TRANSLATIONAL RESEARCH

Parenteral bilirubin in healthy volunteers: a reintroduction in translational research

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AIMS

Preclinical results suggest therapeutic potential of mild hyperbilirubinemia in T2DM and cardiovascular disease. Translational data are limited, because an appropriate bilirubin formulation for parenteral human use is lacking. Considering its use in both clinical practice and medical research in the past, we explored the feasibility to reintroduce parenteral bilirubin for translational experiments.

METHODS

We developed a preparation method in accordance with good manufacturing practice and evaluated the parenteral applicability in healthy volunteers (n = 8). Explorative pharmacokinetic and safety data were compared to the results from a literature study on the former parenteral use of bilirubin. Bilirubin was administered intra-arterially to raise the local plasma concentration in the forearm vascular bed (n = 4) and intravenously to raise the systemic plasma concentration (n = 4). Finally, pharmacokinetic characteristics were studied following a single bolus infusion (n = 3).

RESULTS

During parenteral application, no side effects occurred. Adverse events mentioned during the two-week observation period were in general mild and self-limiting. Three more significant adverse events (appendicitis, asymptomatic cardiac arrhythmia and atopic eczema) were judged unrelated by independent physicians.

A dose-concentration relationship appeared sufficiently predictable for both intra-arterial and intravenous administration. In line with existing knowledge, bilirubin pharmacokinetics could be described best according to a two-compartment model with a volume of distribution of 9.9 (\pm 2.0) l and a total plasma clearance of 36 (\pm 16) ml per minute.

CONCLUSIONS

Supported by previous reports, our data suggest that it is both feasible and safe to perform translational experiments with parenteral albumin bound bilirubin.



WHAT IS ALREADY KNOWN ABOUT THIS SUBJECT

- Multiple preclinical experiments demonstrate beneficial effects of bilirubin administration for both type 2 diabetes mellitus and cardiovascular disease.
- Parenteral administration of bilirubin used to be part of daily clinical practice and medical research in the past.
- Reintroduction of bilirubin for parenteral human use is needed for the translation of promising preclinical findings.

WHAT THIS STUDY ADDS

- If produced under Good Manufacturing Practice (GMP) guidelines and applied in an albumin-bound formulation, parenterally administered bilirubin appears safe up to plasma concentrations of 86 μmol l⁻¹.
- Exploratory pharmacokinetic experiments demonstrate sufficiently predictable dose-concentration curves.

Introduction

Though commonly seen as a harmful residual product from the breakdown of haeme, bilirubin was already demonstrated to possess antioxidant capacities in 1954 [1]. Ever since, a wide variety of *in vitro* and animal studies have been published on the protecting antioxidant, anti-inflammatory and antifibrotic effects of both bilirubin and haeme oxygenase (HO), the rate-limiting enzyme in bilirubin formation [2, 3]. Numerous observational studies have demonstrated an inverse association between bilirubin concentrations and cardiovascular disease in humans [4]. Therefore, both the induction of a mild hyperbilirubinaemia and the enhancement of HO activity have been suggested for the treatment of a variety of diseases, including type 2 diabetes mellitus (T2DM) and related cardiovascular disorders [2, 5, 6]. In the case of cardiovascular disease, translational research is enabled by several well-established measures of endothelium function as surrogate endpoints [7]. Using the perfused forearm technique, we previously succeeded to improve endothelial function and plasma antioxidant capacity in subjects with T2DM by a short-term treatment with the plasma bilirubin-increasing drug atazanavir [8]. Additional experiments are needed to invigorate these promising findings in the atazanavir model.

Though opportune for translational and clinical studies that require a steady elevation of bilirubin plasma levels, in proof of concept studies the use of atazanavir hampers the attribution of observed effects to bilirubin as atazanavir itself might influence the outcome too. In addition, the atazanavir model does not enable the dose–response relationship of bilirubin to be investigated. The availability of a bilirubin formulation that is suitable for parenteral human use would significantly facilitate the design of such experiments.

In the past, intravenous administration of bilirubin was commonly applied in human research and clinical practice (see Table 1). After 1981, however, no new data on the experimental use of bilirubin in humans was published. Subsequent to an extensive literature study on the former preparation and parenteral use of bilirubin, we developed a production method meeting current standards of good manufacturing practice (GMP). In addition to the intravenous use of the preparation, we also studied its applicability for intra-arterial administration in the perfused forearm model. Here, we present the method of preparation and the first results on its intravenous and intra-arterial application in healthy volunteers, including data on its pharmacokinetics and safety. For their exploratory nature, these data are supported by the context of extensive pharmacokinetic and safety data from former reports on the parenteral human use of bilirubin.

Methods

Preparation

Bilirubin concentrate for infusion (6 mg ml $^{-1}$, 10 ml vials) for parenteral use in the clinical trial was produced compliant with GMP at the Department of Pharmacy of the Radboud University Medical Center, Nijmegen, The Netherlands. All chemicals were commercially available and used as obtained. All materials were approved on the basis of in-house quality controls and their suppliers were qualified. The aseptic production process was validated by means of media fills. The drug substance was acquired from Sigma Aldrich (Zwijndrecht, The Netherlands) as bilirubin $\geq 98\%$ powder. The drug substance was tested for identity, purity, content with spectrophotometric and chemical analysis, and endotoxins with the Limulus Amoebocyte Lysate (LAL) test. The drug product was prepared by dissolving 6 g of bilirubin and 4 g of sodium hydroxide (Spruyt Hillen, IJsselstein, The Netherlands) in 1 l of water for injections. After dissolving, the solution was purged with nitrogen, filtered through a Sartopore 2 XLI (0.35 + 0.2 µm) sterile filter (Sartorius AG, Goettingen, Germany) and, subsequently, sterile vials were filled with 10.2 ml of liquid under a continuous flow of nitrogen. This was carried out at the Department of Pharmacv at the Erasmus Medical Center in Rotterdam, The Netherlands. Quality control and release of the product was performed at the Department of Clinical Pharmacy, Radboud University Medical Center. The drug product was tested for the microbiological aspects bioburden, sterility and endotoxins. Furthermore, labelling, volume, pH, identity and content as well particulate contamination were assessed compliant with European Pharmacopoeia requirements for parenteral preparations. The identity and content was assessed by means of a validated high performance liquid chromatography assay with ultraviolet detection (HPLC-UV). Prospective stability studies showed a stability of the product for 12 months at -20°C. The final formulation consisted of bilirubin at 6 mg ml^{-1} in 0.1 M sodium hydroxide with a pH >10. Shortly before administration, the product was passed through a $0.2 \ \mu m$ filter and diluted with human albumin 200 mg ml⁻¹ intravenous solution (Albuman[®], Sanquin Plasma Products,



Table 1

Overview of publications on parenteral human use of bilirubin

	Author(s)	Year	Number of subjects	Number of infusions	Dose	Bilirubin level	AE
1	Von Bergmann [9, 32]	1927	108	120	70 mg	31 (24–38)	Y ^b
2	Boshamer [42]	1928	20	> 46	50 mg	NR	NR
3	Harrop and Barron [43]	1931	27	27	1 mg kg^{-1}	NR	NR
4	Stroebe [44]	1932	16	16	50 mg	NR	NR
5	Zadek [45]	1932	10	10	50 mg	NR	NR
6	Hofbauer [46]	1933	20	20	1 mg kg^{-1}	NR	NR
7	Dietel [47, 48]	1933	45	45	50 mg	NR	NR
8	Binswanger [49]	1933	111	111	50 mg	NR	NR
9	Fuge [50]	1933	44	54	50 mg	31 (12–63)	NR
10	Soffer [35]	1933	21	31	1 mg kg^{-1}	NR	Ν
11	Nürnberger [51]	1933	25	25	50 mg	15 (9–22)	NR
12	Schindel [52]	1934	NR	50	50 mg	NR (12–44)	NR
13	Sullivan [36]	1934	101	147	1 mg kg^{-1}	NR	Ν
14	Bröchner [53]	1935	65	68	50 mg	27 (11–48)	NR
15	Bröchner [54]	1935	50	52	50 mg	31 (17–45)	NR ^b
16	Ruhbaum [55]	1935	44	44	50 mg	NR	NR
17	Fulde [56]	1935	35	36	50 mg	NR	NR
18	Lin [57]	1937	26	26	10 mg 3 mg kg ⁻¹	NR	NR
19	Thompson [33]	1938	10	85	5–20 mg kg $^{-1}$	250 (38–607)	Y ^b
20	Lyon [58]	1938	20	20	1 mg kg^{-1}	NR	Y ^b
21	Rawls [59]	1938	NR	NR	<3 mg kg ⁻¹	NR	NR
22	Hench [34]	1938	13	>78	1–4 g	NR (342–599)	Y ^b
23	Weech [37]	1941	35	35	5 mg kg^{-1}	NR (103–151) ^a	Ν
24	Kornberg [38]	1942	65	65	1.5 mg kg^{-1}	63 (37–123)	Ν
25	With [25]	1943	63	56	50 mg	65 (22–380)	NR
				1	75 mg		
				4	150 mg		
26	Arpe [60]	1946	19	19	NR	NR	NR
27	Tisdale [61]	1959	11	11	15 mg kg^{-1}	268 (96–323)	NR
28	Billing [26]	1964	38	38	2 mg kg^{-1}	NR (51–68) ^a	NR
29	Schmidt [62]	1966	8	8	4–6 mg	46 (19–82)	NR
30	Raymond [63]	1966	10	10	NR	NR	NR ^b
31	Nixon [64]	1967	1	1	2 mg kg^{-1}	84	NR
32	Engstedt [65]	1967	21	21	50 mg	NR	NR
33	Barrett [31]	1968	29	29	<0.4 mg	NR	NR
				8	2 mg kg^{-1}	NR	
34	Berk [11, 24]	1969–1970	53	53	<0.4 mg	NR	NR
35	Okolicsanyi [27]	1978	119	119	2 mg kg ⁻¹	NR	Yb
				14	540 mg	INK (00-104)	

(continues)



Table 1

(Continued)

	Author(s)	Year	Number of subjects	Number of infusions	Dose	Bilirubin level	AE			
36	Goresky [28]	1978	11	11 8 5	2 mg kg ⁻¹ 6 mg kg ⁻¹ 12 mg kg ⁻¹	NR (75–91) ^a NR (221–275) ^a NR (426–559) ^a	NR			
37	Okolicsanyi [29]	1981	29	36	2 mg kg^{-1}	NR	NR			
38	Zenerolli [30]	1982	100	100	2 mg kg^{-1}	NR	NR			
	Total		1423	>1770						
Additi	Additional information on adverse events									
1	Fever and chills, 30–45 min after injection, avoidable by use of freshly prepared bilirubin									
15	10 patients died of seve	10 patients died of severe infections								
19	Co-administration of bile salts; 15 reactions in six patients; Local reactions after repetitive infusions; Systemic reactions both immediate (flushing, tachycardia and headache) and delayed (nausea, vomiting, chills and fever 1 h after administration)									
20	Two deaths (days 6 and 32 after administration), likely related to a complicated pregnancy									
22	Venous thrombosis; Stro	Venous thrombosis; Strong alkaline mixture; 6–30 infusions per subject								
30	Authors state that 'The	Authors state that 'The procedure is safe'								
35	Well tolerated except for mild soreness in the arm after high dose in a few instances									

Bilirubin plasma levels are expressed in μ mol l⁻¹ and depicted as average (range). AE, adverse event; N, no; NR, not reported; Y, yes. ^aEstimated from incompletely published data.

^bAdditional information on adverse events.

Amsterdam, The Netherlands) to a concentration of 1.2 mg ml⁻¹ for intravenous infusion (pH 8.1) and to a concentration of 0.6 mg ml⁻¹ for intra-arterial infusion (pH 7.6). Based on the molecular weight of bilirubin (584.66 g mol⁻¹) and albumin (66.5 kg mol⁻¹), the molarity for bilirubin and albumin was 2.05 and 2.41 mmol l⁻¹, respectively, for the intravenous solution, and 1.03 and 2.71 mmol l⁻¹, respectively, for the intra-arterial solution.

Study design

The applicability of our final bilirubin formulation was evaluated in eight healthy volunteers and included the exploration of its pharmacokinetics and safety to verify the results from our review of former reports. Subjects had to be at least 18 and no older than 65 years of age and were not allowed to have a positive history of smoking, drug abuse or liver disease. Women of child-bearing potential were included only if adequate contraceptive measures had been taken. Except for contraceptives, any use of medication was prohibited. After signing informed consent, all volunteers were screened by means of a complete medical history, physical examination and a limited laboratory evaluation including blood count, renal function and liver enzymes. Subjects with any sign of active disease or laboratory results exceeding twice the upper limit of normal range were excluded. To avoid inclusion of subjects with Gilbert syndrome, baseline plasma levels of total bilirubin were not allowed to surpass 10 μ mol l⁻¹ (0.58 mg dl⁻¹).

The experimental design is depicted in Figure 1. For safety reasons, the study protocol initially consisted of a two-stage design. Both stages were separated by a safety evaluation to enable adjustment of the second stage protocol if needed.

During the experiment, a third stage was added to explore bilirubin plasma kinetics after a short high dose intravenous infusion. Intra-arterial and intravenous dose schedules were adjusted to forearm volume and total body weight, respectively. In all three stages of the protocol, subjects were studied in succession with at least 1 week in between.

During the first intra-arterial stage of the protocol, we aimed for a local increase in bilirubin concentration in the forearm vascular bed without a significant rise of systemic levels. Four increasing doses of bilirubin were infused in the brachial artery, each during 20 min. Based on the assumption of an average brachial artery flow of 2 ml⁻¹ min dl⁻¹ of forearm volume, the lowest intra-arterial dosage of 1.8 µg min⁻¹ dl⁻¹ of forearm volume was estimated to give an ipsilateral rise of plasma bilirubin concentration of 3 µmol l⁻¹ (0.18 mg dl⁻¹). For the highest intra-arterial dosage of 30 µg min⁻¹ dl⁻¹ of forearm volume, we expected an ipsilateral rise of plasma bilirubin concentration of 50 µmol l⁻¹ (2.92 mg dl⁻¹). Venous plasma samples were taken every 5 min from both the ipsilateral and the contralateral arm to assess local as well as systemic changes of bilirubin plasma concentration.

During the second stage we aimed for a systemic hyperbilirubinemia. Each of the three intravenous bolus injections was administered over a period of 5 min, followed by a 15 min pause. As others have reported an increase of approximately 30 (range 24–38) μ mol l⁻¹ after a single bolus injection of 70 mg [9], we estimated that a single bolus of 233 mg bilirubin would cause an increase in plasma bilirubin level of 100 μ mol l⁻¹. For safety reasons and the reported range, we lowered this maximum dose to 200 mg corresponding to an expected average rise of 88 μ mol l⁻¹. To enable adjustment to body weight we redefined this maximum dose



Figure 1

Study design. Bilirubin dosage was adjusted to individual forearm volume in phase 1 and body weight in phases 2 and 3. Grey bars indicate the time of infusion of bilirubin. Venous blood was sampled from the contralateral arm and in phase 1 also from the ipsilateral arm

to 2.9 mg kg⁻¹ body weight assuming an average body weight of 70 kg. The lowest intravenous dosage of 0.086 mg kg⁻¹ body weight was estimated to cause a systemic rise of 2.6 μ mol l⁻¹ (0.15 mg dl⁻¹). For the highest intravenous dosage during the second phase, we expected a rise of 26 μ mol l⁻¹ (1.52 mg dl⁻¹). Every 5 min, a plasma sample was taken from the contralateral arm.

During the third phase, we explored the pharmacokinetics of bilirubin upon a single intravenous bolus of the maximum dosage of 2.9 mg kg⁻¹ body weight administered over 15 min. Subsequently, plasma bilirubin concentrations were measured frequently over a total period of 6 h. The length of interval between sampling was increased during observation and varied from 5 min immediately upon administration to a maximum of 30 min from the second hour of observation.

During all three stages, heart rate, blood pressure, respiratory rate and body temperature were monitored. All complaints mentioned during participation were listed. After treatment, subjects visited the research unit on days 1, 2 and 7 to check for side effects. Two weeks after participation, subjects underwent a complete medical screening.

Haematological parameters were assessed using an ADVIA 120 Hemalog (Bayer Diagnostic, Tarrytown, NY, USA) and chemical parameters including both direct (i.e. conjugated) and total bilirubin concentrations were determined using an Aeroset (Abbott Laboratories, Abbott Park, IL, USA). For bilirubin concentrations above 5 μ mol l⁻¹, precision of the measurement was characterized by a coefficient of variation (CV) of 20%. Below 5 μ mol l⁻¹, which concerned seven samples of total bilirubin, the CV was much higher than 20% and therefore these results were excluded.

The initial study protocol and all interim modifications were approved by the local medical research ethics committee (CMO Regio Arnhem-Nijmegen) and consistent with the Declaration of Helsinki. In addition, we appointed an independent safety monitor and scheduled frequent meetings to discuss all safety data.

Statistics

In general, descriptive data are presented as mean (±SD). All plasma concentrations of bilirubin presented are total (conjugated and unconjugated) bilirubin plasma concentrations. Changes in plasma concentration of total bilirubin upon intra-arterial or intravenous infusion are depicted as delta. For the intra-arterial infusion in the first phase of our experiment, the delta reflects the local rise in plasma bilirubin in the forearm vascular bed per dose calculated by the difference between the venous bilirubin concentration in the forearm of administration and the contemporary systemic bilirubin concentration (i.e. the venous bilirubin concentration in the contralateral forearm) at the end of the 20 min infusion period of the dose concerned. For the intravenous infusion in the second phase of our experiment, the delta reflects the systemic rise in plasma bilirubin upon each of the three bolus infusions calculated by the difference between the highest plasma bilirubin concentration observed after each bolus infusion and the baseline concentration observed at the start of the 5 min infusion period of the dose concerned. The individual plasma concentration-time profiles obtained during the third phase of our experiment are depicted in Figure 2.

For pharmacokinetic analysis, plasma concentration–time data of total bilirubin were analysed with use of WinNonlin (version 6.3; Pharsight, Mountain View, CA, USA) by weighted (1/y) nonlinear regression analysis according to a two-compartment model with short intravenous infusion, first-order elimination, and a constant endogenous baseline concentration of bilirubin. The following pharmacokinetic parameters were obtained (Table 4): total area under the curve (AUC), total area under the first moment curve (AUMC), volume of the central compartment (V₁), half-life of the distribution phase ($t_{1/2,\alpha}$), half-life of the elimination phase ($t_{1/2,\alpha}$), mean residence time (MRT = AUMC/AUC – $T_{inf}/2$, where T_{inf} = infusion time), total plasma clearance (CL = D/AUC), and volume of distribution at steady-state (Vss = CL × MRT).

A summary of supporting pharmacokinetic and safety information from the prior parenteral human use of bilirubin is provided in Table 1.

Nomenclature of targets and ligands

Key ligands in this article are hyperlinked to corresponding entries in http://www.guidetopharmacology.org, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY [10].





Figure 2

Plasma concentration–time curves of bilirubin after short intravenous infusion of 2.9 mg kg^{-1} in three healthy male subjects (F, G and H)

Results

Subject screening

As a result of the interim amendment of our protocol, subjects were recruited on two occasions prior to both the first and the additional third phase of the study. In total, 13 subjects were screened, four of which were excluded because of an elevated plasma bilirubin concentration at baseline. The remaining nine subjects were all suitable for inclusion in our study. Eight of them actually participated in the experiments. Five subjects (2 male, 3 female) participated in the intra-arterial dosing experiment (phase 1) and four



(1 male, 3 female) also participated in the intravenous dosing experiment (phase 2). Subject B (male) was willing but, for logistical reasons, unable to participate in phase 2. The course of the plasma concentration of bilirubin upon a single intravenous bolus infusion (phase 3) was studied in three other subjects (all male). All participating subjects were without any active disease. All three female participants used oral contraceptive medication. The average age of the subjects was 21.5 years (range 19–27).

Intra-arterial (phase 1) and intravenous (phase 2) dosing studies

Although both total and direct bilirubin levels were measured, parenteral administration of our formulation consisting of unconjugated bilirubin bound to albumin, almost exclusively increased total bilirubin levels and hardly affected the plasma levels of direct (conjugated) bilirubin. The local rise of plasma total bilirubin concentration in the forearm vascular bed in response to intra-arterial administration is depicted in Table 2. During the first intra-arterial experiments, we experienced technical difficulties. Because of unsuccessful cannulation of the brachial artery during the very first experiment, we decided to administer bilirubin intravenously instead. Consequently, only systemic concentrations and safety data are presented for the first subject. Dislocation of the intra-arterial needle (second experiment) and temporary obstruction of the sampling system (third experiment) accounted for the remainder of missing values. The intra-arterial bilirubin dosage needed to increase the local plasma bilirubin concentration in the forearm vascular bed by 1 µmol l^{-1} amounted to 1.44, 0.75, 0.82 and 0.71 µg min⁻¹ dl⁻¹ of forearm tissue on average for the four subsequent intra-arterial infusions, respectively. The systemic rise of plasma bilirubin concentration in response to intravenous bolus infusion is depicted in Table 3. The intravenous dosage needed to increase the systemic plasma concentration by 1 µmol l^{-1} amounted to 0.041 (0.005) mg bilirubin per kg body weight on average.

Pharmacokinetic parameters (phase 3)

The pharmacokinetic characteristics of total bilirubin obtained during the third and final phase of our experiments are depicted in Figure 2 and Table 4. The plasma concentration–time curves could be described best by an open two-compartment model, taking the endogenous baseline concentration of total bilirubin into account. Figure 2

Table 2

Individual bilirubin plasma concentrations during intra-arterial administration

	Dose	Expected rise	Delta bilirubin concentration (μ mol I ⁻¹)					Average delta	Increase nor
Subject	$(\mu g \min^{-1} dl^{-1})$	(µmol I ⁻¹)	A	В	c	D	E	(SD; µmol I ⁻¹)	$1 \mu \text{g min}^{-1} \text{dI}^{-1}$
FAV (dL)			10	13	11	11	10		
Dose 1	1.8	3		1		0	3	1.3 (1.5)	0.7
Dose 2	6	10				5	11	8.0 (4.2)	1.3
Dose 3	18	30			23	18	25	22.0 (3.6)	1.2
Dose 4	30	50			45	27	54	42.0 (13.7)	1.4

The delta resembles the local rise of plasma bilirubin concentration in the forearm per intra-arterial dose and is depicted by the difference between the venous bilirubin concentration in the forearm of administration and the contemporary systemic bilirubin concentration in the contralateral forearm at the end of the 20 min infusion period of the dose concerned. FAV, forearm volume.

Table 3

Individual bilirubin plasma concentrations during intravenous administration

	Dose	Expected rice	Delta bili	rubin concent	ration (µmol	Average delta	Increase per	
Subject	$(mg kg^{-1})$	(µmol I ⁻¹)	A	c	D	E	(SD, μ mol I ⁻¹)	1 mg kg ⁻¹
BW (kg)			62	63.5	64	61.5		
Dose 1	0.086	2.6	3		2	2	2.3 (0.6)	27.1
Dose 2	0.29	8.8	8	6	7	7	7.0 (0.8)	24.1
Dose 3	0.86	26.4	24	21	22	19	21.5 (2.1)	25.0

The delta resembles the systemic rise of plasma bilirubin per intravenous dose and is depicted by the difference between the highest plasma bilirubin concentration observed upon each bolus infusion and the baseline bilirubin concentration observed prior to the administration of the intravenous bolus infusion concerned. BW, body weight.



Table 4

Pharmacokinetic parameters of bilirubin after a single intravenous bolus infusion of 2.9 mg kg⁻¹ of body weight

	Individual subjects			
Subject (all male)	F	G	н	Mean (SD)
BW (kg)	82	87	71	
Dosage (µmol)	406	429	351	
Tinf (min)	15	15	12	
Cbaseline (µmol I ⁻¹)	8	8	8	
AUC (μmol*min l ⁻¹)	1.17 × 10 ⁴	0.97 × 10 ⁴	1.19 × 10 ⁴	1,11 (0.12) × 10 ⁴
AUMC (μmol*min ⁻² l ⁻¹)	11.0×10^{6}	2.41 × 10 ⁶	3.72×10^{6}	5.71(4.63) × 10 ⁶
V ₁ (I)	5.8	5.2	4.0	5.0 (0.9)
$t_{1/2,a}$ (min)	43	34	42	40 (5)
t _{1/2,β} (min)	280	184	343	270 (80)
MRT (min)	315	204	333	280 (70)
CL (ml min ⁻¹)	35	44	29	36 (8)
Vss (I)	10.9	9.0	9.8	9.9 (1.0)

Data are presented as mean (SD). AUC, total area under the plasma concentration–time curve; AUMC, total area under the first moment of the plasma concentration–time curve; BW, body weight; CL, total plasma clearance; $t_{1/2,\alpha}$ half-life of the distribution phase; $t_{1/2,\beta}$, half-life of the elimination phase; MRT, mean residence time; V₁, volume of the central compartment; and Vss, volume of distribution at steady-state.

displays the fitted individual curves of total plasma bilirubin concentrations upon the administration of the single intravenous bolus infusion of 2.9 mg bilirubin per kg body weight. Maximum total bilirubin plasma concentrations of 72, 76 and 86 μ mol l⁻¹ were observed in the three subjects studied, respectively. Pharmacokinetic parameters obtained from this analysis are listed in Table 4.

Safety

In general, the administration of bilirubin was well tolerated. None of the experiments, including the one that was complicated by displacement of the intra-arterial needle and consequent extravasation of bilirubin, was associated with side effects during the administration. During the observation period, several adverse events were noted. An overview of all 13 adverse events is provided in the supporting information (Table S1). The majority of these events was mild, selflimiting and/or clearly unrelated. Three events deserve additional attention. First, one of the subjects suffered from a necrotizing appendicitis 10 days after the intra-arterial administration of bilirubin. The subject underwent surgery and fully recovered. The event was considered unrelated to our study by both the safety monitor and the independent surgeon. Although the possibility to withdraw informed consent was explicitly discussed, the subject decided to continue participation and did not show any adverse event during the second phase of the experiments. Second, one of the subjects demonstrated an asymptomatic self-limiting atrial rhythm with a right bundle branch block during the post-infusion observation period in the second phase of the study. An independent cardiology consultant considered the arrhythmia to be physiologic and unrelated to our study. Additional visits did not reveal any sign of side effects. Third, one of the

subjects developed a rash 50 h after a single high dose intravenous bolus injection of bilirubin and albumin. An independent dermatology consultant considered the eruption due to atopic eczema and unrelated to our study. Additional haematological tests and clinical chemistry did not show any sign of systemic side effects. The rash vanished within days after topical application of corticosteroids and symptomatic treatment with cetirizine.

Discussion

Promising preclinical and observational human reports on bilirubin encourage a reintroduction of its parenteral human use for the purpose of translational research [6]. Despite a reasonable amount of data on pharmacokinetics and safety of parenteral human use of bilirubin from the past, the compound is currently not commercially available in a proper formulation for human use. Our data demonstrate that it is nevertheless feasible to prepare and apply bilirubin solutions for parenteral human use in accordance with GMP standards. Due to the exploratory nature of our experiments and the associated low number of subjects studied, our pharmacokinetics and safety results should be discussed in the context of previously reported data. Table 1 provides an overview of all available data on the former human use of bilirubin.

Production

The majority of previous publications on the human use of bilirubin report the use of sodium carbonate solutions with high alkalinity. In contrast to these reports, we dissolved bilirubin in sodium hydroxide and diluted this product in a solution with albumin in excess. Similar procedures have been



published before for animal experiments and a few human studies [11–19]. Apart from the beneficial reduction of alkalinity, this strategy is consistent with normal *in vivo* physiologic conditions in which unconjugated bilirubin is tightly bound to albumin [20]. In addition, albumin binding has been shown to augment the antioxidant activity of unconjugated bilirubin *in vitro* [21, 22].

Pharmacokinetics

In preparation of future translational studies on the supposed beneficial impact of bilirubin, we aimed to explore the intra-arterial use of unconjugated bilirubin and verify the previously reported pharmacokinetic characteristics of intravenously administered unconjugated bilirubin. Our data demonstrate a predictable dose–concentration relationship for both routes of administration, facilitating the design of future translational studies on the therapeutic potential of bilirubin.

During the third phase of our experiments, we studied the pharmacokinetics of unconjugated bilirubin upon a single intravenous bolus infusion. The disposition of unconjugated bilirubin is highly determined by its strong binding to albumin. Its exchange between albumin in plasma and albumin in the interstitial fluid is allowed by a small free fraction, which is estimated to amount only 1/50000 of total serum bilirubin [20]. The volume of distribution (Vss) found in our experiments (Table 4) corresponds to a normal albumin space of 91 [23]. The two phases of the two-compartment model that was fitted to the plasma concentration-time curve relate to the initial distribution within the albumin space and the true bilirubin clearance by the liver. The corresponding halflives are in agreement with values recently predicted by a theoretical model based on the consequences of its strong albumin binding [20]. In the past, multiple studies on the clearance of intravenously administered bilirubin have been performed using either small trace-labelled amounts [11, 24], larger non-radioactive dosages [25–30], such as in our experiments, or both [31]. Two- and three-compartment models with varying pharmacokinetic characteristics have been proposed. Comparison of data is complicated by the fact that the interpretation of the disappearance curve and selection of the most appropriate compartment model in these older reports was hampered by the lack of current insight into bilirubin physiology. Most authors falsely assumed the presence of a hepatic bilirubin pool [24, 26, 28] and did not consider the importance of the small free fraction of unconjugated bilirubin enabling exchange to albumin within the interstitial space. Notably, our experiments were not designed to gain detailed insight into the pharmacokinetics of bilirubin, but aimed to assess the applicability of our formulation for parenteral human use and explore its pharmacokinetic characteristics in comparison with existing data.

Safety

Whereas the actual administration of bilirubin was well tolerated, several adverse events occurred during the subsequent observation including an appendicitis, an asymptomatic cardiac arrhythmia and a pruritic rash. Although independent expert clinicians considered these events to be unrelated to our experiment, it should be stated that we cannot exclude

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a causal association between the events and the combined administration of bilirubin and albumin. Moreover, the limited size of our study does not allow any definite conclusion on safety issues. We therefore performed an extensive review of the literature covering over 1750 individual bilirubin infusions over the last century (Table 1). The earliest reports on the human use of bilirubin mentioned the occurrence of side effects consisting of transient chills and fever [32, 33]. Hinting at microbial contamination, these effects could be avoided by a switch to the exclusive use of fresh sterile solutions [32]. Moreover, immediate effects like flushing, tachycardia and headache have been mentioned upon the combined infusion of bilirubin and bile acids [33]. The contemporary use of bile acids obviously complicates the definite attribution of the side effects reported in this study. Importantly, no other study on the human use of bilirubin has mentioned the occurrence of these immediate effects. Finally, the use of large amounts of bilirubin dissolved in an unbuffered alkaline solution has been associated with the frequent occurrence of venous thrombosis [34]. These authors reported up to 30 infusions per subject and bilirubin plasma concentrations up to 500 μ mol l⁻¹. Though less severe, others have reported mild soreness at the injection site in a few cases of highly dosed bilirubin infusions [27]. Notably, the vast majority of reports on the parenteral use of bilirubin have not mentioned any side effect related to its infusion. Several of these reports explicitly mentioned the absence of side effects, covering a total of 278 bilirubin infusions with plasma concentrations up to 150 μ mol l⁻¹ [35–39].

In view of the available data, we feel confident to state that parenteral use of bilirubin is safe if produced under GMP conditions and applied in its current dosage and albumin bound formulation. Obviously, future studies should maintain focus on the early detection of side effects and it should be noted that bilirubin pigments currently available on the market are purified from either porcine or bovine gallstones. In our opinion only products of porcine origin should be used to nullify the theoretical risk of prion transmission.

Limitations

Several limitations of this study should be mentioned. First, the number of subjects included to test the applicability of our formulation is too low to permit conclusions on pharmacokinetics and safety characteristics on its own. In view of the yet available knowledge from former experiments and a recent theoretical model, we felt it was more appropriate to perform a thorough review of prior reports than to increase the sample size purely to confirm existing knowledge on these topics.

Second, we adjusted the bilirubin dosage to forearm volume for intra-arterial administration and to body weight for intravenous administration in order to minimize variability. This strategy resulted in fairly predictable and consistent bilirubin plasma concentrations. It should be noted though that both the forearm volumes and body weights of our subjects were quite similar and within normal range. Additional experiments are needed to confirm the applicability of our dose adjustment strategies in a study population with a larger inter-individual variability in forearm volume and body weight.



Third, the clearance of bilirubin demonstrates a significant inter-individual variability which is related to several genetic polymorphisms [39]. Most common is the Gilbert syndrome, which is due to a TA-insertion in the UGT1A1 promotor region and accompanied by a significantly reduced glucuronidation activity to approximately 30% of normal [40]. To avoid outliers as a result of the Gilbert syndrome, we excluded subjects with a plasma bilirubin concentration exceeding $10 \,\mu\text{mol}\,l^{-1}$ during the screening visit. Though less sensitive than the assessment of individual genetic polymorphisms, this approach sufficiently excludes subjects with the Gilbert syndrome [41]. In view of the relatively high prevalence of the Gilbert syndrome in the general population, the avoidance of outliers by the exclusion of subjects with high baseline bilirubin concentrations may have caused our clearance data to overestimate the average clearance capacity within the general population.

Finally, we did not assess the impact of our formulation on endothelial function. Additional experiments in healthy volunteers are needed to compare endothelial function during the intra-arterial infusion of bilirubin and saline as control. To discern the effect of bilirubin from possible interference of co-administered albumin, subsequent proof of concept studies in subjects with endothelial dysfunction should ideally be performed with an albumin solution as control.

Conclusion

In conclusion, our study reopens the gate to the human parenteral use of bilirubin for the purpose of translational research. Larger studies are needed to evaluate the usefulness of adjusting dose regimens to either forearm volume or body weight and to study the influence of genetic variation on the elimination rate of bilirubin from plasma. Nevertheless, it appears feasible to corroborate the promising preclinical and observational data of bilirubin in human disease models. As such, our data provide a basis for further experiments on the beneficial antioxidant and anti-inflammatory effects attributed to bilirubin.

Competing Interests

There are no competing interests to declare.

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References

- 1 von Bernhard K, Ritzel G, Steiner KU. Über eine biologische Bedeutung der Gallenfarbstoffe. Bilirubin und Biliverdin als Antioxydantien für das Vitamin A und die essentiellen Fettsäuren. Helv Chim Acta 1954; 37: 306–13.
- **2** Abraham NG, Kappas A. Pharmacological and clinical aspects of heme oxygenase. Pharmacol Rev 2008; 60: 79–127.
- **3** Wagener FA, Volk HD, Willis D, Abraham NG, Soares MP, Adema GJ, *et al.* Different faces of the heme-heme oxygenase system in inflammation. Pharmacol Rev 2003; 55: 551–71.
- **4** Vitek L. The role of bilirubin in diabetes, metabolic syndrome, and cardiovascular diseases. Front Pharmacol 2012; 3: 55.
- 5 McCarty MF. 'Iatrogenic Gilbert syndrome' a strategy for reducing vascular and cancer risk by increasing plasma unconjugated bilirubin. Med Hypotheses 2007; 69: 974–94.
- **6** Durante W. Bilirubin: striking gold in diabetic vasculopathy? Diabetes 2015; 64: 1506–8.
- 7 Anderson TJ, Phillips SA. Assessment and prognosis of peripheral artery measures of vascular function. Prog Cardiovasc Dis 2015; 57: 497–509.
- **8** Dekker D, Dorresteijn MJ, Pijnenburg M, Heemskerk S, Rasing-Hoogveld A, Burger DM, *et al.* The bilirubin-increasing drug atazanavir improves endothelial function in patients with type 2 diabetes mellitus. Arterioscler Thromb Vasc Biol 2011; 31: 458–63.
- **9** Eilbott W, von Bergmann G. Funktionsprüfung der Leber mittels Bilirubinbelastung. Z klinische Med 1927; 106: 520–60.
- **10** Southan C, Sharman JL, Benson HE, Faccenda E, Pawson AJ, Alexander SPH, *et al.* The IUPHAR/BPS Guide to PHARMACOLOGY in 2016: towards curated quantitative interactions between 1300 protein targets and 6000 ligands. Nucl Acids Res 2016; 44: D1–68.
- 11 Berk PD, Howe RB, Bloomer JR, Berlin NI. Studies of bilirubin kinetics in normal adults. J Clin Invest 1969; 48: 2176–90.
- 12 Bjornbeth BA, Villanger O, Christensen T, Lyberg T, Raeder MG. Large intravenous loads of bilirubin photoconversion products, in contrast to bilirubin, do not cause cholestasis in bile aciddepleted pigs. Scand J Gastroenterol 1997; 32: 246–53.
- **13** Bjornbeth BA, Endresen M, Hvattum E, Lyberg T, Villanger O, Raeder MG. Large intravenous bilirubin loads increase the cytotoxicity of bile and lower the resistance of the canalicular membrane to cytotoxic injury and cause cholestasis in pigs. Scand J Gastroenterol 1998; 33: 201–8.
- **14** Bjornbeth BA, Labori KJ, Hvattum E, Lyberg T, Christensen T, Woldseth B, *et al.* Effect of intravenous bilirubin infusion on biliary phospholipid secretion, hepatic P-glycoprotein expression, and biliary cytotoxicity in pigs. Scand J Gastroenterol 1999; 34: 1042–9.
- **15** Labori KJ, Bjornbeth BA, Hvattum E, Lyberg T, Raeder MG. Intravenous bilirubin, dibromosulfophthalein, and bromosulfophthalein infusions uncouple biliary phospholipid and cholesterol secretion from bile acid secretion by inhibiting hepatic phosphoglycoprotein-3 activity in pigs. Scand J Gastroenterol 2000; 35: 873–82.
- **16** Labori KJ, Arnkvaern K, Bjornbeth BA, Press CM, Raeder MG. Cholestatic effect of large bilirubin loads and cholestasis

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protection conferred by cholic acid co-infusion: a molecular and ultrastructural study. Scand J Gastroenterol 2002; 37: 585-96

- 17 Rozdilsky B. Toxicity of bilirubin in adult animals. An experimental study. Arch Pathol 1961; 72: 8–17.
- 18 Veel T, Villanger O, Holthe MR, Skjorten FS, Raeder MG. Intravenous bilirubin infusion causes vacuolization of the cytoplasm of hepatocytes and canalicular cholestasis. Acta Physiol Scand 1991; 143: 421-9.
- 19 Villanger O, Bjornbeth BA, Lyberg T, Raeder MG. Bile acids protect the liver against the cholestatic effect of large bilirubin loads. Scand J Gastroenterol 1995; 30: 1186-93.
- **20** Levitt DG, Levitt MD. Quantitative assessment of the multiple processes responsible for bilirubin homeostasis in health and disease. Clin Exp Gastroenterol 2014; 7: 307-28.
- 21 Stocker R, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN. Bilirubin is an antioxidant of possible physiological importance. Science 1987; 235: 1043-6.
- 22 Stocker R. Glazer AN. Ames BN. Antioxidant activity of albuminbound bilirubin. Proc Natl Acad Sci U S A 1987; 84: 5918-22.
- 23 Levitt DG. The pharmacokinetics of the interstitial space in humans. BMC Clin Pharmacol 2003; 3: 3.
- 24 Berk PD, Bloomer JR, Howe RB, Berlin NI. Constitutional hepatic dysfunction (Gilbert's syndrome). A new definition based on kinetic studies with unconjugated radiobilirubin. Am J Med 1970; 49: 296-305.
- 25 With TK. The bilirubin excretion test as a functional liver test. With remarks on the course of the curve of the serum bilirubin. Acta Med Scand 1943; 116: 96-114.
- 26 Billing BH, Williams R, Richards TG. Defects in hepatic transport of bilirubin in congenital hyperbilirubinaemia: an analysis of plasma bilirubin disappearance curves. Clin Sci (Lond) 1954; 27: 245-57.
- 27 Okolicsanyi L, Ghidini O, Orlando R, Cortelazzo S, Benedetti G, Naccarato R, et al. An evaluation of bilirubin kinetics with respect to the diagnosis of Gilbert's syndrome. Clin Sci Mol Med 1978; 54: 539-47.
- 28 Goresky CA, Gordon ER, Shaffer EA, Pare P, Carassavas D, Aronoff A. Definition of a conjugation of dysfunction in Gilbert's syndrome: studies of the handling of bilirubin loads and of the pattern of bilirubin conjugates secreted in bile. Clin Sci Mol Med 1978; 55: 63-71.
- 29 Okolicsanyi L, Orlando R, Venuti M, Dal BG, Cobelli C, Ruggeri A, et al. A modeling study of the effect of fasting on bilirubin kinetics in Gilbert's syndrome. Am J Physiol 1981; 240: R266-R71.
- 30 Zeneroli ML, Piaggi V, Cremonini C, Gozzi C, Ventura E. Sources of bile pigment overproduction in Gilbert's syndrome: studies with non-radioactive bilirubin kinetics and with delta-[3,5-3H] aminolaevulinic acid and [2-14C]glycine. Clin Sci (Lond) 1982; 62: 643-9.
- 31 Barrett V, Berk PD, Menken M, Berlin NI. Bilirubin turnover studies in normal and pathologic states using bilirubin-14C. Ann Intern Med 1968; 68: 355-77.
- 32 von Bergmann G. Der functionellen Pathologie der Leber. Klin Wochenschr 1927; 16: 776-80.
- 33 Thompson HE, Wyatt BL. Experimentally induced jaundice (hyperbilirubinaemia). Report of animal experimentation and of

the physiologic effect of jaundice in patients with atrophic arthritis. Arch Intern Med 1938; 61: 481-95.

- 34 Hench PS. Effect of spontaneous jaundice on rheumatoid (atrophic) arthritis. Attempts to reproduce the phenomenon. BMJ 1938; 2: 394-8.
- 35 Soffer L. Bilirubin excretion as a test for liver function during normal pregnancy. Bull Johns Hopkins Hosp 1933; 52: 365-75
- 36 Sullivan CF, Tew WP, Watson EM. The bilirubin excretion test of liver function in pregnancy. J Obstet Gynaecol 1934; 41: 347-67.
- 37 Weech AA, Vann D, Grillo RA. The clearance of bilirubin from the plasma. A measure of the excreting power of the liver. J Clin Invest 1941; 20: 323-32.
- **38** Kornberg A. Latent liver disease in persons recovered from catarrhal jaundice and in otherwise normal medical students as revealed by the bilirubine excretion test. J Clin Invest 1942; 21: 299 - 308
- 39 Kringen MK, Piehler AP, Grimholt RM, Opdal MS, Haug KB, Urdal P. Serum bilirubin concentration in healthy adult North-Europeans is strictly controlled by the UGT1A1 TA-repeat variants. PLoS One 2014; 9: e90248.
- 40 Hirschfield GM, Alexander GJ. Gilbert's syndrome: an overview for clinical biochemists. Ann Clin Biochem 2006; 43: 340-3.
- 41 Borlak J, Thum T, Landt O, Erb K, Hermann R. Molecular diagnosis of a familial nonhemolytic hyperbilirubinemia (Gilbert's syndrome) in healthy subjects. Hepatology 2000; 32: 792-5.
- 42 Boshamer K. Untersuchungen über die Einwirkung von Äthernarkosen auf die Leberfunktion. J Mol Med 1928; 7: 445-7.
- 43 Harrop GA, Barron ESG. The excretion of intravenously injected bilirubin as a test of liver function. J Clin Invest 1931; 9: 577-87.
- 44 Stroebe F. Latente Hepatopathie nach Schangerschaftstoxikose. Klin Wochenschr 1932; 11: 495-7.
- 45 Zadek E, Tietze A, Gebert K. Zur Wertung der Leberfunktionsprüfungen am Krankenbett. Klin Wochenschr 1933; 12: 60-2.
- 46 Hofbauer J. Fortschritte in der Ergründung des Wesens und der ätiologischen Behandlung der Graviditätstoxikosen. Klin Wochenschr 1933; 12: 366-73.
- 47 Dietel FG, Polak A. Latente Störung von Leberfunktion und Reticuloendothel nach Eklampsie. Arch Gynecol Obstet 1933; 152: 469-78.
- 48 Dietel FG, Polak A. Funktionszustand von Leber und Reticuloendothel nach Nephropathie. Arch Gynecol Obstet 1933; 154: 122-8.
- 49 Binswanger H. Leberuntersuchungen bei Alkoholpsychosen, gleichzeitig ein Beitrag zur funktionellen Pathologie der Leber. Eur Arch Psychiatry Clin Neurosci 1933; 100: 619-69.
- 50 Fuge K. Die Bilirubinbelastungsprobe der Leber bei der Diagnose der Hepatopathia gravidarum. Arch Gynecol Obstet 1933; 154: 507-21.
- 51 Nürnberger L. Zur Kenntnis der Leberfunktion am Ende der normalen Schwangerschaft. Arch Gynecol Obstet 1933; 153: 1 - 25.
- 52 Schindel L, Barth E. Die Bedeutung der Takata-Reaktion für die Diagnose der Lebererkrankungen in ihrem Verhältnis zur

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Galaktose- und Bilirubin-Belastung. Klin Wochenschr 1934; 13: 1355–9.

- **53** Bröchner-Mortensen K. Über Bilirubinbelastung als Leberfunktionsprobe. Acta Med Scand 1935; 85: 1–32.
- **54** Bröchner-Mortensen K. Untersuchung der Leberfunktion bei Patienten mit akuten febrilen Leiden. Acta Med Scand 1935; 14: 229–43.
- **55** Ruhbaum W, Matheja W. Leberfunktionsproben bei latenter Leberschädigung. Klin Wochenschr 1935; 14: 1568–70.
- **56** Fulde W. Wert und Methodik verschiedener Leberfunktionsprüfungen für Klinik und Praxis. Klin Wochenschr 1935; 14: 1201–6.
- **57** Lin H, Eastman N. The behavior of intravenously injected bilirubin in newborn infants. Am J Obstet Gynecol 1937; 33: 317–24.
- **58** Lyon RA. The bilirubin liver function test in the toxemias of pregnancy. Am J Obstet Gynecol 1938; 36: 99–103.
- **59** Rawls WB. Experimentally induced jaundice (comment). Arch Intern Med 1938; 61: 495–6.
- 60 Arpe H. Vergleichende Untersuchungen über Leberfunktionsprüfungen nach Felix-Teske und Von Bergmann-Eilbott (Belastung mittels p-Oxyphenylbrenztraubensäure bzw. Bilirubin). Klin Wochenschr 1946; 24–25: 364–6.

- **61** Tisdale WA, Klatskin G, Kinsella ED. The significance of the direct-reacting fraction of serum bilirubin in hemolytic jaundice. Am J Med 1959; 26: 214–27.
- **62** Schmidt D, Grauel EL, Syllm-Rapoport I. Bilirubinbelastugen bei prämaturen Neugeborenen. Z Kinderheilkd 1966; 69: 19–29.
- **63** Raymond GD, Galambos JT. Study of bilirubin clearance of the human liver. Gastroenterology 1966; 50: 862–3.
- **64** Nixon JC, Monahan GJ. Gilbert's disease and the bilirubin tolerance test. Can Med Assoc J 1967; 96: 370–3.
- **65** Engstedt L, Johansson S, Nyberg A. Estimation of human erythrocyte life-span from bilirubin turnover. J Lab Clin Med 1967; 70: 195–203.

Supporting Information

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Table S1 Safety data including all adverse events noted during the 2-week observation period following each phase of the experiments