

# Combating Intracellular Pathogens with Repurposed Host-Targeted Drugs

Stanford Schor and Shirit Einav\*

Department of Medicine, Division of Infectious Diseases and Geographic Medicine, and Department of Microbiology and Immunology, School of Medicine, Stanford University, 300 Pasteur Drive, Lane Building Rm L127, Stanford, California 94305, United States

**ABSTRACT:** There is a large, global unmet need for the development of countermeasures to combat intracellular pathogens. The development of novel antimicrobials is expensive and slow and typically focuses on selective inhibition of proteins encoded by a single pathogen, thereby providing a narrow spectrum of coverage. The repurposing of approved drugs targeting host functions required for microbial infections represents a promising alternative. This review summarizes progress and challenges in the repurposing of approved drugs as host-targeted broad-spectrum agents for the treatment of intracellular pathogens. These strategies include targeting both cellular factors required for infection by various viruses, intracellular bacteria, and/or protozoa as well as factors that modulate the host immune response to these microbial infections. The repurposed approach offers complementary means to develop therapeutics against existing and emerging intracellular microbial threats.

There is an urgent need for new strategies to combat hundreds of human disease-causing intracellular pathogens, including viruses, intracellular bacteria, and protozoa. Currently approved antiviral drugs treat fewer than ten viral infections. While antibacterial agents are more widely available, only a minority of them penetrate mammalian cells and achieve a therapeutic level in the appropriate intracellular compartment to treat intracellular bacterial infections.<sup>1</sup> The narrow spectrum of coverage of most antimicrobials that target microbial functions, particularly viruses, is another major limitation. Given the high average cost (over two billion dollars) and long timeline (8–12 years) to develop a new drug, the scalability of targeting microbes individually is limited. Lastly, resistance typically emerges rapidly when conventional drugs that target microbial functions are used as monotherapy.

An attractive solution to overcome these limitations is to repurpose already approved drugs that target host functions required for the lifecycle of intracellular pathogens as antimicrobial agents (Figure 1). Off-label use of approved drugs requires significantly less capital and time and diminishes the clinical risks, because such drugs were already rigorously tested (toxicity, pharmacokinetics, pharmacodynamics dosing, etc.) for their primary indication. Since some host functions are hijacked by various viruses, intracellular bacteria, and/or protozoa, they may represent targets for broad-spectrum antimicrobials. This approach can therefore increase the efficiency of antimicrobial development, improve drug access for patients, and facilitate readiness for future outbreaks of emerging pathogens. Additionally, host-targeted approaches may have the added benefit of a higher genetic barrier to the emergence of resistance.

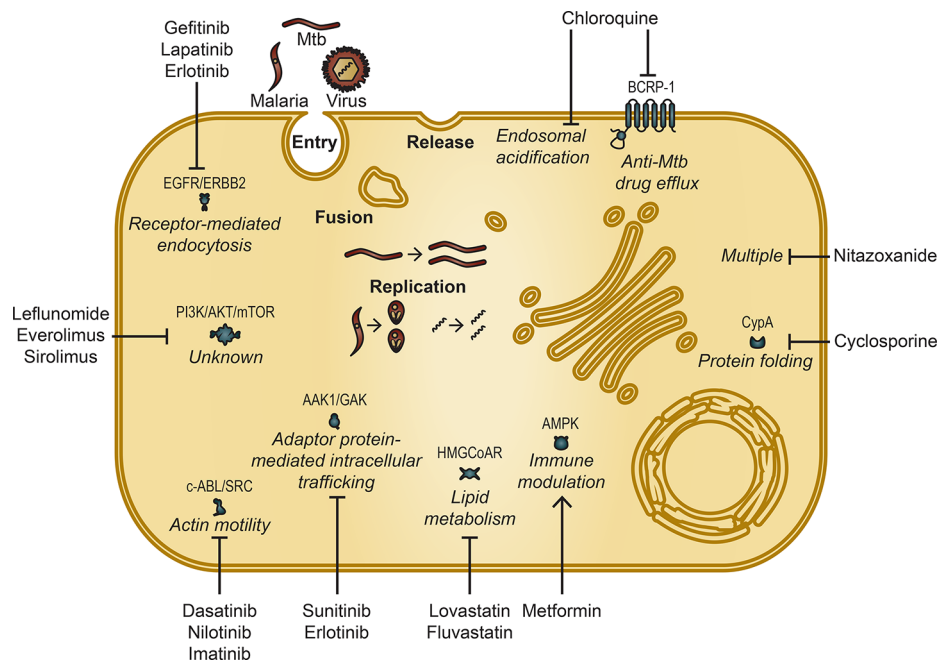
One promising drug category for repurposing as antimicrobials is inhibitors of host kinases, such as those widely approved for the treatment of cancer and inflammatory conditions.<sup>2</sup> A large number of host kinases are co-opted by intracellular pathogens and thus represent attractive targets for broad-spectrum antimicrobial therapy.<sup>3</sup> Indeed, recent efforts

have demonstrated the therapeutic potential of kinase inhibitors to combat various intracellular microbial infections. The anticancer epidermal growth factor receptor (EGFR) and/or ERBB2 kinase inhibitors gefitinib, erlotinib, and lapatinib demonstrate *in vitro* activity against a number of viruses including hepatitis C virus (HCV), human cytomegalovirus (HCMV), and poxvirus.<sup>4,5</sup> Gefitinib and lapatinib also inhibit HCV and HCMV infections in a mouse and guinea pig model, respectively,<sup>4,5</sup> and reduce replication of *Mycobacterium tuberculosis* (Mtb) in the lungs of infected mice.<sup>6</sup> The antiviral effect of these drugs is mediated in part by inhibition of signaling in the entry stage and/or in F-actin-mediated intracellular viral trafficking.<sup>7,8</sup> Enhanced antimicrobial immune effect via inhibition of EGFR/p38 mitogen associated protein kinase signaling plays a role in their anti-Mtb activity.<sup>6</sup>

Another class of approved kinase inhibitors demonstrating a broad antimicrobial coverage is the cellular Abelson tyrosine kinase (c-Abl) inhibitors, such as imatinib and nilotinib. These drugs inhibit replication of Ebola virus (EBOV), dengue virus (DENV), Middle Eastern respiratory syndrome coronavirus (MERS-CoV), and severe acute respiratory syndrome coronavirus (SARS-CoV) in cultured cells and reduce vaccinia virus load, spread, and overall mortality in a mouse model.<sup>9–12</sup> Moreover, imatinib treatment impairs entry and intracellular survival of Mtb and *Mycobacterium marinum* in infected macrophages and reduces the bacterial load and associated pathologies in mice infected with antibiotic-sensitive or resistant Mtb.<sup>13</sup> Notably, when coadministered with rifamycins, imatinib displays synergy, indicating its potential utility in combination anti-Mtb drug regimens.<sup>13</sup> The molecular targets underlying the antimicrobial effect of c-Abl inhibitors remain to be validated. Moreover, while inhibition of actin motility is proposed as a mechanism of antiviral action of these drugs,<sup>12</sup>

Received: December 18, 2017

Published: January 3, 2018



**Figure 1.** Approved host-targeted drugs can be repurposed as countermeasures against intracellular pathogens including viruses, intracellular bacteria such as *Mtb*, and protozoa such as malaria. Depicted are representative stages of the intracellular pathogen lifecycle that are often targeted by antimicrobials. Shown are examples of repurposed drugs and the relevant corresponding pathway and target protein(s), if known. Only targets that are validated as either antimicrobial and/or molecular targets underlying the antimicrobial effect are shown. CypA (Cyclophilin A). Blunt arrows represent inhibition, whereas pointed arrow represents activation.

the precise mechanism and mode of antibacterial action remain unknown.

The approved anticancer drugs sunitinib and erlotinib display potent activity against viruses from six viral families, including DENV and EBOV in cultured cells.<sup>14</sup> Combinations of these drugs reduce morbidity and mortality of dengue and Ebola infected mice and are advancing into clinical studies for dengue and possibly Ebola in future outbreaks (<https://clinicaltrials.gov/NCT02380625>).<sup>14</sup> Inhibition of the cellular kinases adaptor protein 2 (AP2)-associated protein kinase 1 (AAK1) and cyclin G-associated kinase (GAK), regulators of adaptor protein-mediated intracellular viral trafficking, is an important mechanism of action (MOA) of this antiviral approach.<sup>14</sup>

Three approved immunosuppressive drugs that inhibit the phosphatidylinositol 3'-kinase–Akt–mammalian target of rapamycin (PI3K/AKT/mTOR) pathway, leflunomide, everolimus, and sirolimus, reduce viremia and nephropathy progression in patients infected with BK virus, a member of the polyomavirus family, when used individually or in combination.<sup>15</sup> Additionally, leflunomide demonstrates activity against HCMV both *in vitro* and in a rat model.<sup>16</sup> While direct suppression of viral replication is proposed as one underlying mechanism of this approach, the precise MOA and molecular targets remain unknown.

Inhibitors of cyclophilin, a host factor that facilitates protein folding, represent another class of drugs, beyond kinase inhibitors, that shows promise as antimicrobials. Cyclosporine, an immunosuppressant cyclophilin inhibitor, disrupts replication of multiple viruses, including HCV, DENV, other flaviviruses, and human immunodeficiency virus (HIV) *in vitro*.<sup>17,18</sup> Two experimental, nonimmunosuppressive cyclophilin inhibitors, alisporivir and SCY-635, also inhibit multiple RNA viruses in cultured cells, yet their effect in mouse models

is variable.<sup>19</sup> Alisporivir significantly reduced HCV viral load in chronically infected patients;<sup>19</sup> however, a clinical hold was placed on the Phase III trial due to toxicity. Cyclosporine also inhibits the erythrocytic stages of *Plasmodium falciparum in vitro* and rodent malaria in a mouse model.<sup>20</sup> Nevertheless, it remains unknown whether this effect results in part from inhibition of the host cyclophilin or solely the parasitic protein.

Another candidate for repurposing is nitazoxanide, a drug approved for the treatment of parasite-induced diarrhea. It inhibits replication of multiple viruses including HCV, flaviviruses, respiratory viruses, HIV, and hepatitis B virus (HBV) in cultured cells.<sup>21</sup> Nitazoxanide blocks maturation of the influenza hemagglutinin, but the target remains unknown. In the case of HCV, it is thought to activate protein kinase R leading to phosphorylation of eukaryotic initiation factor 2 $\alpha$ , which then blocks viral replication.<sup>21</sup> In a phase II trial, nitazoxanide modestly reduced the time to resolution of symptoms of flu and is currently being evaluated in a phase III trial for this indication.<sup>22</sup> Whereas addition of nitazoxanide to peginterferon-ribavirin improved the sustained virologic response in HCV patients in a phase II trial, no such improvement was observed in a phase III trial in genotype 4 infected patients.<sup>23</sup> Additionally, nitazoxanide is effective against clinical *Mtb* isolates, including those that are resistant to standard antituberculosis drugs.<sup>24</sup> Disruption of membrane potential and pH homeostasis in *Mtb* is one proposed MOA.<sup>24</sup> A phase II clinical trial is currently ongoing to determine the efficacy of 14-day nitazoxanide treatment prior to initiation of standard antituberculosis therapy.

Metformin, a drug approved for the treatment of type 2 diabetes, inhibits the intracellular growth of *Mtb* (including drug resistant strains) and enhances the efficacy of approved anti-*Mtb* drugs in cell culture.<sup>25</sup> In a murine model, metformin mildly reduces *Mtb* load in target organs and improves lung

pathology when used individually or in combination drug treatment.<sup>25</sup> Notably, metformin demonstrated efficacy in decreasing disease severity and improving clinical outcome as an adjunct therapy in retrospective cohorts of tuberculosis patients with diabetes and also reduced the incidence of latent tuberculosis in diabetic patients.<sup>25</sup> Activation of the adenosine monophosphate-activated protein kinase (AMPK), which regulates cellular immune functions, is involved in mediating the antibacterial effect.<sup>25</sup> Beyond tuberculosis, a clinical study that aims to evaluate the size of HIV reservoirs upon addition of metformin to antiretroviral regimens is currently ongoing.

Statins, drugs that inhibit 3-hydroxy-3-methylglutaryl coenzyme A reductase (HMG-CoAR), represent another potential class for repurposing as host-targeted antimicrobials. Various statins inhibit replication of HCV and DENV *in vitro*.<sup>26,27</sup> Nevertheless, while several retrospective studies demonstrated that statins reduce relapse rates in patients infected with HCV when added to peginterferon-ribavirin, this effect was quite variable.<sup>27</sup> The mechanism of anti-HCV action of statins is mediated in part by their effect on lipid biosynthesis, as their antiviral activity in cells is reversed upon addition of mevalonate or geranylgeraniol, and resistance to these drugs coincides with an increase in HMG-CoAR level.<sup>27</sup> Due to their ability to restore endothelial stability, statins were also proposed as a strategy to treat infections associated with endothelial dysfunction. While no formal study has been conducted, it was reported that atorvastatin in combination with an angiotensin receptor blocker reduced mortality in 100 Ebola patients in Sierra Leone.<sup>28</sup>

The antimalarial and anti-inflammatory drug chloroquine also shows some promise as an antimicrobial agent. In a nonhuman primate dengue model, chloroquine reduces viremia, cytokine production, and organ damage.<sup>29</sup> However, when studied in dengue patients, it reduced pain and improved daily activity performance, yet it did not reduce disease duration or viremia.<sup>30,31</sup> In mice infected with Zika virus, chloroquine increases the life span and reduces vertical transmission and fetal brain infection.<sup>32</sup> Moreover, it reduces acute lung injury, morbidity, and mortality in a mouse model of avian influenza A H5N1.<sup>33</sup> The mechanism of antiviral activity of chloroquine is multifaceted and thought to involve alterations in the intracellular pH causing impaired fusion and cleavage of the prM protein during flaviviral release and/or inhibition of autophagy.<sup>32,33</sup> Chloroquine may also show utility as an add-on to antituberculosis regimens, as it enhances the intracellular killing of Mtb in human macrophages by inhibiting the BCRP-1 (breast cancer resistance protein-1)-mediated active efflux of the antituberculosis drugs isoniazid and pyrazinamide.<sup>34</sup>

Collectively, these data provide a proof-of-concept for the useful expansion of antimicrobial drug targets by focusing on host functions and for the therapeutic potential of repurposed drugs targeting these functions to combat intracellular pathogens. Further benefit is provided by the higher genetic barrier to resistance associated with many of these host-targeted approaches. Whereas emergence of resistance can complicate treatment with host-targeted approaches, as in the case of the chemokine receptor 5 antagonist maraviroc,<sup>35</sup> the time to resistance with other host-targeted approaches is longer and the level of resistance is lower relative to typical direct-acting antimicrobials monotherapy. This is exemplified by treatment of DENV with sunitinib/erlotinib combinations, Mtb with nitazoxanide, and HCV with cyclophilin inhib-

itors.<sup>14,19,24</sup> Targeting host proteins that are not under the genetic control of microbes and simultaneously inhibiting several host targets with the same drug or drug combination are thought to facilitate this increase in the resistance barrier of these approaches.

Nevertheless, repurposing of host-targeted drugs as antimicrobials faces its own challenges. Cellular proteins function in a complex network of interactions, and their inhibitors are often nonselective. Thus, the mechanism of antimicrobial action of host-targeted approaches often remains elusive, and the molecular targets underlying this effect are unvalidated. For example, erlotinib's effect on HCV infection was first attributed solely to its effect on EGFR, its cancer target, yet it was later demonstrated that inhibition of GAK, another target that is inhibited by erlotinib with a comparable potency to EGFR, also plays a role.<sup>8,14</sup> For other drugs, such as nitazoxanide and chloroquine, the MOA is even less clear and appears to be pathogen specific.<sup>21,24,32–34</sup>

Another challenge is that the antimicrobial effect observed *in vitro* often cannot be reproduced *in vivo*. For example, several multi kinase inhibitors, such as lestaurtinib, display *in vitro* activity against *Plasmodium falciparum* but no significant effect in malaria-infected mice.<sup>36</sup> Moreover, as exemplified by chloroquine for dengue treatment, a promising effect in animal models cannot guarantee an effect in patients.<sup>29–31</sup> Toxicity is another major concern. The Src- and c-Abl-inhibitor dasatinib demonstrates broad-spectrum antiviral activity in cultured cells, yet in a murine model of vaccinia virus, it induces immunosuppression rather than protection.<sup>12,37</sup> It is thus important to investigate the antimicrobial activity and safety of these drugs in animal models of other infections. Nevertheless, for some drugs, it may be feasible to find a therapeutic window where the drug level is sufficient to inhibit microbial replication with minimal cellular toxicity. Shifting from indications requiring long-term therapy (e.g., months to years for cancer) to a shorter duration sufficient to treat most acute infections should help limit toxicity. In contrast, the utility of some of these approaches for treating chronic infections requiring longer duration treatment may be more limited. This is exemplified by the unexpected occurrence of pancreatitis during the Phase III trial with alisporivir in chronically infected HCV patients. Nevertheless, drugs that are extensively used in humans, such as metformin and chloroquine, are more likely to be safe add-ons to standard regimens even for chronic infections.

In summary, these examples provide a proof-of-concept for the potential feasibility of repurposing approved drugs as host-targeted broad-spectrum therapies to combat intracellular microbial pathogens. To effectively meet the clinical needs posed by emerging intracellular pathogens, such approaches may find utility in combination with novel host-targeted approaches and microbial-targeted strategies.

## ■ AUTHOR INFORMATION

### Corresponding Author

\*E-mail: [seinav@stanford.edu](mailto:seinav@stanford.edu).

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

This research was supported by grants from the NIH (1U19 AI10966201), DoD/CDMRP (PRMRP, PR151090), Ameri-

can Cancer Society (RSG-14-11 0-01-MPC), Stanford Bio-X, and Stanford SPARK program to S.E. The authors acknowledge all the contributions in the field that could not be included in the review.

## REFERENCES

- (1) McOrist, S. (2000) Obligate intracellular bacteria and antibiotic resistance. *Trends Microbiol.* 8, 483–486.
- (2) Gross, S., Rahal, R., Stransky, N., Lengauer, C., and Hoeflich, K. P. (2015) Targeting cancer with kinase inhibitors. *J. Clin. Invest.* 125, 1780–1789.
- (3) Keating, J. A., and Striker, R. (2012) Phosphorylation events during viral infections provide potential therapeutic targets. *Rev. Med. Virol.* 22, 166–181.
- (4) Langhammer, S., Koban, R., Yue, C., and Ellerbrok, H. (2011) Inhibition of poxvirus spreading by the anti-tumor drug Gefitinib (Iressa). *Antiviral Res.* 89, 64–70.
- (5) Schleiss, M., Eickhoff, J., Auerochs, S., Leis, M., Abele, S., Rechter, S., Choi, Y., Anderson, J., Scott, G., Rawlinson, W., Michel, D., Ensminger, S., Klebl, B., Stamminger, T., and Marschall, M. (2008) Protein kinase inhibitors of the quinazoline class exert anticytomegaloviral activity in vitro and in vivo. *Antiviral Res.* 79, 49–61.
- (6) Stanley, S. A., Barczak, A. K., Silvis, M. R., Luo, S. S., Sogi, K., Vokes, M., Bray, M. A., Carpenter, A. E., Moore, C. B., Siddiqi, N., Rubin, E. J., and Hung, D. T. (2014) Identification of host-targeted small molecules that restrict intracellular Mycobacterium tuberculosis growth. *PLoS Pathog.* 10, e1003946.
- (7) Wang, X., Huang, S. M., Chiu, M. L., Raab-Traub, N., and Huang, E. S. (2003) Epidermal growth factor receptor is a cellular receptor for human cytomegalovirus. *Nature* 424, 456–461.
- (8) Lupberger, J., Zeisel, M., Xiao, F., Thumann, C., Fofana, I., Zona, L., Davis, C., Mee, C., Turek, M., Gorke, S., Royer, C., Fischer, B., Zahid, M., Lavillette, D., Fresquet, J., Cosset, F.-L., Rothenberg, S. M., Pietschmann, T., Patel, A., Pessaux, P., Doffol, M., Raffelsberger, W., Poch, O., McKeating, J., Brino, L., and Baumert, T. (2011) EGFR and EphA2 are host factors for hepatitis C virus entry and possible targets for antiviral therapy. *Nat. Med.* 17, 589–595.
- (9) Garcia, M., Cooper, A., Shi, W., Bornmann, W., Carrion, R., Kalman, D., and Nabel, G. J. (2012) Productive replication of Ebola virus is regulated by the c-Abl1 tyrosine kinase. *Sci. Transl. Med.* 4, 123ra24.
- (10) Dyall, J., Coleman, C. M., Hart, B. J., Venkataraman, T., Holbrook, M. R., Kindrachuk, J., Johnson, R. F., Olinger, G. G., Jahrling, P. B., Laidlaw, M., Johansen, L. M., Lear-Rooney, C. M., Glass, P. J., Hensley, L. E., and Frieman, M. B. (2014) Repurposing of clinically developed drugs for treatment of Middle East respiratory syndrome coronavirus infection. *Antimicrob. Agents Chemother.* 58, 4885–4893.
- (11) Clark, M. J., Miduturu, C., Schmidt, A. G., Zhu, X., Pitts, J. D., Wang, J., Potisopon, S., Zhang, J., Wojciechowski, A., Hann Chu, J. J., Gray, N. S., and Yang, P. L. (2016) GNF-2 Inhibits Dengue Virus by Targeting Abl Kinases and the Viral E Protein. *Cell Chem. Biol.* 23, 443–452.
- (12) Reeves, P. M., Smith, S. K., Olson, V. A., Thorne, S. H., Bornmann, W., Damon, I. K., and Kalman, D. (2011) Variola and monkeypox viruses utilize conserved mechanisms of virion motility and release that depend on abl and SRC family tyrosine kinases. *J. Virol.* 85, 21–31.
- (13) Napier, R. J., Rafi, W., Cheruvu, M., Powell, K. R., Zaunbrecher, M. A., Bornmann, W., Salgame, P., Shinnick, T. M., and Kalman, D. (2011) Imatinib-sensitive tyrosine kinases regulate mycobacterial pathogenesis and represent therapeutic targets against tuberculosis. *Cell Host Microbe* 10, 475–485.
- (14) Bekerman, E., Neveu, G., Shulla, A., Brannan, J., Pu, S.-Y., Wang, S., Xiao, F., Barouch-Bentov, R., Bakken, R. R., Mateo, R., Govero, J., Nagamine, C. M., Diamond, M. S., De Jonghe, S., Herdewijn, P., Dye, J. M., Randall, G., and Einav, S. (2017)

Anticancer kinase inhibitors impair intracellular viral trafficking and exert broad-spectrum antiviral effects. *J. Clin. Invest.* 127, 1338.

(15) Williams, J. W., Javadi, B., Kadambi, P. V., Gillen, D., Harland, R., Thistlewaite, J. R., Garfinkel, M., Foster, P., Atwood, W., Millis, J. M., Meehan, S. M., and Josephson, M. A. (2005) Leflunomide for polyomavirus type BK nephropathy. *N. Engl. J. Med.* 352, 1157–1158.

(16) Waldman, W. J., Knight, D. A., Blinder, L., Shen, J., Lurain, N. S., Miller, D. M., Sedmak, D. D., Williams, J. W., and Chong, A. S. (2000) Inhibition of cytomegalovirus in vitro and in vivo by the experimental immunosuppressive agent leflunomide. *Intervirology* 42, 412–418.

(17) Qing, M., Yang, F., Zhang, B., Zou, G., Robida, J. M., Yuan, Z., Tang, H., and Shi, P. Y. (2009) Cyclosporine inhibits flavivirus replication through blocking the interaction between host cyclophilins and viral NS5 protein. *Antimicrob. Agents Chemother.* 53, 3226–3235.

(18) Billich, A., Hammerschmid, F., Peichl, P., Wenger, R., Zenke, G., Quesniaux, V., and Rosenwirth, B. (1995) Mode of action of SDZ NIM 811, a nonimmunosuppressive cyclosporin A analog with activity against human immunodeficiency virus (HIV) type 1: interference with HIV protein-cyclophilin A interactions. *J. Virol.* 69, 2451–2461.

(19) Lin, K., and Gallay, P. (2013) Curing a viral infection by targeting the host: the example of cyclophilin inhibitors. *Antiviral Res.* 99, 68–77.

(20) Murphy, J. R., Baqar, S., Baker, R. H., Roberts, E., Nickell, S. P., and Cole, G. A. (1988) Stage-selective inhibition of rodent malaria by cyclosporine. *Antimicrob. Agents Chemother.* 32, 462–466.

(21) Rossignol, J. F. (2014) Nitazoxanide: a first-in-class broad-spectrum antiviral agent. *Antiviral Res.* 110, 94–103.

(22) Haffizulla, J., Hartman, A., Hoppers, M., Resnick, H., Samudrala, S., Ginocchio, C., Bardin, M., Rossignol, J. F., and US Nitazoxanide Influenza Clinical Study Group (2014) Effect of nitazoxanide in adults and adolescents with acute uncomplicated influenza: a double-blind, randomised, placebo-controlled, phase 2b/3 trial. *Lancet Infect. Dis.* 14, 609–618.

(23) Kohla, M. A., El-Said, H., El-Fert, A., Ehsan, N., Ezzat, S., and Taha, H. (2016) Impact of nitazoxanide on sustained virologic response in Egyptian patients with chronic hepatitis C genotype 4: a double-blind placebo-controlled trial. *Eur. J. Gastroenterol. Hepatol.* 28, 42–47.

(24) Shigyo, K., Ocheretina, O., Merveille, Y. M., Johnson, W. D., Pape, J. W., Nathan, C. F., and Fitzgerald, D. W. (2013) Efficacy of nitazoxanide against clinical isolates of Mycobacterium tuberculosis. *Antimicrob. Agents Chemother.* 57, 2834–2837.

(25) Singhal, A., Jie, L., Kumar, P., Hong, G. S., Leow, M. K.-S., Paleja, B., Tsenova, L., Kurepina, N., Chen, J., Zolezzi, F., Kreiswirth, B., Poidinger, M., Chee, C., Kaplan, G., Wang, Y. T., and De Libero, G. (2014) Metformin as adjunct antituberculosis therapy. *Sci. Transl. Med.* 6, 263ra159–263ra160.

(26) Rothwell, C., Lebreton, A., Young Ng, C., Lim, J. Y., Liu, W., Vasudevan, S., Labow, M., Gu, F., and Gaither, L. A. (2009) Cholesterol biosynthesis modulation regulates dengue viral replication. *Virology* 389, 8–19.

(27) Simon, T. G., and Butt, A. A. (2015) Lipid dysregulation in hepatitis C virus, and impact of statin therapy upon clinical outcomes. *World J. Gastroenterol.* 21, 8293–8303.

(28) Fedson, D. S., Jacobson, J. R., Rordam, O. M., and Opal, S. M. (2015) Treating the Host Response to Ebola Virus Disease with Generic Statins and Angiotensin Receptor Blockers. *mBio* 6, e00716-15.

(29) Farias, K. J., Machado, P. R., Muniz, J. A., Imbeloni, A. A., and da Fonseca, B. A. (2015) Antiviral activity of chloroquine against dengue virus type 2 replication in Aotus monkeys. *Viral Immunol.* 28, 161–169.

(30) Borges, M. C., Castro, L. A., and da Fonseca, B. A. L. (2013) Chloroquine use improves dengue-related symptoms. *Memorias do Instituto Oswaldo Cruz* 108, 596–599.

(31) Tricou, V., Minh, N. N., Van, T. P., Lee, S. J., Farrar, J., Wills, B., Tran, H. T., and Simmons, C. P. (2010) A Randomized

Controlled Trial of Chloroquine for the Treatment of Dengue in Vietnamese Adults. *PLoS Neglected Trop. Dis.* 4, e785.

(32) Shiryayev, S. A., Mesci, P., Pinto, A., Fernandes, I., Sheets, N., Shresta, S., Farhy, C., Huang, C.-T., Strongin, A. Y., Muotri, A. R., and Terskikh, A. V. (2017) Repurposing of the anti-malaria drug chloroquine for Zika Virus treatment and prophylaxis. *Sci. Rep.* 7, 15771.

(33) Yan, Y., Zou, Z., Sun, Y., Li, X., Xu, K. F., Wei, Y., Jin, N., and Jiang, C. (2013) Anti-malaria drug chloroquine is highly effective in treating avian influenza A H5N1 virus infection in an animal model. *Cell Res.* 23, 300–302.

(34) Matt, U., Selchow, P., Dal Molin, M., Strommer, S., Sharif, O., Schilcher, K., Andreoni, F., Stenzinger, A., Zinkernagel, A. S., Zeitlinger, M., Sander, P., and Nemeth, J. (2017) Chloroquine enhances the antimycobacterial activity of isoniazid and pyrazinamide by reversing inflammation-induced macrophage efflux. *Int. J. Antimicrob. Agents* 50, 55–62.

(35) Haqqani, A. A., and Tilton, J. C. (2013) Entry inhibitors and their use in the treatment of HIV-1 infection. *Antiviral Res.* 98, 158–170.

(36) Lotharius, J., Gamo-Benito, F. J., Angulo-Barturen, I., Clark, J., Connelly, M., Ferrer-Bazaga, S., Parkinson, T., Viswanath, P., Bandodkar, B., Rautela, N., Bharath, S., Duffy, S., Avery, V. M., Mohrle, J. J., Guy, R. K., and Wells, T. (2014) Repositioning: the fast track to new anti-malarial medicines? *Malar. J.* 13, 143.

(37) de Wispelaere, M., LaCroix, A. J., and Yang, P. L. (2013) The small molecules AZD0530 and dasatinib inhibit dengue virus RNA replication via Fyn kinase. *J. Virol* 87, 7367–7381.