

Alterations in oral microbial flora induced by waterpipe tobacco smoking

Muhamad Ali K Shakhathreh¹
Omar F Khabour¹
Karem H Alzoubi²
Majed M Masadeh³
Emad I Hussein⁴
George N Bshara¹

¹Department of Medical Laboratory Sciences, Faculty of Applied Medical Sciences, Irbid, Jordan; ²Department of Clinical Pharmacy, Faculty of Pharmacy, Jordan University of Science and Technology, Irbid, Jordan; ³Department of Pharmaceutical Technology, Faculty of Pharmacy, Jordan University of Science and Technology, Irbid, Jordan; ⁴Department of Biological Sciences, Yarmouk University, Irbid, Jordan

Background: Waterpipe smoking is a global health problem and a serious public concern. Little is known about the effects of waterpipe smoking on oral health. In the current study, we examined the alterations of oral microbial flora by waterpipe smoking.

Methods: One hundred adult healthy subjects (59 waterpipe smokers and 41 non-smokers) were recruited into the study. Swabs were taken from the oral cavity and subgingival regions. Standard culturing techniques were used to identify types, frequency, and mean number of microorganisms in cultures obtained from the subjects.

Results: It was notable that waterpipe smokers were significantly associated with a history of oral infections. In subgingiva, *Acinetobacter* and *Moraxella* species were present only in waterpipe smokers. In addition, the frequency of *Candida albicans* was higher in the subgingiva of waterpipe smokers ($p = 0.023$) while the frequency of *Fusobacterium nucleatum* was significantly lower in the subgingiva of waterpipe smokers ($p = 0.036$). However, no change was observed in other tested bacteria, such as *Campylobacter* species; Viridans group streptococci, *Enterobacteriaceae*, and *Staphylococcus aureus*. In oral cavity and when colony-forming units were considered, the only bacterial species that showed significant difference were the black-pigmented bacteria ($p < 0.001$).

Conclusion: This study provides evidence indicating that some of the oral microflora is significantly altered by waterpipe smoking.

Keywords: waterpipe, smoking, oral microflora, hookah, tobacco

Introduction

Waterpipe is a way of tobacco consumption in which the smoke passes through the water before it is inhaled.¹ The use of waterpipes is increasing all over the world, especially among young people and women.² A waterpipe machine has four major parts: a head, stem, vase, and hose (Figure 1). Smoking using this machine includes the use of flavored and hydrated, tobacco known as “moassel.” A charcoal is placed on top of the tobacco to provide the heat needed to burn the moassel.¹ The bottom of the head has holes in it that passes the produced smoke to the stem, which is submerged in water that half-fills the vase. The hose is not submerged, exits from the bowl’s top, and ends with a mouthpiece, from which the user inhales.¹

The health effects of cigarette smoking are well documented; however, knowledge regarding the impact of waterpipe smoking on body health is still lacking.³ Previous literature has shown that smoke produced by a waterpipe contains a similar toxicant profile to that produced by cigarettes with different magnitude. For example, the tar

Correspondence: Muhamad Ali K Shakhathreh
Department of Medical Laboratory Sciences, Faculty of Applied Medical Sciences, Jordan University of Science and Technology, PO Box 3030, Irbid 22110, Jordan
Tel + 962 2 7201000 ext 23874
Email mkshakhathreh@just.edu.jo



Figure 1 A waterpipe machine. The major components of a waterpipe machine are labeled and include the head, stem, vase, and hose.

produced by a single episode of waterpipe is about five times that produced by a single cigarette.⁴ Similarly, exposure to carbon monoxide is at least several folds higher during waterpipe smoking compared with that of cigarette smoking.⁵ Furthermore, the polycyclic aromatic hydrocarbons in waterpipe smoke are many times more than that of cigarette smoke.⁶ In addition, the style of waterpipe smoking results in a dramatically higher exposure volume to smoke, more tobacco consumption per smoking event, and longer smoke inhalation periods.⁷ Finally, tobacco in waterpipes is usually mixed with sugar, glycerol, and flavors, this mixture is burned by charcoal.¹ Thus, it is expected that waterpipe smoking will have a distinct effect on oral microbial flora.

The effects of cigarette smoking on oral health show that cigarette smoking is associated with oral cancer, periodontal disease, oral infections, and interference with the taste and modulation of normal flora.^{8,9} Several studies have also investigated the effect of smoking on oral microbiota and showed significant differences in the subgingival bacteria between smokers and non-smokers.¹⁰ For example, Zambon reported that smokers harbored significantly higher levels of *Bacteroides forsythus* subgingivally.¹¹ In addition, the prevalence of several oral pathogens such as *Prevotella nigrescens*, *Prevotella intermedia*, *Porphyromonas gingivalis*, and *Tannerella* was significantly higher in smokers than in non-smokers.¹² Regarding waterpipe smoking, few studies have examined effects of waterpipe smoking on oral health. A recent study has shown a strong association between waterpipe smoking and periodontal disease.¹³ In addition, waterpipe smoking has been shown to significantly increase potentially malignant oral mucosal lesions¹⁴ and lower lip squamous cell carcinoma and keratoacanthoma.¹⁵ Moreover, waterpipe smoking has been shown to induce DNA damage in buccal cells.¹⁶

Several bacterial species were identified in the human oral cavity including many anaerobic periodontal pathogens which are associated with periodontal infections such as *P. gingivalis*, *Tannerella forsythia*, *P. intermedia*, *Eikenella corrodens*, *Campylobacter rectus*, *Aggregatibacter actinomycetemcomitans*, *Treponema denticola*, and *Fusobacterium nucleatum*.¹⁷ The frequency of these pathogens in the oral cavity has been shown to be altered by cigarette smoking.¹⁸ Since the profile of toxicants and behavior of smoking are different between waterpipe and cigarette smoking, in this study, we investigated the effect of waterpipe smoking on the profile of normal flora in the oral cavity and subgingiva which is still undefined and unclear. The results of this study might be used to derive policies and interventions that target waterpipe smokers.

Materials and methods

Subjects

Fifty-nine healthy waterpipe smokers were recruited to participate in the study. As a control, 41 healthy non-smokers that matched the smokers group in gender and age were also recruited from the same geographical area. Smokers and their matched controls were recruited from customers of waterpipe cafes in Irbid city, which is the largest urban population north of Jordan. Usual customers of waterpipe cafes are young adults of both genders, who are either waterpipe smokers or waterpipe non-smokers who accompany their smoking friends, relatives, or family members to these places. Those who used tobacco products other than waterpipe were excluded from the study. Additionally, participants with previous history of oral diseases/infections or who were taking medications during the past 2 months were also excluded from the study. This study was approved by the Institutional Review Board of Jordan University of Science and Technology (Approval number 152/2012), and written informed consent was obtained from all subjects according to Institutional Review Board approval.

Sample collection

Subgingival samples were collected by inserting and rotating absorbent sterile paper-points (Meta Biomed Co Ltd, Cheongju, South Korea) for 10–15 seconds between the front upper and lower teeth to get high quantities of bacteria. Subjects with bleeding gums were excluded. In addition, any participant with a history of oral infection in the past 2 months was excluded. Oral cavity samples (teeth, tongue, and cheeks) were collected using sterile cotton transport swabs. Occasionally, bleeding occurred while taking the

sample after inserting and rotating the sterile paper points (one for lower part and one for the upper part of teeth). In this case, the sample was discarded and sampling was repeated from a non-bleeding site. Samples were transported under anaerobic conditions using liquid dental transport medium (LDTM) (Anaerobe Systems, Morgan Hill, CA, USA). All samples were processed for microbiology techniques within 1–2 hours.

Culture conditions

Isolation of microorganisms was carried out by methods previously reported.¹⁹ Isolated strains were characterized using standard microbiological methods as described in Clinical and Laboratory Standards Institute (CLSI) ML35-A2²⁰ document (Table 1). Each sample was vortexed at high speed for 60 seconds and then subjected to a series of 10-fold dilutions (up to 10⁻⁴) by using sterile Dulbecco's phosphate buffered saline. Thereafter, aliquots of 100 µL from each different dilution were spread onto different differential and selective media including: crystal violet erythromycin (CVE), Wolinella agar, MacConkey agar, mannitol salt agar (MSA), kanamycin–vancomycin laked blood-2 (KVLB-2), tryptic soy agar supplemented with hemin 5 mg/mL and vitamin K3 (Menadione) 0.5 µg/mL, Mitis Salivarius agar supplemented with tellurite solution, and Sabouraud dextrose agar (SDA). For each microbial species, colony-forming units (CFUs/mL) were recorded for each plate by CFU enumeration assay. Total counts were determined on Columbia blood agar, aerobically and anaerobically. The following media were inoculated and incubated anaerobically at 37°C for 7–10 days by using the Oxoid™ Anaerobic Atmosphere Generation System, Anaero-Gen™ 2.5 L Sachets: 1; ThermoFisher Scientific, Waltham, MA, USA) CVE agar (trypticase soy agar, yeast extract 5 g/L, sodium chloride 5 g/L, glucose 2 g/L, tryptophan 0.2 g/L,

crystal violet 5 mg/L, erythromycin 4 mg/L, defibrinated sheep blood 50 mL/L) was used to assess *F. nucleatum*;²¹ 2) Kanamycin Vancomycin Laked Blood-2 (KVLB-2: Kanamycin 75 µg/mL, Vancomycin 2 µg/mL, laked blood) supplemented with hemin 5 µg/mL, vitamin K1 1 µg/mL), and trypticase soy agar supplemented with (hemin 5 mg/mL, Vitamin K3 0.5 µg/mL, sheep blood) were used to assess the black-pigmented *P. intermedia* and *P. gingivalis*, respectively; and 3) Wolinella agar (trypticase soy agar, vancomycin 9 µg/mL, ferrous sulfate 0.2 g/L, sodium thiosulphate 0.3 g/L, sodium fumarate 3 g/L, sodium formate 2 g/L) was used for the isolation of *C. rectus*.²²

MSA was used to identify aerobic species such as *Staphylococcus aureus* and *S. epidermidis*. Yeast cells *Candida albicans* were identified by SDA. Species of *Enterobacteriaceae* family were identified on MacConkey agar and depending on the following tests: Gram stain, citrate test, motility test, urease test, indole test, oxidase test, catalase test, and triple sugar iron (TSI) agar. Mitis Salivarius Agar with 1% potassium tellurite was used to detect oral viridans streptococci, which consists of *Streptococcus mutans*, *S. salivarius*, and *S. mitis*.

For the determination of total aerobic and anaerobic bacteria counts for each microbial species, CFUs/mL were recorded for each plate by using the CFU enumeration assay. Bacteria were grown on Columbia blood agar under: 1) aerobic condition: 35°C ± 2°C for 18–72 hours under appropriate atmospheric conditions; and 2) anaerobic condition: 37°C for 7–10 days by using AnaeroGen 2.5 L sachets.

Statistical analysis

Statistical analyses were conducted using SPSS software (version 17). Comparison of frequency of bacteria and pathogens between waterpipe smokers and control groups was

Table 1 Culture media used in the current study

Culture media	Identified microorganisms	Additional identification procedures
CBA	Aerobic and anaerobic	None
CVE	<i>Fusobacterium nucleatum</i>	Distinct colonial morphology of <i>F. nucleatum</i> on CVE agar
KVLB-2	The black-pigmented <i>Prevotella intermedia</i>	Characteristic colonial growth and morphology, special-potency antibiotic disks,
MacConkey agar	<i>Enterobacteriaceae</i>	Gram stain, citrate test, motility test, urease test, indole test, oxidase test, catalase test, and TSI agar
MSA	<i>Staphylococcus aureus</i> and <i>Staphylococcus epidermidis</i>	Colony morphology and colors
Mitis Salivarius agar	Oral viridans streptococci	Colony morphology and colors
SDA	<i>Candida albicans</i>	Colony morphology and colors
Tryptic soy agar	<i>Porphyromonas gingivalis</i>	Colony morphology, biochemical tests, fluorescence test

Abbreviations: CBA, Columbia blood agar; CVE, crystal violet erythromycin; KVLB, kanamycin–vancomycin laked blood; MSA, mannitol salt agar; SDA, Sabouraud dextrose agar; TSI, triple sugar iron.

conducted using the Mann–Whitney *U*-test (as data were not normally distributed), and chi-square test. CFU values were expressed as mean \pm standard error of the mean (SEM) and were compared between different groups using the Mann–Whitney *U*-test (as data was also not normally distributed). Significant differences were examined at $p < 0.05$. Power analysis was carried out using G power version 3.0.10 (Franz Faul, Universtat Kiel, Germany). Sample size analysis was performed at 80% and 5% α -level of significance.

Results

The characteristics of study participants are shown in Table 2. The mean age of the waterpipe group was 23.98 ± 2.77 years versus 24.14 ± 4.37 years in the control group ($p = 0.8$). Males represented 74.5% of the waterpipe group and 65.9% of the control group. The average smoke sessions in waterpipe group

per week were 4.98 ± 2.12 . It was notable that waterpipe smokers were significantly associated with a history of oral infections ($p = 0.01$). Table 3 shows the distribution of bacterial species isolated from subgingiva of participants. The most abundant bacteria from examined species were viridans streptococci, *C. rectus* and *F. nucleatum* in waterpipe users (range 84%–94%) and in non-smokers (range: 85%–97%). *Acinetobacter* and *Moraxella* species were present only in waterpipe smokers with a frequency of 5.1% and a mean of 31.7. The frequency of *F. nucleatum* was significantly lower in subgingiva of waterpipe smokers ($p = 0.036$, Figure 2A) while the frequency of *C. albicans* was higher in waterpipe smokers ($p = 0.023$, Figure 2B). Finally, the profile of the remaining examined bacteria was similar between the two groups ($p > 0.05$).

Table 4 shows the distribution of bacterial species isolated from the oral cavity of participants. The most abundant

Table 2 Characteristics of participants

Characteristics	Waterpipe group	Control group	p-value
Age, years (mean \pm SD)	23.98 \pm 2.77	24.14 \pm 4.37	0.8 (using Mann–Whitney <i>U</i> -test)
Age range, years	19–32	19–35	
Gender			
Male	44 (74.5%)	27 (65.9%)	0.4 (using χ^2 test)
Female	15 (25.5%)	14 (34.1%)	
History of oral infection			
Yes	38 (64.4%)	16 (39.0%)	0.01 (using χ^2 test)
No	21 (35.6%)	25 (61.0%)	
Waterpipe use: sessions per week (mean \pm SD)	4.98 \pm 2.12	–	
Duration of WP session, minutes			
Less than 30	12 (20.3%)	–	
30–60	29 (49.2%)		
61–90	15 (25.4%)		
More than 90	3 (5.1%)		
Age of initiation, years			
Less than 14	3 (5.1%)	–	
15–17	14 (23.7%)		
18–21	30 (50.9%)		
More than 21	12 (20.3%)		

Abbreviations: SD, standard deviation; WP, waterpipe.

Table 3 Frequency and CFUs of isolates of detected microorganisms from subgingival plaque of participants

Microorganisms (oral)	Waterpipe frequency, N (%)	Controls frequency, N (%)	p-value, χ^2 test	Waterpipe CFU, mean \pm SD	Controls CFU, mean \pm SD	Mann–Whitney <i>U</i> -test p-value
<i>Fusobacterium nucleatum</i>	50 (84.7)	36 (97.6)	0.036	3.9 \pm 8.9	3.1 \pm 7.6	0.49
Black-pigmented bacteria	43 (68.3)	29 (78.4)	0.31	3.3 \pm 3.6	11.0 \pm 31.9	0.69
<i>Campylobacter</i> spp.	52 (88.1)	38 (92.7)	0.52	5.16 \pm 26.8	83.6 \pm 209.2	0.33
Viridans group streptococci	55 (93.2)	35 (85.4)	0.20	99.7 \pm 320.9	130.1 \pm 408	0.58
<i>Acinetobacter</i> and <i>Moraxella</i>	3 (5.1)	0 (0)	0.14	31.7 \pm 38.8	0	–
<i>Enterobacteriaceae</i>	7 (11.9)	3 (7.35)	0.456	0.02 \pm 0.03	0.03 \pm 0.02	0.25
<i>Staphylococcus aureus</i>	2 (3.4)	4 (9.8)	0.19	0.01 \pm 0.02	0.01 \pm 0.01	0.10
Yeast (<i>Candida albicans</i>)	10 (16.9)	1 (2.45)	0.023	0.1 \pm 0.4	0.01 \pm 0.0	0.63

Abbreviations: CFU, colony-forming units; SD, standard deviation.

bacteria from examined species were viridans streptococci, *Campylobacter* spp. *F. nucleatum* and black-pigmented bacteria in waterpipe users (range 94%–100%) and in non-smokers (range: 92%–96%). From these abundant bacteria, the mean of CFUs of black-pigmented bacteria was significantly lower ($p = 0.001$, Figure 3) in waterpipe smokers. Finally, no significant variations were detected in the distribution or mean of the rest of examined bacteria between the two groups ($p > 0.05$).

Discussion

The popularity of waterpipe smoking is growing in the Eastern Mediterranean and throughout the world, including

the USA and other Western countries, especially among youth.²³ This spread is due, in part, to the use of tobacco that is sweetened and flavored²⁴ and the misperception that the waterpipe “filters” the smoke, rendering it less harmful and less dependent than cigarette smoke.²⁵ While prevalence increases, science lags behind: little is known about harmful effects of the waterpipe on different body organs and whether or not it causes dependence.

The current study was performed to investigate the effect of waterpipe tobacco smoking (WTS) on oral and subgingival microbial flora. Variations were shown in the microbial profile of waterpipe smokers as compared to that of non-smokers, with significant differences in the prevalence and abundance of health-compatible organisms.

The limited information concerning the waterpipe effects on health could represent a major reason for its massive spread globally.²⁶ It has been reported that waterpipe smokers inhale similar toxicants to that of cigarettes smoking⁴ including polycyclic aromatic hydrocarbons,⁶ carbon monoxide, heavy metals, and aldehydes.⁵ In addition, WTS has been shown to increase DNA damage in the lymphocytes and buccal mucosa cells of the users.²⁷ Lately, waterpipe tobacco smoking was reported to interfere with respiratory

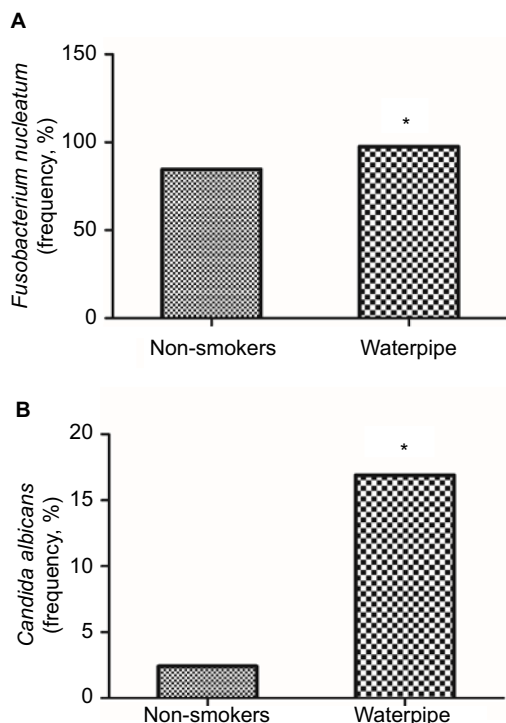


Figure 2 Frequency of some microbes in the waterpipe smokers isolated from the subgingiva of participants. Significant changes in the frequency of (A) *Fusobacterium nucleatum* and (B) *Candida albicans* in the subgingiva of participants. *Indicates significant changes ($p < 0.05$) using the chi-square test.

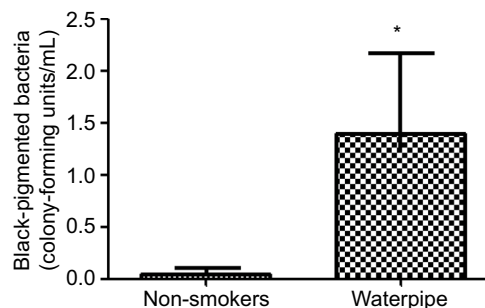


Figure 3 Frequency of some microbes in the waterpipe smokers isolated from the oral cavity of the participants. Significant changes in the frequency of black-pigmented bacteria. *Indicates significant changes ($p < 0.05$) using the chi-square test.

Table 4. Frequency and CFUs of isolates of detected microorganisms from oral cavity of participants

Microorganisms (oral)	Waterpipe frequency, N (%)	Controls frequency, N (%)	p-value, X ² test	Waterpipe CFU, mean ± SD	Controls CFU, mean ± SD	Mann-Whitney U-test p-value
<i>Fusobacterium nucleatum</i>	55 (93.2)	40 (97.6)	0.327	0.42 ± 0.43	6.44 ± 36.4	0.20
Black-pigmented bacteria	40 (63.5)	18 (48.7)	0.15	1.4 ± 0.7	0.05 ± 0.04	0.001
<i>Campylobacter</i> spp.	57 (96.1)	38 (92.7)	0.38	0.60 ± 0.74	0.72 ± 0.74	0.38
Viridans group streptococci	59 (100)	39 (95.1)	0.087	37.6 ± 30.4	36.9 ± 39.0	0.69
<i>Acinetobacter</i> and <i>Moraxella</i>	4 (6.8)	4 (9.8)	0.59	0.26 ± 0.30	0.13 ± 0.25	0.49
<i>Enterobacteriaceae</i>	15 (25.4)	4 (9.8)	0.50	0.01 ± 0.01	0.05 ± 0.08	0.06
<i>Staphylococcus aureus</i>	5 (8.5)	4 (9.8)	0.83	0.01 ± 0.01	0.01 ± 0.01	0.29
Yeast (<i>Candida albicans</i>)	6 (10.2)	6 (14.6)	0.50	0.01 ± 0.01	0.01 ± 0.02	0.31

Note: Significant p-value <0.05 is shown in bold.

Abbreviations: CFU, colony-forming units; SD, standard deviation.

and vascular functions,²⁸ and to enhance oral diseases.¹³ The results reported in the current study showing changes in oral microflora are in accordance with the above findings. This is particularly true because most of the bacteria that appeared or were enhanced among waterpipe smokers, can be pathogenic to human.^{29,30}

Current results indicated the appearance of *Acinetobacter* and *Moraxella* species in subgingiva of waterpipe smokers. Both of these bacterial species are a common cause of human respiratory diseases. For example, a common *Acinetobacter* species is *Acinetobacter baumannii*, which can cause community-acquired pneumonia.^{29,31} Another common *Moraxella* species is *Moraxella catarrhalis*, which causes upper and lower respiratory infections, sinusitis and otitis media in susceptible humans.^{32,33} Notably, for both former mentioned species, tobacco consumption is a listed risk factor for infections.²⁹

The results of this study showed increased frequency of *C. albicans* in the subgingiva of waterpipe smokers. *C. albicans* is a part of the normal oral microflora;³⁴ however, any increase in its number will lead to oral *Candida* infection, which is known as oral thrush.³⁰ In fact, it has been reported that cigarette smoking increases the incidence of *Candida* infections in healthy humans.³⁵ Additionally, it was shown that exposure to tobacco smoke enhances the virulence of *C. albicans*.³⁶

In this study, black-pigmented bacteria such as *P. gingivalis* and *P. intermedia* had higher CFUs in isolates from the oral cavity of waterpipe smokers. These species are common in the oral cavity and are associated with periodontitis.³⁷ Another bacterial species that was reduced in the gingival flora of waterpipe smokers was *F. nucleatum*, which is associated with limited pathogenesis.³⁸ In fact, *F. nucleatum* is one of the most abundant anaerobic species in the oral cavity, in both diseased and healthy individuals. It may normally present in the healthy human oral cavity or considered as periodontal pathogenic. *F. nucleatum* proportions may be higher in current cigarette smokers.³⁹ However, in a study from Jordan, *F. nucleatum* was less prevalent among cigarettes smokers.⁴⁰ Notably, toxicant profile of waterpipe smoke has been shown to be different from that of cigarette smoke in terms of quantity and types. This could explain such variations observed in the current study as compared to cigarette literature.

Cigarettes smoking was also found to cause alteration in subgingival and oral bacterial profiles.⁴¹ For example, periodontitis in smokers is associated with greater depletion of beneficial bacteria such as *Veillonella*, *Neisseria*, and *Streptococcus* species. On the other hand, the abundance of harmful

bacteria such as *Parvimonas*, *Campylobacter*, *Treponema*, *Fusobacterium*, and *Bacteroides* was greatly enhanced in smokers.^{41,42} Only a few studies assessed the effect of WTS on periodontal disease using different outcome measurement such as periodontal bone height loss, plaque index, and gingivitis showing a statistically significant association of periodontal disease with WTS.⁴³ This is in support of the findings of the current study.

It is worth mentioning that the waterpipe group showed significantly higher history of oral infection than the control group. This is expected as a previous study showed the presence of a spectrum of pathogenic bacteria in the hoses of waterpipe smokers.⁴⁴ In addition, waterpipe smoking has been shown to be associated with periodontal disease,⁴⁵ and oral cancer.⁴⁶ Moreover, cigarette smoking has also been shown to be associated with oral infection.⁴⁷ Since the sample of the current study was random, the higher history of oral infection in the waterpipe group supported the imbalance in microbial flora induced by this type of smoking.

Public health policy specific to WTS is lacking in many countries.^{48,49} For example, in Jordan legislation requires health warnings on all tobacco products, but these warnings appear on cigarette packs only and not on WTS tobacco or waterpipes. The same is true in the US, where WTS is not yet regulated nationally. The current study highlights the importance of adopting strong policy regarding waterpipes. The knowledge presented in this study might also be used in interventions that target this type of smoking.

One of the limitations of the study was that a good fraction of oral microflora is uncultivable, and cultural techniques cannot differentiate between close bacteria such as *Campylobacter* spp., the use of other cultivation-independent approaches such as DNA hybridization, q-PCR, and 16S rRNA gene sequencing would be valuable in future investigations. Finally, it is recommended in future investigations to associate altered pathogens by waterpipe smoking with oral diseases by examining their resistance to commonly used antibiotics.

Conclusion

The study provided preliminary evidence for the effect of waterpipe smoking on oral microbiota.

Acknowledgments

The abstract of this paper was presented at the 2nd World Congress on Pediatric Care and Pediatric Infectious Diseases and the International conference on Pediatric Care and Pediatric Infectious Diseases as an abstract presentation

with interim findings. The poster's abstract was published in "Poster Abstracts" in *J Infect Dis Ther* 2016, 4:4(Suppl).

This work was supported by a grant from the Deanship of Research at Jordan University of Science and Technology (grant number 152/2012 to MAK and OFK).

Disclosure

The authors report no conflicts of interest in this work.

References

- Shihadeh A. Investigation of mainstream smoke aerosol of the argileh water pipe. *Food Chem Toxicol*. 2003;41(1):143–152.
- Jawad M, Abass J, Hariri A, et al. Waterpipe smoking: prevalence and attitudes among medical students in London. *Int J Tuberc Lung Dis*. 2013;17(1):137–140.
- Maziak W, Taleb ZB, Bahelah R, et al. The global epidemiology of waterpipe smoking. *Tob Control*. 2015;24 (Suppl 1):i3–i12.
- Sepetdjian E, Shihadeh A, Saliba NA. Measurement of 16 polycyclic aromatic hydrocarbons in narghile waterpipe tobacco smoke. *Food Chem Toxicol*. 2008;46(5):1582–1590.
- Shihadeh A, Saleh R. Polycyclic aromatic hydrocarbons, carbon monoxide, "tar", and nicotine in the mainstream smoke aerosol of the narghile water pipe. *Food Chem Toxicol*. 2005;43(5):655–661.
- Shihadeh A, Azar S, Antonios C, Haddad A. Towards a topographical model of narghile water-pipe cafe smoking: a pilot study in a high socioeconomic status neighborhood of Beirut, Lebanon. *Pharmacol Biochem Behav*. 2004;79(1):75–82.
- Cobb C, Ward KD, Maziak W, Shihadeh AL, Eissenberg T. Waterpipe tobacco smoking: an emerging health crisis in the United States. *Am J Health Behav*. 2010;34(3):275–285.
- Haukioja A, Asunta M, Söderling E, Syrjanen S. Persistent oral human papillomavirus infection is associated with smoking and elevated salivary immunoglobulin G concentration. *J Clin Virol*. 2014;61(1):101–106.
- Sherwin GB, Nguyen D, Friedman Y, Wolff MS. The relationship between smoking and periodontal disease. Review of literature and case report. *NY State Dent J*. 2013;79(6):52–57.
- Eggert FM, McLeod MH, Flowerdew G. Effects of smoking and treatment status on periodontal bacteria: evidence that smoking influences control of periodontal bacteria at the mucosal surface of the gingival crevice. *J Periodontol*. 2001;72(9):1210–1220.
- Zambon JJ. Periodontal diseases: microbial factors. *Ann Periodontol*. 1996;1(1):879–925.
- Kononen E, Paju S, Pussinen PJ, et al. Population-based study of salivary carriage of periodontal pathogens in adults. *J Clin Microbiol*. 2007;45(8):2446–2451.
- Bibars AR, Obeidat SR, Khader Y, Mahasneh AM, Khabour OF. The effect of waterpipe smoking on periodontal health. *Oral Health Prev Dent*. 2015;13(3):253–259.
- Warnakulasuriya S. Waterpipe smoking, oral cancer and other oral health effects. *Evid Based Dent*. 2011;12(2):44–45.
- El-Hakim IE, Uthman MA. Squamous cell carcinoma and keratoacanthoma of the lower lip associated with "Goza" and "Shisha" smoking. *Int J Dermatol*. 1999;38(2):108–110.
- Al-Amrah HJ, Aboznada OA, Alam MZ, ElAssouli MZ, Mujallid MI, ElAssouli SM. Genotoxicity of waterpipe smoke in buccal cells and peripheral blood leukocytes as determined by comet assay. *Inhal Toxicol*. 2014;26(14):891–896.
- D'Ercole S, Catamo G, Piccolomini R. Diagnosis in periodontology: a further aid through microbiological tests. *Crit Rev Microbiol*. 2008;34(1):33–41.
- Riep B, Edesi-Neuss L, Claessen F, et al. Are putative periodontal pathogens reliable diagnostic markers? *J Clin Microbiol*. 2009;47(6):1705–1711.
- D'Ercole S, Catamo G, Tripodi D, Piccolomini R. Comparison of culture methods and multiplex PCR for the detection of periodontopathogenic bacteria in biofilm associated with severe forms of periodontitis. *New Microbiol*. 2008;31(3):383–391.
- Clinical and Laboratory Standards Institute (CLSI). *Abbreviated Identification of Bacteria and Yeast, Approved guidelines, 2nd ed.* CLSI document ML35-A2. Wayne, PA: CLSI; 2008.
- Walker CB, Ratliff D, Muller D, Mandell R, Socransky SS. Medium for selective isolation of *Fusobacterium nucleatum* from human periodontal pockets. *J Clin Microbiol*. 1979;10(6):844–849.
- Kamma JJ, Nakou M, Baehni PC. Clinical and microbiological characteristics of smokers with early onset periodontitis. *J Periodontol Res*. 1999;34(1):25–33.
- Primack BA, Sidani J, Agarwal AA, Shadel WG, Donny EC, Eissenberg TE. Prevalence of and associations with waterpipe tobacco smoking among U.S. university students. *Ann Behav Med*. 2008;36(1):81–86.
- Rastam S, Ward KD, Eissenberg T, Maziak W. Estimating the beginning of the waterpipe epidemic in Syria. *BMC Public Health*. 2004;4:32.
- Kandela P. Narghile smoking keeps Arabs in Wonderland. *Lancet*. 2000;356(9236):1175.
- Akl EA, Jawad M, Lam WY, Co CN, Obeid R, Irani J. Motives, beliefs and attitudes towards waterpipe tobacco smoking: a systematic review. *Harm Reduct J*. 2013;10:12.
- Khabour OF, Alsatari ES, Azab M, Alzoubi KH, Sadiq MF. Assessment of genotoxicity of waterpipe and cigarette smoking in lymphocytes using the sister-chromatid exchange assay: a comparative study. *Environ Mol Mutagen*. 2011;52(3):224–228.
- Alomari MA, Khabour OF, Alzoubi KH, Shqair DM, Eissenberg T. Central and peripheral cardiovascular changes immediately after waterpipe smoking. *Inhal Toxicol*. 2014;26(10):579–587.
- Dexter C, Murray GL, Paulsen IT, Peleg AY. Community-acquired *Acinetobacter baumannii*: clinical characteristics, epidemiology and pathogenesis. *Expert Rev Anti Infect Ther*. 2015;13(5):567–573.
- Hebecker B, Naglik JR, Hube B, Jacobsen ID. Pathogenicity mechanisms and host response during oral *Candida albicans* infections. *Expert Rev Anti Infect Ther*. 2014;12(7):867–879.
- Doughari HJ, Ndakidemi PA, Human IS, Benade S. The ecology, biology and pathogenesis of *Acinetobacter* spp.: an overview. *Microbes Environ*. 2011;26(2):101–112.
- de Vries SP, Bootsma HJ, Hays JP, Hermans PW. Molecular aspects of *Moraxella catarrhalis* pathogenesis. *Microbiol Mol Biol Rev*. 2009;73(3):389–406.
- Perez Vidakovic ML, Riesbeck K. Virulence mechanisms of *Moraxella* in the pathogenesis of infection. *Curr Opin Infect Dis*. 2009;22(3):279–285.
- Cannon RD, Chaffin WL. Oral colonization by *Candida albicans*. *Crit Rev Oral Biol Med*. 1999;10(3):359–383.
- Muzurovic S, Hukic M, Babajic E, Smajic R. The relationship between cigarette smoking and oral colonization with *Candida* species in healthy adult subjects. *Med Glas (Zenica)*. 2013;10(2):397–399.
- Alanazi H, Semlali A, Perraud L, Chmielewski W, Zakrzewski A, Rouabhia M. Cigarette smoke-exposed *Candida albicans* increased chitin production and modulated human fibroblast cell responses. *Biomed Res Int*. 2014;2014:963156.
- Takeuchi K, Furuta M, Takeshita T, et al. Serum antibody to *Porphyromonas gingivalis* and periodontitis progression: the Hisayama Study. *J Clin Periodontol*. 2015.
- Parahitiyawa NB, Scully C, Leung WK, Yam WC, Jin LJ, Samaranyake LP. Exploring the oral bacterial flora: current status and future directions. *Oral Dis*. 2010;16(2):136–145.
- Eick S, Ramseier CA, Rothenberger K, Bragger U, Buser D, Salvi GE. Microbiota at teeth and implants in partially edentulous patients. A 10-year retrospective study. *Clin Oral Implants Res*. 2016;27(2):218–225.
- Karasneh JA, Al Habashneh RA, Marzouka NA, Thornhill MH. Effect of cigarette smoking on subgingival bacteria in healthy subjects and patients with chronic periodontitis. *BMC Oral Health*. 2017;17(1):64.

41. Brook I. The impact of smoking on oral and nasopharyngeal bacterial flora. *J Dent Res*. 2011;90(6):704–710.
42. Shchipkova AY, Nagaraja HN, Kumar PS. Subgingival microbial profiles of smokers with periodontitis. *J Dent Res*. 2010;89(11):1247–1253.
43. Natto S, Baljoon M, Abanny A, Bergstrom J. Tobacco smoking and gingival health in a Saudi Arabian population. *Oral Health Prev Dent*. 2004;2(4):351–357.
44. Masadeh MM, Hussein EI, Alzoubi KH, Khabour O, Shakhatreh MA, Gharaibeh M. Identification, characterization and antibiotic resistance of bacterial isolates obtained from waterpipe device hoses. *Int J Environ Res Public Health*. 2015;12(5):5108–5115.
45. Obeidat SR, Khabour OF, Alzoubi KH, et al. Prevalence, social acceptance, and awareness of waterpipe smoking among dental university students: a cross sectional survey conducted in Jordan. *BMC Res Notes*. 2014;7:832.
46. Waziry R, Jawad M, Ballout RA, Al Akeel M, Akl EA. The effects of waterpipe tobacco smoking on health outcomes: an updated systematic review and meta-analysis. *Int J Epidemiol*. 2017;46(1):32–43.
47. Chen PC, Pan CC, Kuo C, Lin CP. Risk of oral nonmalignant lesions associated with human papillomavirus infection, betel quid chewing, and cigarette smoking in Taiwan: an integrated molecular and epidemiologic study. *Arch Pathol Lab Med*. 2006;130(1):57–61.
48. Jawad M, El Kadi L, Mugharbil S, Nakkash R. Waterpipe tobacco smoking legislation and policy enactment: a global analysis. *Tob Control*. 2015;24 Suppl 1:i60–i65.
49. Ward KD, Siddiqi K, Ahluwalia JS, Alexander AC, Asfar T. Waterpipe tobacco smoking: The critical need for cessation treatment. *Drug Alcohol Depend*. 2015;153:14–21.

International Journal of General Medicine

Publish your work in this journal

The International Journal of General Medicine is an international, peer-reviewed open-access journal that focuses on general and internal medicine, pathogenesis, epidemiology, diagnosis, monitoring and treatment protocols. The journal is characterized by the rapid reporting of reviews, original research and clinical studies across all disease areas.

Submit your manuscript here: <https://www.dovepress.com/international-journal-of-general-medicine-journal>

Dovepress

The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.