

SHORT COMMUNICATION

Soil protist communities form a dynamic hub in the soil microbiome

Wu Xiong^{1,2,3}, Alexandre Jousset^{1,2}, Sai Guo¹, Ida Karlsson^{2,4}, Qingyun Zhao³, Huasong Wu³, George A Kowalchuk², Qirong Shen¹, Rong Li¹ and Stefan Geisen^{5,6}

¹Jiangsu Provincial Key Lab for Organic Solid Waste Utilization, National Engineering Research Center for Organic-based Fertilizers, Jiangsu Collaborative Innovation Center for Solid Organic Waste Resource Utilization, Nanjing Agricultural University, Nanjing, China; ²Ecology and Biodiversity Group, Department of Biology, Institute of Environmental Biology, Utrecht University, Utrecht, Netherlands; ³Spice and Beverage Research Institute, Chinese Academy of Tropical Agricultural Science, Wanning, Hainan, China; ⁴Department of Forest Mycology and Plant Pathology, Swedish University of Agricultural Sciences, Uppsala, Sweden; ⁵Department of Terrestrial Ecology, Netherlands Institute for Ecology (NIOO-KNAW), Wageningen, Netherlands and ⁶Laboratory of Nematology, Wageningen University, Wageningen, Netherlands

Soil microbes are essential for soil fertility. However, most studies focus on bacterial and/or fungal communities, while the top-down drivers of this microbiome composition, protists, remain poorly understood. Here, we investigated how soil amendments affect protist communities and inferred potential interactions with bacteria and fungi. Specific fertilization treatments impacted both the structure and function of protist communities. Organic fertilizer amendment strongly reduced the relative abundance of plant pathogenic protists and increased bacterivorous and omnivorous protists. The addition of individual biocontrol bacteria and fungi further altered the soil protist community composition, and eventually function. Network analysis integrating protist, bacterial and fungal community data, placed protists as a central hub in the soil microbiome, linking diverse bacterial and fungal populations. Given their dynamic response to soil management practices and key position in linking soil microbial networks, protists may provide the leverage between soil management and the enhancement of bacterial and fungal microbiota at the service of improved soil health.

The ISME Journal (2018) 12, 634–638; doi:10.1038/ismej.2017.171; published online 13 October 2017

Protists are an often overlooked component of the soil microbiome. They are abundant and extremely diverse in soil, where they carry out a range of functions (Foissner, 1997; Geisen, 2016a). Protists are among the main consumers of soil bacteria and fungi, but also algae and nematodes (Geisen, 2016a, b; Seppey *et al.*, 2017). Protist activity directly increases plant performance by enhancing the microbial loop (Bonkowski, 2004) and stimulating plant growth-promoting rhizobacteria (Rosenberg *et al.*, 2009; Jousset, 2012). Further, soil protist communities encompass a range of plant- and animal-pathogenic species (Geisen *et al.*, 2015b). Given their functional diversity, protists exert control over various soil organisms and are likely of

critical importance for soil fertility. However, we lack an understanding of how protist communities are structured, how targeted soil management can alter protist communities, how such changes might affect protist functioning, and how they are in turn linked to their potential bacterial and fungal prey. Here, we experimentally examined under controlled greenhouse conditions if applications of organic versus conventional fertilizers could modify the taxonomic and functional composition of protists, as well as their putative interactions within the soil microbiome. Furthermore, we added either a bacterial (*Bacillus amyloliquefaciens*) or fungal (*Trichoderma guizhouense*) plant-protective agent (Wang *et al.*, 2013; Zhang *et al.*, 2016) to organic fertilizer (subsequently termed OF+B and OF+T, respectively) to study if these biocontrol agents also impact protist communities (Xiong *et al.*, 2017 and Supplementary Information for additional details).

Protist communities differed significantly between all treatments one-year post application. Organic fertilizer-treated soil contained a fundamentally different protist community structure compared

Correspondence: R Li, Jiangsu Provincial Key Lab for Organic Solid Waste Utilization, Nanjing Agricultural University, No. 1 Weigang, Nanjing 210095, China. E-mail: lirong@njau.edu.cn or S Geisen, Department of Terrestrial Ecology, Netherlands Institute for Ecology (NIOO-KNAW), Droevendaalsesteeg 10, 6708 PB Wageningen, Netherlands. E-mail: S.Geisen@nioo.knaw.nl
Received 13 April 2017; revised 28 August 2017; accepted 5 September 2017; published online 13 October 2017

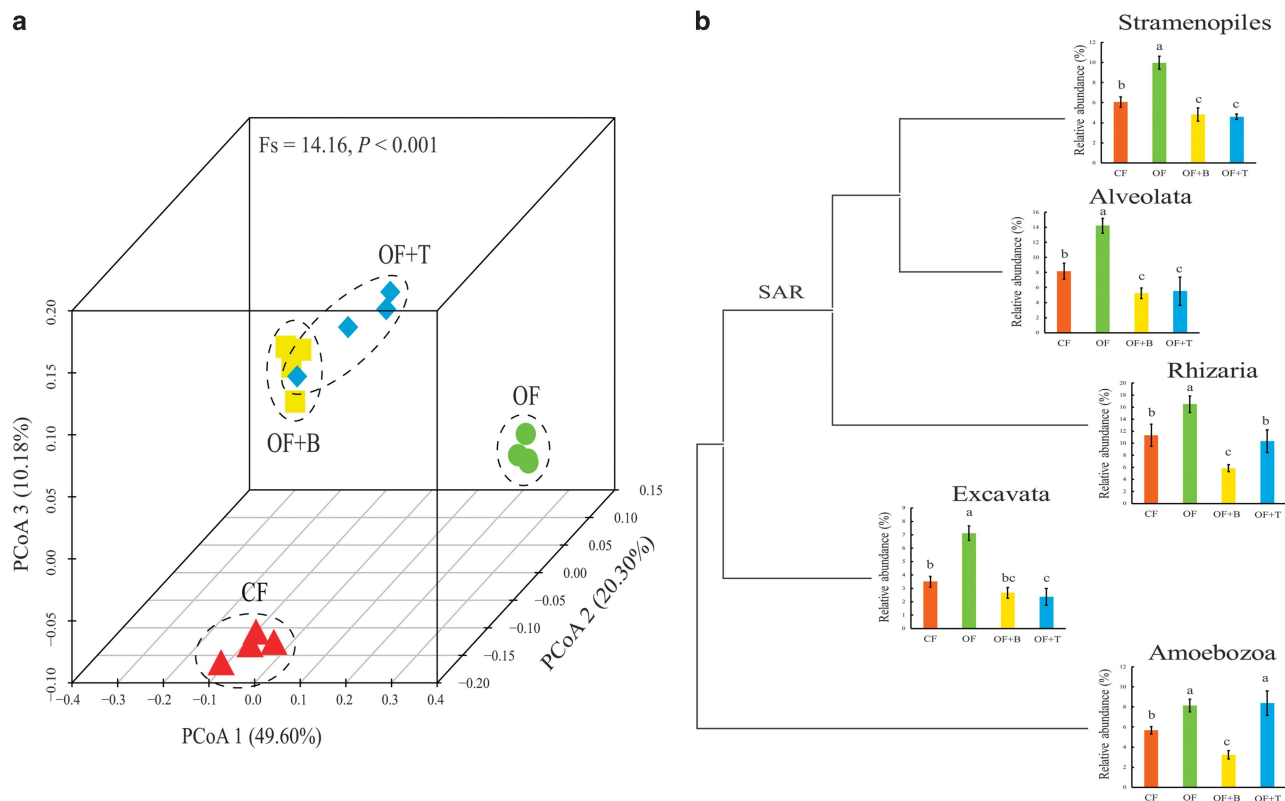


Figure 1 Protist community changes induced by fertilizations. **(a)** UniFrac-weighted principle coordinate analysis (PCoA) of soil protist community in the four fertilizer treatments and **(b)** relative abundance of five main protist taxonomic groups (Stramenopiles, Alveolata, Rhizaria, Excavata and Amoebozoa) with coarse phylogenetic affinities in the four fertilizer treatments. CF, chemical fertilizer; OF, organic matter fertilizer; OF+B, *Bacillus* enriched organic fertilizer; OF+T, *Trichoderma* enriched organic fertilizer. Different letters above the bars indicate a significant difference at the 0.05 probability level according to the Tukey's test.

with the chemical fertilizer (CF) treatment (UniFrac-weighted and -unweighted PCoA: Figure 1a; Supplementary Figure 2; RDA: Supplementary Figure 6), which is in line with former studies (Heger *et al.*, 2012; Murase *et al.*, 2015). Organic fertilizer (OF) amendment enhanced the relative abundance of the most abundant protist taxonomic groups, that is, Stramenopiles, Alveolata, Rhizaria, Excavata and Amoebozoa (Figure 1b), most of which are predators of other microbes. This can be explained by the fact that organic fertilizer provides a wider resource spectrum than chemical fertilizer, which thereby promotes a higher biomass and diversity of bacteria and fungi (Xiong *et al.*, 2017). This impact on primary consumers may foster diverse and active protist consumers, as previously observed in studies of paddy rice (Murase *et al.*, 2015). Organic fertilizer enriched with beneficial microbes (*Bacillus* and *Trichoderma*) caused a further shift in the protist community composition by reducing Stramenopiles, Alveolata and Excavata, and most strongly Rhizaria and Amoebozoa in the OF+B treatment (Figure 1b). As a result, protist richness (observed OTUs), diversity (phylogenetic and Shannon) and evenness (Shannon) were lower in the OF+B and OF+T treatments as compared with the OF treatment (Supplementary Figure 1). These results suggest that the added microbes differentially

affected specific protist taxonomic groups (Jousset *et al.*, 2006; Pedersen *et al.*, 2011), possibly by producing inhibitory compounds. *Bacillus* species are known to produce a broad range of secondary metabolites such as cyclic lipopeptide antibiotics (Alvarez *et al.*, 2012) and various volatile compounds that can deter or even kill potential protist predators (Mazzola *et al.*, 2009; Schulz-Bohm *et al.*, 2017). *Trichoderma* may also have affected protists via the production of antimicrobial compounds (Cai *et al.*, 2013) or by reducing fungal prey, but information on protist-fungal interactions is too scarce for reliably interpreting these data (Geisen, 2016a). Introduced microbes may also exert indirect effects; the lower diversity of protists and lower relative abundances of Stramenopiles, Alveolata and Excavata in the two microbe-enriched organic fertilizer treatments may be linked to the bacterial genus *Lysobacter*, which increased ~6-fold in OF+B and OF+T treatments compared with the CF treatment (Xiong *et al.*, 2017). Members of this genus are known to produce a broad range of bioactive secondary metabolites that can inhibit soil organisms (Expósito *et al.*, 2015).

Changes in protist taxonomic community compositions induced by fertilizer regime subsequently resulted in shifts in the relative proportion of protist functional groups (Figure 2). Several potentially

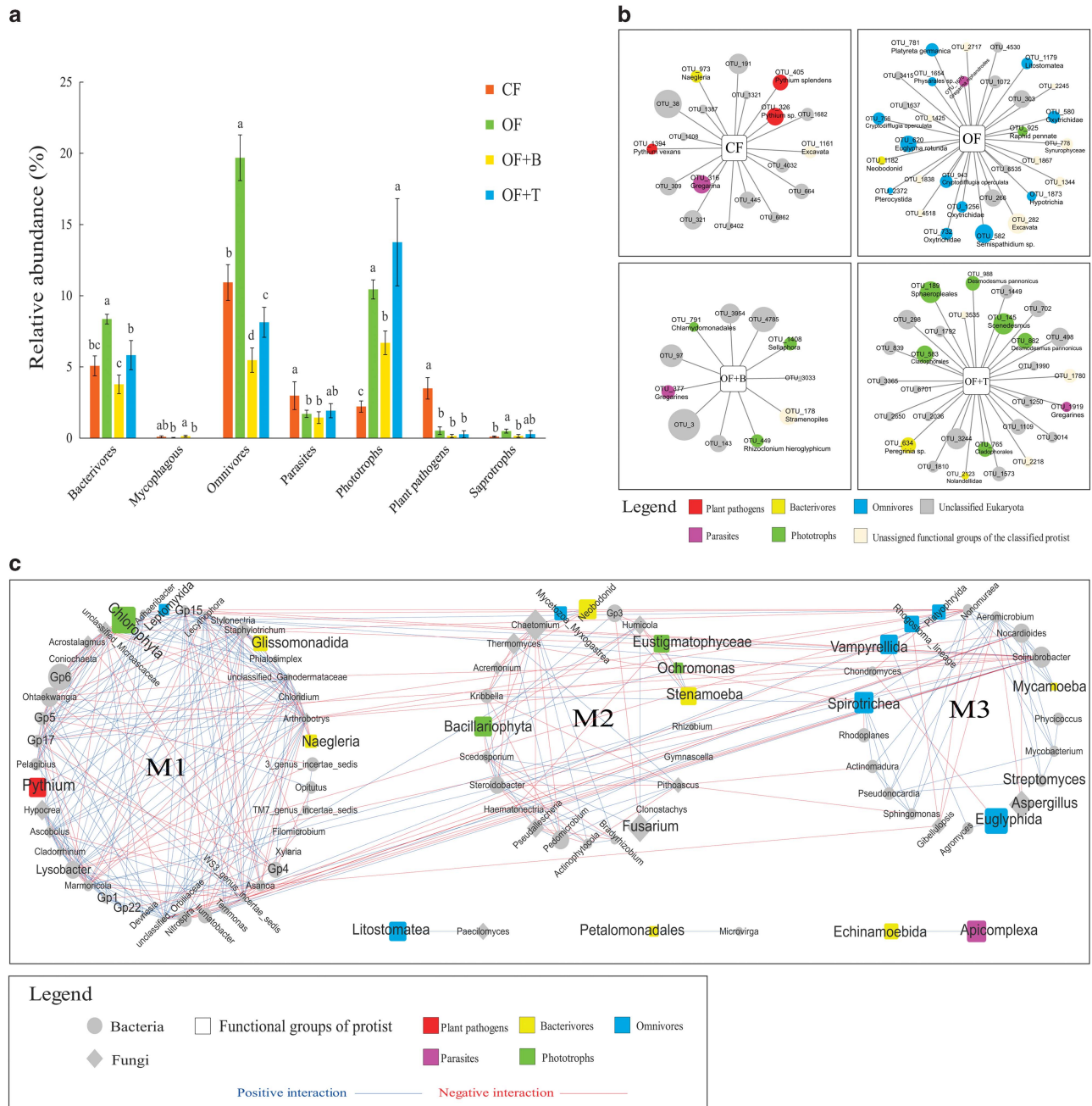


Figure 2 Protists form a functional and dynamic hub in the soil microbiome induced by fertilizations. (a) Overview of the relative abundance of protist functional groups in the four fertilizer treatments. CF, chemical fertilizer; OF, organic matter fertilizer; OF+B, *Bacillus* enriched organic fertilizer; OF+T, *Trichoderma* enriched organic fertilizer. Different letters above the bars indicate a significant difference at the 0.05 probability level according to the Tukey's test. (b) Protist indicator OTUs for the four fertilizer treatments. Circles represent protist OTUs and circle sizes correspond to their average relative abundance (log transformation) across all the samples. (c) Correlation-based network analysis showing potential interactions between abundant bacterial and fungal genera as well as protist functional groups. The node size is proportional to a taxon's average relative abundance (log transformation) across all the samples. Lines connecting nodes (edges) represent positive (blue) or negative (red) co-occurrence relationships.

plant pathogenic *Pythium* spp., a group of widely distributed pathogens of thousands of plant species (Schroeder *et al.*, 2012), were indicative for CF treatment (Figure 2b). In line, putative protist plant pathogens, including *Pythium* spp., were significantly reduced in all three organic fertilizer treatments (OF, OF+B and OF+T) compared with CF treatment (Figure 2a; Supplementary Table 2). This

may be due either to direct inhibition by the introduced beneficial microbes, the stimulation of antagonistic microbes, or simply the promotion of a range of protists that consume or outcompete *Pythium* in organic matter. Apicomplexa, known as obligate parasites of vertebrates and invertebrate hosts (Kopečná *et al.*, 2006), were lower in all organic fertilizer treatments as compared with the

CF treatment (Supplementary Table 2). These results suggest a relatively 'pathogen and parasite' driven food-web in chemical fertilizer-treated soils. In contrast, soils treated with OF were not only higher in saprotrophs, but also in phototrophic algae (Figure 2a), indicative of less disturbed soils and increased soil carbon inputs (Zancan *et al.*, 2006). The indicator taxa for OF were mainly omnivores consuming other eukaryotes (Figure 2b), suggesting a more complex food-web. The addition of *Bacillus* into OF, but not of *Trichoderma*, significantly reduced the saprotrophic and phototrophic protists. More targeted studies would be necessary to examine the functional changes in the protist communities after applying different biocontrol agents and their long-term impact on soil functioning. It has to be mentioned that we used a conservative approach to assign the classified protists into different functions, focusing merely on feeding mode. As any inference tool, our database approach should be interpreted with caution. It is necessarily reliant on the limited number of characterized reference species (Berney *et al.*, 2017), and may therefore be biased toward specific functions as some elements of functional variation. Current efforts to expand and improve this taxonomic and functional database (Berney *et al.*, 2017) will further improve this tool, and we anticipate that this approach will become more useful in the future to advance studies that seek to more meaningfully integrate protists in soil microbiome research.

Finally, we examined the role of protists as an integral part of soil ecological networks. By constructing a combined co-occurrence network encompassing bacteria, fungi and functional groups of protists, we found that protists form distinct hubs in the soil network, linking a range of bacterial and fungal taxa (Supplementary Information; Figure 2c). Three main modules emerged in our constructed network, with protists present in all modules. Module 1 was dominated by bacteria such as *Lysobacter* and several Acidobacteria groups and contained five protist nodes from distinct taxonomic and functional groups (the mainly plant pathogenic *Pythium*, the bacterivorous Glissomonadida and *Naegleria*). Module 2 was phototroph-dominated (such as Bacillariophyta and Eustigmatophyceae). Module 3 was dominated by omnivorous protists (such as Spirotrichea, Vampyrellida and Euglyphida). Each module appeared to be generally associated with a specific range of functions (Zhou *et al.*, 2011), suggesting interactions between similarly functioning microbes that provide either a stimulatory or inhibitory loop for soil functioning. Parasitic protist taxa, including the potential plant pathogenic *Pythium* and animal parasitic Apicomplexa, were present, but poorly connected to other microorganisms in the network. This disconnected position is likely related to the dependence of these organisms on plants (Xu *et al.*, 2012) and soil animals (Geisen *et al.*, 2015a) rather than other soil microbes.

Our study highlights the multi-trophic nature of the soil microbiome. This study is one of the first to link taxonomically assigned protist taxa to functional groups that are embedded within soil food webs. Soil amendments strongly impacted protist communities 1-year after application, with addition of organic material and beneficial microbes leading to profound changes of protist community composition, and eventually function. This study also serves as a plea to the scientific community to better integrate protists into microbiome studies. Given their large impacts on multiple soil functions, we propose that manipulation of soil protist communities offers new avenues to promote soil health, plant performance and other ecosystem services.

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgements

We thank Steven Lindow and four anonymous reviewers for their constructive comments on the manuscript. This research was supported by the National Key Basic Research Program of China (2015CB150506), the National Natural Science Foundation of China (31572212, 31672242 and 31501824), the Royal Netherlands Academy of Arts and Sciences (KNAW) joint network grant, the Natural Science Foundation of Jiangsu (BK20150059), National Key Scientific Research Project (2016YFD0800605, 2016YFD0200106 and 2016YFE0101100), the Priority Academic Program Development of the Jiangsu Higher Education Institutions (PAPD), and Qing Lan Project. Stefan Geisen was supported by the ERC advanced grant SPECIALS (ERC-Adv 260-55290) awarded to Wim H. van der Putten. Alexandre Jousset was supported by the Netherlands Organisation for Scientific Research (NWO) (ALW.870.15.050) and the Royal Netherlands Academy of Arts and Sciences (KNAW) (530-5CDP18). Wu Xiong was supported by the Sino-Dutch Bilateral Exchange Scholarship.

References

- Alvarez F, Castro M, Príncipe A, Borioli G, Fischer S, Mori G *et al.* (2012). The plant-associated *Bacillus amyloliquefaciens* strains MEP218 and ARP23 capable of producing the cyclic lipopeptides iturin or surfactin and fengycin are effective in biocontrol of sclerotinia stem rot disease. *J Appl Microbiol* **112**: 159–174.
- Berney C, Ciuprina A, Bender S, Brodie J, Edgcomb V, Kim E *et al.* (2017). UniEuk: time to speak a common language in protistology!. *J Eukaryot Microbiol* **64**: 407–411.
- Bonkowski M. (2004). Protozoa and plant growth: the microbial loop in soil revisited. *New Phytol* **162**: 617–631.
- Cai F, Yu G, Wang P, Wei Z, Fu L, Shen Q *et al.* (2013). Harzianolide, a novel plant growth regulator and

- systemic resistance elicitor from *Trichoderma harzianum*. *Plant Physiol Biochem* **73**: 106–113.
- Expósito RG, Postma J, Raaijmakers JM, De Bruijn I. (2015). Diversity and activity of *Lysobacter* species from disease suppressive soils. *Front Microbiol* **6**: 1243.
- Foissner W. (1997). Protozoa as bioindicators in agroecosystems, with emphasis on farming practices, biocides, and biodiversity. *Agric Ecosyst Environ* **62**: 93–103.
- Geisen S. (2016a). The bacterial-fungal energy channel concept challenged by enormous functional versatility of soil protists. *Soil Biol Biochem* **102**: 22–25.
- Geisen S. (2016b). Thorough high-throughput sequencing analyses unravels huge diversities of soil parasitic protists. *Environ Microbiol* **18**: 1669–1672.
- Geisen S, Laros I, Vizcaíno A, Bonkowski M, de Groot GA. (2015a). Not all are free-living: high-throughput DNA metabarcoding reveals a diverse community of protists parasitizing soil metazoa. *Mol Ecol* **24**: 4556–4569.
- Geisen S, Tveit AT, Clark IM, Richter A, Svenning MM, Bonkowski M et al. (2015b). Metatranscriptomic census of active protists in soils. *ISME J* **9**: 2178–2190.
- Heger TJ, Straub F, Mitchell EAD. (2012). Impact of farming practices on soil diatoms and testate amoebae: a pilot study in the DOK-trial at Therwil, Switzerland. *Eur J Soil Biol* **49**: 31–36.
- Jousset A. (2012). Ecological and evolutive implications of bacterial defences against predators. *Environ Microbiol* **14**: 1830–1843.
- Jousset A, Lara E, Wall LG, Valverde C. (2006). Secondary metabolites help biocontrol strain *Pseudomonas fluorescens* CHA0 to escape protozoan grazing. *Appl Environ Microbiol* **72**: 7083–7090.
- Kopečná J, Jirků M, Oborník M, Tokarev YS, Lukeš J, Modrý D. (2006). Phylogenetic analysis of coccidian parasites from invertebrates: search for missing links. *Protist* **157**: 173–183.
- Mazzola M, de Bruijn I, Cohen MF, Raaijmakers JM. (2009). Protozoan-induced regulation of cyclic lipopeptide biosynthesis is an effective predation defense mechanism for *Pseudomonas fluorescens*. *Appl Environ Microbiol* **75**: 6804–6811.
- Murase J, Hida A, Ogawa K, Nonoyama T, Yoshikawa N, Imai K. (2015). Impact of long-term fertilizer treatment on the microeukaryotic community structure of a rice field soil. *Soil Biol Biochem* **80**: 237–243.
- Pedersen AL, Winding A, Altenburger A, Ekelund F. (2011). Protozoan growth rates on secondary-metabolite-producing *Pseudomonas* spp. correlate with high-level protozoan taxonomy. *FEMS Microbiol Lett* **316**: 16–22.
- Rosenberg K, Bertaux J, Krome K, Hartmann A, Scheu S, Bonkowski M. (2009). Soil amoebae rapidly change bacterial community composition in the rhizosphere of *Arabidopsis thaliana*. *ISME J* **3**: 675–684.
- Schroeder KL, Martin FN, de Cock AWAM, Lévesque CA, Spies CFJ, Okubara PA et al. (2012). Molecular detection and quantification of pythium species: evolving taxonomy, new tools, and challenges. *Plant Dis* **97**: 4–20.
- Schulz-Bohm K, Geisen S, Wubs ERJ, Song C, de Boer W, Garbeva P. (2017). The prey's scent – volatile organic compound mediated interactions between soil bacteria and their protist predators. *ISME J* **11**: 817–820.
- Sepey CVW, Singer D, Dumack K, Fournier B, Belbahri L, Mitchell EAD et al. (2017). Distribution patterns of soil microbial eukaryotes suggests widespread algivory by phagotrophic protists as an alternative pathway for nutrient cycling. *Soil Biol Biochem* **112**: 68–76.
- Wang B, Yuan J, Zhang J, Shen Z, Zhang M, Li R et al. (2013). Effects of novel bioorganic fertilizer produced by *Bacillus amyloliquefaciens* W19 on antagonism of Fusarium wilt of banana. *Biol Fertil Soils* **49**: 435–446.
- Xiong W, Guo S, Jousset A, Zhao Q, Wu H, Li R et al. (2017). Bio-fertilizer application induces soil suppressiveness against Fusarium wilt disease by reshaping the soil microbiome. *Soil Biol Biochem* **114**: 238–247.
- Xu L, Ravnskov S, Larsen J, Nilsson RH, Nicolaisen M. (2012). Soil fungal community structure along a soil health gradient in pea fields examined using deep amplicon sequencing. *Soil Biol Biochem* **46**: 26–32.
- Zancan S, Trevisan R, Paoletti MG. (2006). Soil algae composition under different agro-ecosystems in North-Eastern Italy. *Agric Ecosyst Environ* **112**: 1–12.
- Zhang J, Bayram Akcapinar G, Atanasova L, Rahimi MJ, Przylucka A, Yang D et al. (2016). The neutral metalloproteinase NMP1 of *Trichoderma guizhouense* is required for mycotrophy and self-defence. *Environ Microbiol* **18**: 580–597.
- Zhou J, Deng Y, Luo F, He Z, Yang Y. (2011). Phylogenetic molecular ecological network of soil microbial communities in response to elevated CO₂. *mBio* **2**: e00122–11.

Supplementary Information accompanies this paper on The ISME Journal website (<http://www.nature.com/ismej>)