonance imaging (MRI) where significant oxygen diffusion restriction is detected, consistent with absent or defective blood flow [20,24,25]. Molecular markers of hypoxia, such as hypoxia inducible factor 1 (HIF-1) and VEGF staining, as well as tumor vascularity, can be detected on dynamic contrast enhanced MRI and correlate with worse progression-free and overall patient survival [24,26]. Microscopy analyses of GBMs reveal multiple hypoxic regions and a widespread invasion in the growing periphery of these tumors [23]. Hypoxia occurs in GBM due to increased cell proliferation/tumor growth linked to an erratic tumor neovascularization that leads to poor oxygen diffusion [27]. GBM vessels are tortuous, disorganized, highly permeable, and characterized by

abnormalities in their endothelial walls due to the lack of pericyte coverage [28–30]. These vessels also have significantly larger diameters and thicker basement membranes compared to those of the normal brain [31]. At the tumor tissue level, the occurrence of microvascular thromboses that lead to vessel occlusion is commonly observed, further promoting intratumoral hypoxia [25]. Intratumoral blood flow is thus impeded, creating a heterogeneous tumor environment with respect to oxygenation and interstitial fluid pressure [32]. Inconsistent intratumoral oxygenation leads to hypoxia, acidosis, and necrosis, whereas the increased hydrostatic pressure outside of the GBM vasculature promotes intratumoral edema, a major cause of morbidity for GBM patients [33].

3. HIF Transcription Factors

HIF transcription factors constitute the master regulators of the hypoxia adaptive response. HIFs are heterodimeric complexes constituted by O₂-regulated α subunits (HIF-1 α , EPAS1/HIF-2 α , or HIF-3 α) and a constitutively expressed β subunit, HIF-1 β , also known as an aryl hydrocarbon receptor nuclear translocator (ARNT) [34,35]. Of the three α subunits, HIF-1 α and HIF-2 α are the best studied and considered to be the main regulators of the hypoxia response. HIF-1 α is ubiquitously expressed, whereas HIF-2 α is selectively expressed in distinct cell populations [34]. HIF-1 α and HIF-2 α have both overlapping and distinct target genes [35] and are differentially regulated under various physiological and pathological conditions [36]. For instance, they play different roles in tumorigenesis depending on specific tumor microenvironments [34,35]. The existence of multiple variants of HIF-3 α has made it very challenging to elucidate HIF-3 functions. Distinct HIF-3 α variants are expressed in different tissues and are differentially regulated by hypoxia. While the full-length HIF-3 α protein has been shown to function as a transcription activator, triggering a unique transcriptional program in response to hypoxia [37], some HIF-3 α variants act as dominant-negative regulators of HIF-1/2 α functions [38]. The protein structures of HIF-1 α , HIF-2 α , and HIF-1 β subunits contain a basic helix-loop-helix (bHLH), a Per-Arnt-Sim (PAS) domain, and a C-terminal (C-TAD) domain, while HIF- α subunits contain an additional oxygen-dependent degradation domain (ODD) and an N-terminal domain (N-TAD). The bHLH and PAS domains are involved in heterodimer formation and binding to hypoxia responsive elements (HRE) within the promoters of HIF target genes, whereas the N-TAD and C-TAD domains are involved in transactivation through interactions with the transcriptional coactivators p300/CBP [39–41]. HIF-3 α can exist in many different variants, with distinct deletions of the domains described above (for a detailed description, see [38]). Under normoxic conditions (normal oxygen levels), two prolyl residues within the HIF- α subunits are hydroxylated by prolyl hydroxylases 1-3 (PHD1-3). This allows the binding of the von Hippel-Lindau (VHL) protein to the HIF- α subunit. The VHL protein then recruits E3 ubiquitin ligases, which target HIF- α for proteasomal degradation [42–44]. PHDs are 2-oxoglutarate- and iron-dependent dioxygenases, whose activities are positively regulated by O₂. Hence, hypoxia leads to PHDs inhibition and subsequent stabilization of the HIF- α subunits. HIF- α then translocates to the nucleus and binds to the HIF-1 β subunit and cofactors such as p300/CBP; this complex subsequently binds to HRE within promoters of target genes and orchestrates a concerted transcriptional response to hypoxia [45–47]. Factor-inhibiting HIF (FIH) is also an O₂ regulated hydroxylase that blocks the interaction between HIF- α and the transcriptional activators, p300/CBP, through asparagine hydroxylation of HIF α , attenuating HIF transactivation activity [48,49].

Genetic alterations that lead to HIF activation have been reported in GBM, further exacerbating the hypoxia response in this type of tumor. The activation of the epidermal growth factor receptor (EGFR) and the loss of tumor suppressor function (p53, PTEN) are common in GBM and can affect HIF expression. *EGFR* gene mutation and/or amplification is frequent in GBM [50]. The most common *EGFR* gene mutation (*EGFRvIII*) consists of the deletion of exons 2–7, resulting in a constitutively active and ligand independent receptor [51]. Initiation of EGFR signaling by ligand binding (which is exacerbated in GBM cells over-expressing EGFR) or gene mutation (i.e., *EGFRvIII*) results in activation of the PI3K/AKT/mTOR pathway with the subsequent up-regulation of HIF-1 α (Figure 2A) [52,53]. GBM has a 20% to 40% incidence of phosphatase and tensin homolog (*PTEN*) deletions [54]. *PTEN* is

the main inhibitor of the PI3K/AKT signaling pathway. Consequently, the loss of PTEN leads to increased HIF-1 α via the PI3K/AKT/mTOR pathway (summarized in Figure 2A). It has been proposed that p53 protein may lead to the inhibition of HIF activity in hypoxia by promoting MDM2-mediated ubiquitination and the degradation of HIF-1 α [55]. Therefore, the loss of the *p53* gene, which is common in GBM, will lead to HIF-1 α stabilization (Figure 2B). Other proteins that have been shown to regulate HIF-1 α expression under hypoxic conditions leading to increased GBM cell migration and invasion include FAT atypical cadherin 1, integrins α v β 3 and α v β 5, Hypoxia-inducible protein 2 (HIG2), and geranylgeranyltransferase I (GGTI) [56–59].

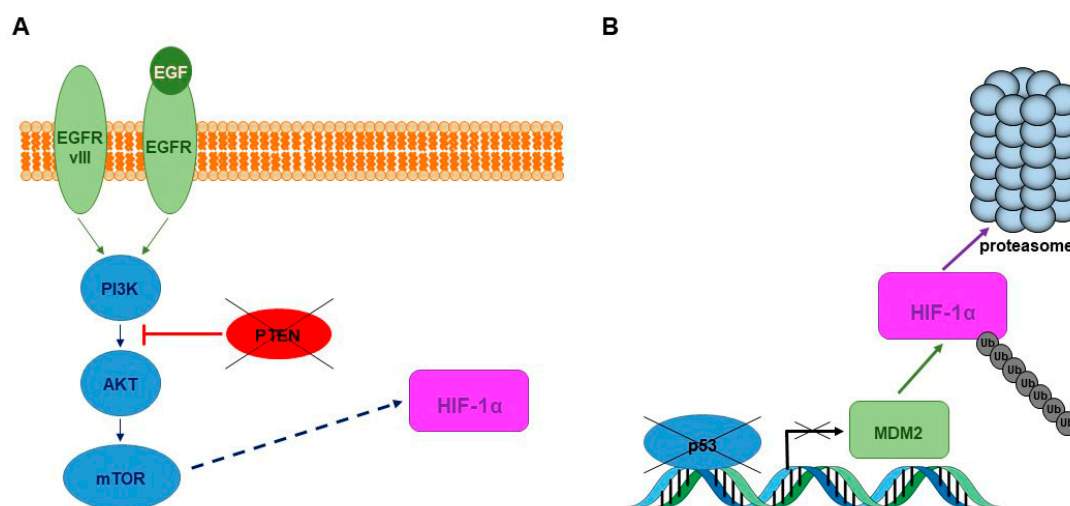


Figure 2. Genetic alterations that lead to HIF activation in GBM. (A) *EGFR* gene amplification and/or overexpression is frequent in GBM. The most common *EGFR* gene mutation (EGFRvIII) consists in the deletion of exons 2–7, resulting in a constitutively active and ligand independent receptor. Initiation of EGFR/EGFRvIII signaling by ligand binding, gene amplification, or mutation results in activation of the PI3K/AKT/mTOR pathway with the subsequent up-regulation of HIF-1 α . *PTEN* gene deletion is common in GBM. *PTEN* protein is the main inhibitor of the PI3K/AKT signaling pathway, as such loss of *PTEN* function leads to increased HIF-1 α via the PI3K/AKT/mTOR pathway; (B) It has been proposed that p53 may lead to inhibition of HIF activity in hypoxia by promoting MDM2-mediated ubiquitination and degradation of HIF-1 α . Therefore, the loss of the *p53* gene, which is common in GBM, will lead to HIF-1 α stabilization.

As a result of our understanding of the deregulation of the PI3K/AKT/mTOR signaling pathway in GBM, its targeting has attracted significant attention for the development of anti-cancer therapeutics. Initially, rapamycin analogs (mTOR inhibitors) were tested, although these demonstrated little impact on overall patient survival [60,61]. BKM120 (Novartis) is a pan-class I PI3K inhibitor without mTOR or Vps34 activity that can cross the blood brain barrier (BBB), which is critical for clinical applications against GBM. Preclinical studies demonstrated that the survival of NOD/SCID mice harboring intracerebral U87MG tumor xenografts significantly improved following BKM120 treatment [62]. To date, there is an ongoing phase II clinical trial with this agent in patients with first or second GBM recurrence (clinical trial #NCT01339052) that is being coordinated by the Dana-Farber/Brigham and Women’s Cancer Center, U.S.A. Beyond single agent inhibitors, the dual inhibition of both the PI3K and mTOR pathway is considered a more effective therapeutic strategy, the most extensively studied being BEZ235 (Novartis), a dual PI3K/mTORC1/2 inhibitor [63]. BEZ235 has been used in a wide range of in vitro studies [64] and tested against glioma cells where BEZ235 treatment induced G₁ cell-cycle arrest and autophagy, and reduced VEGF expression [65]. Furthermore, BEZ235 significantly increased the survival of tumor bearing mice [66]. Recent studies have highlighted conflicting results regarding the effectiveness of this agent in vivo where BEZ235 demonstrated antitumor efficacy with

improved survival against U87MG orthotopic gliomas in one study [67]. However, very little to no efficacy was observed in another independent orthotopic xenograft model study [68]. It is important to note that many PI3K/mTOR inhibitors demonstrate weak affinities for ABC transporters, but in spite of this, can still achieve target inhibition in GBM albeit with modest single-agent efficacy. For this reason, it is likely that these drugs will require combination-treatments with other (BBB penetrable) inhibitors and agents.

HIF-induced transcription regulates hundreds of genes that promote angiogenesis, erythropoiesis, migration, cell survival, proliferation, epithelial to mesenchymal transition (EMT), the recruitment of inflammatory cells, invasion, metastasis and metabolic reprogramming, and of particular importance, the shifting of cellular metabolism from oxidative phosphorylation to glycolysis [36,45,69–71]. However, the alterations of specific genes and their clinical significance in GBM remain to be better explored. Hypoxia is a main promoter of GBM invasion which is tightly linked with poor patients' survival and chemoresistance. For this reason, identifying and characterizing hypoxia induced proteins that regulate GBM invasion is key for the development of novel and more effective therapies against this deadly type of tumor.

4. Hypoxia Driven Up-Regulation of Invasion Proteins in GBM

In this review, we will perform a thorough examination of the literature focused in identifying hypoxia responsive invasion genes and proteins and characterizing the molecular mechanisms by which these proteins contribute to GBM progression.

4.1. Extracellular Matrix Degradation and Remodeling

Degradation and remodeling of the extracellular matrix (ECM) that surrounds the tumor are essential processes for cancer cell invasion, creating spaces and scaffolding that allow tumor cell proliferation/growth and migration (spreading) away from the initial tumor site [72].

Carbonic anhydrase (CA) IX has been shown to be over-expressed in GBM [73–75] and CAIX gene expression has been shown to be regulated by hypoxia [76]. High expression of CAIX was identified as an independent prognostic factor for poor survival in patients with GBM [77]. An *in vitro* study using an siRNA approach to knockdown CAIX expression in the GBM cell lines, U251 and Ln 18, showed that CAIX was not only important for GBM cell attachment and invasion, but also for providing radio- and chemo-resistance [77]. CAIX is a membrane bound, zinc dependent enzyme that catalyzes CO₂ hydration into bicarbonate with a proton release [78]. Since CO₂ is a main by-product of oxidation, CAs play a main role in maintaining acid/base homeostasis in cells. CAIX has the highest catalytic activity among all membrane bound CAs due to the presence of a unique proteoglycan fragment located close to its active site [79,80]. The CAIX active site is located at the outer side of the plasma membrane, so protons are generated on the extracellular space. The extrusion of hydrogen ions into the extracellular space by CAIX thus contributes to the regulation of the intracellular pH, and at the same time, to the acidification of the extracellular space surrounding the GBM, thus leading to ECM and surrounding tissue degradation and creating an environment conducive to enhanced invasion.

Acetazolamide (a CAIX inhibitor) has been shown to effectively treat 2D or 3D GBM models [81]. These results indicate that CAIX constitutes a promising therapeutic target for GBM. Furthermore, the effectiveness of this agent was significantly enhanced when acetazolamide was incorporated into a polymeric poly(ethylene glycol)-block-poly(lactide-co-glycolide) nano-carrier (that readily crosses the BBB) and used to treat 3D spheroid models [81]. A second CAIX inhibitor, Indisulam, is currently in Phase II of clinical trials for the treatment of a range of cancers, including metastatic melanoma, lung, and metastatic breast cancer where brain metastasis is frequently observed [82].

Epidermal growth factor receptor (EGFR) is amplified in approximately 50% of all GBMs. Expression of the mutant variant of EGFR that lacks exons 2–7 (EGFRvIII), leading to the constitutive activation of this receptor, is particularly common and has been shown to be tightly linked with poor patients' prognosis [83]. A recent study using U87MG and LN229 GBM cell lines over-expressing

EGFRvIII showed that hypoxia stimulates the interaction of EGFRvIII with the integrin $\beta 3$ in GBM cells, leading to EGFRvIII protein stabilization. Tumor cells adhere to the ECM via integrin receptors which induce the activation of focal adhesion signaling and the subsequent degradation of the ECM, creating a path for cancer cell invasion of adjacent tissues [84]. Interaction of integrin $\beta 3$ with EGFRvIII stimulated the activation of an integrin $\beta 3$ /SRC/FAK/EGFRvIII signaling axis that led to the activation of ERK1/2 (MAPK), AKT, and STAT3 signaling pathways in the GBM cells and the up-regulation of the cancer cell invasion markers, matrix metalloproteinase-2 (MMP-2) and MMP-9, as well as integrin $\beta 3$, subsequently promoting GBM cell invasion [85].

EGFR is an extremely compelling therapeutic target for GBM treatment. In particular, mutant EGFRvIII [86–88], which is reported in approximately 25–33% of all GBM cases [89]. The exclusive expression of EGFRvIII can be targeted by multiple approaches including CAR T-cell therapy, therapeutic vaccines, specific antibodies, or bi-specific T-cell engager approaches, (all extensively reviewed in [90,91]). While there are clinical trials proposed for these agents, there are still significant difficulties waiting to be solved in order to achieve the clinical translation of these agents into GBM patients. These include off-tumor toxicity, potential cross reactivity complications, and a limited tumor penetration capacity. However, the combination of targeting EGFRvIII and other treatment modalities is considered to have significant potential to improve GBM patient prognosis.

Integrins $\alpha v\beta 3$ and $\alpha v\beta 5$ have also been shown to be recruited to the cell membrane of GBM cells (e.g., U87MG and SF763) in response to hypoxia, leading to the activation of the focal adhesion kinase (FAK) protein to promote tumor invasion [58]. This study also showed that the inhibition of integrins $\alpha v\beta 3$ or $\alpha v\beta 5$, either by using a specific inhibitor or by siRNA, led to the decrease of HIF-1 α intracellular levels. The integrin-dependent HIF-1 α regulation was mediated by FAK/RhoB dependent inhibition of GSK3- β which promotes HIF-1 α degradation in a VHL-independent manner [92].

The proteins of the integrin family constitute promising therapeutic targets. Cilengitide, the most advanced specific integrin inhibitor, has demonstrated anti-tumor activity in both phase I and phase II clinical trials for recurrent GBM [93–95]. Treatment of newly diagnosed GBM patients with Cilengitide in combination with conventional therapy (radio-therapy and temozolomide) has been encouraging [96], suggesting synergy of this drug with concomitant chemo- and radiation therapy. A pivotal phase III study (CENTRIC) in newly diagnosed GBM patients was conducted [97]. Within this large phase III study, 3471 GBM patients were screened and 3060 had their *MGMT* status tested. 926 tumours exhibited a methylated *MGMT* promoter, and from this cohort, 545 were randomly assigned to the cilengitide ($N = 272$) or control ($N = 273$) groups. Patient median overall survival was 26.3 months in the cilengitide treatment group and 26.3 months in the control group [97]. Unfortunately, none of the predefined clinical subgroups showed a benefit from cilengitide treatment although the clinical trial did report that there was no overall additional toxic effects with cilengitide treatment [97]. The addition of cilengitide to radiotherapy and temozolomide chemotherapy did not improve patient outcome and the authors state that they will no further develop cilengitide as an anti-GBM drug. Alongside the CENTRIC study, a second independent CORE clinical trial has been conducted (clinical trials identifier NCT00813943). CORE is a Phase II clinical trial for newly diagnosed GBM that recruited 265 GBM patients with the unmethylated *MGMT* gene promoter and incorporated a two-times weekly or an intense, five-times weekly 2000 milligram cilengitide regimen in combination with conventional treatment (temozolomide with concomitant radiation therapy, followed by temozolomide maintenance therapy) for comparison to conventional treatment alone. Temozolomide and radiotherapy treatment alone ($N = 89$) resulted in an overall survival (OS) time of 13.4 months. In comparison, two-time weekly cilengitide treatment ($N = 88$) revealed an OS of 16.3 months. In contrast, intense cilengitide treatment ($N = 88$, at the same 2000 milligram dose) had an overall survival of 14.5 months. When the cilengitide (two-times weekly, in combination with temozolomide and radiotherapy) treatment was compared to temozolomide and radiotherapy alone, and log rank calculated, a p value of 0.0328 was reported, indicating that bi-weekly cilengitide alongside conventional treatment significantly improved overall patient survival. It is important to

note, that while this CORE study has been completed and data is available (<https://clinicaltrials.gov/ct2/show/NCT00813943>), this data has not yet been published. Despite the CENTRIC study set back, promising data from the CORE study suggest that the integrins remain potential treatment targets for GBM.

Procollagenlysinase 2-oxoglutarate 5-dioxygenase 2 (PLOD2) is an enzyme that catalyzes collagen cross-linking. A meta-analysis of seven independent GBM datasets, consisting of 861 GBM samples, revealed that *PLOD2* was significantly overexpressed across all GBM datasets analyzed compared to a normal brain [98]. Analysis of *PLOD2* mRNA expression from the Repository of Molecular Brain Neoplasia Data (REMBRANDT) database ($N = 178$), further showed that *PLOD2* mRNA expression was significantly increased in GBM compared to lower grade gliomas and normal brain tissue [98]. Moreover, immunohistochemistry studies demonstrated increased *PLOD2* expression associated with increasing tumor grade. Staining for *PLOD2* protein was absent or weak in non-neoplastic brain tissue samples and grade II astrocytomas, increased in grade III astrocytomas and enhanced to an even greater extent in GBM [98]. Analysis of The Cancer Research Atlas (TCGA) and the REMBRANDT databases indicated that *PLOD2* can constitute an effective diagnostic marker to distinguish lower grade gliomas from GBM. Moreover, Kaplan-Meier plots based on the TCGA dataset showed significant lower overall survival (OS) and progression free survival (PFS) in GBM patients with high *PLOD2* expression compared to patients with low *PLOD2* [98]. In vitro studies using U87MG and U251 GBM cell lines showed that hypoxia regulates *PLOD2* protein expression in a HIF-1 α dependent manner and that the knockdown of *PLOD2* by shRNA significantly inhibits GBM cell migration and invasion [98]. The molecular mechanism by which *PLOD2* contributed to GBM cell migration and invasion involved the phosphorylation of FAK (Tyr 397), leading to increased focal adhesion plaques under hypoxic conditions [98]. Cancer cell invasion is closely related to its ability to stably adhere to the ECM [99] and this process is regulated by a key protein, FAK [100,101]. In vivo studies using subcutaneous and orthotopic xenograft mouse models further showed that *PLOD2* promoted cross-linking of collagen fibers around the tumors and enhanced ECM stiffness [98]. The formation of collagen cross-links is initiated by *PLOD2*, which specifically hydroxylates lysine residues of the collagen telopeptide area [102]. Enhanced ECM stiffness is commonly reported in many types of solid tumors, including gliomas [103,104]. ECM stiffness increases principally as a result of collagen deposition and crosslinking, and this has been shown to induce cancer cell invasion and metastasis [103–105].

There has been a concerted effort to target *PLOD2*, particularly in metastatic disease. *PLOD2* inhibition not only affects the cancer cells themselves, but it also impacts on the tumor microenvironment, thus impairing metastasis. To date, several pharmacologic inhibitors have been confirmed to have an anti-metastatic effect, while some compounds could be direct *PLOD2* inhibitors (reviewed in [106]). Minoxidil (a *PLOD2* inhibitor) suppresses the expression of the *PLOD2* protein, and significantly reduces cancer cell migration, resulting in an anti-metastatic effect both in vitro and in vivo [107,108]. Berberine can also suppress the expression of *PLOD2* and has been shown to inhibit pulmonary metastasis in melanoma [109]. Amentoflavone [110] and beta-carotene [111] also decrease *PLOD2* expression, reducing metastasis. Even though there have been substantial advances in targeting *PLOD2* in different types of cancer, to date, the effectiveness of these anti-*PLOD2* agents in GBM models remains unknown.

4.2. Epithelial to Mesenchymal Transition (EMT)

Increasing evidence supports that detachment of cancer cells from the initial tumor site is initiated by the activation of an embryonic development program referred to as epithelial to mesenchymal transition (EMT), whereby epithelial cells lose apicobasal polarity and cell–cell contact and acquire a more motile and invasive mesenchymal phenotype [112].

CBF1 (also known as Recombination signal Binding Protein for immunoglobulin kappa J, RBPJ) constitutes a key transcriptional regulator of the Notch signaling network. The Notch pathway not only regulates stem cell maintenance, but is also a potent inducer of EMT in a variety of

tumors [113]. A recent study showed that down regulation of CBF1 expression by shRNA in GBM cells (e.g., GBM1, 407p, JHH) leads to the efficient suppression of EMT activators, including zinc finger E-box-binding protein 1 (ZEB1) protein, *CD44*, and *SNAIL1* genes. CBF1 depletion also correlated with the inhibition of invasion in GBM cells [114], which could be due, at least in part, to the down-regulation of the EMT inducing proteins and genes observed in these cells. To characterize the clinical importance of CBF1, the authors assessed its predictive value by analyzing clinical data sets. This analysis showed that *CBF1* gene expression (assessed by RNA seq) was principally high in pseudopalisades and the peri-necrotic regions of GBM, which are known to be hypoxic areas of the tumor that contain highly invasive cells [114]. Examination of the expression levels of CBF1 in various brain tumor types (as classified by the WHO), showed that GBM has relatively low CBF1 expression [114]. The authors also reported that in two independent data sets (French and TCGA) there was a significant correlation between high CBF1 and patient overall survival; however, this was not observed in another two cohorts (German Glioma Network and Chinese cohort). CBF1 levels were significantly increased in proneural glioblastoma as compared with any other molecular sub-groups as classified by Verhaak et al. [114,115]. Importantly, GBM with mutant IDH1 showed significantly increased *CBF1* expression compared to tumors with wild-type IDH1 [114]. In another study, in silico analysis of the TCGA data set showed that CBF1 was highly expressed in GBM compared to non-neoplastic brain tissue [115]. This study also showed that high expression of CBF1 in *IDH* wild-type GBM correlated with a slightly worse patient's prognosis [115]. Taking into account these two studies, it is currently still unclear if CBF1 could be a prognostic factor for GBM.

Zinc finger E-box binding Homebox 1 (ZEB1) is a transcriptional repressor which has been shown to promote tumor invasion and metastasis [116]. Analysis of the TCGA database showed that ZEB1 is a short survival indicator in GBM patients and a potential therapeutic target [117]. Immunofluorescence and immunohistochemistry studies using human clinical samples showed that ZEB1 is expressed along the tumor invasive front, in pseudopalisades, which are associated with hypoxic areas of the GBM [117,118]. In vitro studies, using a combination of inhibitory drugs and shRNA approaches, showed that hypoxia induces ZEB1 expression and nuclear localization in GBM cells in a HIF-1 α dependent manner leading to EMT and increased migration/invasion [118,119]. ZEB1 inhibits the expression of the miRNA-200 family, which suppresses stem cell factors [116]. In GBM, the ZEB1-miRNA-200 feedback loop targets specific stem cell regulators, namely SOX2, OLIG2, and CD133, leading to the activation of the EMT program (Figure 3). Knockdown of ZEB1 in three cell lines generated from primary glioblastoma specimens (hGBM L0, L1, and L2) inhibited invasion [117]. Mechanistically, ZEB1 positively regulated the ROBO1 protein that has been shown to sever the anchorage of N-cadherin to the cytoskeleton leading to increased GBM cell motility and invasion (Figure 3) [117,120]. ZEB1 was also found to play an important role in temozolomide resistance in GBM cells, via up-regulation of c-MYB by the ZEB1-miR-200 feedback loop. c-MYB, in its turn, binds to the *MGMT* promoter in GBM cells, leading to increased expression of the *MGMT* protein that repairs DNA damage caused by alkylating agents such as temozolomide (Figure 3) [117].

TWIST1 transcription factor is also a critical hypoxia induced invasion protein [121]. Physiologically, TWIST1 constitutes a master regulator of embryogenic processes, such as gastrulation and mesoderm differentiation, via the induction of EMT [122,123]. However, TWIST1 has also been involved in pathological processes such as tumor metastasis [121,124,125]. Analysis of *TWIST1* mRNA expression in the publically available databases, REMBRANDT and ATCG, showed that *TWIST1* was more highly expressed in GBM compared to lower grade gliomas [126]. Furthermore, within GBM, the highest *TWIST1* expression was observed in the mesenchymal subtype and lowest expression in the proneural subtype. Elevated *TWIST1* expression was also associated with poor patients' overall survival [126]. Immunohistochemistry analysis of TWIST1 in 269 infiltrating astrocytoma samples showed a stronger expression of TWIST1 associated with a higher malignancy grade [126]. In addition, positive TWIST1 expression predicted a worse patients' outcome, which was particularly striking in GBM patients [126]. In vitro studies showed that *TWIST1* mRNA transcription is regulated by tumor

hypoxia in a HIF-1 α dependent way [127]. The TWIST1 protein was shown to promote an invasive phenotype in GBM cells [128,129], ex vivo using organotypic brain slice cultures and in vivo using a orthotopic xenotransplant mouse model [128]. Nevertheless, TWIST1 did not stimulate the canonical switch between E-cadherin and N-cadherin in GBM cells [128]. Instead, TWIST1 induced changes in cellular interactions (cell-cell and cell-substrate) and cytoskeleton reorganization through alternative pathways in GBM to induce migration and invasion [128], described in detail in Table 1.

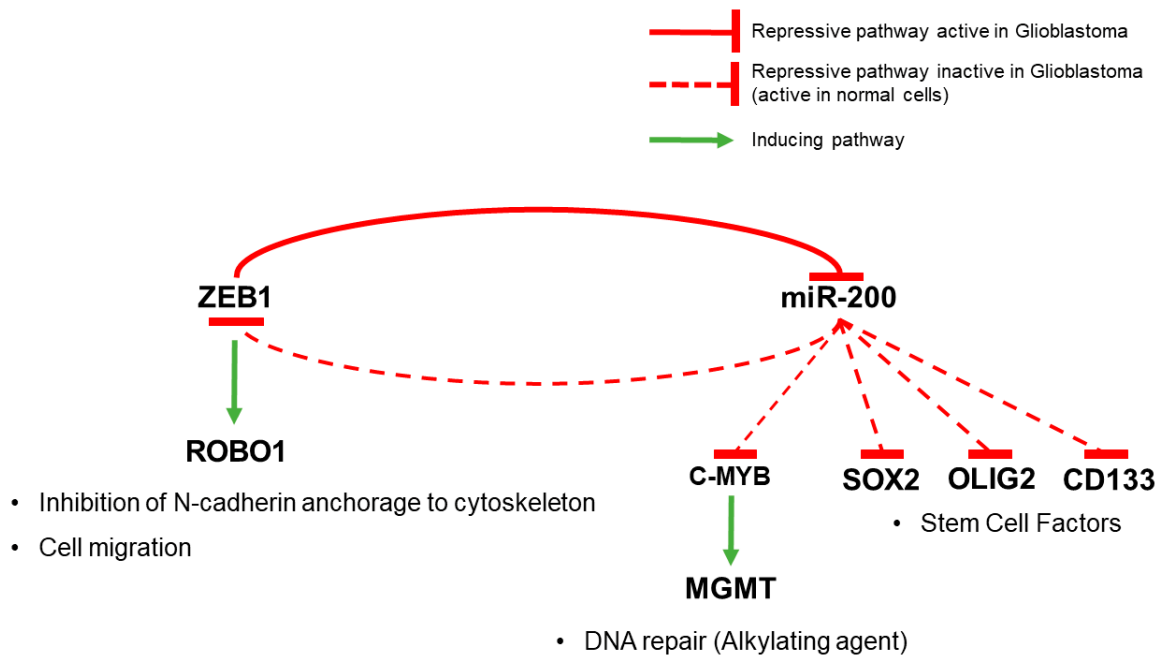


Figure 3. Glioblastoma ZEB1-miRNA-200 feedback loop interactions. In GBM, the ZEB1-miRNA-200 feedback loop targets specific stem cell regulators, namely SOX2, OLIG2, and CD133. ZEB1 up-regulation of c-MYB by the ZEB1-miRNA-200 feedback loop leads to increased expression of the MGMT protein that repairs DNA damage caused by alkylating agents such as temozolomide. ZEB1 positively regulates the ROBO1 protein that has been shown to sever the anchorage of N-cadherin to the cytoskeleton leading to increased GBM cell motility.

Table 1. TWIST1 induced pathways and proteins in GBM.

Protein/Pathway	Role
Periostin	Recruitment of M2 tumor-associated macrophages which are tumor-supportive and immunosuppressive [130].
Fibronectin 1	Promotes cell cohesion and invasion of basement membrane [131].
SPARC	Upholds ECM degradation through the uPA-uPAR pathway and stimulates survival, proliferation and invasion via the PI3K pathway [132].
SNAI2	Inhibits miR-34 activity, a regulatory microRNA for differentiation, promoting stemness [133,134].
ID1	Activates tumor progression pathways, such as Snail, ERK1/2 and AKT, and promotes stem cell self-renewal transcription factors Sox2, Oct3/4 and Nanog [135].
HGF	Promotes survival, proliferation, transformation and invasion through activation of the PI3K/AKT, STAT3/JNK, SOS/RAS/ERK/MAPK pathways [136].
LOX	Activates HIF-1 α (via AKT pathway in a positive feed-back loop), FAK and VEGF gene expression [137].
Cadherin 11	Regulates cell-cell interactions and survival, and promotes cell migration [138].
BMI1/EZH2	Part of the Polycomb repressive complex 1 and 2, in that order, promotes gene silencing of the P16 and P14 tumor suppressors, and inhibits cancer stem cells differentiation [139].

4.3. Chemokine Receptors

Several studies have shown that C-X-C chemokine receptor type 4 (CXCR4) is highly expressed in GBM, particularly in hypoxic pseudopalisading areas, and plays a critical role in GBM cell invasion [140–143]. Importantly, no CXCR4 staining was observed in healthy brain tissue [140,141]. These results suggest that CXCR4 expression in GBM might be modulated by hypoxia. In fact, an in vitro study showed that hypoxia leads to the up-regulation of CXCR4 in GBM cell lines (e.g., U87MG and LN308) in a HIF-1 α dependent manner [140]. Migration of both U87MG and LN308 cells under hypoxic conditions was significantly increased compared to control normoxic cells and the inhibition of CXCR4 significantly repressed the hypoxia-stimulated migration of GBM cells [140]. CXCR4 is a chemokine receptor for stromal cell-derived factor-1 α (SDF-1 α), also known as CXCL12. HIF-1 induces SDF-1 α expression on hypoxic endothelial cells [144]. Therefore, over-expression of CXCR4 in cancer cells could lead to their migration towards blood vessels.

Inhibiting the CXCR4/SDF-1 α axis following local tumor irradiation has shown significant clinical promise and can complement standard GBM treatment. AMD3100 (Plerixafor), an inhibitor of the CXCR4/SDF-1 α interaction [145], has been shown to significantly interfere with the vasculogenesis pathway. Treatment of GBM-bearing mice with AMD3100 immediately after irradiation prevented the radiation-induced recruitment of bone marrow-derived cells into the tumor site and critically, AMD3100 exposed tumors could be treated using radio-therapy regimens which were otherwise ineffective [146]. More recently, peptide R, a new specific CXCR4 antagonist, has been developed using a ligand-based screening approach and was shown to impair the metabolic activity and proliferation of U87MG GBM cells. Furthermore, this agent stably reduced CXCR4 expression and cell migration in response to CXCL12 in vitro. Peptide R was shown to reduce tumor cellularity, promote anti-tumor features of glioma-associated microglia/macrophages and astrogliosis, and hinder intra-tumoral vasculature [147]. Overall, CXCR4 targeted therapy has the potential to improve GBM prognosis by supporting conventional therapeutic treatments, as well as potentially treat GBM where standard frontline treatments fail.

C-C chemokine receptor type 5 (CCR5) mRNA levels have been shown to be significantly and gradually up-regulated with tumor grade, grade II ($N = 15$), grade III ($N = 15$), and grade IV ($N = 70$) gliomas compared to adjacent non-neoplastic brain tissues [148]. Moreover, immunohistochemistry staining revealed that CCR5 protein expression was gradually stronger with increasing tumor grade [148]. Kaplan–Meier analysis further showed that patients with high levels of CCR5 expression have significantly shorter disease free survival and overall survival, indicating that CCR5 may be a potential prognostic marker for GBM [148]. A recent study showed that hypoxia induces CCR5 up-regulation in the GBM cell line, U87MG [149]. Interestingly, hypoxia also enhanced the expression of the CCR5 ligand, CCL4, in THP-1-derived macrophages. Knocking down CCR5 by shRNA in U87MG cells led to a significant decrease in the invasive capability of hypoxic U87MG cells which was concomitant with the decreased transcription of MMP-9 [149]. Even though these results are interesting and seem to suggest a potentially important role for the CCL4-CCR5 axis in GBM invasion, reproducibility of these experiments using other GBM model cell lines is paramount to strengthening the significance and broadness of these findings.

Interestingly, both CXCR4 and CCR5 chemokine receptors (that have been shown to be up-regulated by hypoxia in GBM), are co-receptors for Human Immunodeficiency Virus 1 (HIV-1). Namely, CXCR4 is the co-receptor for strains that infect T-cells (T-tropic or X4 strains), while CCR5 is the co-receptor for HIV-1 strains that infect macrophages (M-tropic or R5 strains). For this reason, pharmacological drugs targeting these proteins have been developed for the treatment of HIV patients. This is in fact the case for AMD3100 [150] that has been repurposed for cancer therapy as described above.

Studies have also been made to evaluate the use of the FDA approved HIV-1 drug, maraviroc (Pfizer) [151], for the treatment of GBM. Maraviroc is a CCR5 receptor antagonist [152]. Treatment of cancer patients with maraviroc has shown promising results in inhibiting invasion and metastasis in

different types of cancer [153–156]. Unfortunately, to date, the results regarding the use of this drug for GBM treatment are disappointing. A study tested several currently marketed non-chemotherapeutic agents for their ability to enhance/synergize the effect of temozolomide, the current standard chemotherapeutic drug used in the clinic for GBM treatment [157]. This study found that maraviroc (20–180 μ M) had no cytotoxic effect against the GBM cell line, GAMG.

4.4. Cytoskeleton Dynamics

Microarray-based gene expression studies using glioma or neuroblastoma cells have shown that knockdown of HIF-1 α inhibits the expression of the cyclin G2 encoding gene, *ccng2* [76,158,159]. Furthermore, immunohistochemistry studies showed the expression of cyclin G2 in hypoxic regions of GBM, principally in pseudopalisades [160]. In addition, in vitro studies using U87MG, U251MG, and LN2308 GBM cell lines showed a significant up-regulation of cyclin G2 expression in response to hypoxia [160]. This study also showed that HIF-1 α binds to the *ccng2* promoter and that knockdown of *HIF-1 α* by shRNA inhibited the hypoxia-induced expression of cyclin G2. These results indicate that hypoxia is a main regulator of cyclin G2 expression in GBM. Cyclin G2 was first identified as a negative regulator of the cell cycle [161] and later it was shown to induce cell cycle arrest in a p53-dependent manner [162]. Moreover, cyclin G2 was also shown to bind to and stabilize microtubules [162]. A more recent study demonstrated that cyclin G2 plays an important role in the regulation of hypoxia induced migration and invasion in GBM cells (e.g., U87MG and U251MG) [160]. This study revealed that mechanistically cyclin G2 is able to recruit cortactin to the leading edge of migrating GBM cells, promoting the subsequent tyrosine phosphorylation of cortactin, which is essential for ruffle formation and tumor cell invasion [160].

4.5. Hemostasis

Tissue factor (TF) has been shown to be upregulated in GBM cells in response to hypoxia [25,163,164]. TF is a main regulator of hemostasis, initiating the blood coagulation cascade upon binding to its ligand, factor VII (FVII) [165]. Expression of TF has been shown to positively correlate with the histological grade of gliomas, as well as with the extent of necrosis [166]. A more recent analysis of a large patient cohort of the TCGA data platform ($N = 424$) revealed that *TF* mRNA levels are significantly upregulated in GBM compared to normal brain tissue samples [167]. Kaplan–Meier survival analysis of the REMBRANDT database ($N = 213$) further determined a significant reduction in overall survival in GBM patients with high levels of *TF* expression [167]. Moreover, immunohistochemistry studies revealed that TF is predominantly expressed in perinecrotic and perivascular areas of human GBM [167]. Given the key role of TF in the regulation of blood clotting, increased expression of this protein in tumor cells has been shown to correlate with hyper-coagulation in malignant gliomas, further promoting a hypoxic environment [25,165]. In addition to its pro-thrombotic function, TF has also been shown to constitute a transmembrane receptor that regulates intracellular signaling pathways [165,168,169]. TF/Factor VII (FVII) signaling has been shown to play an important role in glioma cell growth, migration, and invasion [170]. This study suggested that the effects of TF/FVII were mediated through the downstream activation of PAR-2 and the ERK1/2 MAPK signaling pathways.

There has been some limited research focusing on targeting TF directly (as opposed to ERK 1/2 MAPK inhibitors). A murine xenograft model where human GBM cells of MZ-18 were transplanted into nude mice brains showed that treatment of these mice with a monoclonal antibody against TF (mAb TF9-10H10), using an intracranial osmotic pump system for delivery, significantly inhibited MZ-18 cell invasion compared to mock-treated control animals [167]. The extent of activated blood vessels was also reduced upon anti-TF treatment. These results suggest that targeting TF might be a promising treatment strategy for GBM therapy, by inhibiting both tumor invasion and tumor vasculature. However, clinical trials still need to be performed to test these encouraging results in GBM patients.

4.6. Others

Over-expression of the pro-motility receptor protein tyrosine kinase, ephrin type-A receptor 2 (EphA2), has been observed in GBM and associated with poor prognosis/lower overall survival [171–173]. In vitro studies have further shown that EphA2 induces GBM cell motility and invasion [174–176]. EphA2 activity is antagonized by the EphA2 ligand, ephrin A1 [151,154,155], which is frequently suppressed in GBM, further promoting EphA2 signaling [172,177]. EphA2 coordinates signaling from a variety of receptor tyrosine kinases (RTKs) via growth factor mediated activation of AKT, which initiates AKT-dependent EphA2 phosphorylation at residue S897 [175]. The resulting P-S897-EphA2 is required for lamellipodia formation and subsequent GBM cell motility and invasion [175]. The eHsp90/LRP1 complex was identified as a key regulator of EphA2-dependent GBM cell motility and invasion through its ability to sustain AKT-dependent phosphorylation of EphA2 at residue S897 [178]. Interestingly, hypoxia elicited increased expression and cell surface levels of both eHsp90 and LRP1 in G48a, U87MG, and U251 GBM cells. Moreover, hypoxia-mediated up-regulation of eHsp90 and LRP1 led to amplification of the eHsp90-LRP1 signaling axis, as assessed by robust activation of AKT and EphA2, and concomitantly induced cell motility and invasion [178].

Efforts have been made in the development of HSP90-directed therapies. Several pharmacological inhibitors that target the N-terminal ATP-binding domain of HSP90 have been developed, including ansamycins, purine analogs, and resorcinol derivatives [179,180]. Preclinical and clinical trials have demonstrated the antitumor efficacy of these drugs in different cancer types [180,181]. However, to date, no clinical trials with HSP90 inhibitors have been performed in GBM patients.

Transient receptor potential 6 (TRPC6) mRNA and protein levels have been shown to be up-regulated in GBM samples compared to normal brain tissue [182,183]. TRPC6 is a member of the Transient Receptor Potential (TRP) superfamily of cation channels [184]. In vitro studies using a combination of inhibitory drugs and shRNAs for Notch1 or TRPC6, showed that hypoxia induces the expression of TRPC6 in a Notch1 dependent manner in U373MG GBM cells [182]. Mechanistically, hypoxia induced TRPC6 expression caused a sustained elevation of intracellular calcium levels leading to the activation of the calcineurin-nuclear factor of the activated T-cell (NFAT) signaling pathway and subsequent promotion of GBM cell growth, invasion, and angiogenesis [182].

5. Conclusions

GBM, the most common and deadly type of brain tumor, is characterized by extensive hypoxic foci that trigger an invasive cancer cell phenotype. Tumor invasion is a major contributor to GBM chemoresistance and patients' mortality. For this reason, the identification and characterization of proteins involved in hypoxia induced invasion in GBM is critical for the development of novel and more effective therapies against this deadly disease. The treatment protocol for GBM patients has not changed since 2005, and temozolomide treatment in particular has been shown to be primarily limited to tumors with *MGMT* promoter methylation [5]. In this review, we have compiled, to the best of our knowledge, the progress made in identifying hypoxia responsive invasive proteins in GBM, the molecular mechanisms by which these proteins contribute to GBM migration and invasion, and the advancements made regarding the development of pharmacological drugs against these proteins. Proteins involved in ECM degradation and remodeling (e.g., CAIX, integrins $\beta 3$, $\alpha v \beta 3$ and $\alpha v \beta 5$, PLOD2, MMP-2, MMP-9); EMT (e.g., CBF1, ZEB1, TWIST1); chemokine receptors (e.g., CXCR4, CCR5); cytoskeleton dynamics (e.g., cyclin G2); hemostasis (e.g., TF, factor VII); the pro-motility receptor, EphA2; and the cation channel protein, TRPC6, have been shown to contribute to GBM migration and invasion in response to hypoxia. Of these, CAIX, integrins, PLOD2, CXCR4, EGFRvIII (that induces up-regulation of MMP-2, MMP-9, and integrin $\beta 3$ in a HIF-1 α /hypoxia dependent manner), HSP90 (which induces hypoxic activation of EphA2) and to a limited extent TF have been shown to be potentially promising targets for GBM treatment. Phase I and II trials are currently being pursued for CAIX, EGFRvIII, PLOD2, and EphA2 targeting chemotherapeutics in different types of cancer. However, clinical trials in GBM patients have not yet been reported for these

drugs. Even though a phase III study for the integrins targeting drug, cilengitide (CENTRIC), in newly diagnosed GBM patients showed disappointing results, another independent phase II study with this drug (CORE) showed that bi-weekly treatment with cilengitide alongside conventional therapy significantly improved overall patient survival. While the CENTRIC study was performed in patients with GBM tumors containing a methylated *MGMT* promoter, the CORE study was done in patients with an un-methylated *MGMT* promoter. These results suggest that cilengitide might be beneficial in the treatment of tumors with an un-methylated *MGMT* promoter.

A major obstacle for brain tumor treatment is the BBB. Many of the chemotherapeutics that have shown promising in vitro results against GBM cells do not have the capacity to cross the BBB. For this reason, the development of drug delivery systems that can transport these drugs through the BBB is equally imperative.

Acknowledgments: Patrícia A. Madureira (P.A.M.) is the recipient of an FCT Investigator contract from the Foundation for Science and Technology (Fundação para a Ciência e a Tecnologia, FCT) of Portugal, ref: IF/00614/2014, and an FCT exploratory grant, ref: IF/00614/2014/CP12340006. A.R.M. is the recipient of an ERASMUS+ fellowship. The Centre for Biomedical Research (CBMR) is financed by an FCT Research Center Grant, ref: UID/BIM/04773/2013 CBMR 1334. Research in the Portsmouth Brain Tumour Research Centre is core funded by the charity, Brain Tumor Research.

Author Contributions: P.A.M., A.R.M., and R.H. wrote the manuscript. P.A.M. and A.R.M. designed the figures. P.A.M. conceptualized the structure and edited the manuscript. P.A.M. and G.J.P. reviewed the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Gilbert, M.R.; Wang, M.; Aldape, K.D.; Stupp, R.; Hegi, M.E.; Jaeckle, K.A.; Armstrong, T.S.; Wefel, J.S.; Won, M.; Blumenthal, D.T.; et al. Dose-dense temozolomide for newly diagnosed glioblastoma: A randomized phase III clinical trial. *J. Clin. Oncol.* **2013**, *31*, 4085–4091. [[CrossRef](#)] [[PubMed](#)]
- Stupp, R.; Hegi, M.E.; Mason, W.P.; van den Bent, M.J.; Taphoorn, M.J.; Janzer, R.C.; Ludwin, S.K.; Allgeier, A.; Fisher, B.; Belanger, K.; et al. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-Year analysis of the EORTC-NCIC trial. *Lancet Oncol.* **2009**, *10*, 459–466. [[CrossRef](#)]
- Wang, M.; Dignam, J.J.; Won, M.; Curran, W.; Mehta, M.; Gilbert, M.R. Variation over time and interdependence between disease progression and death among patients with glioblastoma on RTOG 0525. *Neuro-Oncology* **2015**, *17*, 999–1006. [[CrossRef](#)] [[PubMed](#)]
- Stupp, R.; Mason, W.P.; van den Bent, M.J.; Weller, M.; Fisher, B.; Taphoorn, M.J.B.; Belanger, K.; Brandes, A.A.; Marosi, C.; Bogdahn, U.; et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N. Engl. J. Med.* **2005**, *352*, 987–996. [[CrossRef](#)] [[PubMed](#)]
- Hegi, M.E.; Diserens, A.-C.; Gorlia, T.; Hamou, M.-F.; de Tribolet, N.; Weller, M.; Kros, J.M.; Hainfellner, J.A.; Mason, W.; Mariani, L.; et al. *MGMT* gene silencing and benefit from temozolomide in glioblastoma. *N. Engl. J. Med.* **2005**, *352*, 997–1003. [[CrossRef](#)] [[PubMed](#)]
- Felsberg, J.; Thon, N.; Eigenbrod, S.; Hentschel, B.; Sabel, M.C.; Westphal, M.; Schackert, G.; Kreth, F.W.; Pietsch, T.; Löffler, M.; et al. Promoter methylation and expression of *MGMT* and the DNA mismatch repair genes *MLH1*, *MSH2*, *MSH6* and *PMS2* in paired primary and recurrent glioblastomas. *Int. J. Cancer* **2011**, *129*, 659–670. [[CrossRef](#)] [[PubMed](#)]
- McLendon, R.; Friedman, A.; Bigner, D.; Van Meir, E.G.; Brat, D.J.; Olson, J.J.; Mikkelsen, T.; Lehman, N.; Aldape, K.; Alfred Yung, W.K.; et al. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* **2008**, *455*, 1061–1068. [[CrossRef](#)] [[PubMed](#)]
- Verhaak, R.G.W.; Hoadley, K.A.; Purdom, E.; Wang, V.; Qi, Y.; Wilkerson, M.D.; Miller, C.R.; Ding, L.; Golub, T.; Mesirov, J.P.; et al. Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in *PDGFRA*, *IDH1*, *EGFR*, and *NF1*. *Cancer Cell* **2010**, *17*, 98–110. [[CrossRef](#)] [[PubMed](#)]
- Phillips, H.S.; Kharbanda, S.; Chen, R.; Forrester, W.F.; Soriano, R.H.; Wu, T.D.; Misra, A.; Nigro, J.M.; Colman, H.; Soroceanu, L.; et al. Molecular subclasses of high-grade glioma predict prognosis, delineate a pattern of disease progression, and resemble stages in neurogenesis. *Cancer Cell* **2006**, *9*, 157–173. [[CrossRef](#)] [[PubMed](#)]

10. Brennan, C.W.; Verhaak, R.G.W.; McKenna, A.; Campos, B.; Nounshmehr, H.; Salama, S.R.; Zheng, S.; Chakravarty, D.; Sanborn, J.Z.; Berman, S.H.; et al. The somatic genomic landscape of glioblastoma. *Cell* **2013**, *155*, 462–477. [[CrossRef](#)] [[PubMed](#)]
11. Nounshmehr, H.; Weisenberger, D.J.; Diefes, K.; Phillips, H.S.; Pujara, K.; Berman, B.P.; Pan, F.; Pelloski, C.E.; Sulman, E.P.; Bhat, K.P.; et al. Identification of a CpG island methylator phenotype that defines a distinct subgroup of glioma. *Cancer Cell* **2010**, *17*, 510–522. [[CrossRef](#)] [[PubMed](#)]
12. Philpott, C.; Tovell, H.; Frayling, I.M.; Cooper, D.N.; Upadhyaya, M. The NF1 somatic mutational landscape in sporadic human cancers. *Hum. Genom.* **2017**, *11*, 13. [[CrossRef](#)] [[PubMed](#)]
13. Louis, D.N.; Perry, A.; Reifenberger, G.; von Deimling, A.; Figarella-Branger, D.; Cavenee, W.K.; Ohgaki, H.; Wiestler, O.D.; Kleihues, P.; Ellison, D.W. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: A summary. *Acta Neuropathol.* **2016**, *131*, 803–820. [[CrossRef](#)] [[PubMed](#)]
14. Nørøxe, D.S.; Poulsen, H.S.; Lassen, U. Hallmarks of glioblastoma: A systematic review. *ESMO Open* **2016**, *1*, e000144. [[CrossRef](#)] [[PubMed](#)]
15. Mahase, S.; Rattenni, R.N.; Wesseling, P.; Leenders, W.; Baldotto, C.; Jain, R.; Zagzag, D. Hypoxia-Mediated Mechanisms Associated with Antiangiogenic Treatment Resistance in Glioblastomas. *Am. J. Pathol.* **2017**, *187*, 940–953. [[CrossRef](#)] [[PubMed](#)]
16. Brat, D.J.; Van Meir, E.G. Vaso-occlusive and prothrombotic mechanisms associated with tumor hypoxia, necrosis, and accelerated growth in glioblastoma. *Lab. Invest.* **2004**, *84*, 397–405. [[CrossRef](#)] [[PubMed](#)]
17. Brat, D.J.; Castellano-Sanchez, A.A.; Hunter, S.B.; Pecot, M.; Cohen, C.; Hammond, E.H.; Devi, S.N.; Kaur, B.; Van Meir, E.G. Pseudopalisades in glioblastoma are hypoxic, express extracellular matrix proteases, and are formed by an actively migrating cell population. *Cancer Res.* **2004**, *64*, 920–927. [[CrossRef](#)] [[PubMed](#)]
18. Zagzag, D.; Zhong, H.; Scalzitti, J.M.; Laughner, E.; Simons, J.W.; Semenza, G.L. Expression of hypoxia-inducible factor 1alpha in brain tumors: Association with angiogenesis, invasion, and progression. *Cancer* **2000**, *88*, 2606–2618. [[CrossRef](#)]
19. Brat, D.J.; Castellano-Sanchez, A.; Kaur, B.; Van Meir, E.G. Genetic and biologic progression in astrocytomas and their relation to angiogenic dysregulation. *Adv. Anat. Pathol.* **2002**, *9*, 24–36. [[CrossRef](#)] [[PubMed](#)]
20. Brat, D.J.; Van Meir, E.G. Glomeruloid microvascular proliferation orchestrated by VPF/VEGF: A new world of angiogenesis research. *Am. J. Pathol.* **2001**, *158*, 789–796. [[CrossRef](#)]
21. Carmeliet, P.; Jain, R.K. Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nat. Rev. Drug Discov.* **2011**, *10*, 417–427. [[CrossRef](#)] [[PubMed](#)]
22. Yang, L.; Lin, C.; Wang, L.; Guo, H.; Wang, X. Hypoxia and hypoxia-inducible factors in glioblastoma multiforme progression and therapeutic implications. *Exp. Cell Res.* **2012**, *318*, 2417–2426. [[CrossRef](#)] [[PubMed](#)]
23. Kaur, B.; Khwaja, F.W.; Severson, E.A.; Matheny, S.L.; Brat, D.J.; Van Meir, E.G. Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. *Neuro-Oncology* **2005**, *7*, 134–153. [[CrossRef](#)] [[PubMed](#)]
24. Jensen, R.L.; Mumert, M.L.; Gillespie, D.L.; Kinney, A.Y.; Schabel, M.C.; Salzman, K.L. Preoperative dynamic contrast-enhanced MRI correlates with molecular markers of hypoxia and vascularity in specific areas of intratumoral microenvironment and is predictive of patient outcome. *Neuro-Oncology* **2014**, *16*, 280–291. [[CrossRef](#)] [[PubMed](#)]
25. Rong, Y.; Durden, D.L.; Van Meir, E.G.; Brat, D.J. “Pseudopalisading” necrosis in glioblastoma: A familiar morphologic feature that links vascular pathology, hypoxia, and angiogenesis. *J. Neuropathol. Exp. Neurol.* **2006**, *65*, 529–539. [[CrossRef](#)] [[PubMed](#)]
26. Clara, C.A.; Marie, S.K.N.; de Almeida, J.R.W.; Wakamatsu, A.; Oba-Shinjo, S.M.; Uno, M.; Neville, M.; Rosenberg, S. Angiogenesis and expression of PDGF-C, VEGF, CD105 and HIF-1 α in human glioblastoma. *Neuropathology* **2014**, *34*, 343–352. [[CrossRef](#)] [[PubMed](#)]
27. Bar, E.E.; Lin, A.; Mahairaki, V.; Matsui, W.; Eberhart, C.G. Hypoxia increases the expression of stem-cell markers and promotes clonogenicity in glioblastoma neurospheres. *Am. J. Pathol.* **2010**, *177*, 1491–1502. [[CrossRef](#)] [[PubMed](#)]
28. Winkler, F.; Kozin, S.V.; Tong, R.T.; Chae, S.-S.; Booth, M.F.; Garkavtsev, I.; Xu, L.; Hicklin, D.J.; Fukumura, D.; di Tomaso, E.; et al. Kinetics of vascular normalization by VEGFR2 blockade governs brain tumor response to radiation: Role of oxygenation, angiopoietin-1, and matrix metalloproteinases. *Cancer Cell* **2004**, *6*, 553–563. [[CrossRef](#)] [[PubMed](#)]

29. Yuan, F.; Salehi, H.A.; Boucher, Y.; Vasthare, U.S.; Tuma, R.F.; Jain, R.K. Vascular permeability and microcirculation of gliomas and mammary carcinomas transplanted in rat and mouse cranial windows. *Cancer Res.* **1994**, *54*, 4564–4568. [[PubMed](#)]
30. Plate, K.H.; Mennel, H.D. Vascular morphology and angiogenesis in glial tumors. *Exp. Toxicol. Pathol.* **1995**, *47*, 89–94. [[CrossRef](#)]
31. Bullitt, E.; Zeng, D.; Gerig, G.; Aylward, S.; Joshi, S.; Smith, J.K.; Lin, W.; Ewend, M.G. Vessel tortuosity and brain tumor malignancy: A blinded study. *Acad. Radiol.* **2005**, *12*, 1232–1240. [[CrossRef](#)] [[PubMed](#)]
32. Jain, R.K.; di Tomaso, E.; Duda, D.G.; Loeffler, J.S.; Sorensen, A.G.; Batchelor, T.T. Angiogenesis in brain tumours. *Nat. Rev. Neurosci.* **2007**, *8*, 610–622. [[CrossRef](#)] [[PubMed](#)]
33. Dubois, L.G.; Campanati, L.; Righy, C.; D’Andrea-Meira, I.; Spohr, T.C.L.D.S.; Porto-Carreiro, I.; Pereira, C.M.; Balça-Silva, J.; Kahn, S.A.; DosSantos, M.F.; et al. Gliomas and the vascular fragility of the blood brain barrier. *Front. Cell. Neurosci.* **2014**, *8*, 418. [[CrossRef](#)] [[PubMed](#)]
34. Majmundar, A.J.; Wong, W.J.; Simon, M.C. Hypoxia-Inducible Factors and the Response to Hypoxic Stress. *Mol. Cell* **2010**, *40*, 294–309. [[CrossRef](#)] [[PubMed](#)]
35. Keith, B.; Johnson, R.S.; Simon, M.C. HIF1 α and HIF2 α : Sibling rivalry in hypoxic tumour growth and progression. *Nat. Rev. Cancer* **2011**, *12*, 9–22. [[CrossRef](#)] [[PubMed](#)]
36. Semenza, G.L. Hypoxia-inducible factors: Mediators of cancer progression and targets for cancer therapy. *Trends Pharmacol. Sci.* **2012**, *33*, 207–214. [[CrossRef](#)] [[PubMed](#)]
37. Zhang, P.; Yao, Q.; Lu, L.; Li, Y.; Chen, P.-J.; Duan, C. Hypoxia-inducible factor 3 is an oxygen-dependent transcription activator and regulates a distinct transcriptional response to hypoxia. *Cell Rep.* **2014**, *6*, 1110–1121. [[CrossRef](#)] [[PubMed](#)]
38. Duan, C. Hypoxia-inducible factor 3 biology: Complexities and emerging themes. *Am. J. Physiol. Cell Physiol.* **2016**, *310*, C260–C269. [[CrossRef](#)] [[PubMed](#)]
39. Huang, L.E.; Bunn, H.F. Hypoxia-inducible factor and its biomedical relevance. *J. Biol. Chem.* **2003**, *278*, 19575–19578. [[CrossRef](#)] [[PubMed](#)]
40. Schofield, C.J.; Ratcliffe, P.J. Oxygen sensing by HIF hydroxylases. *Nat. Rev. Mol. Cell Biol.* **2004**, *5*, 343–354. [[CrossRef](#)] [[PubMed](#)]
41. Semenza, G.L. Regulation of mammalian O₂ homeostasis by hypoxia-inducible factor 1. *Annu. Rev. Cell Dev. Biol.* **1999**, *15*, 551–578. [[CrossRef](#)] [[PubMed](#)]
42. Cockman, M.E.; Masson, N.; Mole, D.R.; Jaakkola, P.; Chang, G.-W.; Clifford, S.C.; Maher, E.R.; Pugh, C.W.; Ratcliffe, P.J.; Maxwell, P.H. Hypoxia Inducible Factor- α Binding and Ubiquitylation by the von Hippel-Lindau Tumor Suppressor Protein. *J. Biol. Chem.* **2000**, *275*, 25733–25741. [[CrossRef](#)] [[PubMed](#)]
43. Maxwell, P.H.; Wiesener, M.S.; Chang, G.W.; Clifford, S.C.; Vaux, E.C.; Cockman, M.E.; Wykoff, C.C.; Pugh, C.W.; Maher, E.R.; Ratcliffe, P.J. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature* **1999**, *399*, 271–275. [[CrossRef](#)] [[PubMed](#)]
44. Srinivas, V.; Zhang, L.P.; Zhu, X.H.; Caro, J. Characterization of an oxygen/redox-dependent degradation domain of hypoxia-inducible factor alpha (HIF-alpha) proteins. *Biochem. Biophys. Res. Commun.* **1999**, *260*, 557–561. [[CrossRef](#)] [[PubMed](#)]
45. Ebert, B.L.; Bunn, H.F. Regulation of transcription by hypoxia requires a multiprotein complex that includes hypoxia-inducible factor 1, an adjacent transcription factor, and p300/CREB binding protein. *Mol. Cell. Biol.* **1998**, *18*, 4089–4096. [[CrossRef](#)] [[PubMed](#)]
46. Huang, L.E.; Arany, Z.; Livingston, D.M.; Bunn, H.F. Activation of hypoxia-inducible transcription factor depends primarily upon redox-sensitive stabilization of its alpha subunit. *J. Biol. Chem.* **1996**, *271*, 32253–32259. [[CrossRef](#)] [[PubMed](#)]
47. Kallio, P.J.; Pongratz, I.; Gradin, K.; McGuire, J.; Poellinger, L. Activation of hypoxia-inducible factor 1alpha: Posttranscriptional regulation and conformational change by recruitment of the Arnt transcription factor. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 5667–5672. [[CrossRef](#)] [[PubMed](#)]
48. Mahon, P.C.; Hirota, K.; Semenza, G.L. FIH-1: A novel protein that interacts with HIF-1alpha and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev.* **2001**, *15*, 2675–2686. [[CrossRef](#)] [[PubMed](#)]
49. Lando, D.; Peet, D.J.; Gorman, J.J.; Whelan, D.A.; Whitelaw, M.L.; Bruick, R.K. FIH-1 is an asparaginyl hydroxylase enzyme that regulates the transcriptional activity of hypoxia-inducible factor. *Genes Dev.* **2002**, *16*, 1466–1471. [[CrossRef](#)] [[PubMed](#)]

50. Frederick, L.; Wang, X.Y.; Eley, G.; James, C.D. Diversity and frequency of epidermal growth factor receptor mutations in human glioblastomas. *Cancer Res.* **2000**, *60*, 1383–1387. [[PubMed](#)]
51. Holland, E.C.; Hively, W.P.; DePinho, R.A.; Varmus, H.E. A constitutively active epidermal growth factor receptor cooperates with disruption of G1 cell-cycle arrest pathways to induce glioma-like lesions in mice. *Genes Dev.* **1998**, *12*, 3675–3685. [[CrossRef](#)] [[PubMed](#)]
52. Clarke, K.; Smith, K.; Gullick, W.J.; Harris, A.L. Mutant epidermal growth factor receptor enhances induction of vascular endothelial growth factor by hypoxia and insulin-like growth factor-1 via a PI3 kinase dependent pathway. *Br. J. Cancer* **2001**, *84*, 1322–1329. [[CrossRef](#)] [[PubMed](#)]
53. Zhong, H.; Chiles, K.; Feldser, D.; Laughner, E.; Hanrahan, C.; Georgescu, M.M.; Simons, J.W.; Semenza, G.L. Modulation of hypoxia-inducible factor 1alpha expression by the epidermal growth factor/phosphatidylinositol 3-kinase/PTEN/AKT/FRAP pathway in human prostate cancer cells: Implications for tumor angiogenesis and therapeutics. *Cancer Res.* **2000**, *60*, 1541–1545. [[PubMed](#)]
54. Sansal, I.; Sellers, W.R. The biology and clinical relevance of the PTEN tumor suppressor pathway. *J. Clin. Oncol.* **2004**, *22*, 2954–2963. [[CrossRef](#)] [[PubMed](#)]
55. Ravi, R.; Mookerjee, B.; Bhujwala, Z.M.; Sutter, C.H.; Artemov, D.; Zeng, Q.; Dillehay, L.E.; Madan, A.; Semenza, G.L.; Bedi, A. Regulation of tumor angiogenesis by p53-induced degradation of hypoxia-inducible factor 1alpha. *Genes Dev.* **2000**, *14*, 34–44. [[PubMed](#)]
56. Madan, E.; Dikshit, B.; Gowda, S.H.; Srivastava, C.; Sarkar, C.; Chattopadhyay, P.; Sinha, S.; Chosdol, K. FAT1 is a novel upstream regulator of HIF1 α and invasion of high grade glioma. *Int. J. Cancer* **2016**, *139*, 2570–2582. [[CrossRef](#)] [[PubMed](#)]
57. Zhou, X.; Liu, Z.; Shi, Q.; Jiao, J.; Bian, W.; Song, X.; Mo, J.; Sang, B.; Xu, Y.; Qian, J.; et al. Geranylgeranyltransferase I regulates HIF-1 α promoting glioblastoma cell migration and invasion. *J. Neurooncol.* **2013**, *112*, 365–374. [[CrossRef](#)] [[PubMed](#)]
58. Skuli, N.; Monferran, S.; Delmas, C.; Favre, G.; Bonnet, J.; Toulas, C.; Cohen-Jonathan Moyal, E. Alfvbeta3/alfvbeta5 integrins-FAK-RhoB: A novel pathway for hypoxia regulation in glioblastoma. *Cancer Res.* **2009**, *69*, 3308–3316. [[CrossRef](#)] [[PubMed](#)]
59. Mao, X.-G.; Wang, C.; Liu, D.-Y.; Zhang, X.; Wang, L.; Yan, M.; Zhang, W.; Zhu, J.; Li, Z.-C.; Mi, C.; et al. Hypoxia upregulates HIG2 expression and contributes to bevacizumab resistance in glioblastoma. *Oncotarget* **2016**, *7*, 47808–47820. [[CrossRef](#)] [[PubMed](#)]
60. Cloughesy, T.F.; Yoshimoto, K.; Nghiemphu, P.; Brown, K.; Dang, J.; Zhu, S.; Hsueh, T.; Chen, Y.; Wang, W.; Youngkin, D.; et al. Antitumor activity of rapamycin in a Phase I trial for patients with recurrent PTEN-deficient glioblastoma. *PLoS Med.* **2008**, *5*, e8. [[CrossRef](#)] [[PubMed](#)]
61. Galanis, E.; Buckner, J.C.; Maurer, M.J.; Kreisberg, J.I.; Ballman, K.; Boni, J.; Peralba, J.M.; Jenkins, R.B.; Dakhil, S.R.; Morton, R.F.; et al. Phase II trial of temsirolimus (CCI-779) in recurrent glioblastoma multiforme: A North Central Cancer Treatment Group Study. *J. Clin. Oncol.* **2005**, *23*, 5294–5304. [[CrossRef](#)] [[PubMed](#)]
62. Koul, D.; Fu, J.; Shen, R.; LaFortune, T.A.; Wang, S.; Tiao, N.; Kim, Y.-W.; Liu, J.-L.; Ramnarian, D.; Yuan, Y.; et al. Antitumor activity of NVP-BKM120—A selective pan class I PI3 kinase inhibitor showed differential forms of cell death based on p53 status of glioma cells. *Clin. Cancer Res.* **2012**, *18*, 184–195. [[CrossRef](#)] [[PubMed](#)]
63. Maira, S.-M.; Stauffer, F.; Brueggen, J.; Furet, P.; Schnell, C.; Fritsch, C.; Brachmann, S.; Chène, P.; De Pover, A.; Schoemaker, K.; et al. Identification and characterization of NVP-BEZ235, a new orally available dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor with potent in vivo antitumor activity. *Mol. Cancer Ther.* **2008**, *7*, 1851–1863. [[CrossRef](#)] [[PubMed](#)]
64. Hill, R.; Madureira, P.A.; Ferreira, B.; Baptista, I.; Machado, S.; Colaço, L.; dos Santos, M.; Liu, N.; Dopazo, A.; Ugurel, S.; et al. TRIB2 confers resistance to anti-cancer therapy by activating the serine/threonine protein kinase AKT. *Nat. Commun.* **2017**, *8*, 14687. [[CrossRef](#)] [[PubMed](#)]
65. Wen, P.Y.; Lee, E.Q.; Reardon, D.A.; Ligon, K.L.; Alfred Yung, W.K. Current clinical development of PI3K pathway inhibitors in glioblastoma. *Neuro-Oncology* **2012**, *14*, 819–829. [[CrossRef](#)] [[PubMed](#)]
66. Liu, T.-J.; Koul, D.; LaFortune, T.; Tiao, N.; Shen, R.J.; Maira, S.-M.; Garcia-Echeverria, C.; Yung, W.K.A. NVP-BEZ235, a novel dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor, elicits multifaceted antitumor activities in human gliomas. *Mol. Cancer Ther.* **2009**, *8*, 2204–2210. [[CrossRef](#)] [[PubMed](#)]

67. Lin, F.; de Gooijer, M.C.; Hanekamp, D.; Chandrasekaran, G.; Buil, L.C.M.; Thota, N.; Sparidans, R.W.; Beijnen, J.H.; Würdinger, T.; van Tellingen, O. PI3K-mTOR Pathway Inhibition Exhibits Efficacy Against High-grade Glioma in Clinically Relevant Mouse Models. *Clin. Cancer Res.* **2017**, *23*, 1286–1298. [[CrossRef](#)] [[PubMed](#)]
68. Netland, I.A.; Førde, H.E.; Sleire, L.; Leiss, L.; Rahman, M.A.; Skeie, B.S.; Gjerde, C.H.; Enger, P.Ø.; Goplen, D. Dactolisib (NVP-BEZ235) toxicity in murine brain tumour models. *BMC Cancer* **2016**, *16*, 657. [[CrossRef](#)] [[PubMed](#)]
69. Huss, W.J.; Hanrahan, C.F.; Barrios, R.J.; Simons, J.W.; Greenberg, N.M. Angiogenesis and prostate cancer: Identification of a molecular progression switch. *Cancer Res.* **2001**, *61*, 2736–2743. [[PubMed](#)]
70. Castaldo, S.A.; Freitas, J.R.; Conchinha, N.V.; Madureira, P.A. The Tumorigenic Roles of the Cellular REDOX Regulatory Systems. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 8413032. [[CrossRef](#)] [[PubMed](#)]
71. Klimova, T.; Chandel, N.S. Mitochondrial complex III regulates hypoxic activation of HIF. *Cell Death Differ.* **2008**, *15*, 660–666. [[CrossRef](#)] [[PubMed](#)]
72. Brösicke, N.; Faissner, A. Role of tenascins in the ECM of gliomas. *Cell Adhes. Migr.* **2015**, *9*, 131–140. [[CrossRef](#)] [[PubMed](#)]
73. Bache, M.; Rot, S.; Kessler, J.; Güttler, A.; Wichmann, H.; Greither, T.; Wach, S.; Taubert, H.; Söling, A.; Bilkenroth, U.; et al. mRNA expression levels of hypoxia-induced and stem cell-associated genes in human glioblastoma. *Oncol. Rep.* **2015**, *33*, 3155–3161. [[CrossRef](#)] [[PubMed](#)]
74. Said, H.M.; Polat, B.; Staab, A.; Hagemann, C.; Stein, S.; Flentje, M.; Theobald, M.; Katzer, A.; Vordermark, D. Rapid detection of the hypoxia-regulated CA-IX and NDRG1 gene expression in different glioblastoma cells in vitro. *Oncol. Rep.* **2008**, *20*, 413–419. [[CrossRef](#)] [[PubMed](#)]
75. Said, H.M.; Hagemann, C.; Staab, A.; Stojic, J.; Kühnel, S.; Vince, G.H.; Flentje, M.; Roosen, K.; Vordermark, D. Expression patterns of the hypoxia-related genes osteopontin, CA9, erythropoietin, VEGF and HIF-1 α in human glioma in vitro and in vivo. *Radiother. Oncol.* **2007**, *83*, 398–405. [[CrossRef](#)] [[PubMed](#)]
76. Marotta, D.; Karar, J.; Jenkins, W.T.; Kumanova, M.; Jenkins, K.W.; Tobias, J.W.; Baldwin, D.; Hatzigeorgiou, A.; Alexiou, P.; Evans, S.M.; et al. In vivo profiling of hypoxic gene expression in gliomas using the hypoxia marker EF5 and laser-capture microdissection. *Cancer Res.* **2011**, *71*, 779–789. [[CrossRef](#)] [[PubMed](#)]
77. Proescholdt, M.A.; Merrill, M.J.; Stoerr, E.-M.; Lohmeier, A.; Pohl, F.; Brawanski, A. Function of carbonic anhydrase IX in glioblastoma multiforme. *Neuro-Oncology* **2012**, *14*, 1357–1366. [[CrossRef](#)] [[PubMed](#)]
78. Kobliakov, V.A. Role of Proton Pumps in Tumorigenesis. *Biochemistry* **2017**, *82*, 401–412. [[CrossRef](#)] [[PubMed](#)]
79. Kivelä, A.J.; Parkkila, S.; Saarnio, J.; Karttunen, T.J.; Kivelä, J.; Parkkila, A.K.; Pastoreková, S.; Pastorek, J.; Waheed, A.; Sly, W.S.; et al. Expression of transmembrane carbonic anhydrase isoenzymes IX and XII in normal human pancreas and pancreatic tumours. *Histochem. Cell Biol.* **2000**, *114*, 197–204. [[PubMed](#)]
80. Hilvo, M.; Baranauskienė, L.; Salzano, A.M.; Scaloni, A.; Matulis, D.; Innocenti, A.; Scozzafava, A.; Monti, S.M.; Di Fiore, A.; De Simone, G.; et al. Biochemical Characterization of CA IX, One of the Most Active Carbonic Anhydrase Isozymes. *J. Biol. Chem.* **2008**, *283*, 27799–27809. [[CrossRef](#)] [[PubMed](#)]
81. Amiri, A.; Le, P.U.; Moquin, A.; Machkalyan, G.; Petrecca, K.; Gillard, J.W.; Yoganathan, N.; Maysinger, D. Inhibition of carbonic anhydrase IX in glioblastoma multiforme. *Eur. J. Pharm. Biopharm.* **2016**, *109*, 81–92. [[CrossRef](#)] [[PubMed](#)]
82. Porporato, P.E.; Dhup, S.; Dadhich, R.K.; Copetti, T.; Sonveaux, P. Anticancer targets in the glycolytic metabolism of tumors: A comprehensive review. *Front. Pharmacol.* **2011**, *2*, 49. [[CrossRef](#)] [[PubMed](#)]
83. Mao, H.; Lebrun, D.G.; Yang, J.; Zhu, V.F.; Li, M. Deregulated signaling pathways in glioblastoma multiforme: Molecular mechanisms and therapeutic targets. *Cancer Invest.* **2012**, *30*, 48–56. [[CrossRef](#)] [[PubMed](#)]
84. Burden-Gulley, S.M.; Qutaish, M.Q.; Sullivant, K.E.; Lu, H.; Wang, J.; Craig, S.E.L.; Basilion, J.P.; Wilson, D.L.; Brady-Kalnay, S.M. Novel cryo-imaging of the glioma tumor microenvironment reveals migration and dispersal pathways in vivid three-dimensional detail. *Cancer Res.* **2011**, *71*, 5932–5940. [[CrossRef](#)] [[PubMed](#)]
85. Liu, Z.; Han, L.; Dong, Y.; Tan, Y.; Li, Y.; Zhao, M.; Xie, H.; Ju, H.; Wang, H.; Zhao, Y.; et al. EGFRvIII/integrin β 3 interaction in hypoxic and vitronectin-enriching microenvironment promote GBM progression and metastasis. *Oncotarget* **2016**, *7*, 4680–4694. [[CrossRef](#)] [[PubMed](#)]
86. Sugawa, N.; Ekstrand, A.J.; James, C.D.; Collins, V.P. Identical splicing of aberrant epidermal growth factor receptor transcripts from amplified rearranged genes in human glioblastomas. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 8602–8606. [[CrossRef](#)] [[PubMed](#)]

87. Ekstrand, A.J.; Sugawa, N.; James, C.D.; Collins, V.P. Amplified and rearranged epidermal growth factor receptor genes in human glioblastomas reveal deletions of sequences encoding portions of the N- and/or C-terminal tails. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 4309–4313. [[CrossRef](#)] [[PubMed](#)]
88. Malden, L.T.; Novak, U.; Kaye, A.H.; Burgess, A.W. Selective amplification of the cytoplasmic domain of the epidermal growth factor receptor gene in glioblastoma multiforme. *Cancer Res.* **1988**, *48*, 2711–2714. [[PubMed](#)]
89. Aldape, K.D.; Ballman, K.; Furth, A.; Buckner, J.C.; Giannini, C.; Burger, P.C.; Scheithauer, B.W.; Jenkins, R.B.; James, C.D. Immunohistochemical detection of EGFRvIII in high malignancy grade astrocytomas and evaluation of prognostic significance. *J. Neuropathol. Exp. Neurol.* **2004**, *63*, 700–707. [[CrossRef](#)] [[PubMed](#)]
90. Gan, H.K.; van den Bent, M.; Lassman, A.B.; Reardon, D.A.; Scott, A.M. Antibody-drug conjugates in glioblastoma therapy: The right drugs to the right cells. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 695–707. [[CrossRef](#)] [[PubMed](#)]
91. Yang, J.; Yan, J.; Liu, B. Targeting EGFRvIII for glioblastoma multiforme. *Cancer Lett.* **2017**, *403*, 224–230. [[CrossRef](#)] [[PubMed](#)]
92. Flügel, D.; Görlach, A.; Michiels, C.; Kietzmann, T. Glycogen synthase kinase 3 phosphorylates hypoxia-inducible factor 1alpha and mediates its destabilization in a VHL-independent manner. *Mol. Cell. Biol.* **2007**, *27*, 3253–3265. [[CrossRef](#)] [[PubMed](#)]
93. Nabors, L.B.; Mikkelsen, T.; Rosenfeld, S.S.; Hochberg, F.; Akella, N.S.; Fisher, J.D.; Cloud, G.A.; Zhang, Y.; Carson, K.; Wittemer, S.M.; et al. Phase I and correlative biology study of cilengitide in patients with recurrent malignant glioma. *J. Clin. Oncol.* **2007**, *25*, 1651–1657. [[CrossRef](#)] [[PubMed](#)]
94. MacDonald, T.J.; Stewart, C.F.; Kocak, M.; Goldman, S.; Ellenbogen, R.G.; Phillips, P.; Lafond, D.; Poussaint, T.Y.; Kieran, M.W.; Boyett, J.M.; et al. Phase I clinical trial of cilengitide in children with refractory brain tumors: Pediatric Brain Tumor Consortium Study PBTC-012. *J. Clin. Oncol.* **2008**, *26*, 919–924. [[CrossRef](#)] [[PubMed](#)]
95. MacDonald, T.J.; Vezina, G.; Stewart, C.F.; Turner, D.; Pierson, C.R.; Chen, L.; Pollack, I.F.; Gajjar, A.; Kieran, M.W. Phase II study of cilengitide in the treatment of refractory or relapsed high-grade gliomas in children: A report from the Children’s Oncology Group. *Neuro-Oncology* **2013**, *15*, 1438–1444. [[CrossRef](#)] [[PubMed](#)]
96. Reardon, D.A.; Fink, K.L.; Mikkelsen, T.; Cloughesy, T.F.; O’Neill, A.; Plotkin, S.; Glantz, M.; Ravin, P.; Raizer, J.J.; Rich, K.M.; et al. Randomized phase II study of cilengitide, an integrin-targeting arginine-glycine-aspartic acid peptide, in recurrent glioblastoma multiforme. *J. Clin. Oncol.* **2008**, *26*, 5610–5617. [[CrossRef](#)] [[PubMed](#)]
97. Stupp, R.; Hegi, M.E.; Gorlia, T.; Erridge, S.C.; Perry, J.; Hong, Y.-K.; Aldape, K.D.; Lhermitte, B.; Pietsch, T.; Grujicic, D.; et al. Cilengitide combined with standard treatment for patients with newly diagnosed glioblastoma with methylated MGMT promoter (CENTRIC EORTC 26071-22072 study): A multicentre, randomised, open-label, phase 3 trial. *Lancet Oncol.* **2014**, *15*, 1100–1108. [[CrossRef](#)]
98. Xu, Y.; Zhang, L.; Wei, Y.; Zhang, X.; Xu, R.; Han, M.; Huang, B.; Chen, A.; Li, W.; Zhang, Q.; et al. Procollagen-lysine 2-oxoglutarate 5-dioxygenase 2 promotes hypoxia-induced glioma migration and invasion. *Oncotarget* **2017**, *8*, 23401–23413. [[CrossRef](#)] [[PubMed](#)]
99. Stracke, M.L.; Liotta, L.A. Multi-step cascade of tumor cell metastasis. *In Vivo* **1992**, *6*, 309–316. [[PubMed](#)]
100. Schlaepfer, D.D.; Mitra, S.K. Multiple connections link FAK to cell motility and invasion. *Curr. Opin. Genet. Dev.* **2004**, *14*, 92–101. [[CrossRef](#)] [[PubMed](#)]
101. Mañes, S.; Mira, E.; Gómez-Moutón, C.; Lacalle, R.A.; Martínez, C. Cells on the move: A dialogue between polarization and motility. *IUBMB Life* **2000**, *49*, 89–96. [[CrossRef](#)] [[PubMed](#)]
102. van der Slot, A.J.; Zuurmond, A.-M.; Bardoel, A.F.J.; Wijmenga, C.; Pruijs, H.E.H.; Sillence, D.O.; Brinckmann, J.; Abraham, D.J.; Black, C.M.; Verzijl, N.; et al. Identification of PLOD2 as telopeptide lysyl hydroxylase, an important enzyme in fibrosis. *J. Biol. Chem.* **2003**, *278*, 40967–40972. [[CrossRef](#)] [[PubMed](#)]
103. Ulrich, T.A.; de Juan Pardo, E.M.; Kumar, S. The mechanical rigidity of the extracellular matrix regulates the structure, motility, and proliferation of glioma cells. *Cancer Res.* **2009**, *69*, 4167–4174. [[CrossRef](#)] [[PubMed](#)]
104. Barcus, C.E.; Keely, P.J.; Eliceiri, K.W.; Schuler, L.A. Stiff Collagen Matrices Increase Tumorigenic Prolactin Signaling in Breast Cancer Cells. *J. Biol. Chem.* **2013**, *288*, 12722–12732. [[CrossRef](#)] [[PubMed](#)]

105. Levental, K.R.; Yu, H.; Kass, L.; Lakins, J.N.; Egeblad, M.; Erler, J.T.; Fong, S.F.T.; Csiszar, K.; Giaccia, A.; Weninger, W.; et al. Matrix crosslinking forces tumor progression by enhancing integrin signaling. *Cell* **2009**, *139*, 891–906. [[CrossRef](#)] [[PubMed](#)]
106. Du, H.; Pang, M.; Hou, X.; Yuan, S.; Sun, L. PLOD2 in cancer research. *Biomed. Pharmacother.* **2017**, *90*, 670–676. [[CrossRef](#)] [[PubMed](#)]
107. Eisinger-Mathason, T.S.K.; Zhang, M.; Qiu, Q.; Skuli, N.; Nakazawa, M.S.; Karakasheva, T.; Mucaj, V.; Shay, J.E.S.; Stangenberg, L.; Sadri, N.; et al. Hypoxia-dependent modification of collagen networks promotes sarcoma metastasis. *Cancer Discov.* **2013**, *3*, 1190–1205. [[CrossRef](#)] [[PubMed](#)]
108. Zuurmond, A.-M.; van der Slot-Verhoeven, A.J.; van Dura, E.A.; De Groot, J.; Bank, R.A. Minoxidil exerts different inhibitory effects on gene expression of lysyl hydroxylase 1, 2, and 3: Implications for collagen cross-linking and treatment of fibrosis. *Matrix Biol.* **2005**, *24*, 261–270. [[CrossRef](#)] [[PubMed](#)]
109. Hamsa, T.P.; Kuttan, G. Berberine inhibits pulmonary metastasis through down-regulation of MMP in metastatic B16F-10 melanoma cells. *Phytother. Res.* **2012**, *26*, 568–578. [[CrossRef](#)] [[PubMed](#)]
110. Guruvayoorappan, C.; Kuttan, G. Amentoflavone inhibits experimental tumor metastasis through a regulatory mechanism involving MMP-2, MMP-9, prolyl hydroxylase, lysyl oxidase, VEGF, ERK-1, ERK-2, STAT-1, NM23 and cytokines in lung tissues of C57BL/6 mice. *Immunopharmacol. Immunotoxicol.* **2008**, *30*, 711–727. [[CrossRef](#)] [[PubMed](#)]
111. Guruvayoorappan, C.; Kuttan, G. Beta-carotene inhibits tumor-specific angiogenesis by altering the cytokine profile and inhibits the nuclear translocation of transcription factors in B16F-10 melanoma cells. *Integr. Cancer Ther.* **2007**, *6*, 258–270. [[CrossRef](#)] [[PubMed](#)]
112. Zheng, H.; Kang, Y. Multilayer control of the EMT master regulators. *Oncogene* **2014**, *33*, 1755–1763. [[CrossRef](#)] [[PubMed](#)]
113. Timmerman, L.A.; Grego-Bessa, J.; Raya, A.; Bertrán, E.; Pérez-Pomares, J.M.; Díez, J.; Aranda, S.; Palomo, S.; McCormick, F.; Izpisua-Belmonte, J.C.; et al. Notch promotes epithelial-mesenchymal transition during cardiac development and oncogenic transformation. *Genes Dev.* **2004**, *18*, 99–115. [[CrossRef](#)] [[PubMed](#)]
114. Maciaczyk, D.; Picard, D.; Zhao, L.; Koch, K.; Herrera-Rios, D.; Li, G.; Marquardt, V.; Pauck, D.; Hoerbelt, T.; Zhang, W.; et al. CBF1 is clinically prognostic and serves as a target to block cellular invasion and chemoresistance of EMT-like glioblastoma cells. *Br. J. Cancer* **2017**, *117*, 102–112. [[CrossRef](#)] [[PubMed](#)]
115. Xie, Q.; Wu, Q.; Kim, L.; Miller, T.E.; Liau, B.B.; Mack, S.C.; Yang, K.; Factor, D.C.; Fang, X.; Huang, Z.; et al. RBPJ maintains brain tumor-initiating cells through CDK9-mediated transcriptional elongation. *J. Clin. Investig.* **2016**, *126*, 2757–2772. [[CrossRef](#)] [[PubMed](#)]
116. Wellner, U.; Schubert, J.; Burk, U.C.; Schmalhofer, O.; Zhu, F.; Sonntag, A.; Waldvogel, B.; Vannier, C.; Darling, D.; zur Hausen, A.; et al. The EMT-activator ZEB1 promotes tumorigenicity by repressing stemness-inhibiting microRNAs. *Nat. Cell Biol.* **2009**, *11*, 1487–1495. [[CrossRef](#)] [[PubMed](#)]
117. Siebzehnruhl, F.A.; Silver, D.J.; Tugertimur, B.; Deleyrolle, L.P.; Siebzehnruhl, D.; Sarkisian, M.R.; Devers, K.G.; Yachnis, A.T.; Kupper, M.D.; Neal, D.; et al. The ZEB1 pathway links glioblastoma initiation, invasion and chemoresistance. *EMBO Mol. Med.* **2013**, *5*, 1196–1212. [[CrossRef](#)] [[PubMed](#)]
118. Joseph, J.V.; Conroy, S.; Pavlov, K.; Sontakke, P.; Tomar, T.; Eggens-Meijer, E.; Balasubramaniyan, V.; Wagemakers, M.; den Dunnen, W.F.A.; Kruijff, F.A.E. Hypoxia enhances migration and invasion in glioblastoma by promoting a mesenchymal shift mediated by the HIF1 α -ZEB1 axis. *Cancer Lett.* **2015**, *359*, 107–116. [[CrossRef](#)] [[PubMed](#)]
119. Kahlert, U.D.; Suwala, A.K.; Raabe, E.H.; Siebzehnruhl, F.A.; Suarez, M.J.; Orr, B.A.; Bar, E.E.; Maciaczyk, J.; Eberhart, C.G. ZEB1 Promotes Invasion in Human Fetal Neural Stem Cells and Hypoxic Glioma Neurospheres. *Brain Pathol.* **2015**, *25*, 724–732. [[CrossRef](#)] [[PubMed](#)]
120. Rhee, J.; Mahfooz, N.S.; Arregui, C.; Lilien, J.; Balsamo, J.; VanBerkum, M.F.A. Activation of the repulsive receptor Roundabout inhibits N-cadherin-mediated cell adhesion. *Nat. Cell Biol.* **2002**, *4*, 798–805. [[CrossRef](#)] [[PubMed](#)]
121. Yang, M.-H.; Wu, K.-J. TWIST activation by hypoxia inducible factor-1 (HIF-1): Implications in metastasis and development. *Cell Cycle* **2008**, *7*, 2090–2096. [[CrossRef](#)] [[PubMed](#)]
122. Ansieau, S.; Morel, A.-P.; Hinkal, G.; Bastid, J.; Puisieux, A. TWISTing an embryonic transcription factor into an oncoprotein. *Oncogene* **2010**, *29*, 3173–3184. [[CrossRef](#)] [[PubMed](#)]
123. Castanon, I.; Baylies, M.K. A Twist in fate: Evolutionary comparison of Twist structure and function. *Gene* **2002**, *287*, 11–22. [[CrossRef](#)]

124. Malek, R.; Gajula, R.P.; Williams, R.D.; Nghiem, B.; Simons, B.W.; Nugent, K.; Wang, H.; Taparra, K.; Lemtiri-Chlieh, G.; Yoon, A.R.; et al. TWIST1-WDR5-*Hottip* Regulates *Hoxa9* Chromatin to Facilitate Prostate Cancer Metastasis. *Cancer Res.* **2017**, *77*, 3181–3193. [[CrossRef](#)] [[PubMed](#)]
125. Wang, J.; Nikhil, K.; Viccaro, K.; Chang, L.; Jacobsen, M.; Sandusky, G.; Shah, K. Aurora A-Twist1 axis promotes highly aggressive phenotypes in pancreatic carcinoma. *J. Cell Sci.* **2017**, *130*, 1078–1093. [[CrossRef](#)] [[PubMed](#)]
126. Nordfors, K.; Haapasalo, J.; Mäkelä, K.; Granberg, K.J.; Nykter, M.; Korja, M.; Paavonen, T.; Haapasalo, H.; Soini, Y. Twist predicts poor outcome of patients with astrocytic glioma. *J. Clin. Pathol.* **2015**, *68*, 905–912. [[CrossRef](#)] [[PubMed](#)]
127. Yang, M.-H.; Wu, M.-Z.; Chiou, S.-H.; Chen, P.-M.; Chang, S.-Y.; Liu, C.-J.; Teng, S.-C.; Wu, K.-J. Direct regulation of TWIST by HIF-1 α promotes metastasis. *Nat. Cell Biol.* **2008**, *10*, 295–305. [[CrossRef](#)] [[PubMed](#)]
128. Mikheeva, S.A.; Mikheev, A.M.; Petit, A.; Beyer, R.; Oxford, R.G.; Khorasani, L.; Maxwell, J.-P.; Glackin, C.A.; Wakimoto, H.; González-Herrero, I.; et al. TWIST1 promotes invasion through mesenchymal change in human glioblastoma. *Mol. Cancer* **2010**, *9*, 194. [[CrossRef](#)] [[PubMed](#)]
129. Elias, M.C.; Tozer, K.R.; Silber, J.R.; Mikheeva, S.; Deng, M.; Morrison, R.S.; Manning, T.C.; Silbergeld, D.L.; Glackin, C.A.; Reh, T.A.; et al. TWIST is expressed in human gliomas and promotes invasion. *Neoplasia* **2005**, *7*, 824–837. [[CrossRef](#)] [[PubMed](#)]
130. Zhou, W.; Ke, S.Q.; Huang, Z.; Flavahan, W.; Fang, X.; Paul, J.; Wu, L.; Sloan, A.E.; McLendon, R.E.; Li, X.; et al. Periostin secreted by glioblastoma stem cells recruits M2 tumour-associated macrophages and promotes malignant growth. *Nat. Cell Biol.* **2015**, *17*, 170–182. [[CrossRef](#)] [[PubMed](#)]
131. Serres, E.; Debarbieux, F.; Stanchi, F.; Maggiora, L.; Grall, D.; Turchi, L.; Burel-Vandenbos, F.; Figarella-Branger, D.; Virolle, T.; Rougon, G.; et al. Fibronectin expression in glioblastomas promotes cell cohesion, collective invasion of basement membrane in vitro and orthotopic tumor growth in mice. *Oncogene* **2014**, *33*, 3451–3462. [[CrossRef](#)] [[PubMed](#)]
132. Kunigal, S.; Gondi, C.S.; Gujrati, M.; Lakka, S.S.; Dinh, D.H.; Olivero, W.C.; Rao, J.S. SPARC-induced migration of glioblastoma cell lines via uPA-uPAR signaling and activation of small GTPase RhoA. *Int. J. Oncol.* **2006**, *29*, 1349–1357. [[CrossRef](#)] [[PubMed](#)]
133. Yang, H.W.; Menon, L.G.; Black, P.M.; Carroll, R.S.; Johnson, M.D. SNAI2/Slug promotes growth and invasion in human gliomas. *BMC Cancer* **2010**, *10*, 301. [[CrossRef](#)] [[PubMed](#)]
134. Casas, E.; Kim, J.; Bendesky, A.; Ohno-Machado, L.; Wolfe, C.J.; Yang, J. Snail2 is an essential mediator of Twist1-induced epithelial mesenchymal transition and metastasis. *Cancer Res.* **2011**, *71*, 245–254. [[CrossRef](#)] [[PubMed](#)]
135. Soroceanu, L.; Murase, R.; Limbad, C.; Singer, E.; Allison, J.; Adrados, I.; Kawamura, R.; Pakdel, A.; Fukuyo, Y.; Nguyen, D.; et al. Id-1 is a key transcriptional regulator of glioblastoma aggressiveness and a novel therapeutic target. *Cancer Res.* **2013**, *73*, 1559–1569. [[CrossRef](#)] [[PubMed](#)]
136. Cruickshanks, N.; Zhang, Y.; Yuan, F.; Pahuski, M.; Gibert, M.; Abounader, R. Role and Therapeutic Targeting of the HGF/MET Pathway in Glioblastoma. *Cancers* **2017**, *9*, 87. [[CrossRef](#)] [[PubMed](#)]
137. Da Silva, R.; Uno, M.; Marie, S.K.N.; Oba-Shinjo, S.M. LOX expression and functional analysis in astrocytomas and impact of IDH1 mutation. *PLoS ONE* **2015**, *10*, e0119781. [[CrossRef](#)] [[PubMed](#)]
138. Kaur, H.; Phillips-Mason, P.J.; Burden-Gulley, S.M.; Kerstetter-Fogle, A.E.; Basilion, J.P.; Sloan, A.E.; Brady-Kalnay, S.M. Cadherin-11, a marker of the mesenchymal phenotype, regulates glioblastoma cell migration and survival in vivo. *Mol. Cancer Res.* **2012**, *10*, 293–304. [[CrossRef](#)] [[PubMed](#)]
139. Abdouh, M.; Facchino, S.; Chatoo, W.; Balasingam, V.; Ferreira, J.; Bernier, G. BMI1 sustains human glioblastoma multiforme stem cell renewal. *J. Neurosci.* **2009**, *29*, 8884–8896. [[CrossRef](#)] [[PubMed](#)]
140. Zagzag, D.; Lukyanov, Y.; Lan, L.; Ali, M.A.; Esencay, M.; Mendez, O.; Yee, H.; Voura, E.B.; Newcomb, E.W. Hypoxia-inducible factor 1 and VEGF upregulate CXCR4 in glioblastoma: Implications for angiogenesis and glioma cell invasion. *Lab. Investig.* **2006**, *86*, 1221–1232. [[CrossRef](#)] [[PubMed](#)]
141. Rempel, S.A.; Dudas, S.; Ge, S.; Gutiérrez, J.A. Identification and localization of the cytokine SDF1 and its receptor, CXC chemokine receptor 4, to regions of necrosis and angiogenesis in human glioblastoma. *Clin. Cancer Res.* **2000**, *6*, 102–111. [[PubMed](#)]
142. Ehtesham, M.; Winston, J.A.; Kabos, P.; Thompson, R.C. CXCR4 expression mediates glioma cell invasiveness. *Oncogene* **2006**, *25*, 2801–2806. [[CrossRef](#)] [[PubMed](#)]

143. Hong, X.; Jiang, F.; Kalkanis, S.N.; Zhang, Z.G.; Zhang, X.-P.; DeCarvalho, A.C.; Katakowski, M.; Bobbitt, K.; Mikkelsen, T.; Chopp, M. SDF-1 and CXCR4 are up-regulated by VEGF and contribute to glioma cell invasion. *Cancer Lett.* **2006**, *236*, 39–45. [[CrossRef](#)] [[PubMed](#)]
144. Burger, J.A.; Kipps, T.J. CXCR4: A key receptor in the crosstalk between tumor cells and their microenvironment. *Blood* **2006**, *107*, 1761–1767. [[CrossRef](#)] [[PubMed](#)]
145. Gerlach, L.O.; Skerlj, R.T.; Bridger, G.J.; Schwartz, T.W. Molecular Interactions of Cyclam and Bicyclam Non-peptide Antagonists with the CXCR4 Chemokine Receptor. *J. Biol. Chem.* **2001**, *276*, 14153–14160. [[CrossRef](#)] [[PubMed](#)]
146. Kioi, M.; Vogel, H.; Schultz, G.; Hoffman, R.M.; Harsh, G.R.; Brown, J.M. Inhibition of vasculogenesis, but not angiogenesis, prevents the recurrence of glioblastoma after irradiation in mice. *J. Clin. Investig.* **2010**, *120*, 694–705. [[CrossRef](#)] [[PubMed](#)]
147. Mercurio, L.; Ajmone-Cat, M.A.; Cecchetti, S.; Ricci, A.; Bozzuto, G.; Molinari, A.; Manni, I.; Pollo, B.; Scala, S.; Carpinelli, G.; et al. Targeting CXCR4 by a selective peptide antagonist modulates tumor microenvironment and microglia reactivity in a human glioblastoma model. *J. Exp. Clin. Cancer Res.* **2016**, *35*, 55. [[CrossRef](#)] [[PubMed](#)]
148. Zhao, L.; Wang, Y.; Xue, Y.; Lv, W.; Zhang, Y.; He, S. Critical roles of chemokine receptor CCR5 in regulating glioblastoma proliferation and invasion. *Acta Biochim. Biophys. Sin.* **2015**, *47*, 890–898. [[CrossRef](#)] [[PubMed](#)]
149. Wang, Y.; Liu, T.; Yang, N.; Xu, S.; Li, X.; Wang, D. Hypoxia and macrophages promote glioblastoma invasion by the CCL4-CCR5 axis. *Oncol. Rep.* **2016**, *36*, 3522–3528. [[CrossRef](#)] [[PubMed](#)]
150. Schols, D.; Struyf, S.; Van Damme, J.; Esté, J.A.; Henson, G.; De Clercq, E. Inhibition of T-tropic HIV strains by selective antagonization of the chemokine receptor CXCR4. *J. Exp. Med.* **1997**, *186*, 1383–1388. [[CrossRef](#)] [[PubMed](#)]
151. Westby, M.; van der Ryst, E. CCR5 Antagonists: Host-Targeted Antivirals for the Treatment of HIV Infection. *Antivir. Chem. Chemother.* **2005**, *16*, 339–354. [[CrossRef](#)] [[PubMed](#)]
152. Lieberman-Blum, S.S.; Fung, H.B.; Bandres, J.C. Maraviroc: A CCR5-receptor antagonist for the treatment of HIV-1 infection. *Clin. Ther.* **2008**, *30*, 1228–1250. [[CrossRef](#)]
153. Velasco-Velázquez, M.; Xolalpa, W.; Pestell, R.G. The potential to target CCL5/CCR5 in breast cancer. *Expert Opin. Ther. Targets* **2014**, *18*, 1265–1275. [[CrossRef](#)] [[PubMed](#)]
154. Tanabe, Y.; Sasaki, S.; Mukaida, N.; Baba, T. Blockade of the chemokine receptor, CCR5, reduces the growth of orthotopically injected colon cancer cells via limiting cancer-associated fibroblast accumulation. *Oncotarget* **2016**, *7*, 48335–48345. [[CrossRef](#)] [[PubMed](#)]
155. Halvorsen, E.C.; Hamilton, M.J.; Young, A.; Wadsworth, B.J.; LePard, N.E.; Lee, H.N.; Firmino, N.; Collier, J.L.; Bennewith, K.L. Maraviroc decreases CCL8-mediated migration of CCR5(+) regulatory T cells and reduces metastatic tumor growth in the lungs. *Oncoimmunology* **2016**, *5*, e1150398. [[CrossRef](#)] [[PubMed](#)]
156. Halama, N.; Zoernig, I.; Berthel, A.; Kahlert, C.; Klupp, F.; Suarez-Carmona, M.; Suetterlin, T.; Brand, K.; Krauss, J.; Lasitschka, F.; et al. Tumoral Immune Cell Exploitation in Colorectal Cancer Metastases Can Be Targeted Effectively by Anti-CCR5 Therapy in Cancer Patients. *Cancer Cell* **2016**, *29*, 587–601. [[CrossRef](#)] [[PubMed](#)]
157. Kast, R.E.; Ramiro, S.; Lladó, S.; Toro, S.; Coveñas, R.; Muñoz, M. Antitumor action of temozolomide, ritonavir and aprepitant against human glioma cells. *J. Neurooncol.* **2016**, *126*, 425–431. [[CrossRef](#)] [[PubMed](#)]
158. Ragel, B.T.; Couldwell, W.T.; Gillespie, D.L.; Jensen, R.L. Identification of hypoxia-induced genes in a malignant glioma cell line (U-251) by cDNA microarray analysis. *Neurosurg. Rev.* **2007**, *30*, 181–187. [[CrossRef](#)] [[PubMed](#)]
159. Méndez, O.; Zavadil, J.; Esencay, M.; Lukyanov, Y.; Santovasi, D.; Wang, S.-C.; Newcomb, E.W.; Zagzag, D. Knock down of HIF-1 α in glioma cells reduces migration in vitro and invasion in vivo and impairs their ability to form tumor spheres. *Mol. Cancer* **2010**, *9*, 133. [[CrossRef](#)] [[PubMed](#)]
160. Fujimura, A.; Michiue, H.; Cheng, Y.; Uneda, A.; Tani, Y.; Nishiki, T.; Ichikawa, T.; Wei, F.-Y.; Tomizawa, K.; Matsui, H. Cyclin G2 promotes hypoxia-driven local invasion of glioblastoma by orchestrating cytoskeletal dynamics. *Neoplasia* **2013**, *15*, 1272–1281. [[CrossRef](#)] [[PubMed](#)]
161. Bennin, D.A.; Don, A.S.A.; Brake, T.; McKenzie, J.L.; Rosenbaum, H.; Ortiz, L.; DePaoli-Roach, A.A.; Horne, M.C. Cyclin G2 associates with protein phosphatase 2A catalytic and regulatory B' subunits in active complexes and induces nuclear aberrations and a G1/S phase cell cycle arrest. *J. Biol. Chem.* **2002**, *277*, 27449–27467. [[CrossRef](#)] [[PubMed](#)]

162. Arachchige Don, A.S.; Dallapiazza, R.F.; Bennin, D.A.; Brake, T.; Cowan, C.E.; Horne, M.C. Cyclin G2 is a centrosome-associated nucleocytoplasmic shuttling protein that influences microtubule stability and induces a p53-dependent cell cycle arrest. *Exp. Cell Res.* **2006**, *312*, 4181–4204. [[CrossRef](#)] [[PubMed](#)]
163. Rong, Y.; Post, D.E.; Pieper, R.O.; Durden, D.L.; Van Meir, E.G.; Brat, D.J. PTEN and hypoxia regulate tissue factor expression and plasma coagulation by glioblastoma. *Cancer Res.* **2005**, *65*, 1406–1413. [[CrossRef](#)] [[PubMed](#)]
164. Rong, Y.; Hu, F.; Huang, R.; Mackman, N.; Horowitz, J.M.; Jensen, R.L.; Durden, D.L.; Van Meir, E.G.; Brat, D.J. Early growth response gene-1 regulates hypoxia-induced expression of tissue factor in glioblastoma multiforme through hypoxia-inducible factor-1-independent mechanisms. *Cancer Res.* **2006**, *66*, 7067–7074. [[CrossRef](#)] [[PubMed](#)]
165. Belting, M.; Ahamed, J.; Ruf, W. Signaling of the tissue factor coagulation pathway in angiogenesis and cancer. *Arterioscler. Thromb. Vasc. Biol.* **2005**, *25*, 1545–1550. [[CrossRef](#)] [[PubMed](#)]
166. Hamada, K.; Kuratsu, J.; Saitoh, Y.; Takeshima, H.; Nishi, T.; Ushio, Y. Expression of tissue factor correlates with grade of malignancy in human glioma. *Cancer* **1996**, *77*, 1877–1883. [[CrossRef](#)]
167. Harter, P.N.; Dützmänn, S.; Drott, U.; Zachskorn, C.; Hattingen, E.; Capper, D.; Gessler, F.; Senft, C.; Seifert, V.; Plate, K.H.; et al. Anti-tissue factor (TF9–10H10) treatment reduces tumor cell invasiveness in a novel migratory glioma model. *Neuropathology* **2013**, *33*. [[CrossRef](#)] [[PubMed](#)]
168. Schaffner, F.; Ruf, W. Tissue factor and PAR2 signaling in the tumor microenvironment. *Arterioscler. Thromb. Vasc. Biol.* **2009**, *29*, 1999–2004. [[CrossRef](#)] [[PubMed](#)]
169. Schaffner, F.; Ruf, W. Tissue factor and protease-activated receptor signaling in cancer. *Semin. Thromb. Hemost.* **2008**, *34*, 147–153. [[CrossRef](#)] [[PubMed](#)]
170. Gessler, F.; Voss, V.; Dützmänn, S.; Seifert, V.; Gerlach, R.; Kögel, D. Inhibition of tissue factor/protease-activated receptor-2 signaling limits proliferation, migration and invasion of malignant glioma cells. *Neuroscience* **2010**, *165*, 1312–1322. [[CrossRef](#)] [[PubMed](#)]
171. Wykosky, J.; Gibo, D.M.; Stanton, C.; Debinski, W. Interleukin-13 receptor alpha 2, EphA2, and Fos-related antigen 1 as molecular denominators of high-grade astrocytomas and specific targets for combinatorial therapy. *Clin. Cancer Res.* **2008**, *14*, 199–208. [[CrossRef](#)] [[PubMed](#)]
172. Wykosky, J.; Gibo, D.M.; Stanton, C.; Debinski, W. EphA2 as a novel molecular marker and target in glioblastoma multiforme. *Mol. Cancer Res.* **2005**, *3*, 541–551. [[CrossRef](#)] [[PubMed](#)]
173. Wang, L.-F.; Fokas, E.; Bieker, M.; Rose, F.; Rexin, P.; Zhu, Y.; Pagenstecher, A.; Engenhardt-Cabillic, R.; An, H.-X. Increased expression of EphA2 correlates with adverse outcome in primary and recurrent glioblastoma multiforme patients. *Oncol. Rep.* **2008**, *19*, 151–156. [[CrossRef](#)] [[PubMed](#)]
174. Annamalai, B.; Liu, X.; Gopal, U.; Isaacs, J.S. Hsp90 is an essential regulator of EphA2 receptor stability and signaling: Implications for cancer cell migration and metastasis. *Mol. Cancer Res.* **2009**, *7*, 1021–1032. [[CrossRef](#)] [[PubMed](#)]
175. Miao, H.; Li, D.-Q.; Mukherjee, A.; Guo, H.; Petty, A.; Cutter, J.; Basilion, J.P.; Sedor, J.; Wu, J.; Danielpour, D.; et al. EphA2 mediates ligand-dependent inhibition and ligand-independent promotion of cell migration and invasion via a reciprocal regulatory loop with Akt. *Cancer Cell* **2009**, *16*, 9–20. [[CrossRef](#)] [[PubMed](#)]
176. Miao, H.; Burnett, E.; Kinch, M.; Simon, E.; Wang, B. Activation of EphA2 kinase suppresses integrin function and causes focal-adhesion-kinase dephosphorylation. *Nat. Cell Biol.* **2000**, *2*, 62–69. [[CrossRef](#)] [[PubMed](#)]
177. Macrae, M.; Neve, R.M.; Rodriguez-Viciana, P.; Haqq, C.; Yeh, J.; Chen, C.; Gray, J.W.; McCormick, F. A conditional feedback loop regulates Ras activity through EphA2. *Cancer Cell* **2005**, *8*, 111–118. [[CrossRef](#)] [[PubMed](#)]
178. Gopal, U.; Bohonowych, J.E.; Lema-Tome, C.; Liu, A.; Garrett-Mayer, E.; Wang, B.; Isaacs, J.S. A novel extracellular Hsp90 mediated co-receptor function for LRP1 regulates EphA2 dependent glioblastoma cell invasion. *PLoS ONE* **2011**, *6*, e17649. [[CrossRef](#)] [[PubMed](#)]
179. Powers, M.V.; Workman, P. Inhibitors of the heat shock response: Biology and pharmacology. *FEBS Lett.* **2007**, *581*, 3758–3769. [[CrossRef](#)] [[PubMed](#)]
180. Neckers, L.; Workman, P. Hsp90 molecular chaperone inhibitors: Are we there yet? *Clin. Cancer Res.* **2012**, *18*, 64–76. [[CrossRef](#)] [[PubMed](#)]
181. Wang, X.; Chen, M.; Zhou, J.; Zhang, X. HSP27, 70 and 90, anti-apoptotic proteins, in clinical cancer therapy (Review). *Int. J. Oncol.* **2014**, *45*, 18–30. [[CrossRef](#)] [[PubMed](#)]

182. Chigurupati, S.; Venkataraman, R.; Barrera, D.; Naganathan, A.; Madan, M.; Paul, L.; Pattisapu, J.V.; Kyriazis, G.A.; Sugaya, K.; Bushnev, S.; et al. Receptor channel TRPC6 is a key mediator of Notch-driven glioblastoma growth and invasiveness. *Cancer Res.* **2010**, *70*, 418–427. [[CrossRef](#)] [[PubMed](#)]
183. Alptekin, M.; Eroglu, S.; Tutar, E.; Sencan, S.; Geyik, M.A.; Ulasli, M.; Demiryurek, A.T.; Camci, C. Gene expressions of TRP channels in glioblastoma multiforme and relation with survival. *Tumor Biol.* **2015**, *36*, 9209–9213. [[CrossRef](#)] [[PubMed](#)]
184. Venkatachalam, K.; Montell, C. TRP channels. *Annu. Rev. Biochem.* **2007**, *76*, 387–417. [[CrossRef](#)] [[PubMed](#)]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).