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Biomimetic Polyurethane/TiO_2 Nanocomposite Scaffolds Capable of Promoting Biomineralization and Mesenchymal Stem Cell Proliferation

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Abstract

Scaffolds with extracellular matrix-like fibrous morphology, suitable mechanical properties, biomineralization capability, and excellent cytocompatibility are desired for bone regeneration. In this work, fibrous and degradable poly(ester urethane)urea (PEUU) scaffolds reinforced with titanium dioxide nanoparticles ($nTiO_2$) were fabricated to possess these properties. To increase the interfacial interaction between PEUU and $nTiO_2$, poly(ester urethane) (PEU) was grafted onto the $nTiO_2$. The scaffolds were fabricated by electrospinning and exhibited fiber diameter of <1 μm. SEM and EDX mapping results demonstrated that the PEU modified $nTiO_2$ was homogeneously distributed in the fibers. In contrast, severe agglomeration was found in the scaffolds with unmodified $nTiO_2$. PEU modified $nTiO_2$ significantly increased Young's modulus and tensile stress of the PEUU scaffolds while unmodified $nTiO_2$ significantly decreased Young's modulus and tensile stress. The greatest reinforcement effect was observed for the scaffold with 1:1 ratio of PEUU and PEU modified $nTiO_2$. When incubating in the simulated body fluid over an 8-week period, biomineralization was occurred on the fibers. The scaffolds with PEU modified $nTiO_2$ showed the highest Ca and P deposition than pure PEUU scaffold and PEUU scaffold with unmodified $nTiO_2$. To examine scaffold cytocompatibility, bone marrow-derived mesenchymal stem cells were cultured on the scaffold. The PEUU scaffold with PEU modified $nTiO_2$ demonstrated significantly higher cell proliferation compared to pure PEUU scaffold and PEUU scaffold with unmodified $nTiO_2$. The above results demonstrates that the developed fibrous nanocomposite scaffolds have potential for bone tissue regeneration.

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Keywords

nanocomposite; polyurethane; TiO_2 nanoparticle; mesenchymal stem cells; biocompatibility

1. Introduction

Self-regeneration of critical size bone defects caused by trauma, tumor removal, and infection remains challenging in clinical settings. [1–3] Scaffolds have been widely used to aid the regeneration. A typical scaffold should have appropriate porosity to allow cell ingrowth, be osteoconductive, and possess suitable mechanical properties. [1–3] Among different types of scaffolds, those mimicking the properties of bone tissue extracellular matrix (ECM) have been considered as promising candidates.[4–7] These scaffolds can accelerate the regeneration by preventing fibrous encapsulation, promoting osseointegration, and stimulating cell infiltration, proliferation and osteogenic differentiation. [4–8]

Bone is a hard tissue that also has high toughness and tensile strength. To fabricate scaffolds with suitable toughness, flexible polymers such as polyurethane and polycaprolactone can be used. [9–44] The resulting scaffolds generally have higher toughness than those based on stiffer polymers such as polylactide and polyglycolide. Biodegradable polyurethane is a class of polymer that has attracted great attention in the biomaterials community due to its excellent biocompatibility and robust mechanical properties. Porous thermoplastic and thermoset polyurethane scaffolds have been utilized for bone regeneration in animal and preclinical studies. [9–29] The scaffolds with tailored chemical and mechanical properties can promote osteogenic cells to populate and differentiate within the scaffolds, thus stimulating bone regeneration.[25–29] To further augment the regeneration, growth factors such as BMP-2 and PDGF have been loaded into polyurethane scaffolds.[28, 45–47] One of the limitations for biodegradable polyurethane scaffolds is that their modulus and tensile strength are much lower than those of the bone tissue. Increasing these properties is expected to make polyurethane scaffolds more suitable for bone regeneration. An effective approach is to use stiffer soft segment during the synthesis. For example, replacing polycaprolactone with polyhydroxybutyrate can largely increase polyurethane Young's modulus and tensile strength.[48–52] However, this approach may simultaneously compromise toughness of the polymers.

Polyurethane composite scaffolds may retain toughness of the polyurethane while increasing modulus and tensile strength. Microspheres and nanoparticles can be incorporated into polyurethane scaffolds during the fabrication, such as hydroxyapatite,[14, 53, 54] carbon nanotubes,[55–57] and titanium dioxide (TiO_2). [58, 59] These inorganic materials are much stronger and stiffer than polymers. Compared to microspheres, nanoparticles may better reinforce polyurethanes because of their higher surface area-to-volume ratio.[60] TiO_2 nanoparticles are attractive for polymer reinforcement especially in dental applications.[61–63] These nanoparticles have good biocompatibility and antibacterial property.[61–63] In addition, they can suppress immune response which commonly occurs after scaffold implantation.[64] In this work, we took advantage of these properties to fabricate TiO_2 nanoparticles-reinforced polyurethane scaffolds. A major limitation of using unmodified

TiO_2 nanoparticles to reinforce polymers is the uneven distribution, which compromises the reinforcement effect.[65] In addition, the nanoparticles may readily leach out from the scaffolds when their interactions with polymers are weak. [65] The released nanoparticles may be intaken by cells causing potential damage.[66] To address these limitations, approaches such as surface modification of TiO_2 nanoparticles, [65] and increase of polymer polarity [67] have been developed to augment the physical interactions of the nanoparticles and polymers. In this work, we hypothesized that chemical conjugation of polymers to the TiO_2 nanoparticles can better increase the interactions than simply modifying either the nanoparticles or polymers, thus efficiently increasing scaffold modulus and tensile strength, and decreasing nanoparticle release.

Bone tissue ECM is a nanocomposite consisting of collagen fibers and hydroxyapatite nanoparticles. Thus, scaffolds with fibrous morphology and biomineralization capability are desired for bone regeneration. To fabricate scaffolds with fibrous morphology, commonly used techniques include thermally induced phase separation,[68–71] and electrospinning. [72–75] Thermally induced phase separation technique generates fibers with diameter ranging from few to 100 nm depending on the phase separation temperature and solution concentration. [68–71] Electrospinning of polymer solution is a more convenient approach to fabricate fibrous scaffolds. The resulting scaffolds typically have fiber diameters in the range of 10–1000 nm, within the range of fibrous ECM. [72–75] A major advantage of electrospinning is that reinforcement nanoparticles can be readily incorporated into the fibers during fabrication by mixing with polymer solutions.[76] In this work, we electrospun polyurethane scaffolds with TiO_2 nanoparticles in the fibers. Previous study demonstrated that TiO_2 surface with nanostructure has the ability to promote apatite formation.[8] It is hypothesized that the TiO_2 nanoparticles impart the scaffolds with biomineralization capability. We investigated the capability of TiO_2 nanoparticles in improving scaffold mechanical properties, promoting biomineralization, and supporting osteogenic cell proliferation.

2. Materials and methods

2.1. Materials

All chemicals were purchased from Sigma-Aldrich unless otherwise stated. Hexamethylene diisocyanate (HMDI) was purified by vacuum distillation. Polycaprolactone (PCL) diol with an average molecular weight of 2000 g/mol, and dimethylolpropionic acid (DMPA) were vacuum dried overnight at 60°C before use. TiO_2 nanoparticles comprised of 50% anatase and 50% rutile crystal forms. The average particle size and purity were 21 nm and 99.9%, respectively. Anhydrous toluene, dimethylformamide (DMF), and isopropanol were used as received.

2.2. Functionalization of TiO_2 nanoparticles with reactive hydroxyl groups

TiO_2 nanoparticles were reacted with DMPA to introduce hydroxyl groups (Scheme 1) following a previous report.[65] DMPA was dissolved in 2-propanol. The nanoparticles were then dispersed in the DMPA solution. After ultrasonic agitation for 5 min, the mixture was reacted at 80°C for 12 h under constant stirring with the protection of nitrogen gas. The

molar ratio of DMPA to $nTiO_2$ was controlled at 2.4. After reaction, the nanoparticles were collected by centrifugation at 10000 rpm, and then washed with methanol for 3 times to remove the unreacted DMPA.

2.3 Synthesis of poly(ester urethane) (PEU) grafted TiO_2, and poly(ester urethane)urea (PEUU)

The PEU grafted TiO_2 nanoparticles (PEU-g-$nTiO_2$) were synthesized by a two-step approach (Scheme 2). In the first step, HMDI and PCL diol were dissolved in a mixture of DMF/toluene at 1:1 volume ratio. The molar ratio of HMDI and PCL diol was 2:1. Stannous octoate was then added. The reaction was conducted at 85°C for 2 h with the protection of nitrogen gas. In the second step, the DMPA functionalized $nTiO_2$ was added to the above solution. The molar ratio of HMDI and the functionalized $nTiO_2$ was 1:1. The reaction was conducted at 80°C for 4 h. The mixture was then centrifuged followed by washing with DMF/toluene for 3 times. The PEU-g-$nTiO_2$ was finally vacuum dried at 40°C. To confirm the conjugation of PEU, the material was characterized by FT-IR.

PEUU was synthesized using PCL as soft segment, and HMDI and putrescine as hard segment following our established protocols.[75, 77] The molar ratio of PCL diol, HMDI and putrescine was controlled at 1/2/1. In brief, PCL diol was dissolved in DMSO to form a solution. HMDI was then added under the protection of nitrogen gas. After addition of stannous octoate, the reaction was conducted at 80°C in an oil bath for 3 h to form prepolymer. The solution was cooled down to room temperature. Putrescine solution in DMSO was then added dropwise to the prepolymer solution for chain extension. The mixture was stirred at room temperature overnight. The polymer solution was precipitated in cold NaCl solution. After immersing in the DI water for 24 h, the polymer was vacuum dried at 60°C.

2.4. Fabrication of fibrous PEUU scaffolds reinforced with PEU-g-$nTiO_2$

The fibrous scaffolds were fabricated by electrospinning. PEUU was dissolved in 1,1,1,3,3,3-hexafluoro-2-propanol (HFIP) to form a 6% solution. PEU-g-TiO_2 was then added to the solution. The mixture was sonicated to allow particles to uniformly distribute in the solution. The ratio of PEUU and PEU-g-TiO_2 was controlled at 1/1, 1/2, and 2/1 wt%, respectively. The mixture was charged at +15 kv. The flow rate was 1 ml/h. The fibers were collected on a rotating mandrel with rotation speed of 1000 rpm, and charged at -10 kv. The resulting scaffolds (abbreviated as PEU-g-TiO_2/PEUU) had a thickness of ~100 μm. Pure PEUU scaffold, and PEUU scaffold with unmodified TiO_2 nanoparticles (ratio of 1:1, abbreviated as $nTiO_2$/PEUU) were also fabricated to serve as controls.

2.5 Characterization of PEUU scaffolds reinforced with PEU-g-$nTiO_2$

Morphology of the scaffolds was characterized by a LEO 1530 scanning electron microscopy (SEM). The bulk composition was analyzed using energy-dispersive x-ray spectroscopy (EDX) attached to the SEM. FT-IR spectra were recorded on a Nicolet Magna-IR 750 spectrometer. To measure mechanical properties, dog bone-shaped die with ~20 mm gauge length and ~2 mm gauge width were used to cut 4–5 specimens from each scaffold. The specimens were immersed in 37°C water for 24 h before test. The tensile testing was

performed on a TestResources 1000R load frame (model 1322) equipped with a 222.4 N load cell and a 37°C water bath. [75, 77] A cross-head speed of 10 mm/min was used.

2.6 Biomineralization of PEUU scaffolds reinforced with PEU-g-nTiO$_2$

PEUU, nTiO$_2$/PEUU, and PEU-g-nTiO$_2$/PEUU scaffolds were used for the assessment of biomineralization property. The samples were weighted and then immersed in a simulated body fluid (SBF) at 37°C. SBF was prepared by dissolving 10.806 g NaCl, 0.852 g Na_2CO_3, 1.008 g $NaHCO_3$, 0.144 g Na_2SO_4, 0.450 g KCl, 0.351 g K_2HPO_4, 0.622 g $MgCl_2{\cdot}6H_2O$, 200 ml of 0.2 M NaOH solution, and 0.586 g $CaCl_2$ and 34.784 g HEPES in 1 L of DI water. [78] The inorganic ion concentrations in SBF were equal to those of human blood plasma. [78] After 1, 2, 4, and 8 weeks of incubation, samples (n=5 for each scaffold type at each time point) were collected, freeze dried, and weighted. Weight change was then quantified. To confirm the biomineralization, EDX was used to characterize the scaffolds.

2.7. Mesenchymal stem cell growth on PEUU scaffolds reinforced with PEU-g-nTiO$_2$

To evaluate the ability of PEUU scaffolds reinforced with PEU-g-nTiO$_2$ to support cell growth, rat bone marrow-derived mesenchymal stem cells (MSCs) were seeded on the scaffolds. PEUU and nTiO$_2$/PEUU scaffolds were used as controls. MSCs were cultured in a T-175 flask using Dulbecco's Modified Eagle Medium (DMEM) supplemented with 20% FBS, 2% l-glutamine and 1% penicillin/streptomycin as culture medium.[79, 80] The scaffolds were punched into 6 mm diameter disks. After sterilizing under UV irradiation for 1 h in a laminar flow hood, the disks were placed in a 96-well tissue culture plate. MSCs were seeded onto each disk at a density of 2×10^5 cells/ml. After 1, 3, and 7 days of culture under normal conditions (21% O_2, 5% CO_2), double-stranded DNA (dsDNA) content of the live cells in each sample was measured using PicoGreen assay (Invitrogen).[79, 80]

2.8. Statistical analysis

One way ANOVA test was utilized for data analysis. Data were presented as mean ± standard deviation. Statistical significance was defined as $p<0.05$.

3. Results and discussion

3.1. Synthesis of PEU-g-nTiO$_2$

TiO_2 nanoparticle was first functionalized with DMPA to introduce hydroxyl groups onto the surface before grafting PEU. These hydroxyl groups can readily react with isocyanate groups. The reaction of DMPA and nTiO$_2$ is occurred between carboxyl groups of DMPA and Ti of nTiO$_2$ by forming bidentate chelating type coordination bonding. [81] To graft PEU onto the functionalized TiO_2 nanoparticles, PEU prepolymer with isocyanate groups was first prepared by reacting PCL diol with HMDI at a molar ratio of 1:2. The use of PCL allows the PEU to be degradable. Successful synthesis of PEU-g-nTiO$_2$ was confirmed by FTIR spectrum that exhibited characteristic peaks of PEU and TiO_2 (Figure 3). The absorption at 643 cm^{-1} is from nTiO$_2$. The carbonyl peak at 1725 cm^{-1} is from urethane group. All of the isocyanate groups in the prepolymer were reacted with hydroxyl groups introduced onto the nTiO$_2$ surface since there is no isocyanate peak at 2265cm^{-1}. Consistent with previous report, [65] the coordination bonding between DMPA and nTiO$_2$ is not

obvious in the spectrum, possibly because the absorption level of Ti–O–C coordination is significantly small compared to that of the bonds in PEU.

3.2. Fibrous nanocomposite scaffold fabrication

Fibrous scaffolds based on PEUU and PEU-g-$nTiO_2$ were fabricated by electrospinning. The PEUU was synthesized using the same soft segment and diisocyanate as PEU. Our previous study demonstrated that this polymer supported the growth of cardiosphere-derived cells. [75] HFIP was used as a solvent for PEUU. The benefit of using this high polarity solvent is that it allowed PEU-g-$nTiO_2$ to evenly and stably suspend in the PEUU solution, thus facilitating the fabrication of fibers with uniform distribution of $nTiO_2$.

The fabricated PEUU scaffold without $nTiO_2$ or PEU-g-$nTiO_2$ assumed smooth fibers with a diameter less than 1 μm (Figure 4a). The scaffolds based on unmodified $nTiO_2$ and PEUU ($nTiO_2$/PEUU) exhibited both fibers and beads (Figure 4b). EDX analysis was performed to determine the distribution of $nTiO_2$. Figure 5 demonstrated that nanoparticles were not uniformly distributed in the scaffolds, and beads were nanoparticle aggregation. It is likely that unmodified $nTiO_2$ aggregated during the fabrication due to poor interactions between $nTiO_2$ and PEUU. The modification of $nTiO_2$ with PEU (PEU-g-$nTiO_2$) increased the interfacial interaction of the nanoparticles and PEUU. This allowed nanoparticles to stably suspend in the solution during scaffold fabrication. As a result, the scaffolds contained only fibers without beads (Figure 4d–e). EDX analysis confirmed that nanoparticles were evenly distributed in the scaffolds (Figure 5). Scaffold fiber morphology was dependent on the ratio of PEU-g-$nTiO_2$ to PEUU. When the ratio was 1:2, the fibers were smooth. The increase of the ratio to 1:1 and 2:1 led to forming more rough fibers (Figure 4e).

3.3 Scaffold mechanical properties

One of the purposes in using $nTiO_2$ is to reinforce the PEUU scaffolds thus increasing both Young's modulus and tensile strength for improved performance during bone regeneration. The pure PEUU scaffold had Young's modulus and tensile strength of 31.8±2.3 and 34.3±0.9, respectively. Simply mixing unmodified $nTiO_2$ and PEUU ($nTiO_2$/PEUU scaffold) did not show reinforcement effect. Instead, both Young's modulus and tensile strength were significantly decreased compared with PEUU scaffold ($p<0.001$). This is likely due to the low interfacial interaction between $nTiO_2$ and PEUU. It led to the aggregation of $nTiO_2$ in the solution during the scaffold fabrication process. The scaffolds therefore had $nTiO_2$ aggregates attached to the fibers (Figure 4b and Figure 5a), which cannot efficiently dissipate external force. For those nanoparticles that are in the fibers, even they can dispense uniformly, the weak interfacial interaction between PEUU cannot effectively reinforce the scaffold.

Modification of $nTiO_2$ with PEU (PEU-g-$nTiO_2$) can increase its interfacial interaction with PEUU because of the strong hydrogen bonding between urethane groups and urethane-urea groups in both polymers. This resulted in the reinforcement effect. Figure 6 demonstrated that adding PEU-g-$nTiO_2$ into the PEUU scaffolds significantly increased Young's modulus compared to the pure PEUU scaffold when the ratio of PEU-g-$nTiO_2$ and PEUU was ranged from 1:2 to 2:1 ($p<0.01$). The highest Young's modulus was for the scaffold with the ratio of

1:1 where it was 48.8±3.2 MPa, a 53.5% of increase over pure PEUU scaffold. For the tensile strength, the scaffolds with PEU-g-$nTiO_2$/PEUU ratios of 1:2 and 1:1 were significantly higher than pure PEUU scaffold ($p<0.05$) while the scaffold with the ratio of 2:1 showed similar value. The decrease of Young's modulus and tensile strength when the PEU-g-$nTiO_2$/PEUU ratio was increased from 1:1 to 2:1 is probably attributed to the decrease of interfacial interaction between PEUU and PEU when the content of PEU-g-$nTiO_2$ is higher than PEUU. It is also possible that PEU-g-$nTiO_2$ cannot efficiently distribute in the PEUU fibers when its content is high. Similar trend was found for collagen scaffolds reinforced with carbon nanotubes.[82]

3.4 Scaffold biomineralization

Biomineralization is critical during bone regeneration. Scaffolds capable of stimulating biomineralization may be able to promote the regeneration. [8] The developed fibrous scaffolds have high surface-area-to-volume ratio, thus may facilitate the biomineralization. To investigate the biomineralization capability of TiO_2 nanoparticles reinforced PEUU scaffold, PEU-g-$nTiO_2$/PEUU=1:1 was used since it had the highest Young's modulus. The scaffold was incubated in SBF for 8 weeks. Controls were PEUU and $nTiO_2$/PEUU scaffolds. All scaffolds showed slight weight loss after 1 week of incubation ($p>0.05$ for each scaffold). It is possible that PEUU degradation-induced weight loss is greater than biomineralization-induced weight gaining during this period. The PEUU scaffold exhibited continuous weight loss for 4 weeks. Significant net weight gaining was observed only after 8 weeks ($p<0.05$, week 4 vs. week 8), indicating that biomineralization was dominated for PEUU after 4 weeks. The $nTiO_2$/PEUU scaffold demonstrated net weight gaining after 4 weeks, earlier than PEUU scaffold. After 8 weeks of incubation, the scaffold gained ~13% of weight, significantly greater than the original weight ($p<0.05$, week 0 vs. week 8). These results suggest that TiO_2 nanoparticles in the scaffold accelerated the biomineralization. This is consistent with previous studies where TiO_2 containing materials promoted the absorption of Ca^{2+} and PO_4^{3-}. [83]

The PEU-g-$nTiO_2$/PEUU scaffold showed net weight gaining only after 2 weeks of incubation, sooner than $nTiO_2$/PEUU scaffold. After 8 weeks, the net weight gaining was 10% ($p<0.05$ for weight of week 0 vs. weight of week 8). The Ca and P containing ions were deposited uniformly in the PEU-g-$nTiO_2$/PEUU scaffold (Figure 8), attributing to the even distribution of TiO_2 nanoparticles (Figure 5). Elemental content of Ca and P analyzed from EDX mapping is listed in Table 1. Consistent with weight change results in Figure 7, the PEU-g-$nTiO_2$/PEUU scaffold had the highest Ca and P deposition. In addition, the $nTiO_2$/PEUU scaffold demonstrated greater Ca and P deposition than PEUU scaffold. The above results suggest that PEU modification stimulated $nTiO_2$ biomineralization. The PEU grafted onto the nanoparticles is based on polycaprolactone and HMDI. The chain length of PEU should be shorter than that of PEUU as no chain extension reaction was performed for it. Therefore, PEU possibly degraded faster than PEUU. The hydrolysis of PCL chain may leave −COOH groups on the $nTiO_2$ surface, which then attract cationic species like Ca^{2+} to deposit.

3.5 Mesenchymal stem cell growth on scaffolds

Scaffolds for bone regeneration can be implanted alone to allow endogenous cells including osteoblasts and stem cells to induce regeneration. They can also be transplanted together with osteogenic cells to direct the regeneration. In both approaches, it is necessary for the scaffolds to support cell proliferation. To investigate the capability of PEU-g-nTiO_2/PEUU scaffold to supporting cell growth, bone marrow-derived MSCs were seeded on the scaffold of PEU-g-nTiO_2/PEUU=1:1 since it had the highest Young's modulus. PEUU and nTiO_2/PEUU scaffolds were used as controls. Bone marrow-derived MSCs are known for their ability to promote bone regeneration. Cell dsDNA (for live cells) content was monitored during the culture. Figure 9 demonstrated that MSC dsDNA content was increased on all 3 scaffolds during the 7-day culture period. The highest increase was found for PEU-g-nTiO_2/PEUU scaffold where dsDNA content was 2.5 and 9.9 folds of day 1 at days 3 and 7, respectively ($p<0.01$ for day 3 vs. day 1, and day 7 vs. day 3). The nTiO_2/PEUU scaffold and PEUU scaffold exhibited similar levels of dsDNA increase at day 7 ($p<0.05$ for day 7 vs. day 1 for both scaffolds). The above results demonstrate that incorporation of PEU modified TiO_2 nanoparticles into PEUU scaffold improved MSC proliferation while incorporation of unmodified TiO_2 nanoparticles did not. It is possible that PEU on the nanoparticle surface augmented its hydrophilicity, thus increasing its interaction with cells. Our future work will explore how scaffold properties such as TiO_2 content, fiber diameter, and single fiber modulus can be tuned to induce the differentiation of MSCs into osteogenic phenotype.

One of the concerns for using TiO_2 nanoparticles is that the released nanoparticles may be toxic to bone cells. [66] For example, TiO_2 nanoparticles with size of 15 nm have been shown to impair SOD1 and SOD2 secretion and promote ROS generation after intaking by osteoblasts.[84] TiO_2 nanoparticles can also change the ultrastructure of cells. [84] In this work, the PEU modified TiO_2 nanoparticles may not be readily released from the PEUU fibers due to increased interaction between the nanoparticles and PEUU. In addition, the PEU modified TiO_2 nanoparticles may not be easily intaken by the cells even after PEU and PEUU are degraded. Biomineralization study demonstrated that the modified nanoparticles promoted mineral deposition, which can increase the size of the nanoparticles to an extent that cells cannot readily intake.

4. Conclusion

Fibrous PEUU scaffolds reinforced with TiO_2 nanoparticles were fabricated for bone regeneration. Unmodified TiO_2 nanoparticles cannot uniformly distribute in the fibers, and did not show reinforcement effect. The PEU modified nanoparticles can evenly distribute in the fibers, and significantly increased scaffold Young's modulus and tensile strength. The scaffolds based on modified TiO_2 nanoparticles and PEUU exhibited greater biomineralization capability than PEUU scaffold. In addition, these scaffolds better promoted MSC growth than pure PEUU scaffold and PEUU scaffold with unmodified TiO_2 nanoparticles. These scaffolds alone or combined with osteogenic cells have the potential for bone regeneration.

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Highlights

- Grafted degradable polyurethane into TiO_2 nanoparticles;
- Modified TiO_2 nanoparticles reinforced fibrous polyurethane scaffolds;
- Reinforced scaffolds promoted biomineralization and stem cell proliferation.

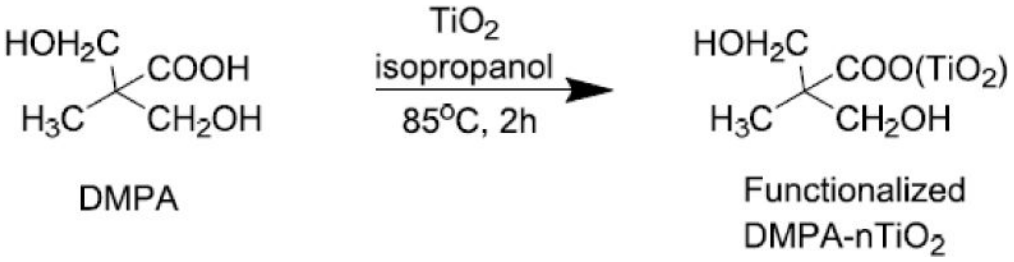

Figure 1.
Synthesis of DMPA functionalized TiO_2 nanoparticles (DMPA-nTiO_2).

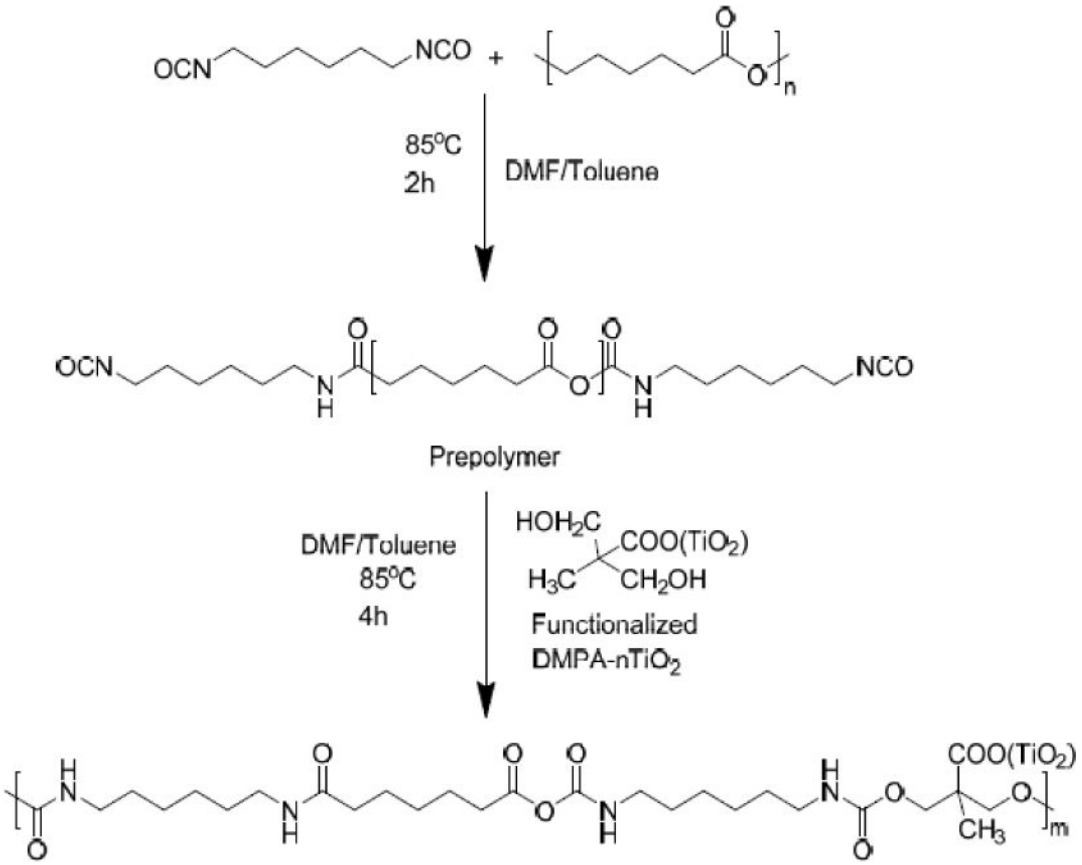

Figure 2.
Synthesis of degradable polyurethane conjugated with TiO_2 nanoparticles (PEU-g-nTiO_2).

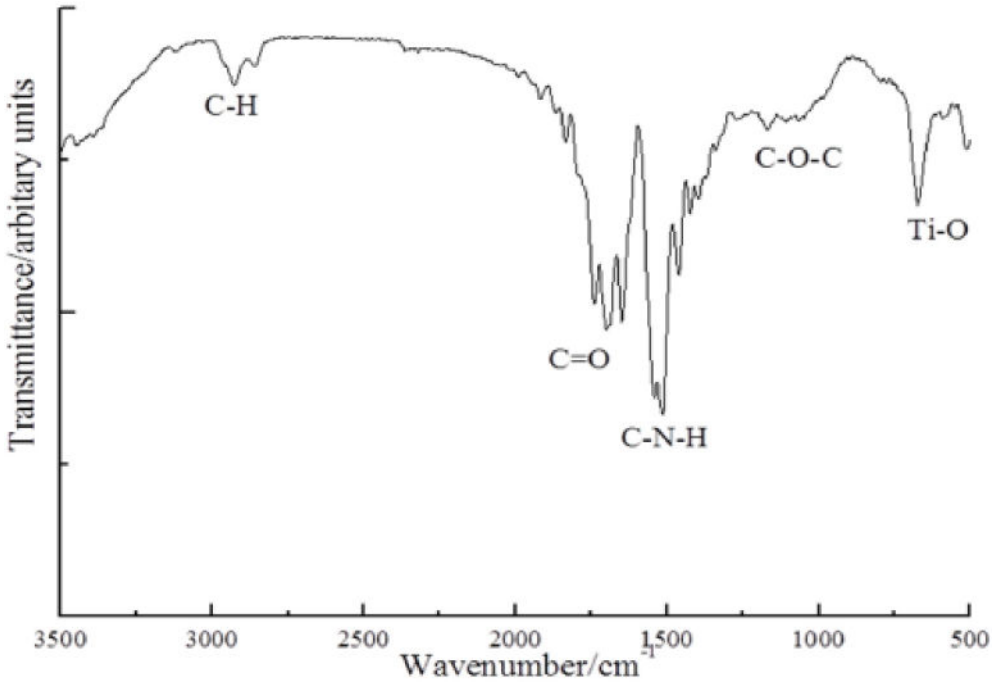

Figure 3.
FTIR spectrum of synthesized PEU-g-nTiO_2.

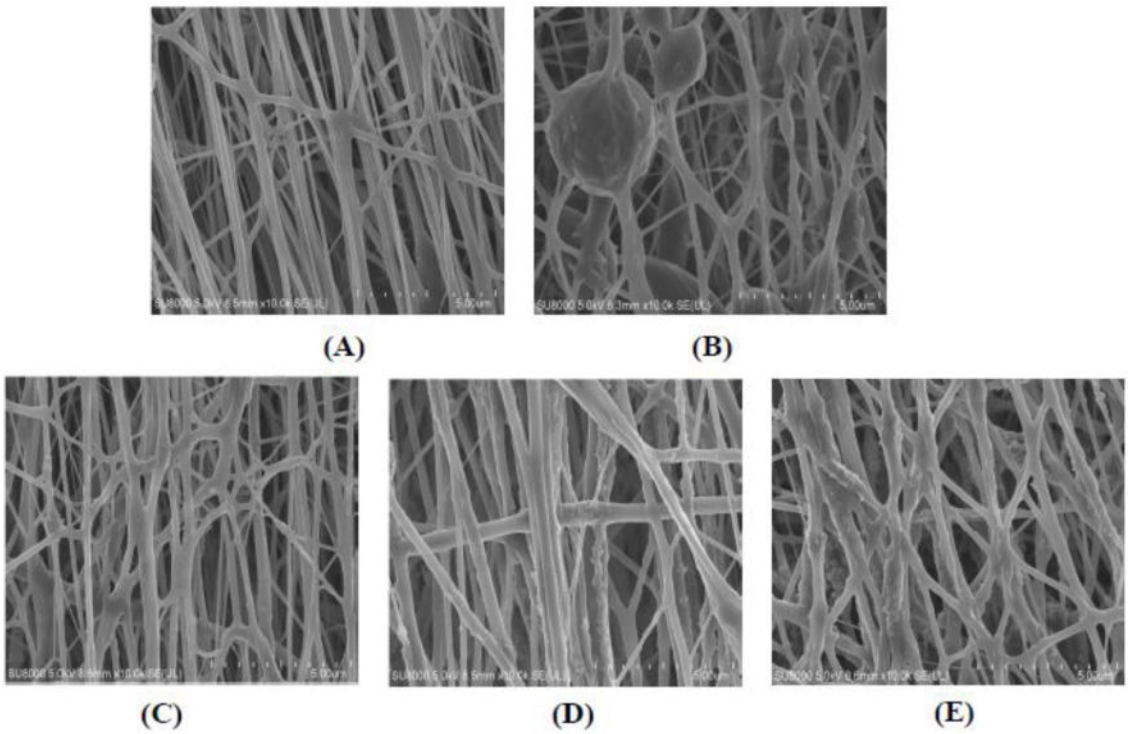

Figure 4.

Morphology of the electrospun scaffolds. (A) PEUU; (B) $nTiO_2$/PEUU; (C) PEU-g–$nTiO_2$/PEUU=1:2; (D) PEU-g–$nTiO_2$/PEUU=1:1; and (E) PEU-g–$nTiO_2$/PEUU=2:1.

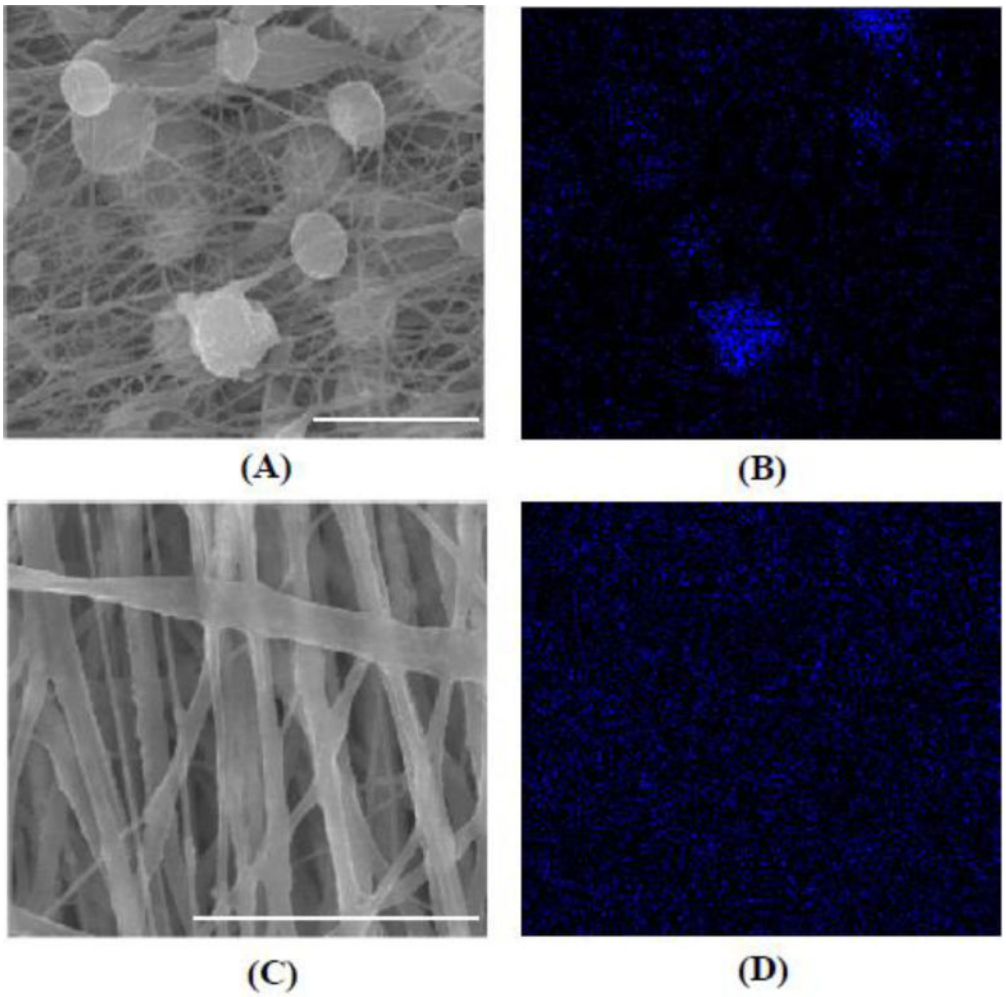

Figure 5.
SEM images (A, C) and corresponding EDX analysis of Ti distribution in the scaffolds (B, D). (A, B) $nTiO_2$/PEUU; (C, D) PEU-g–$nTiO_2$/PEUU=2/1. Scale bar = 5 μm.

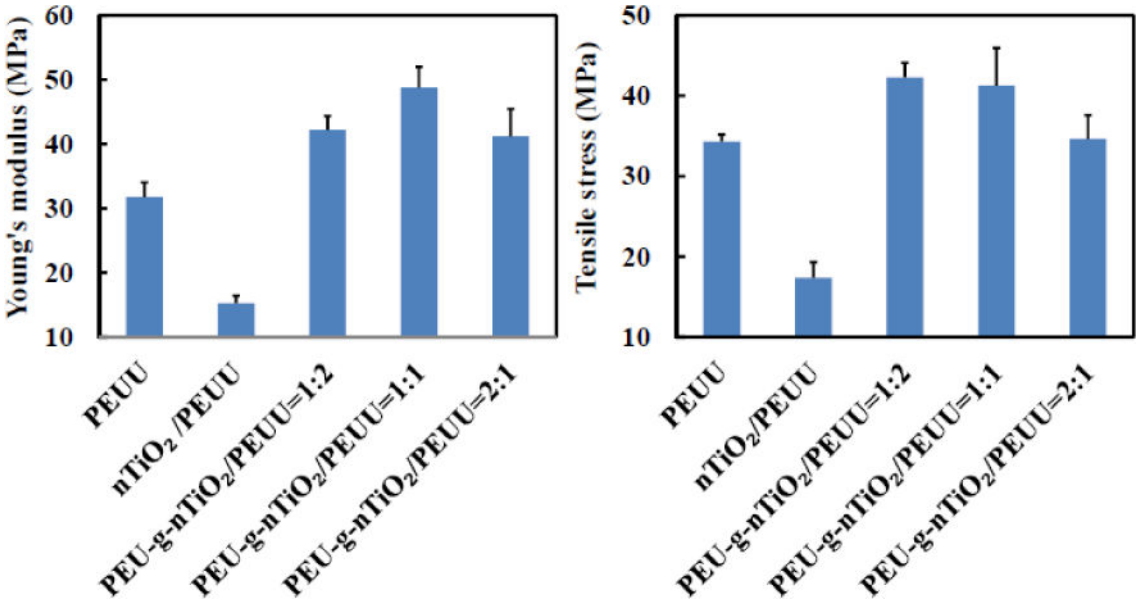

Figure 6.

Mechanical properties of electrospun scaffolds with and without reinforcement with PEU-gnTiO_2.

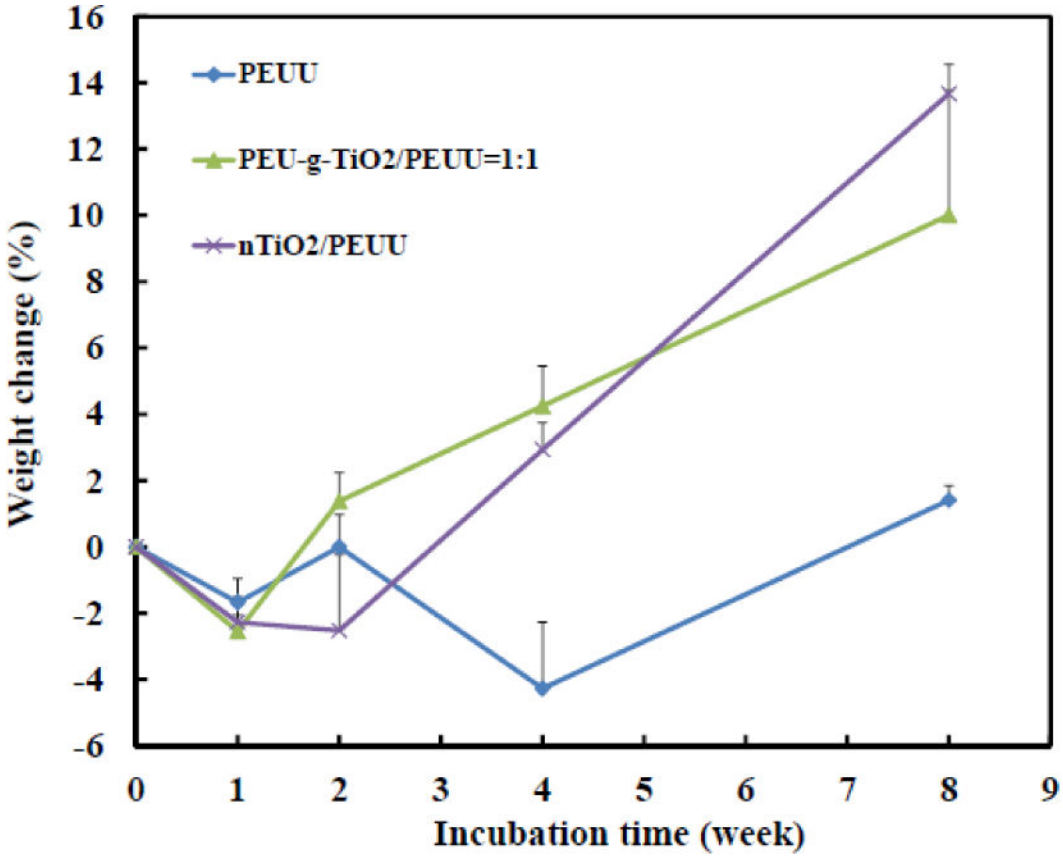

Figure 7.
Weight change of scaffolds incubated in the 37°C simulated body fluid for 8 weeks.

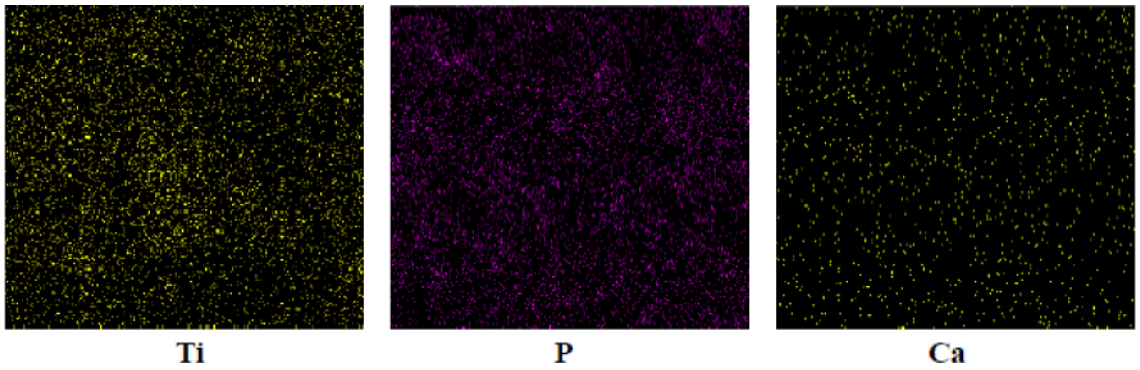

Figure 8.
EDX characterization of Ca and P deposition in the scaffold PEU-g-nTiO_2/PEUU = 1:1.

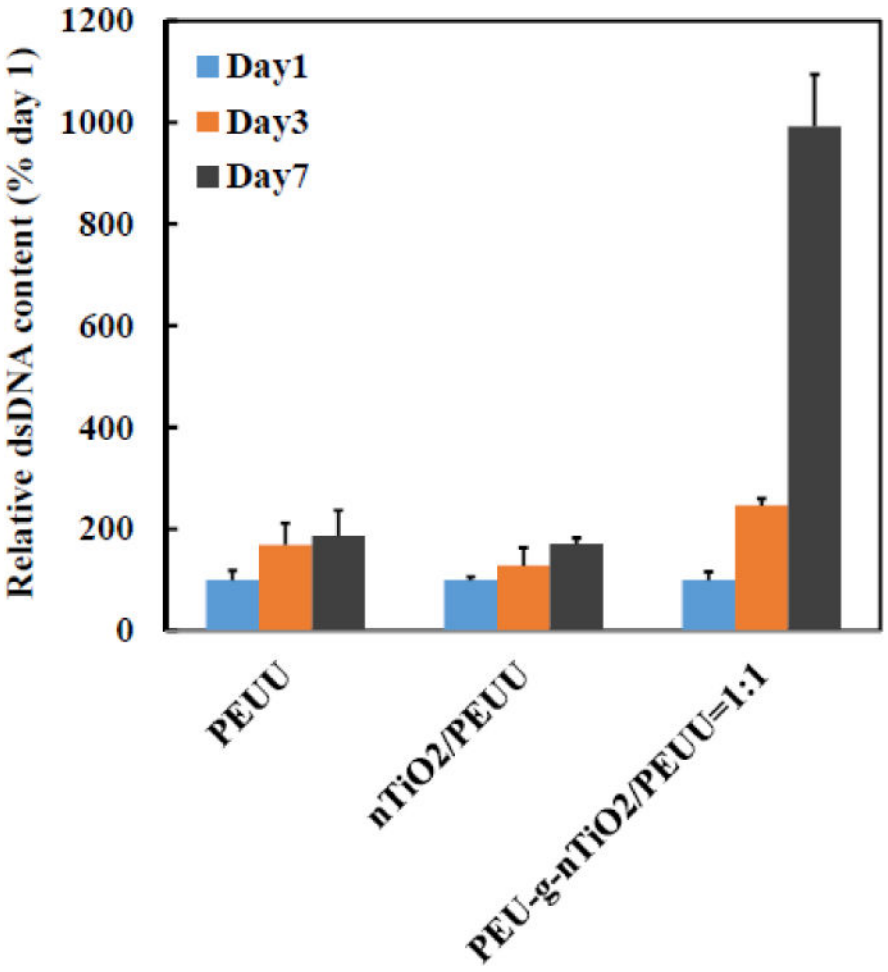

Figure 9.
Mesenchymal stem cell growth on the scaffolds with and without reinforcement with PEU-g$nTiO_2$.

Table 1

EDX analysis of Ca and P content in the scaffolds after 8 weeks of incubation in SBF.

Element (%)	PEUU	$nTiO_2$/PEUU	PEU-g-TiO_2/PEUU
Ca	0.039	0.103	0.203
P	0.143	0.327	1.270