



# Differing associations between A $\beta$ accumulation, hypoperfusion, blood–brain barrier dysfunction and loss of PDGFRB pericyte marker in the precuneus and parietal white matter in Alzheimer’s disease

J Scott Miners, Isabel Schulz and Seth Love

## Abstract

Recent studies implicate loss of pericytes in hypoperfusion and blood–brain barrier (BBB) leakage in Alzheimer’s disease (AD). In this study, we have measured levels of the pericyte marker, platelet-derived growth factor receptor- $\beta$  (PDGFRB), and fibrinogen (to assess blood–brain barrier leakage), and analyzed their relationship to indicators of microvessel density (von Willebrand factor level), ante-mortem oxygenation (myelin-associated glycoprotein:proteolipid protein-I ratio and vascular endothelial growth factor level), A $\beta$  level and plaque load, in precuneus and underlying white matter from 49 AD to 37 control brains. There was reduction in PDGFRB and increased fibrinogen in the precuneus in AD. These changes correlated with reduction in oxygenation and with plaque load. In the underlying white matter, increased fibrinogen correlated with reduced oxygenation, but PDGFRB level was unchanged. The level of platelet-derived growth factor- $\beta\beta$  (PDGF-BB), important for pericyte maintenance, was increased in AD but mainly in the insoluble tissue fraction, correlating with insoluble A $\beta$  level. Loss of the PDGFRB within the precuneus in AD is associated with fibrinogen leakage and reduced oxygenation, and related to fibrillar A $\beta$  accumulation. In contrast, fibrinogen leakage and reduced oxygenation of underlying white matter occur independently of loss of PDGFRB, perhaps secondary to reduced transcortical perfusion.

## Keywords

Alzheimer’s disease, blood–brain barrier, cerebral hypoperfusion, fibrinogen, pericyte

Received 18 October 2016; Revised 30 December 2016; Accepted 2 January 2017

## Introduction

Histological and ultrastructural examination of human post-mortem brain tissue has shown evidence of pericyte degeneration in AD,<sup>1–4</sup> and blood–brain barrier (BBB) breakdown, indicated by the extravasation of serum proteins such as fibrinogen, has also been widely reported.<sup>3–7</sup> Pericyte loss and BBB breakdown were found to be associated with higher A $\beta$  level and to be exacerbated in *APOE*  $\epsilon$ 4 carriers.<sup>3,4</sup> In the present study, we have used biochemical methods to investigate the relationships between the pericyte marker PDGFRB,<sup>4,8,9</sup> fibrinogen, cerebral hypoperfusion, *APOE* genotype, Braak tangle stage, and the pericyte trophic protein PDGF-BB, by analysis of the precuneus

and underlying white matter from post-mortem brain tissue in AD and control brains.

Cerebral blood flow declines within the medial parietal cortex (precuneus) early in the development of Alzheimer’s disease (AD). This is the first affected region of brain to show hypoperfusion in AD, up to 10 years before clinical symptoms.<sup>10–16</sup> Reduced

---

Dementia Research Group, Institute of Clinical Neurosciences, School of Clinical Sciences, University of Bristol, Bristol, UK

### Corresponding author:

Seth Love, School of Clinical Sciences, University of Bristol, Learning & Research level 2, Southmead Hospital, Bristol BS10 5NB, UK.  
Email: seth.love@bris.ac.uk

cerebral blood flow predicts the onset of dementia in AD<sup>17</sup> and occurs well before behavioral or pathological alterations in animal models.<sup>18–20</sup> It causes ischemic damage and may exacerbate AD pathology by increasing the production and reducing the clearance of A $\beta$  (reviewed in Chui et al.<sup>21</sup>). Conversely, there is evidence that the accumulation of A $\beta$  reduces cerebral blood flow not only through the development of cerebral amyloid angiopathy (CAA) but also by inducing both chronic vasoconstriction and interfering with autoregulation and neurovascular coupling (reviewed in Love and Miners<sup>22</sup> and Miners et al.<sup>23</sup>).

We previously reported that chronic reduction in oxygenation of brain tissue could be quantified by comparison of the levels of two myelin proteins: myelin-associated glycoprotein (MAG), which is highly susceptible to reduced tissue oxygenation, and proteolipid protein-1 (PLP-1), which is relatively resistant.<sup>24–27</sup> Both proteins are stable under post-mortem conditions and, as they have half-lives of several months, a decline in MAG:PLP1 reflects reduced oxygenation of oligodendrocytes over a sustained period prior to death (reviewed in Love and Miners<sup>22,28</sup>). Reduced oxygenation of the precuneus is evident at an early stage of AD (i.e. Braak tangle stage III-IV disease),<sup>26</sup> and correlates strongly with the level of endothelin-1 (EDN1), a potent vasoconstrictor, which is increased in AD,<sup>26,27,29</sup> probably as a consequence of A $\beta$ 42-mediated upregulation of endothelin-converting enzyme-2 (ECE-2),<sup>30</sup> but A $\beta$ 40-mediated upregulation of ECE-1 may also contribute.<sup>29</sup>

Several animal models have provided mechanistic insights into the relationship between neurovascular dysfunction and AD. Deletion of the *Meox2* gene in mice resulted in a perfusion deficit that was associated with increased A $\beta$  level (due to impeded LRP-1-mediated clearance).<sup>31</sup> Mice deficient in platelet-derived growth factor receptor- $\beta$  (*Pdgfr $\beta$ <sup>+/-</sup>*), a specific marker of pericytes,<sup>32</sup> showed age-related loss of pericytes, associated with BBB breakdown, impaired neurovascular coupling, reduced capillary density and reduced cerebral blood flow.<sup>9</sup> Accelerated progression of AD-related pathology and neuronal loss was observed when *Pdgfr $\beta$ <sup>+/-</sup>* mice were crossed with APP<sup>sw/0</sup> transgenic mice.<sup>33</sup> BBB breakdown and impaired interstitial drainage of A $\beta$  were thought to be responsible for the accelerated neurodegeneration, as neuronal damage was more marked in these mice than in those with a perfusion deficit alone.

Imaging studies have provided additional evidence linking pericyte loss, BBB breakdown and cerebral hypoperfusion in the early stages of AD. In 21 patients with no cognitive impairment, 21 with mild cognitive impairment and 19 patients with multiple sclerosis, imaging revealed that BBB leakage within the

hippocampus was a feature of normal aging but was exacerbated in the MCI group.<sup>34</sup> The severity of BBB leakage correlated with the CSF level of soluble PDGFRB, a marker of pericyte injury.<sup>35</sup> Van de Haar et al.<sup>36,37</sup> examined a small cohort of patients with early AD and age-matched controls and found that BBB leakage in early AD was associated with cognitive decline and reduced cerebral blood flow. The precise interrelationship between these neurovascular abnormalities in early AD remains unclear. In clinical and experimental studies of acute stroke, hypoperfusion is closely followed by disruption of the BBB.<sup>38,39</sup> However, as noted above,<sup>9</sup> pericyte loss can itself lead to both BBB leakage and hypoperfusion.

In the present study, we have analyzed the relationship between pericyte loss, BBB leakage and hypoperfusion in both the precuneus, as it is a region of early and consistent vascular dysfunction in AD, and the underlying white matter, which is perfused by perforating arterioles that pass through the precuneus. Present findings reveal a strong association between A $\beta$  plaque load, loss of pericyte protein PDGFRB, elevated fibrinogen and hypoperfusion in the cortex in AD. However, in the underlying white matter there is fibrinogen accumulation and hypoperfusion in the absence of loss of PDGFRB. Our findings suggest that fibrillar A $\beta$  accumulation plays a key role in pericyte degeneration in human brain tissue and highlight important differences between cerebral cortex and white matter in the pathophysiology of neurovascular dysfunction in early AD.

## Materials and methods

### Case selection

Brain tissue was obtained from the South West Dementia Brain Bank, University of Bristol, UK, with ethical approval from NRES committee South West-Central Bristol, UK (NRES approval 08/H0106/28+5). The brains had been dissected within 72 h of death. The right cerebral cortex had been fixed in buffered formalin for three weeks before the tissue was processed and paraffin blocks were taken for pathological assessment. The left cerebral hemisphere had been sliced and frozen at  $-80^{\circ}\text{C}$ . We studied 49 brains from patients with AD (ages, mean 77.5 y, SD 8.2 y) with post-mortem delays of 4 to 72 h (mean 31.4 h, SD 19.3 h) and 37 control brains (ages 58 to 94 y (mean 79.8 y, SD 8.9 y) with post-mortem delays from 3 to 67 h (mean 32.7 h, SD 16.3 h). All of the brains had been subjected to detailed neuropathological assessment. In those from patients with AD, the diagnosis had been made according to the NIA-AA guidelines.<sup>40</sup> Control brains were from people with no

history of dementia, few or absent neuritic plaques, a Braak tangle stage of III or less and no other neuropathological abnormalities. The demographic data, neuropathological findings, and MRC identifier numbers in this cohort are summarized in Supplementary Tables 1 and 2.

The cohort overlapped that of a previous study of several determinants of perfusion of the precuneus<sup>26</sup> and parietal white matter<sup>24</sup> in AD, vascular dementia, and control brains. Measurements of MAG:PLP1,<sup>24,26</sup> VEGF,<sup>27</sup> and soluble and insoluble A $\beta$ 40 and A $\beta$ 42<sup>41</sup> were previously reported. A $\beta$  plaque load had been measured by determining the area fraction of cerebral cortex immunopositive for A $\beta$ . Small vessel disease (SVD) had been scored on a 4-point semi-quantitative scale as previously described,<sup>25</sup> according to the extent of thickening of the arteriolar walls and associated narrowing of the vessel lumina: 0 = normal vessel wall thickness, 1 = slightly increased thickness, 2 = moderately increased thickness, and 3 = markedly increased thickness such that for many arterioles the diameter of the lumen was <50% of the outer diameter of the blood vessel. CAA had also been previously graded semi-quantitatively on a 4-point scale by a method adapted from that of Chalmers et al.<sup>42</sup> and Olichney et al.,<sup>43</sup> ranging from “0” for vessels devoid of amyloid to “3” for extensive vascular deposition.

### Preparation of brain tissue

Frozen tissue was dissected from the left medial parietal cortex (Brodmann area 7) and separate samples were dissected from the underlying parietal white matter. Biochemical analyses were performed on 200 mg samples of the dissected tissue that were homogenized in a Precellys homogenizer (Stretton Scientific, Derbyshire, UK) and extracted in 1% sodium dodecyl sulfate lysis buffer or guanidine-HCl, as previously described<sup>24,26,27</sup> and then aliquoted and stored at  $-80^{\circ}\text{C}$  until required. All measurements were made in duplicate and the mean determined.

### Measurement of PDGFRB

PDGFRB level was measured by sandwich ELISA (duoset, Cat no DY385, R&D systems, Oxford, UK). High-binding capacity clear 96-well plates (Costar EIA plates, R&D systems, Oxford, UK) were coated overnight at room temperature with anti-human PDGFRB capture antibody (diluted to 6  $\mu\text{g}/\text{ml}$  in PBS). The plate was washed five-times in PBS:0.05% tween-20 and blocked for 2 h in PBS:1% bovine serum albumin (Sigma Aldrich, Dorset, UK) at room temperature. Following a further wash step, brain tissue samples at (2  $\mu\text{l}$  + 98  $\mu\text{l}$  PBS) or recombinant human PDGFRB

(16,000 – 250 pg/ml) were incubated for 2 h at room temperature without shaking. The plate was washed and biotinylated anti-human PDGFRB detection antibody (diluted to 0.5  $\mu\text{g}/\text{ml}$  in PBS) was added for 2 h at room temperature without shaking. The plate was washed and incubated for 20 min at room temperature with streptavidin:HRP (diluted 1 in 200 in PBS) (R&D systems, UK) and then washed and incubated in the dark with 3,3',5,5'-tetramethylbenzidine (TMB) substrate (R&D systems, UK) for 15 min. Absorbance was read at 450 nm following the addition of 2 N sulfuric acid, in a FLUOstar OPTIMA plate reader (BMG labtech, Aylesbury, UK). The absolute concentration of PDGFRB was interpolated from the standard curve for each case, derived from duplicate measurement of the recombinant PDGFRB.

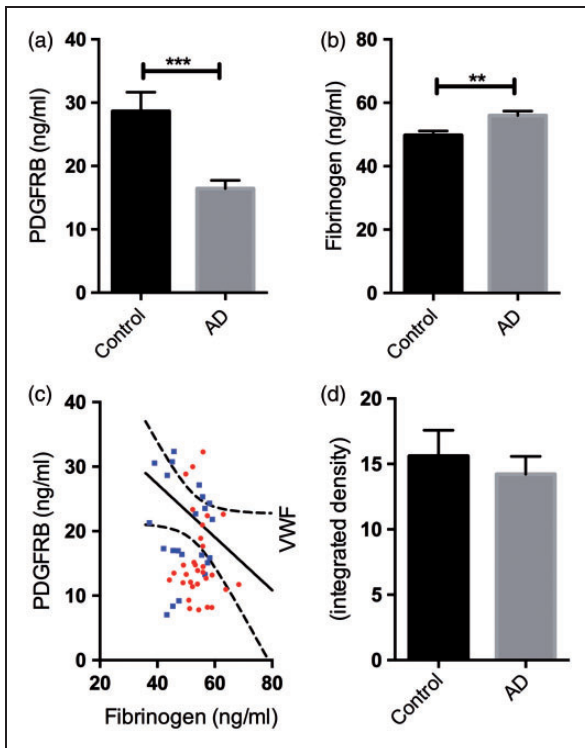
To confirm the specificity of the assay, we performed immunofluorescent labeling of sections of parietal cortex and white matter with the PDGFRB detection antibody used in the ELISA, in combination with antibody to smooth muscle actin or Von Willebrand factor (vWF) (see supplementary information for methodological details). This confirmed the finding of Craggs et al.<sup>44</sup> that PDGFRB is largely restricted to cells associated with capillaries (Supplementary Figure 1). There was negligible overlap of PDGFRB immunofluorescent signal with that of smooth muscle actin in the tunica media of adjacent arterioles, and limited overlap of the PDGFRB signal with that of vWF in the underlying endothelium.

### Measurement of fibrinogen

Fibrinogen content was measured in brain tissue homogenates by use of a commercially available sandwich ELISA (Human Fibrinogen ELISA kit, Cat no EH3057, Wuhan Fine Biological Technology Co, Wuhan City, Hubei Province, China). The plate was pre-coated with an anti-human fibrinogen capture antibody. Recombinant human fibrinogen or brain tissue homogenate (50  $\mu\text{l}$  + 50  $\mu\text{l}$  proprietary dilution buffer) was incubated at  $37^{\circ}\text{C}$  for 90 min. The plate was washed five times in PBS:0.05% tween-20 and biotinylated-detection antibody added at  $37^{\circ}\text{C}$  for 60 min. Bound antibody was detected as above (see Measurement of PDGFRB). The absolute concentration of fibrinogen was interpolated from measurements of serially diluted recombinant human fibrinogen (600–9.375 ng/ml).

### Measurement of hemoglobin

A colorimetric assay kit (Cayman Chemicals, Cat No 700540) (Ann Arbor, MI, USA) was used according to the manufacturer's instructions to measure



**Figure 1.** Platelet-derived growth factor receptor- $\beta$  (PDGFRB) loss and blood-brain barrier (BBB) breakdown in the precuneus in AD. (a) Bar chart showing reduced PDGFRB in AD compared to age-matched controls. (b) Bar chart showing increased fibrinogen level in AD compared to age-matched controls. (c) Scatterplot showing a trend toward a negative correlation between PDGFRB and fibrinogen level in the precuneus ( $r = -0.25$ ,  $P = 0.054$ ). Each point in the scatterplot indicates a single AD (red circle) or control (blue square) brain. The best-fit linear regression line and 95% confidence interval are superimposed. (d) The level of von Willebrand factor (vWF) in the precuneus did not differ significantly between AD and control brains. The bars indicate the mean and SEM. \*\* $P < 0.001$ , \*\*\* $P < 0.0001$ .

the hemoglobin level in the brain tissue homogenates. The proprietary hemoglobin detection reagent was incubated with tissue homogenates and absorbance measured at 560–590 nm in a FLUOstar OPTIMA plate reader. Hemoglobin content was determined by interpolation from measurements of serial dilutions of hemoglobin (0.4–0.016 g/dl) and calculated using the following equation

$$\text{Hemoglobin (g/dl)} = (\text{corrected sample absorbance} - (\text{y} - \text{intercept})/\text{slope}) \times 10 \times \text{sample dilution.}$$

#### Measurement of vWF

vWF level in brain tissue homogenates was determined by dot blot as previously described.<sup>24,27</sup> Samples were

diluted in tris-buffered saline (TBS) (1 in 800) and blotted onto nitrocellulose membrane (GE Healthcare, St. Giles, UK) for 1 h at room temperature. The membrane was blocked in 5% non-fat dried milk protein (NFDMP) in TBS at 4°C overnight, washed, and then incubated for 1 h with polyclonal rabbit anti-human VWF, (0.3  $\mu\text{g/ml}$ ) (Dako, Glostrup, Denmark) at room temperature with agitation. The membrane was washed then incubated with anti-rabbit peroxidase-conjugated secondary antibody (Vector Laboratories, Burlingame, CA, USA) in 5% NFDMP diluted in 0.3% TBS-T for 1 h at room temperature with agitation. The membrane was again washed and then developed using chemiluminescent ECL substrate (Millipore, Billerica, MA, USA) according to the manufacturer's guidelines. Image-J was used to measure the integrated density of each blot. Serial dilutions of a standard reference brain tissue homogenate were used to control for any blot-to-blot variation. We previously demonstrated that vWF is stable under conditions of simulated post-mortem delay for up to 72 h at 4°C or RT and that vWF level, measured by dot blot, is an excellent indicator of microvessel density.<sup>24,45</sup>

#### Measurement of PDGF-BB

PDGFBB level in 1% sodium dodecyl sulfate lysis buffer or guanidine-HCl extracts of the homogenized brain tissue was measured by sandwich ELISA (duoset, Cat no DY220, R&D systems, Oxford, UK). High-binding capacity clear 96-well plates (Costar EIA plates, R&D systems, Oxford, UK) were coated overnight at room temperature with anti-human PDGF-BB capture antibody (diluted to 0.4  $\mu\text{g/ml}$  in PBS). The plate was washed five-times in PBS:0.05% tween-20 and blocked for 2 h in PBS:1% bovine serum albumin (Sigma Aldrich, Dorset, UK) at room temperature. Following a further wash step, brain tissue samples at (5  $\mu\text{l}$  + 95  $\mu\text{l}$  PBS) or recombinant human PDGFRB (2000 – 31.25 pg/ml) were incubated for 2 h at room temperature without shaking. The plate was washed and biotinylated anti-human PDGF-BB detection antibody (diluted to 0.4  $\mu\text{g/ml}$  in PBS) was added for 2 h at room temperature without shaking. The plate was washed and incubated for 20 min at room temperature with streptavidin:HRP (diluted 1:200 in PBS) (R&D systems, UK) and then washed and incubated in the dark with 3,3',5,5'-TMB substrate (R&D systems, UK) for 15 min. Absorbance was read at 450 nm following the addition of 2 N sulfuric acid, in a FLUOstar OPTIMA plate reader (BMG labtech, Aylesbury, UK). The absolute concentration of PDGF-BB was interpolated from the standard curve for each case, derived from duplicate measurement of the recombinant PDGF-BB.

## Statistical analysis

Unpaired two-tailed t-tests or ANOVA with Bonferroni post-hoc analysis were used for comparisons between groups, and Pearson's or Spearman's test to assess linear or rank order correlation, as appropriate, with the help of SPSS version 16 (SPSS, Chicago) and GraphPad Prism version 6 (GraphPad Software, La Jolla, CA).  $P$ -values  $< 0.05$  were considered statistically significant. We also used Wizard version 1.8.22 (<http://www.wizardmac.com/>) to perform multivariable regression analysis of the association of PDGFRB, fibrinogen, and PDGF-BB with age, gender, post-mortem delay, and diagnosis of AD.

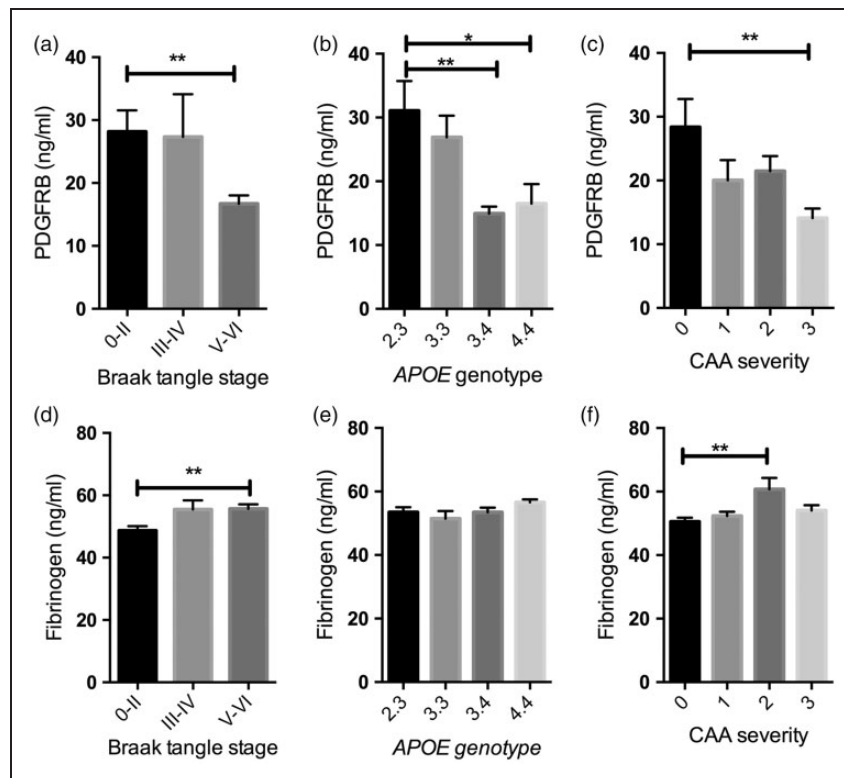
## Results

### PDGFRB loss, BBB breakdown and reduced microvessel density in the precuneus in AD

PDGFRB level was significantly reduced in the precuneus in AD compared to age-matched controls

( $P = 0.0002$ ) (Figure 1(a)), and fibrinogen level, which rises with BBB breakdown<sup>3,4,6</sup> was significantly elevated ( $P = 0.0026$ ) (Figure 1(b)). Fibrinogen level tended to vary inversely with PDGFRB level ( $r = -0.25$ ,  $P = 0.054$ ) (Figure 1(c)), suggesting that BBB breakdown was related to lower pericyte content within the precuneus. There was a small, non-significant reduction in the level of vWF in the precuneus in AD (Figure 1(d)). We previously showed that vWF level is reduced in mid-frontal cortex in AD but was unchanged or elevated in regions of cerebrum that have less A $\beta$  pathology, such as the thalamus.<sup>27</sup>

When grouped according to Braak tangle stage, PDGFRB level was significantly lower (Figure 2(a)) and fibrinogen level higher (Figure 2(d)) in Braak tangle stage V–VI (end-stage) disease than in Braak stage 0–II brains ( $P < 0.01$  for both). The differences in PDGFRB level and fibrinogen between Braak stage 0–II and III–IV, and between Braak stage III–IV and V–VI were not statistically significant. PDGFRB level was significantly lower in *APOE*  $\epsilon 3.4$  ( $P < 0.01$ ) and *APOE*  $\epsilon 4.4$



**Figure 2.** Platelet-derived growth factor receptor- $\beta$  (PDGFRB) loss and blood–brain barrier (BBB) breakdown in relation to disease severity (i.e. Braak tangle stage), *APOE* genotype, and cerebral amyloid angiopathy (CAA) in the precuneus in AD. Bar charts showing (a) reduced PDGFRB in Braak tangle stage V–VI (end-stage) compared to Braak tangle stage 0–II. ( $P < 0.01$ ) (b) reduced PDGFRB level in *APOE*  $\epsilon 3.4$  ( $P < 0.01$ ) and *APOE*  $\epsilon 4.4$  ( $P < 0.05$ ) compared with *APOE*  $\epsilon 2.3$  individuals and (c) reduced PDGFRB level in severe CAA compared to absent CAA ( $P < 0.01$ ). Bar charts showing (d) increased fibrinogen level in Braak tangle stage V–VI (end stage) compared to Braak stage 0–II ( $P < 0.01$ ), (e) no significant difference in fibrinogen level in relation to *APOE* genotype and (f) increased fibrinogen level in moderate CAA compared to absent CAA ( $P < 0.01$ ). The bars indicate the mean and SEM. CAA severity scores adapted from Olichney et al.<sup>42,43</sup> 0 = absent, 1 = mild, 2 = moderate, 3 = severe.

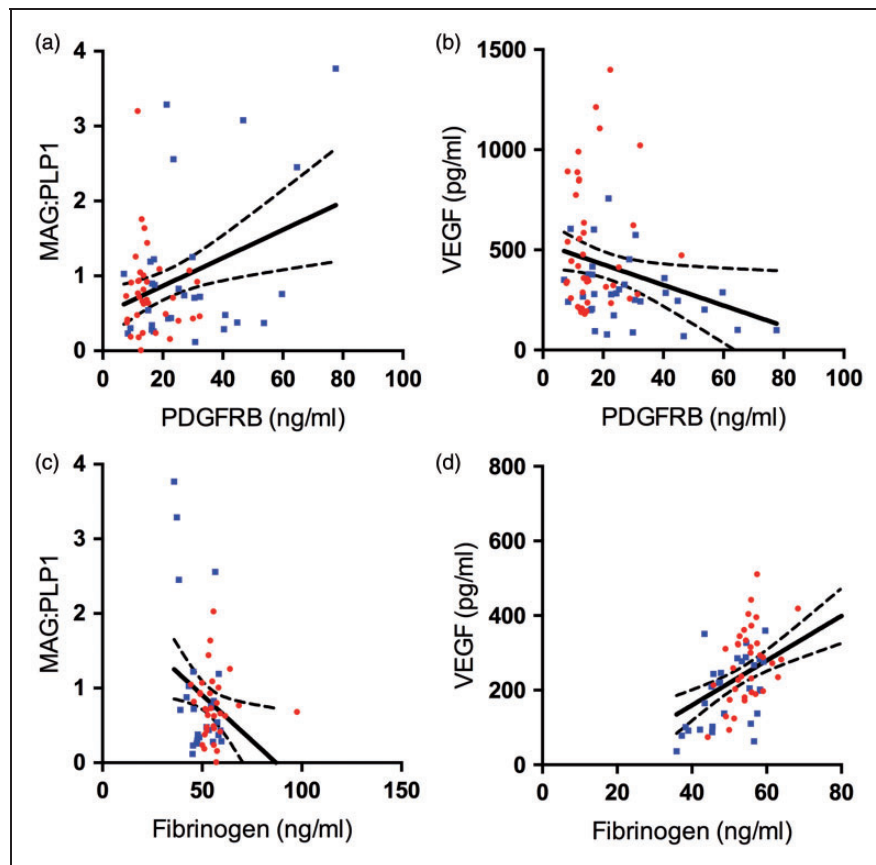
( $P < 0.05$ ) brains than in those with an *APOE*  $\epsilon 2:3$  genotype (Figure 2(b)) but did not vary between *APOE*  $\epsilon 2:3$  and  $\epsilon 3:3$ . PDGFRB level was significantly lower in severe CAA than in brains without CAA ( $P < 0.01$ ) (Figure 2(c)). Fibrinogen level did not vary significantly with *APOE* genotype (Figure 2(e)) but was increased in moderate CAA ( $P < 0.01$ ) (Figure 2(f)).

Finally, to ensure that alterations in fibrinogen level within the brain were not caused by differences in blood content, we compared fibrinogen level after adjusting the values (i) for hemoglobin content, and (ii) for microvessel content/vWF level. The hemoglobin-adjusted values remained higher in AD but the difference did not quite reach significance ( $P = 0.09$ ) (Supplementary Figure 2(a)). The vWF-adjusted fibrinogen values were also elevated in AD but again did not reach statistical significance (Supplementary Figure 2(b)).

There was no significant association between PDGFRB in the precuneus, and gender, age or post-mortem delay in either the AD or control cohort, or when all cases were combined. Fibrinogen in the precuneus did not vary with gender or age but did increase slightly with post-mortem delay in the AD ( $P = 0.018$ , coefficient of correlation 0.163) and combined cohorts ( $P = 0.010$ , coefficient of correlation 0.148). However, the association of PDGFRB with AD remained significant ( $P = 0.004$ ) after incorporating post-mortem delay into a multivariable regression model (results not shown).

### *PDGFRB loss and fibrinogen accumulation in AD, associated with cerebral hypoperfusion of the precuneus*

We previously reported that MAG:PLP1 was reduced in the precuneus from an early stage of AD, indicating



**Figure 3.** Platelet-derived growth factor receptor- $\beta$  (PDGFRB) loss and blood–brain barrier breakdown are associated with hypoperfusion of the precuneus. (a) Scatterplot showing a strong positive correlation between PDGFRB and MAG:PLP1 ( $r = 0.24$ ,  $P = 0.006$ ), i.e. the lowest PDGFRB levels were in samples with least preservation of MAG relative to PLP1. (b) Scatterplot showing a negative correlation between PDGFRB level and VEGF level in the precuneus ( $r = -0.26$ ,  $P = 0.029$ ), i.e. the lowest PDGFRB levels were in samples with greatest elevation in VEGF. (c) Scatterplot showing a negative correlation between fibrinogen level and MAG:PLP1 ( $r = -0.30$ ,  $P = 0.022$ ) and (d) Scatterplot showing a strongly positive correlation between fibrinogen and VEGF level ( $r = 0.49$ ,  $P < 0.0001$ ). Each point in the scatterplots indicates a single AD (red circle) or control (blue square) brain. The best-fit linear regression lines and 95% confidence intervals are superimposed.

a perfusion deficit with respect to the energy requirements.<sup>26</sup> In the present study, PDGFRB level correlated positively with MAG:PLP1 ( $r = +0.34$ ,  $P = 0.006$ ) and negatively with VEGF level ( $r = -0.26$ ,  $P = 0.029$ ), providing evidence that pericyte loss is associated with cerebral hypoperfusion (Figure 3(a) and (b)). Fibrinogen level correlated negatively with MAG:PLP ( $r = -0.30$ ,  $P = 0.022$ ) (Figure 3(c)) and positively with VEGF ( $r = 0.49$ ,  $P < 0.0001$ ) (Figure 3(d)) indicating that BBB breakdown is also associated with chronic cerebral hypoperfusion.

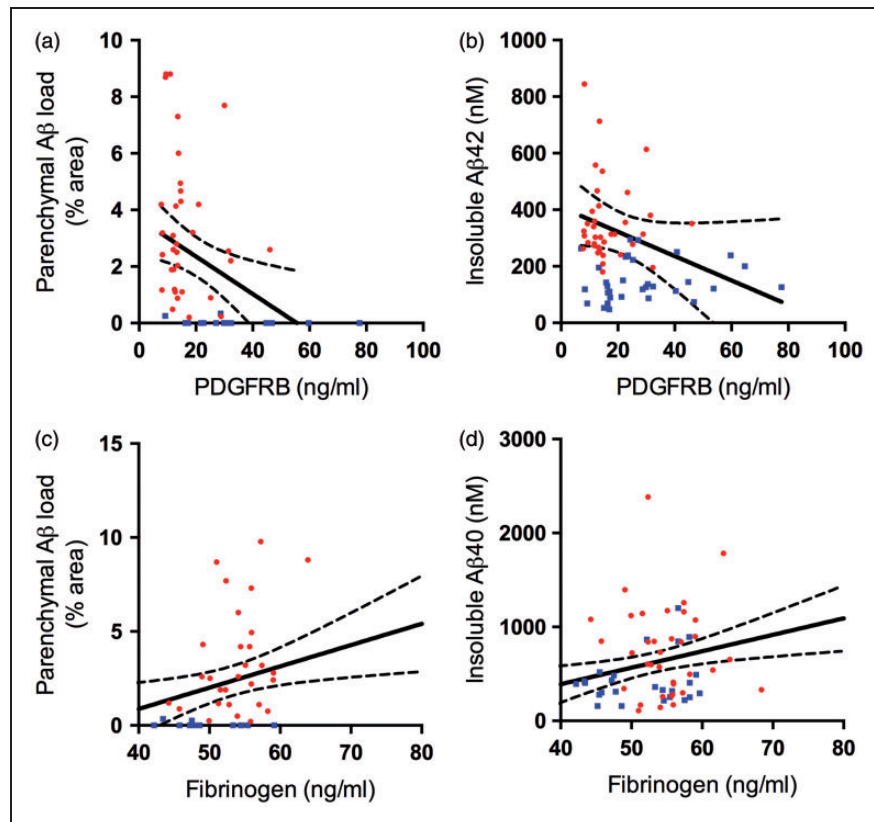
#### **PDGFRB loss and BBB breakdown, associated with accumulation of parenchymal A $\beta$ within the precuneus**

PDGFRB level correlated negatively with parenchymal A $\beta$  plaque load ( $r = -0.36$ ,  $P = 0.010$ ) (Figure 4(a)). PDGFRB level also declined with increasing insoluble A $\beta$ 42 but this relationship did not reach statistical significance ( $r = -0.20$ ,  $P = 0.097$ ) (Figure 4(b)).

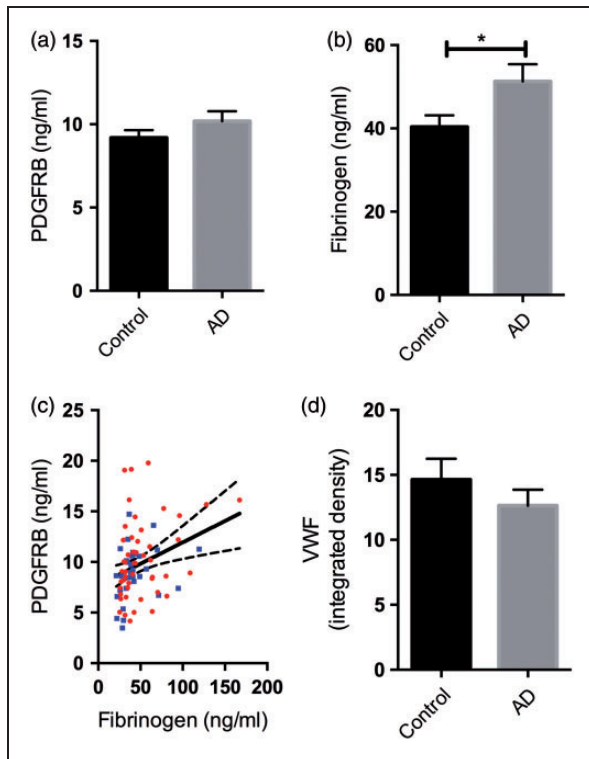
Fibrinogen level correlated positively with parenchymal A $\beta$  load ( $r = 0.37$ ,  $P = 0.015$ ) (Figure 4(c)) and insoluble A $\beta$ 40 ( $r = 0.33$ ,  $P = 0.006$ ) (Figure 4(d)) but not A $\beta$ 42. No correlations were observed between either PDGFRB or fibrinogen with soluble species of A $\beta$ 40 or A $\beta$ 42.

#### **Hypoperfusion of cerebral white matter in AD, associated with fibrinogen accumulation without reduction in PDGFRB**

In contrast to the precuneus, the white matter did not show significant alteration in PDGFRB in AD (Figure 5(a)). Fibrinogen level in the white matter was significantly higher in AD than controls ( $P = 0.05$ ) (Figure 5(b)) and remained higher after adjustment for hemoglobin but not vWF level (Supplementary Figure 2(c) and (d)). Unlike in the precuneus, in underlying white matter, PDGFRB correlated positively (rather than negatively) with fibrinogen level ( $r = 0.36$ ,  $P < 0.0001$ ) (Figure 5(c)). Differences between AD and



**Figure 4.** Platelet-derived growth factor receptor- $\beta$  (PDGFRB) loss and BBB breakdown in the precuneus in AD are related to A $\beta$  level. (a–b) Scatterplots showing negative correlation between PDGFRB and A $\beta$  plaque load (area fraction of cortex immunopositive for A $\beta$ ) ( $r = -0.36$ ,  $P = 0.022$ ) and insoluble A $\beta$ 42 level ( $r = -0.26$ ,  $P = 0.029$ ). (c–d) Scatterplots showing positive correlation between fibrinogen level and A $\beta$  plaque load ( $r = 0.37$ ,  $P = 0.015$ ) and insoluble A $\beta$ 40 ( $r = 0.35$ ,  $P = 0.006$ ). Each point in the scatterplots indicates a single AD (red circle) or control (blue square) brain. The best-fit linear regression lines and 95% confidence intervals are superimposed.



**Figure 5.** Blood–brain barrier (BBB) breakdown in parietal white matter in AD not associated with pericyte loss. (a) There was no significant change in platelet-derived growth factor receptor- $\beta$  (PDGFRB) in the parietal white matter in AD. (b) Bar chart showing increased fibrinogen level in white matter in AD compared to age-matched controls. (c) Scatterplot showing highly significant positive correlation between PDGFRB and fibrinogen level in white matter ( $r=0.36$ ,  $P<0.0001$ ). Each point indicates a single AD (red circle) or control (blue square) brain. The best-fit linear regression line and 95% confidence interval are superimposed. (d) The level of von Willebrand factor (vWF), a marker of vessel density, was not significantly altered in the parietal white matter in AD. The bars indicate the mean and SEM. \* $P<0.05$ .

control brains in vWF level in the parietal white matter were not significant (Figure 5(d)). White matter PDGFRB and fibrinogen level did not vary significantly with Braak tangle stage or *APOE* genotype (Supplementary Figure 3).

We previously reported on chronic hypoperfusion (reduction in MAG:PLP1) in the parietal white matter in AD.<sup>24</sup> Unlike in the precuneus, PDGFRB level in the white matter correlated negatively with MAG:PLP1 ( $r=-0.31$ ,  $P=0.004$ ) (Figure 6(a)) and positively with VEGF ( $r=0.19$ ,  $P=0.019$ ) (Figure 6(b)) indicating an increase in PDGFRB with declining white matter perfusion. Fibrinogen level, however, showed a strong negative correlation with MAG:PLP1 ( $r=-0.48$ ,  $P<0.0001$ ) and a strong

positive correlation with VEGF ( $r=0.42$ ,  $P<0.0001$ ), suggesting a close relationship between breakdown of the BBB and chronic hypoperfusion in the white matter (Figure 6(c) and (d)).

There was no significant association between PDGFRB and fibrinogen in the white matter, and gender, age or post-mortem delay in either the AD or control cohort, or when all cases were combined.

#### *Increased PDGF-BB in AD, associated with hypoperfusion, BBB leakiness, and A $\beta$ accumulation*

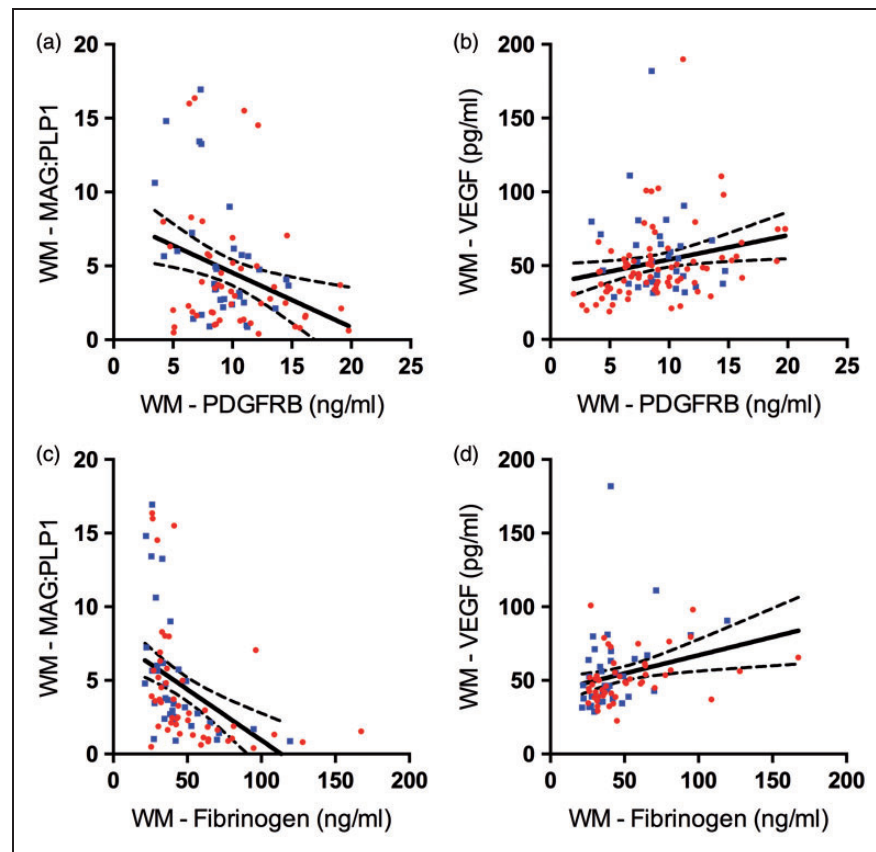
To determine whether the PDGFRB loss in the precuneus was secondary to a decline in PDGF-BB (a key trophic factor for pericytes), we measured the concentration of PDGF-BB in the precuneus and underlying white matter, in both SDS-extracted (soluble) and guanidine-HCl (insoluble) fractions. In AD, the level of PDGF-BB was significantly increased in both regions and in both fractions ( $P<0.0001$  for all comparisons between AD and control brains except in the insoluble white matter fraction where  $P=0.001$ ) (Figure 7). Of possible relevance, the concentration of PDGF-BB was approximately 3- to 4-fold higher in the insoluble than the corresponding soluble fractions. In the precuneus, PDGF-BB concentration showed highly significant positive correlations with the levels of insoluble A $\beta$ 40 ( $r=0.43$ ,  $P<0.01$ ) and A $\beta$ 42 ( $r=0.40$ ,  $P<0.01$ ) (Supplementary Table 3). In contrast, PDGF-BB showed weaker negative correlations with A $\beta$ 40 ( $r=-0.298$ ,  $P<0.05$ ) and A $\beta$ 42 ( $r=-0.212$ ,  $P<0.05$ ) within the GuHCl-extract in the underlying white matter, which contains much less insoluble A $\beta$ .<sup>41</sup>

There was no significant association between PDGF-BB in the precuneus or white matter, and gender, age or post-mortem delay in either the AD or control cohort, or when all cases were combined.

## Discussion

We previously reported that oxygenation of both the precuneus and the underlying white matter was reduced in the early stages of AD.<sup>26</sup> Present findings show the reduced oxygenation of the precuneus to be associated with loss of the pericyte protein PDGFRB, and the accumulation of fibrinogen and A $\beta$  (particularly plaque-associated fibrillar A $\beta$ ). In contrast, hypoperfusion of the underlying white matter (where the amount of A $\beta$  is orders of magnitude lower than in the precuneus in AD<sup>41</sup>) was associated with fibrinogen accumulation in the absence of any reduction in PDGFRB. The pathophysiological processes that underlie pericyte degeneration, BBB breakdown, and cerebral hypoperfusion are thus region-specific and likely to be influenced by A $\beta$  level. In neither cortex nor white matter





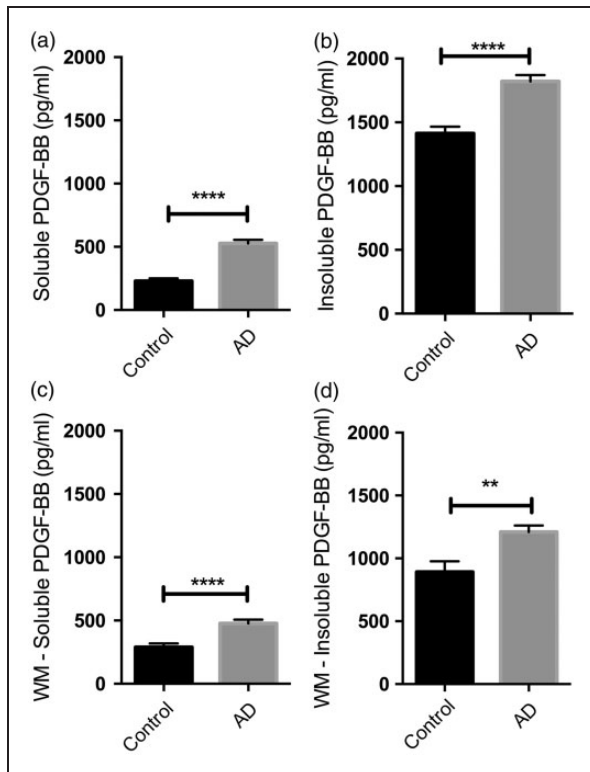
**Figure 6.** Reduced oxygenation in the parietal white matter in AD associated with blood–brain barrier (BBB) breakdown despite concomitant increase in pericytes. (a) Scatterplot showing strong negative correlation between platelet-derived growth factor receptor- $\beta$  (PDGFRB) and myelin-associated glycoprotein:proteolipid protein-1 (MAG:PLP1) ratio in white matter (WM) ( $r = -0.31$ ,  $P = 0.004$ ). (b) Scatterplot showing positive correlation between platelet-derived growth factor receptor- $\beta$  (PDGFRB) level and vascular endothelial growth factor (VEGF) level ( $r = 0.19$ ,  $P = 0.019$ ). (c) Scatterplot showing very strong negative correlation between fibrinogen and MAG:PLP1 ( $r = -0.48$ ,  $P < 0.0001$ ). (d) Scatterplot showing very strong positive correlation between fibrinogen and VEGF level in the white matter ( $r = 0.42$ ,  $P < 0.0001$ ). Each point in the scatterplots indicates a single AD (red circle) or control (blue square) brain. The best-fit linear regression lines and 95% confidence intervals are superimposed.

could loss of PDGFRB be attributed to a reduced level of PDGF-BB, which was elevated in AD. However, as discussed below, the differential distribution of fibrillar A $\beta$  may have influenced the availability of this trophic factor.

Our findings are consistent with previous reports of an association between pericyte loss and BBB breakdown in AD<sup>34</sup> and provide further evidence that pericyte loss is related to A $\beta$  level and influenced by *APOE* genotype.<sup>3,4</sup> At least within the cerebral cortex, there is also a close relationship between PDGFRB content, fibrinogen, and blood flow, in keeping with *in vivo* evidence that loss of BBB integrity is related to pericyte degeneration, cerebral hypoperfusion,<sup>37</sup> and cognitive impairment.<sup>36</sup> Uncertainty persists as to the precise timing of the various vascular abnormalities in AD and the details of the complex causal interrelationships. In *Pdgfr $\beta$ <sup>+/-</sup>* mice, age-related pericyte loss led to loss

of cerebral microvessels, breakdown of the BBB, reduced cerebral blood flow, and neurovascular uncoupling.<sup>9</sup> Evidence that these vascular abnormalities have the potential to exacerbate AD pathology comes from a study in which *APP<sup>sw/0</sup>* mice developed more severe A $\beta$  and tau pathology, neuronal loss, and memory impairment when they were crossed with *Pdgfr $\beta$ <sup>+/-</sup>* mice.<sup>33</sup>

In contrast, several lines of evidence suggest that A $\beta$  contributes to pericyte injury,<sup>33,35,46</sup> BBB dysfunction,<sup>3,4</sup> and hypoperfusion.<sup>26,27,47</sup> In recent years imaging studies, particularly in individuals with familial AD, have shown that A $\beta$  begins to accumulate as early as 20 years before the development of dementia, whereas cerebral hypoperfusion occurs later, 5 to 10 years before disease onset.<sup>11,12</sup> *In vitro* studies indicate that A $\beta$  peptides cause pericyte injury and death.<sup>33,35,46</sup> It is also possible that the damaging effects of A $\beta$  on pericyte survival are partly indirect, through the sequestration of PDGF-BB,



**Figure 7.** Platelet-derived growth factor- $\beta\beta$  (PDGF-BB) level is increased in the precuneus and underlying white matter in AD, and present in greater quantities in the insoluble fraction. (a–b) Bar charts showing increased PDGF-BB in the precuneus in AD in (a) SDS-extracted soluble ( $P < 0.0001$ ) and (b) guanidine-HCl-extracted insoluble fractions of brain tissue homogenates ( $P < 0.0001$ ). (c–d) Bar charts showing increased PDGF-BB in underlying white in AD in (c) soluble ( $P < 0.0001$ ) and (d) insoluble ( $P = 0.001$ ) fractions. The bars indicate the mean and SEM.

upregulation of VEGF, and even through competition for binding to PDGFRB.

PDGF-BB was shown previously to be mainly associated with A $\beta$  plaques in AD brains.<sup>48</sup> Although we found PDGF-BB to be increased in AD, possibly as a physiological response to reduced tissue oxygenation, most of it was in the insoluble fraction of the tissue, which also contained high levels of A $\beta$ . Indeed, there was a close correlation between the levels of PDGF-BB and A $\beta$  within the insoluble fraction, as might be expected if the PDGF-BB was sequestered by fibrillar A $\beta$  and thus biologically inactive – a mechanism proposed to apply to another vascular trophic factor in AD, VEGF.<sup>49</sup> VEGF level is significantly increased in AD, as noted in the present study and also previously.<sup>27</sup> Although VEGF induces vasculogenesis as well as elongation and migration of pericytes, and may promote their interaction with capillaries during development,<sup>50</sup> VEGF also acts as a negative regulator of pericyte function and maturation of vessels, interferes

with the interaction between pericytes and endothelial cells and may promote vascular leakage.<sup>51–53</sup> A final possibility to consider is whether A $\beta$  may interfere with the binding of PDGF-BB to its pericyte receptor, PDGFRB. Again, the VEGF signaling pathway provides an example of this mechanism, in that A $\beta$  was shown to interfere with the binding of VEGF to VEGF receptor-2.<sup>54</sup>

Data from previous studies suggest that there is loss of pericytes and leakage of fibrinogen at a relatively early stage of AD.<sup>34,36,37</sup> We found PDGFRB and fibrinogen levels to vary significantly with Braak tangle stage. On pairwise post hoc testing, only differences between Braak stage 0–II and V–VI disease were significantly different. However, our cohort included relatively few cases in the Braak stage III–IV group and there was greater variation between cases in this group than in the 0–II and V–VI groups. Further analysis of larger cohorts is needed to clarify the precise timing of pericyte loss and BBB leakage in relation to tangle pathology and other pathological features of AD.

We found that PDGFRB level was preserved or even increased in the white matter in AD, possibly because there was insufficient fibrillar A $\beta$  to damage pericytes or to sequester PDGF-BB. Factors other than pericyte loss must account for the white matter hypoperfusion and BBB leakage. Most of the perfusion of the white matter derives from transcortical perforating arterioles, and we reported previously that oxygenation of the cerebral white matter declined with increasing levels of the vasoconstrictor EDN1 in the overlying cortex, suggesting that hypoperfusion of the white matter in AD results partly from vasoconstriction of perforating arterioles as they traverse the cortex.<sup>26,47,55</sup> It is possible that BBB leakage in the white matter is secondary to hypoperfusion.<sup>38,39</sup> Alternatively, it could reflect upregulation of the plasma kallikrein-kinin system in AD,<sup>56</sup> leading to increased production of bradykinin, which increases the permeability of the BBB.<sup>57–59</sup> This merits further study.

Together, our data suggest that pericytes degenerate in regions of brain with elevated fibrillar A $\beta$ , causing or exacerbating breakdown of the BBB, and possibly contributing to loss of microvessels, hypoperfusion, and impaired vascular clearance of A $\beta$ . Factors other than pericyte degeneration are likely to be responsible for hypoperfusion and breakdown of the BBB in the white matter in AD. Further study is needed of the timing, regional distribution, and mechanisms of pericyte degeneration in AD.

### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by Alzheimer's Research

UK (ARUK-PG2015-11). The South West Dementia Brain Bank is part of the Brains for Dementia Research program, jointly funded by Alzheimer's Research UK and Alzheimer's Society, and is supported by BRACE (Bristol Research into Alzheimer's and Care of the Elderly) and the Medical Research Council.

### Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

### Authors' contributions

JSM and SL were responsible for the conception and design of experiments; JSM and IS were responsible for acquisition of data; JSM and SL analyzed and interpreted the data; JSM drafted the manuscript; SL revised and reviewed the final article for intellectual content and final approval.

### Supplementary material

Supplementary material for this paper can be found at the journal website: <http://journals.sagepub.com/home/jcb>.

### References

- Baloyannis SJ and Baloyannis IS. The vascular factor in Alzheimer's disease: A study in Golgi technique and electron microscopy. *J Neurol Sci* 2012; 322: 117–121.
- Farkas E and Luiten PG. Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol* 2001; 64: 575–611.
- Halliday MR, Rege SV, Ma Q, et al. Accelerated pericyte degeneration and blood-brain barrier breakdown in apolipoprotein E4 carriers with Alzheimer's disease. *J Cereb Blood Flow Metab* 2016; 36: 216–227.
- Sengillo JD, Winkler EA, Walker CT, et al. Deficiency in mural vascular cells coincides with blood-brain barrier disruption in Alzheimer's disease. *Brain Pathol* 2013; 23: 303–310.
- Cortes-Canteli M, Mattei L, Richards AT, et al. Fibrin deposited in the Alzheimer's disease brain promotes neuronal degeneration. *Neurobiol Aging* 2015; 36: 608–617.
- Hultman K, Strickland S and Norris EH. The APOE  $\epsilon$ 4/ $\epsilon$ 4 genotype potentiates vascular fibrin(ogen) deposition in amyloid-laden vessels in the brains of Alzheimer's disease patients. *J Cereb Blood Flow Metab* 2013; 33: 1251–1258.
- Zipser BD, Johanson CE, Gonzalez L, et al. Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. *Neurobiol Aging* 2007; 28: 977–986.
- Craggs LJL, Fenwick R, Oakley AE, et al. Immunolocalization of platelet-derived growth factor receptor- $\beta$  (PDGFR- $\beta$ ) and pericytes in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL). *Neuropath Appl Neuro* 2015; 41: 557–570.
- Bell RD, Winkler EA, Sagare AP, et al. Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* 2010; 68: 409–427.
- Asllani I, Habeck C, Scarmeas N, et al. Multivariate and univariate analysis of continuous arterial spin labeling perfusion MRI in Alzheimer's disease. *J Cereb Blood Flow Metab* 2008; 28: 725–736.
- Benzinger TLS, Blazey T, Jack CR, et al. Regional variability of imaging biomarkers in autosomal dominant Alzheimer's disease. *Proc Natl Acad Sci U S A* 2013; 110: E4502–E4509.
- Binnewijzend MAA, Kuijper JPA, Benedictus MR, et al. Cerebral blood flow measured with 3D pseudocontinuous arterial spin-labeling MR imaging in Alzheimer disease and mild cognitive impairment: A marker for disease severity. *Radiology* 2013; 267: 221–230.
- Dai WY, Lopez OL, Carmichael OT, et al. Mild cognitive impairment and Alzheimer disease: Patterns of altered cerebral blood flow at MR imaging. *Radiology* 2009; 250: 856–866.
- Langbaum JBS, Chen KW, Caselli RJ, et al. Hypometabolism in Alzheimer-affected brain regions in cognitively healthy Latino individuals carrying the apolipoprotein E  $\epsilon$ 4 allele. *Arch Neurol* 2010; 67: 462–468.
- Matsuda H. Cerebral blood flow and metabolic abnormalities in Alzheimer's disease. *Ann Nucl Med* 2001; 15: 85–92.
- Sakamoto S, Ishii K, Sasaki M, et al. Differences in cerebral metabolic impairment between early and late onset types of Alzheimer's disease. *J Neurol Sci* 2002; 200: 27–32.
- Ruitenbergh A, den Heijer T, Bakker SLM, et al. Cerebral hypoperfusion and clinical onset of dementia: The Rotterdam study. *Ann Neurol* 2005; 57: 789–794.
- Niwa K, Kazama K, Younkin L, et al. Cerebrovascular autoregulation is profoundly impaired in mice overexpressing amyloid precursor protein. *Am J Physiol Heart Circ Physiol* 2002; 283: H315–H323.
- Niwa K, Kazama K, Younkin SG, et al. Alterations in cerebral blood flow and glucose utilization in mice overexpressing the amyloid precursor protein. *Neurobiol Dis* 2002; 9: 61–68.
- Niwa K, Porter VA, Kazama K, et al. A $\beta$ -peptides enhance vasoconstriction in cerebral circulation. *Am J Physiol Heart Circ Physiol* 2001; 281: H2417–2424.
- Chui HC, Zheng L, Reed BR, et al. Vascular risk factors and Alzheimer's disease: Are these risk factors for plaques and tangles or for concomitant vascular pathology that increases the likelihood of dementia? An evidence-based review. *Alzheimers Research & Therapy* 2012; 4: 1.
- Love S and Miners JS. Cerebrovascular disease in ageing and Alzheimer's disease. *Acta Neuropathol* 2016; 131: 645–658.
- Miners JS, Palmer JC, Tayler H, et al. Abeta degradation or cerebral perfusion? Divergent effects of multifunctional enzymes. *Front Aging Neurosci* 2014; 6: 238.
- Barker R, Ashby EL, Wellington D, et al. Pathophysiology of white matter perfusion in Alzheimer's disease and vascular dementia. *Brain* 2014; 137: 1524–1532.
- Barker R, Wellington D, Esiri MM, et al. Assessing white matter ischemic damage in dementia patients by

- measurement of myelin proteins. *J Cereb Blood Flow Metab* 2013; 33: 1050–1057.
26. Miners JS, Palmer JC and Love S. Pathophysiology of hypoperfusion of the precuneus in early Alzheimer's disease. *Brain Pathol* 2016; 26: 533–541.
  27. Thomas TL, Miners SJ, Kehoe PG, et al. Correlation between levels of VEGF and insoluble A $\beta$ (1-42) in the cerebral cortex in Alzheimer's disease. *Neuropathol Appl Neurobiol* 2015; 41: 22.
  28. Love S and Miners JS. White matter hypoperfusion and damage in dementia: Post-mortem assessment. *Brain Pathol* 2015; 25: 99–107.
  29. Palmer JC, Barker R, Kehoe PG, et al. Endothelin-1 is elevated in Alzheimer's disease and upregulated by amyloid- $\beta$ . *J Alzheimers Dis* 2012; 29: 853–861.
  30. Palmer JC, Baig S, Kehoe PG, et al. Endothelin-converting enzyme-2 is increased in Alzheimer's disease and upregulated by A $\beta$ . *Am J Pathol* 2009; 175: 262–270.
  31. Wu Z, Guo H, Chow N, et al. Role of the ME0X2 homeobox gene in neurovascular dysfunction in Alzheimer disease. *Nat Med* 2005; 11: 959–965.
  32. Winkler EA, Bell RD and Zlokovic BV. Pericyte-specific expression of PDGF  $\beta$  receptor in mouse models with normal and deficient PDGF beta receptor signaling. *Mol Neurodegener* 2010; 5: 32.
  33. Sagare AP, Bell RD, Zhao Z, et al. Pericyte loss influences Alzheimer-like neurodegeneration in mice. *Nat Commun* 2013; 4.
  34. Montagne A, Barnes SR, Sweeney MD, et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 2015; 85: 296–302.
  35. Sagare AP, Sweeney MD, Makshanoff J, et al. Shedding of soluble platelet-derived growth factor receptor- $\beta$  from human brain pericytes. *Neurosci Lett* 2015; 607: 97–101.
  36. van de Haar HJ, Burgmans S, Jansen JF, et al. Blood-brain barrier leakage in patients with early Alzheimer disease. *Radiology* 2016; 281: 527–535.
  37. van de Haar HJ, Jansen JF, van Osch MJ, et al. Neurovascular unit impairment in early Alzheimer's disease measured with magnetic resonance imaging. *Neurobiol Aging* 2016; 45: 190–196.
  38. Garrigue P, Giacomino L, Bucci C, et al. Single photon emission computed tomography imaging of cerebral blood flow, blood-brain barrier disruption, and apoptosis time course after focal cerebral ischemia in rats. *Int J Stroke* 2016; 11: 117–126.
  39. Lorberboym M, Lampl Y and Sadeh M. Correlation of 99mTc-DTPA SPECT of the blood-brain barrier with neurologic outcome after acute stroke. *J Nucl Med* 2003; 44: 1898–1904.
  40. Montine TJ, Phelps CH, Beach TG, et al. National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta Neuropathol* 2012; 123: 1–11.
  41. Miners JS, Clarke P and Love S. Clusterin levels are increased in Alzheimer's disease and influence the regional distribution of A $\beta$ . *Brain Pathol* 2016. Epub ahead of print 8 July 2016. DOI: 10.1111/bpa.12392.
  42. Chalmers K, Wilcock GK and Love S. APOE  $\epsilon$ 4 influences the pathological phenotype of Alzheimer's disease by favouring cerebrovascular over parenchymal accumulation of A $\beta$  protein. *Neuropathol Appl Neurobiol* 2003; 29: 231–238.
  43. Olichney JM, Hansen LA, Galasko D, et al. The apolipoprotein E  $\epsilon$ 4 allele is associated with increased neuritic plaques and cerebral amyloid angiopathy in Alzheimer's disease and Lewy body variant. *Neurology* 1996; 47: 190–196.
  44. Craggs LJ, Fenwick R, Oakley AE, et al. Immunolocalization of platelet-derived growth factor receptor- $\beta$  (PDGFR- $\beta$ ) and pericytes in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL). *Neuropathol Appl Neurobiol* 2015; 41: 557–570.
  45. Miners S, Moulding H, de Silva R, et al. Reduced vascular endothelial growth factor and capillary density in the occipital cortex in dementia with Lewy bodies. *Brain Pathol* 2014; 24: 334–343.
  46. Verbeek MM, Van Nostrand WE, Otte-Holler I, et al. Amyloid- $\beta$ -induced degeneration of human brain pericytes is dependent on the apolipoprotein E genotype. *Ann N Y Acad Sci* 2000; 903: 187–199.
  47. Love S and Miners JS. Cerebral hypoperfusion and the energy deficit in Alzheimer's disease. *Brain Pathol* 2016; 26: 607–617.
  48. Masliah E, Mallory M, Alford M, et al. PDGF is associated with neuronal and glial alterations of Alzheimer's disease. *Neurobiol Aging* 1995; 16: 549–556.
  49. Yang SP, Bae DG, Kang HJ, et al. Co-accumulation of vascular endothelial growth factor with beta-amyloid in the brain of patients with Alzheimer's disease. *Neurobiol Aging* 2004; 25: 283–290.
  50. Hagedorn M, Balke M, Schmidt A, et al. VEGF coordinates interaction of pericytes and endothelial cells during vasculogenesis and experimental angiogenesis. *Dev Dynam* 2004; 230: 23–33.
  51. Greenberg JI, Shields DJ, Barillas SG, et al. A role for VEGF as a negative regulator of pericyte function and vessel maturation. *Nature* 2008; 456: 809–813.
  52. Bergers G and Song S. The role of pericytes in blood-vessel formation and maintenance. *Neuro-Oncology* 2005; 7: 452–464.
  53. Bai Y, Zhu XJ, Chao J, et al. Pericytes contribute to the disruption of the cerebral endothelial barrier via increasing VEGF expression: Implications for stroke. *Plos One* 2015; 10: e0124362.
  54. Patel NS, Mathura VS, Bachmeier C, et al. Alzheimer's beta-amyloid peptide blocks vascular endothelial growth factor mediated signaling via direct interaction with VEGFR-2. *J Neurochem* 2010; 112: 66–76.

55. Charidimou A, Pantoni L and Love S. The concept of sporadic cerebral small vessel disease: A road map on key definitions and current concepts. *Int J Stroke* 2016; 11: 6–18.
56. Ashby EL, Love S and Kehoe PG. Assessment of activation of the plasma Kallikrein-Kinin system in frontal and temporal cortex in Alzheimer's disease and vascular dementia. *Neurobiol Aging* 2012; 33: 1345–1355.
57. Bartus RT, Elliott P, Hayward N, et al. Permeability of the blood brain barrier by the bradykinin agonist, RMP-7: Evidence for a sensitive, auto-regulated, receptor-mediated system. *Immunopharmacology* 1996; 33: 270–278.
58. Bartus RT, Elliott PJ, Dean RL, et al. Controlled modulation of BBB permeability using the bradykinin agonist, RMP-7. *Exp Neurol* 1996; 142: 14–28.
59. Raymond JJ, Robertson DM and Dinsdale HB. Pharmacological modification of bradykinin induced breakdown of the blood-brain barrier. *Can J Neurol Sci* 1986; 13: 214–220.