

The Effects of Onion Consumption on Prevention of Nonalcoholic Fatty Liver Disease

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Abstract It is well known that dietary intakes play a pivotal role in pathogenesis of nonalcoholic fatty liver disease (NAFLD)/nonalcoholic steatohepatitis (NASH); however, the role of each component of diet has not yet been elucidated. Our objective was to evaluate the effects of onion consumption on prevention of NAFLD/NASH development. Sprague–Dawley rats were fed either high-fat, high sugar diet (model group), or high-fat, high sugar diet plus 7% onion powder (model + onion), or chow diet ad libitum for 7 weeks. Serum levels of fasting glucose, triglyceride, cholesterol, liver enzymes, insulin, and hepatic tumor necrosis factor-alpha (TNF- α) gene expression were determined. Hepatic histology was examined by H&E stain. Model + onion group had significantly lower hepatic steatosis, ballooning, lobular inflammation, and portal inflammation ($p < 0.001$), lower hepatic TNF- α gene expression ($p < 0.001$), lower plasma levels of ALT ($p = 0.026$), AST ($p = 0.041$), insulin ($p < 0.001$), TG ($p = 0.041$), and glucose ($p = 0.009$) compared with model group; however, weight gain, food intake, plasma total cholesterol and LDL levels were not significantly different between these two groups. Our data indicate that regular consumption of onion can prevent NAFLD even in the presence of the other risk factors such as obesity,

hypercholesterolemia, and high energy, fat, and sugar intakes.

Keywords Onion · NAFLD · Fatty liver · Prevention · Experimental model

Introduction

Nonalcoholic fatty liver disease (NAFLD) includes a spectrum of hepatic disorders ranging from simple triglyceride (TG) accumulation in hepatocytes through hepatic steatosis with inflammation, which is called as nonalcoholic steatohepatitis (NASH), to cirrhosis [1]. Although the exact pathophysiology of NAFLD has not yet elucidated, there is a consensus on the “multiple hit” hypothesis [2]. According to this hypothesis, multiple factors interact for the development of NAFLD. The first hit is the accumulation of TG in hepatocytes mostly due to insulin resistance (IR) [3]. The subsequent hits include a combination of oxidative stress, lipid peroxidation, and release of inflammatory mediators [4]. Although some dietary interventions have shown promising effects in the treatment of NAFLD [5–9], there are scarce studies evaluating the effects of dietary intakes in prevention of the disease.

Given the high content of phytochemicals, such as flavonoids, fructo-oligosaccharides (FOS), thiosulphinates and other sulphur compounds [10], Onion is an excellent source of antioxidants, immunomodulators, antimicrobs, and prebiotics [11]. Thus, we hypothesized that its regular consumption can inhibit the pathogenesis of NAFLD. To examine this hypothesis, we designed an experimental study evaluating the effects of regular consumption of onion on the development of a high fat, high sugar NAFLD/NASH model.

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Materials and Methods

Animals and Diets

Eighteen male Sprague–Dawley rats (weighted 120–150 g), which were purchased from Pasteur Institute (Karaj, Iran), were housed individually in wire bar-floor cages. Body weights (BW) in grams were recorded on arrival and every 2 week thereafter. Food intake was also monitored twice a week. The animals were allowed 1 week of acclimatization in a standard environment at 22 °C, 50% humidity and 12-h light/dark cycles with free access to food and water. During the first week, all animals were fed a standard laboratory chow diet (Pasture Institute, Iran) and afterwards, they were randomly assigned to three groups: first group fed a standard chow diet (control group) with 10% of energy derived from fat, 30% from protein, and 60% from carbohydrates, second group fed a high-fat, high sugar diet (model group) [12] with 59% of energy derived from fat, 30% from carbohydrates, and 11% from protein and finally the third group fed high-fat, high sugar diet with 7% (w/w) onion powder (model + onion group) including 59% of energy derived from fat, 31% from carbohydrates, and 10% from protein. Onion powder was prepared according to the methods used by Hamazu et al. [13]. All animals were fed ad libitum. The diets were prepared weekly and stored as vacuum packed (500 g) at –20 °C. Packs taken for use were thawed in the refrigerator at 4 °C. The food was offered daily, and the remains were weighed and removed after 48 h.

After 7-week feeding period, animals were killed by exsanguination (under light pentobarbital anesthesia). All animal procedures were carried out in accordance with the National Nutrition and Food Technology Research Institute (NNFTRI). The study protocol was approved at NNFTRI ethics committee with ethics code of NNFTRI 1393-568.

Tissue and Blood Preparation

Animals were sacrificed by exsanguination of blood from their heart under chloroform anesthesia. Five milliliters blood were collected in heparinized tubes; then, centrifuged (3500 rpm, 15 min, at 6 °C) to obtain the plasma. Fasted plasma glucose was measured immediately, and the remaining samples were kept at –80 °C before biochemical analysis.

After blood sampling, the livers were excised, washed with cold physiologic saline (0.9%), and dried. One part of each hepatic lobe was preserved in 10% buffered-formalin solution for histopathologic examination. Other liver samples were placed in liquid nitrogen tank, and then kept at –80 °C for gene expression evaluation.

RNA Extraction and Quantitative RT-PCR

Total RNA was purified using RNeasy Plus Mini Kits (Qiagen) according to the manufacturer's instructions and cDNA synthesized with Superscript II reverse transcriptase (Invitrogen). Quantitative real-time PCR was performed using the Bio-Rad Laboratories MJ mini Opticon real-time PCR system, using IQ SYBR Green Supermix (Bio-Rad).

The PCR mix contained 2 µl cDNA, 1 µl of the appropriate forward and reverse primers, and 2 µl SYBR Green PCR Master mix in a total volume of 25 µl. PCR consisted of 50 cycles of denaturation at 94 °C for 30 s, annealing at melting temperature (T_m) for 30 s, and extension at 72 °C for 60 s. Primer sequences for each target gene, their source as well as their optimal PCR annealing temperatures are as follows: GAPDH forward primer 5'-GTGCTGAGTATGTC GTGGAGTCTA-3' and reverse 5'-TCTCGTGGTTCACA CCCATCAC-3' (T_m 60 °C), and TNF- α forward primer 5'-ACT GAA CTT CGG GGT GAT TG-3' and reverse 5'-GCT TGG TGG TTT GCT ACG AC-3' (T_m 60 °C). Primer specificity was confirmed from the product size by agarose gel electrophoresis and the specificity of the PCR products checked by melt curve analysis.

Biochemical Assessments

Plasma concentrations of alanine aminotransferase (ALT), and aspartate aminotransferase (AST) were measured using optimized UV at 340 nm, and insulin concentrations were measured using a rat insulin radioimmunoassay kit at 4 °C (Linco Research Inc, St Charles, MO). Plasma Glucose and TG were measured calorimetrically, Gamma glutamyl transferase (GGT), and Alkaline phosphatase (ALP) assessed photometrically, total Cholesterol, HDL, and LDL cholesterol were examined enzymatically all by using a commercial kit (Parsazmoon, Tehran, Iran).

Histopathology

Five sections from different lobes of each liver were submitted and processed through ethyl alcohol and xylene series, and embedded in paraffine blocks. Slides were stained with Hematoxyline Eosin and Masson's Trichome and viewed under light microscopy by Nikon E 200. The grading was defined as follow: for hepatic steatosis: grade 0, no fat; grade 1, steatosis occupying less than 33% of the hepatic parenchyma; grade 2, 34–66% of the hepatic parenchyma; grade 3, more than 66% of the hepatic parenchyma; for inflammatory cell infiltration: grade 0: none; grade 1, 1–2 foci/field; grade 2, 3–4 foci/field; grade 3, more than 4 foci/field [14]; for ballooning: minimal, mild, and marked [15].

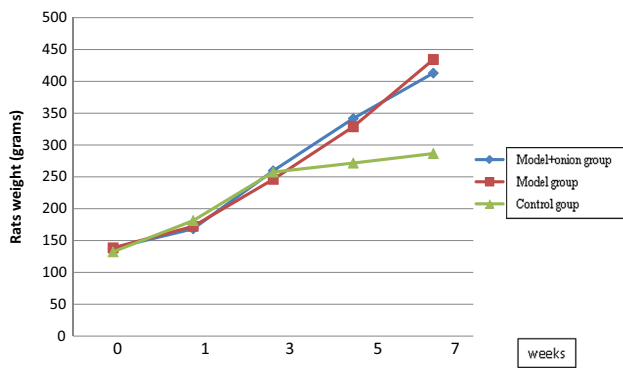


Fig. 1 Mean weight of animals in nonalcoholic fatty liver disease model group, control group, and model +7% onion group (n = 6 in each group) at weeks 1, 3, 5, and 7; from 3rd week, the mean weight of control group was significantly less than two other groups ($p = 0.002$)

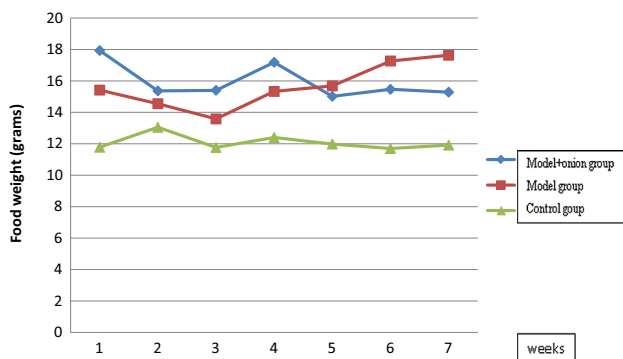


Fig. 2 Mean food intake in animals in nonalcoholic fatty liver disease model group, control group, and model +7% onion group (n = 6 in each group) at the end of each week. At weeks one to seven food intake of control group was significantly less than two other groups ($p < 0.05$)

Statistical Analysis

Results are expressed as median (interquartile range) and using nonparametric tests included Mann–Whitney, Kruskal–Wallis and Chi square tests. $p < 0.05$ was considered for significance level. All statistical analyses were performed with the use of either GraphPad Prism Software Version 5.00 (GraphPad Software, SanDiego, CA), or SPSS 20.0 software (Chicago, IL, USA).

Results

Body Weight, Food Consumption

Total animals weight were measured and compared between three groups at week one, three, five and seven. Animal weights in different groups during the study are shown in Fig. 1. Weight gain differences among the groups were significantly different after week 3 ($p = 0.003$ for week 5, $p = 0.002$ for week 7 and $p = 0.003$ for total weight gain). Weight gain was not significantly different between model group and model + onion group; however, the weight gain was significantly lower in control group in comparison to the other two groups ($p = 0.002$).

Animals’ food intake during the study is shown in Fig. 2. Kruskal–Wallis test showed that food intake was different among three groups in every week ($p = 0.05$); however, this near significant difference was related only to the significant lower food intake in control group ($p < 0.05$), and was not present between the model and model + onion groups.

Table 1 Plasma levels of liver enzymes, glycemic indices, and lipid profiles in study groups

| | Model group [median (IQR)] | Model + onion group [median (IQR)] | Control group [median (IQR)] | <i>p</i> value* |
|---------------------|----------------------------------|------------------------------------|----------------------------------|-----------------|
| ALT (IU/l) | 36 (28.2–65.7) ^b | 26 (20.4–29.2) ^a | 20.5 (15.5–23.7) ^a | 0.01 |
| AST (IU/l) | 45 (42.2–52) ^b | 20 (14.5–30.5) ^a | 21.5 (18.5–24.5) ^a | 0.015 |
| GGT (IU/l) | 12.3 (5.3–18) ^b | 5.7 (4.1–6.9) ^{a,b} | 4 (2.7–7.5) ^a | 0.05 |
| ALP | 895 (498.5–1074) ^b | 980 (827–1255) ^b | 430 (342.5–630) ^a | 0.01 |
| Glucose (mg/dl) | 179.3 (168–191) ^b | 147 (136.2–155.7) ^a | 152.5 (137.7–159.7) ^a | 0.01 |
| Insulin (pmol/l) | 442 (391–505) ^b | 201 (182–229) ^a | 199 (176–231) ^a | 0.01 |
| TG (mg/dl) | 159.5 (143.7–168.7) ^b | 135.5 (106.2–156.5) ^a | 108 (97.7–118.7) ^a | 0.004 |
| Cholesterol (mg/dl) | 145 (120–171.5) ^b | 135.5 (112.5–155.7) ^{a,b} | 104 (96.5–118) ^a | 0.05 |
| HDL-c (mg/dl) | 27.5 (22.2–35.2) ^a | 22 (18.2–35.7) ^a | 30 (26.5–37) ^a | 0.36 |
| LDL-c (mg/dl) | 87.4 (62.2–109) ^b | 80.9 (58.3–101.4) ^b | 39 (31–56.2) ^a | 0.02 |

* Kruskal–Wallis test

^{a,b} In every row different scripts show significant difference

Biochemical Assessments

Model + onion group had significantly lower plasma levels of ALT ($p = 0.026$), AST ($p = 0.041$), Insulin ($p < 0.001$), TG ($p = 0.041$), and glucose ($p = 0.009$) compared with model group; however, total cholesterol and LDL levels were not significantly different between these two groups (Table 1).

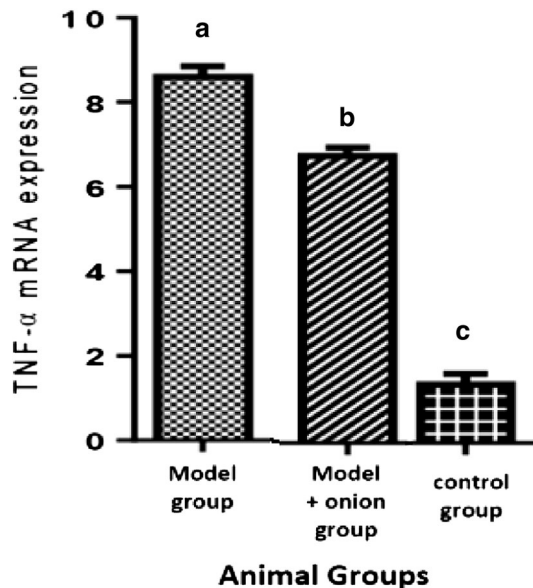


Fig. 3 TNF- α mRNA expression comparison between three groups differences among three groups were statistically significant ($p < 0.05$)

Table 2 Histopathological characteristics of model, control and model + onion groups (n = 6 in each group)

| | Group | Stage | | <i>p</i> value |
|----------------------------|---------------|----------|----------|----------------|
| | | Low | High | |
| Steatosis n (%) | Model | 1 (16.7) | 5 (83.3) | 0.01 |
| | Model + onion | 4 (66.7) | 2 (33.3) | |
| | Control | 6 (100) | 0 | |
| Ballooning n (%) | Model | 1 (16.7) | 5 (83.3) | 0.004 |
| | Model + onion | 2 (33.3) | 4 (66.7) | |
| | Control | 6 (100) | 0 | |
| Lobular inflammation n (%) | Model | 2 (33.3) | 4 (66.7) | 0.05 |
| | Model + onion | 4 (66.7) | 2 (33.3) | |
| | Control | 6 (100) | 0 | |
| Portal inflammation n (%) | Model | 2 (33.3) | 4 (66.7) | 0.05 |
| | Model + onion | 4 (66.7) | 2 (33.3) | |
| | Control | 6 (100) | 0 | |

Low = 0 or 1 stage

High = 2 or 3 stage

Hepatic TNF- α Gene Expression

Hepatic TNF- α gene expression was significantly decreased in model + onion group in comparison to model group (Fig. 3).

Histological Analysis

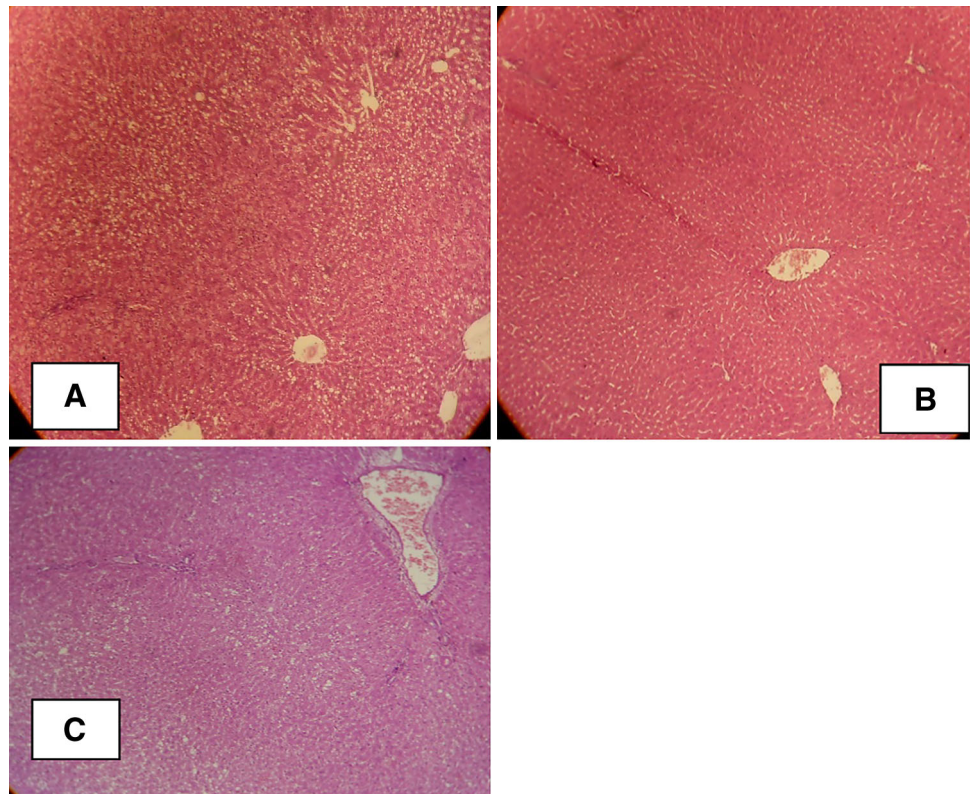
Hepatic histopathological features of the three groups are shown in Table 2, and Fig. 4. Model + onion group had significantly lower hepatic steatosis, ballooning, lobular inflammation, and portal inflammation in comparison to the model group.

Discussion

To our knowledge, this is the first study that has evaluated the effects of onion powder consumption on the prevention of NAFLD. Since it is not convenient to evaluate the effects of dietary intakes in prevention of NAFLD in human, we designed this study in the experimental model of the disease. Our results have shown that regular consumption of onion powder can prevent development of NAFLD even at the presence of other risk factors such as obesity and high energy, fat, and sugar intakes.

The beneficial effects of onion consumption on the NAFLD risk factors such as hyperlipidemia and oxidative stress [16, 17], hyperglycemia [18, 19], and inflammation [20] have been shown previously. These effects are attributed to the high content of flavonoids, and prebiotics in the onion. Two flavonoid subgroups are found in onion,

Fig. 4 Hepatic pathology in rats from control group $\times 100$ (a), model group $\times 100$ (b), and model + onion group $\times 100$ (c). The liver samples were stained with haematoxylin and eosin



the anthocyanins, and flavanols such as quercetin and its derivatives [11, 21].

A recent study has shown that oral administration of quercetin to hyperlipidemia rats is highly effective in decreasing the levels of serum lipids, hepatic enzymes, steatosis, and inflammation through regulating the expressions of Sirt1, NF- κ B p65 and iNOS [22]. Another recent study has reported that quercetin significantly decrease hepatic damage enzymes, lipoperoxidation, DNA damage and macrovesicular steatosis, ballooning and inflammatory process in an experimental model of NASH [23]. Furthermore, Marcolin et al. [24] indicated that oral administration of quercetin attenuates steatohepatitis in an experimental model of NASH via down regulation of profibrotic and proinflammatory gene pathways.

Moreover, the beneficial effects of prebiotics in modulation of gut microbiota, and attenuation of NAFLD have been shown recently [7, 25]. Prebiotics can increase the number of prebiotics in the gut lumen, which results in reduction in inflammation through reduction in pathogenic bacteria, enhancement of intestinal permeability, and immune regulation [7].

This study has several advantages; we used a diet induced model of NAFLD, which is similar to the disease development in the human. We evaluated the effects of onion consumption as a food, which is available in every season with a cheap price, and can be used in every population at risk of NAFLD development. Finally, we have

shown these effects in the prevention of disease, and the results have shown that the effects will remain even in the presence of other risk factors for NAFLD development.

In conclusion, our results indicate that regular consumption of onion can prevent from development of NAFLD at least partially through improvement in serum glycemic indices, and TG, and down regulation of hepatic TNF- α gene expression, which plays a pivotal role in both inflammation and insulin resistance. These beneficial effects were observed even in the presence of main risk factors of NAFLD development such as obesity, hypercholesterolemia, and high energy, fat, and sugar intakes.

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Compliance with Ethical Standards

Conflict of interest All authors declare that they have no conflict of interest.

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