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## Keeping an Eye on Bardet-Biedl Syndrome: A Comprehensive Review of the Role of Bardet-Biedl Syndrome Genes in the Eye

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### Abstract

Upwards of 90% of individuals with Bardet-Biedl syndrome (BBS) display rod-cone dystrophy with early macular involvement. BBS is an autosomal recessive, genetically heterogeneous, pleiotropic ciliopathy for which 21 causative genes have been discovered to date. In addition to retinal degeneration, the cardinal features of BBS include obesity, cognitive impairment, renal anomalies, polydactyly, and hypogonadism. Here, we review the genes, proteins, and protein complexes involved in BBS and the BBS model organisms available for the study of retinal degeneration. We include comprehensive lists for all known BBS genes, their known phenotypes, and the model organisms available. We also review the molecular mechanisms believed to lead to retinal degeneration. We provide an overview of the mode of inheritance and describe the relationships between BBS genes and Joubert syndrome, Leber Congenital Amaurosis, Senior-Løken syndrome, and non-syndromic retinitis pigmentosa. Finally, we propose ways that new advances in technology will allow us to better understand the role of different BBS genes in retinal formation and function.

### Keywords

Bardet-Biedl syndrome; eye; retinal degeneration; ciliopathy; blindness; BBSome; photoreceptor; nyctalopia; Joubert syndrome; Leber Congenital Amaurosis; Senior-Løken syndrome

## 1. Bardet-Biedl Syndrome

Bardet-Biedl syndrome (BBS) is an autosomal recessive, genetically heterogeneous, pleiotropic disorder (1–3) caused by mutations in 21 genes (Table 1) that are involved in proper function of the primary cilium. Eight of these proteins (BBS1, BBS2, BBS4, BBS5, BBS7, BBS8, BBS9, and BBS18) form a complex, the BBSome, which is involved in protein trafficking within cilia, as well as intracellular trafficking (4–13). BBS6, BBS10, and BBS12 are involved in a chaperonin complex required for BBSome assembly (11, 14–16). BBS3, BBS14, and BBS17 are responsible for proper trafficking of the BBSome to and

within cilia (17–21). Other BBS genes are not as closely associated with the BBSome (BBS11, BBS13, BBS15–16, and BBS19–21) (11, 22–29) and their role in cilia has not yet been fully characterized. BBS proteins are also thought to play a role in protein trafficking in the photoreceptor connecting cilium and outer segment (OS), which have similarities to the primary cilium (30, 31).

### 1.1. Clinical Diagnosis

The clinical diagnosis of BBS is typically defined as having either four primary features or three primary and two secondary features (33). Primary features include retinal degeneration, postaxial polydactyly, truncal obesity, cognitive impairment, hypogonadism, and renal anomalies. Secondary features include speech delay, developmental delay, behavioral abnormalities, orodental abnormalities, brachydactyly/syndactyly, ataxia/poor conditioning, mild hypertonia, diabetes mellitus, cardiovascular anomalies, hepatic involvement, craniofacial dysmorphism, Hirschsprung disease, anosmia, and additional eye abnormalities such as strabismus, cataracts, and astigmatism (33).

Obesity occurs in 72–92% of patients beginning in early childhood and worsening with age (34). Polydactyly is present in almost 95% of BBS patients, primarily with a postaxial presentation (69% of BBS cases) (35). Many BBS patients suffer from hypogonadism and genital anomalies such as hypoplastic fallopian tubes and uterus (36). Abnormal spermatozoa have been found in male patients. However, patients of both sexes have been reported to have successfully reproduced (2, 37). Renal failure may not present until late childhood and onward but is a major cause of morbidity and mortality in this disorder (38). Cognitive impairment is the most variable primary phenotype: 29% of BBS patients display severe intellectual disability, 38% display moderately reduced intelligence, and 29% display average intelligence. Some patients even display intellectual performance in the higher range of abilities within the normal population (4%) (35). The role of BBS genes in obesity (39, 40), renal failure (41, 42), and the cilium (42, 43) have been comprehensively reviewed elsewhere; here, we review the role of BBS genes in syndromic and non-syndromic retinal degeneration.

### 1.2. Mode of Inheritance

The autosomal recessive nature of BBS has been clearly established by segregation analysis of large pedigrees and consanguineous families. The prevalence of BBS is population dependent with an estimated incidence of 1:160,000 in Switzerland and Tunisia, 1:18,000 in Newfoundland, and 1:36,000 in the mixed Arab populations in Kuwait (37, 44–46). It has been reported that homozygous or compound heterozygous mutations in 5 BBS genes (BBS1, BBS10, BBS2, BBS9, and MKKS (BBS6)) cause the majority of BBS cases (33, 46). However, there is some controversy about how frequently BBS displays a “triallelic” inheritance pattern caused by three pathogenic alleles at two different loci (47). This hypothesis is based on BBS patients identified with three putative mutant alleles in two different BBS genes (BBS2 and BBS6) and unaffected individuals carrying two BBS2 mutations. The same group reported that BBS1 has a complex inheritance pattern (48). The group identified asymptomatic individuals with homozygous mutations for BBS1. In addition, these researchers reported a higher prevalence of the M390R allele in a control

cohort compared to the number of affected M390R individuals, suggesting an oligogenic model of BBS for this allele (48). Upon further investigation using the exome aggregation consortium data, the minor allele frequency for the M390R allele is known to be three times lower than previously reported by the group and thus does not seem to follow an oligogenic model of inheritance.

In addition, studies by other groups have not supported a triallelic inheritance pattern, and the vast majority of BBS patients reported to date display classic Mendelian autosomal recessive inheritance (49–51). For example, Biesecker *et al.* describe an Amish family with three children affected by homozygous mutations in *BBS6* (50). The father is a carrier and the mother has a non-penetrant homozygous BBS mutation. Sequencing of eight additional BBS genes in this family did not support triallelic inheritance. Another study of eight BBS genes found no families with involvement of two BBS loci consistent with triallelic inheritance (49). Later studies involving 14 BBS genes also failed to detect evidence of a triallelic model of BBS (51). Autosomal recessive inheritance is also consistent with data from multiple BBS mouse models (30, 31, 52–58). One important ramification of the triallelic model of inheritance is the counseling of parents; the 25% recurrence risk of an autosomal recessive disorder differs from the lower than 25% recurrence risk associated with an oligogenic inheritance pattern.

It has also been proposed that a third mutation in a different BBS gene can result in a more severe phenotype in patients with homozygous or compound heterozygous mutations in a BBS gene (59). The ability of BBS genes to cause other ciliopathies and non-syndromic retinal degeneration provides support for the presence of modifier genes, as do data from animal models. For example, the *BBS14* (CEP290) homozygous mutant mouse displays increased obesity and an accelerated rate of retinal degeneration when only a single copy of *BBS4* is present (60). In one family with two affected individuals homozygous for *BBS7* mutations, the individual with a third mutation in *BBS4* had a more severe phenotype (61). In theory, genetic modifiers may act on homozygous BBS gene mutations to affect the phenotypic presentation of BBS. It is important to note that many of the genes reported to cause non-syndromic retinal degeneration code for ciliary proteins and hence, they potentially interact with each other (62).

### 1.3. Retinal Degeneration

Retinal degeneration is the most highly penetrant feature of BBS, making this disorder the second most common syndromic retinal degeneration behind Usher syndrome (46). In a large systematic study of ophthalmic features in BBS adult patients, 60 of 62 patients had clinically evident retinopathy and the other 2 had evidence of retinopathy using electrophysiologic testing (63). BBS patients typically develop symptoms of retinitis pigmentosa (RP) in the first decade of life and often reach legal blindness between the second and third decades of life (64). Night blindness (nyctalopia) is the most common initial visual symptom and is usually first noted around 8.5 years of age (2, 65).

RP causes decreased peripheral vision, which presents as a constricted visual field (65, 66). During this progressive loss of the rods, the patient also loses some cone photoreceptors, which causes loss of visual acuity and a diminution of color discrimination (67).

Electrophysiologically, BBS patients are usually found to have a rod-cone dystrophy with early macular involvement (Figure 1) (46, 65–68).

## 2. Molecular Mechanisms

### 2.1. Animal Models of BBS

Investigating the cause of retinal degeneration has been a focus in many BBS animal models. With the advent and increased availability of knockout, knock-in, and transgenic mice, mouse models for a large number of BBS genes have become available to study the role of BBS proteins in the development and maintenance of the mammalian retina (24, 30, 31, 52–55, 57, 58, 69–81). Zebrafish, an organism with a rapidly developing eye, has also been used to investigate how loss of BBS genes affects eye development and maintenance, as well as the effect on vision (70).

BBS can be caused by mutations in two groups of genes: BBSome/BBSome-related genes and non-BBSome genes. Mutations in BBSome genes are more common in humans, which resulted in earlier identification of these genes compared to most of the non-BBSome genes (1, 4–10, 17). The recent advent of next-generation sequencing methods has allowed for the identification of several non-BBSome genes in smaller pedigrees or isolated cases (11, 26). While most BBS genes now have a mouse model available, the retinal phenotype is most well studied in the BBSome (models available in 7 of the 8 genes and a zebrafish model available for the remaining BBSome gene) (30, 54, 58, 69, 70, 73), chaperonin complex genes (3 of 3 mouse models available) (57, 71, 72), or BBSome-interacting genes (3 of 3) (Table 1) (31, 52, 76, 77, 80). While mouse models of the non-BBSome or BBSome-interacting genes *Bbs11*, *Bbs13*, *Bbs15*, *Bbs16*, *Bbs19*, and *Bbs21* are available (24, 32, 55, 78, 79, 81), with a zebrafish model in *bbs20* available (28), retinal phenotypes in the *Bbs19* mouse and *bbs20* zebrafish are unreported and knockout mice for *Bbs11* and *Bbs13* lack retinal BBS phenotypic features (24, 78). In addition, the *Bbs15* mouse model has anophthalmia, a severe phenotype that is not consistent with patient phenotypes (81).

### 2.2. Molecular Mechanism of BBS in Retinal Degeneration

The majority of BBS model organisms with retinal phenotypes display rhodopsin localization defects, suggesting a common mechanism of disease. Under normal conditions, rhodopsin is localized to the outer segment (OS) of rods and performs the first step of phototransduction. In BBS mutant mice, rhodopsin is mislocalized to the inner segment (IS) and cell bodies of the outer nuclear layer (ONL) (30, 54, 55, 73) diminishing its presence in the OS (57). Apoptosis of photoreceptor cells has also been noted (54, 82). In addition to rhodopsin mislocalization, BBS mouse models display a disorganization of the OS (30, 31) and progressive loss of OS, IS, and ONL with preservation of the inner retinal layers (53, 57). This phenotype is also observed in the zebrafish knockdown model of *bbs9*, which shows a complete loss of the OS (70).

Most of the research pertaining to BBS mouse models has focused on changes to photoreceptor structure and localization of specific proteins like rhodopsin, transducin, and arrestin. However, recent work in the *Bbs17* mouse model has identified global changes to

the OS proteome in mutant mice. Datta *et al.* identified abnormal enrichment of 138 proteins in the OS of *Bbs17* mutant mice, whereas only 8 proteins showed decreased OS localization. Many of these enriched proteins were absent from the WT OS prep. Accumulation of non-OS proteins in mutants was concomitant with a loss of these proteins in the cell body (IS, ONL, synaptic terminal) where they are normally required for regulation of retinal function. One such set of mislocalized IS proteins was the SNARE complex, which mediates membrane fusion events. STX3, a component of the SNARE complex has been linked to rhodopsin trafficking in photoreceptors, is mislocalized to the OS in multiple BBS mouse models (31). As such, the authors suggest that BBS proteins are directly or indirectly required for proper OS protein trafficking and that photoreceptor cell death in BBS mutants may be due to the combination of insufficient protein functions in the cell body and aberrant accumulation of proteins in the OS (Figure 2).

### 2.3. *In vitro* Molecular Mechanisms of BBS

The connecting cilium of photo-receptors is the analogous structure of the transition zone of the primary cilium of non-photoreceptor cell types (83). The connecting cilium derives its name from the fact that it connects the inner segment, which contains the metabolic and trafficking components of photoreceptors, with the outer segment, which contains the photo-transduction apparatus. Like the primary cilium, the connecting cilium consists of an axoneme that is comprised of nine microtubule doublets. There is trafficking of proteins and membrane components through the connecting cilium by a process known as intraflagellar transport (IFT), a process requiring BBS proteins (84, 85).

The BBSome plays a role in transport within the cell, including the known effects of BBSome associated proteins on the leptin receptor (86) and the insulin receptor (87), though the mechanism for this trafficking is not well understood. In addition to intracellular trafficking, in the presence of BBS mutations, an array of ciliary receptors are mislocalized, including: the smooth muscle receptor (19), somatostatin receptor type 3 (88), melanin-concentrating hormone receptor 1 (88), Neuropeptide Y Family Receptors (89), OSM-9 (90), and ODR-10 (90). The majority of these receptor trafficking defects involve G-protein coupled receptors, which would affect cAMP signaling and phosphatidylinositol signaling (91). Due to the shared structure of the connecting cilium and the transition zone of the primary cilium, the retinal degeneration phenotype is likely due to changes in ciliary receptor trafficking and not generalized intracellular trafficking. Additional work is needed to further define the role and interacting partners of the BBSome in photoreceptor cells.

### 2.4. Transcriptional Variation

Consistent with the role of BBS proteins in retinal development and maintenance, retina-specific isoforms have been identified in *BBS3*, *BBS5*, and *BBS8* (77, 92, 93). Loss of the *bbs3* retina-specific isoform (*bbs3l*) in zebrafish causes vision loss that cannot be rescued by the canonical isoform of *Bbs3*. Loss of the *Bbs3l* isoform in the mouse causes disorganization of the IS but does not lead to severe retinal dysmorphology. However, green cone opsin mislocalization can be rescued with *Bbs3l*, suggesting an important role for this isoform specific to eye morphology and physiology. The retinal isoform of *BBS5*, *BBS5L*, has not been studied at a functional level in the eye, but is known to localize to the axonemal

structure of the photoreceptor and is found at the boundary of the IS and OS, in the IS and OS, the inner plexiform layer, and ganglion cell layer but not in the connecting cilium (92). Based on current patient information and known mutations, no specific pathology can currently be ascribed to *BBS5L* (92). Interestingly, a mutation that results in the skipping of the *BBS8* retina-specific isoform has been identified in patients with non-syndromic RP, supporting the role of this isoform in retinal pathology (93).

The presence of these splice variants highlights the unique characteristics of the retina and confirm there is still much to be learned about the retina, its function, and the role of BBS genes in eye development and function. To date, approximately one quarter of retinal genes have been found to express alternate transcripts (94). It is possible that other BBS genes have unidentified retinal specific splice variants that may account for additional cases of non-syndromic RP or may modify the eye phenotype of BBS patients. Future research could pursue identification of additional splice variants and further investigation of the role of known isoforms like BBS8 and *BBS5L*.

### 3. Other Disorders Attributed to BBS Genes

Retinal degeneration is a common feature of many ciliopathies. Mutations in genes that usually cause BBS have currently been identified in three additional ciliopathies that present with retinal degeneration as well as in non-syndromic retinal degeneration (Table 2). Here, we examine the similarities and differences between these disorders.

#### 3.1. Leber Congenital Amaurosis

Leber congenital amaurosis (LCA) is the most severe retinal degeneration of the ciliopathies reviewed here. LCA is usually apparent shortly after birth and is characterized by severe dysfunction of both photoreceptor types, nystagmus, reduced pupillary response, keratoconus, and hyperopia (95–97). LCA patients commonly display excessive eye poking and rubbing (98). This behavior is known as Franceschetti’s oculo-digital sign and contributes to the development of keratoconus in these patients (95, 98). Currently, there have been at least 17 genes implicated in the development of LCA including CEP290 (BBS14) (99).

#### 3.2. Joubert Syndrome

Joubert syndrome is a rare disorder that affects many organ systems. A cardinal feature of Joubert syndrome is the “molar tooth sign” resulting from abnormal development of the mid-brain, which is identified using cranial magnetic resonance imaging (100). Other common characteristics include hypotonia, abnormal eye movements, ataxia, irregular breathing patterns, and mild to severe developmental delay (101, 102). Joubert syndrome may also present with retinal degeneration, cystic kidney disease, nephronophthisis, polydactyly, encephalocele, and endocrine defects (103–107). The age of onset of retinal degeneration is variable, and was seen to only occur in approximately 30% of patients. It can be as severe as neonatal onset of congenital blindness, but it may also present as a stable loss of vision (108). Currently, there are at least 34 genes associated with Joubert syndrome including MKS1 (BBS13) and CEP290 (BBS14)(109).

### 3.3. Senior-Løken Syndrome

Senior-Løken syndrome (SLS), first reported in 1961, is a rare autosomal recessive disease characterized by having both nephronophthisis and LCA (110, 111). Nephronophthisis is characterized by the development of fluid-filled cysts known as cystic dilation within the kidney leading to polyuria, polydipsia, weakness, fatigue, and eventually renal failure (98, 112–115). SLS is classified as infantile, juvenile, or adolescent depending on the manifestation of renal failure. RP and the subsequent blindness that follows may occur as early as birth or it can manifest as a progressive loss of vision during childhood (114–116). Usually all SLS patients have been diagnosed with alterations to their eyes by 10 years of age. Patients with juvenile renal failure, which is linked to mutations in NPHP5 and CEP290, have a greater propensity for RP (116, 117). Milder RP has been observed in patients with mutations in other NPHP genes even though disease symptoms often present during the first two years of life with almost all SLS patients diagnosed with eye changes by 10 years of age (118, 119). Presently, at least 9 genes are associated with SLS including CEP290 (BBS14) and SDCCAG8 (BBS16) (26, 116, 120–123).

### 3.4. Non-Syndromic Retinal Degeneration

The clinical findings in patients with syndromic and non-syndromic BBS-associated retinal degeneration are very similar; both groups experience early central vision loss coupled with a progressive loss of peripheral vision which can ultimately result in complete blindness in some patients. Mutations in five different BBS genes have been reported to cause non-syndromic RP. While the BBS1 M390R mutation causes the greatest percentage of clinically diagnosed BBS, Estrada-Cuzcano *et al.* identified two non-syndromic RP patients that were homozygous for the M390R mutation (124). Shevach identified a set of 4 missense mutations in BBS2 that cosegregated with non-syndromic RP in five independent families (125). A homozygous mutation in ARL6 (BBS3) and a novel splice site mutation in TTC8 (BBS8) have been identified in patients with non-syndromic RP (93, 126). Finally, Estrada-Cuzcano *et al.* reported that homozygous splice and homozygous missense mutations in C8orf37 (BBS21) are associated with non-syndromic autosomal-recessive cone-rod dystrophy (CRD) and RP (127). The combination of low disease prevalence, plus gene- and mutation-level heterogeneity makes the inference of mutation specificity for BBS versus non-syndromic retinal degeneration challenging.

## 4. Future Directions

In order to accurately distinguish between variants that cause BBS, other ciliopathies, and non-syndromic retinal degeneration, more work is required to understand the many different BBS proteins and their functions. This will include the use of transcriptomics, proteomics, animal studies, and cellular studies using patient-derived induced pluripotent stem cells to provide a comprehensive analysis of each gene and its role in the retina and beyond.

The availability of numerous animal models will allow for further in-depth investigation into the development of BBS retinopathy. For example, it is currently unknown whether accumulation of one specific protein in the OS is sufficient to induce photoreceptor degeneration or whether degeneration is caused by a cumulative stress on the OS trafficking

system. It would be interesting to compare the OS proteome of BBS17 mutant mice with proteomes from other BBS models, including BBS models that don't directly interact with the BBSome, to determine whether certain proteins remain consistently enriched in the OS. Additional data could be obtained utilizing RNA sequencing on similar samples to identified transcription differences within the eye. These data could provide vital clues in identifying which proteins require precise localization or splice variants to prevent retinal degeneration and may provide a shared mechanism of OS degeneration between BBSome and non-BBSome genes. Protein mislocalization could also be used to identify novel BBS or retinal ciliopathy genes.

However, developing mouse models of every specific patient mutation is time consuming, expensive, and impractical. As such, other methods will need to be utilized to assess the pathogenicity of these mutations. Recent efforts to characterize the three-dimensional structure of the BBSome as well as *in silico* modeling of selected mutations observed in patients (128) could be used to predict the effect of novel mutations on protein structure and BBSome assembly. The functional effects of patient mutations could also be assessed *in vitro*. The CRISPR/Cas9 system greatly facilitates the creation of cell lines harboring patient mutations for further study. Additionally, induced pluripotent stem cells (iPSCs) derived from patient fibroblasts could be differentiated into photoreceptor precursor cells (129). These cells would allow researchers to study the effect of patient mutations in a disease-relevant cell type in the context of the patient's specific genetic background. Of course, the ultimate goal is to be able to treat patients with BBS. Patients in the early stages of disease can potentially be treated with gene therapy (130), whereas patients with advanced disease could be treated with photoreceptor precursor cells derived from autologous patient derived iPSCs. Investigative studies of such work is underway (131).

Some patients with the clinical diagnosis of BBS have not yet had the specific molecular cause of their disease identified. It is possible that causal mutations lie in regulatory or non-exonic regions, as well as in alternatively spliced isoforms of known BBS genes; additional sequencing and molecular characterization will be required to identify these variants. It is also possible that these patients harbor mutations in genes that cause BBS that have yet to be identified as such. It may be worth cataloging interactors of the BBSome to identify potential sequencing targets for patients. Future work could involve identifying cargos of the BBSome, other interactors, or other genes with ciliary function in order to determine their likelihood of causing BBS.

## 5. Conclusions

In summary, we have discussed the diagnostic criteria for BBS, the mode of inheritance of this disorder, and the primary phenotype seen in BBS patients, retinal degeneration. We also reviewed the molecular mechanisms that lead to retinal degeneration in these patients. Animal models have proven to be essential for the study of the outer segment and the role of protein mislocalization in the development of this phenotype. In addition to the animal models, *in vitro* studies have isolated specific receptors in the cilium that are affected by changes to BBS genes, suggesting a possible role of BBS on their ciliary localization and ability to respond normally to extracellular signaling. Retina-specific transcriptional



isoforms have been identified in three BBS genes, with two showing an association to retinal degeneration. We have also reviewed the role of mutations in BBS genes in associated but clinically distinct ciliopathies such as Leber Congenital Amaurosis, Joubert syndrome, Senior-Løken syndrome, and non-syndromic retinitis pigmentosa. Furthermore, we have discussed potential approaches to answer current gaps in knowledge surrounding BBSome-associated and non-BBSome associated genes and how these two separate pools of genes can lead to shared phenotypes. As we continue to study this disease and its current proteins, we will gain a deeper understanding of the molecular mechanisms, potentially identify additional genes that cause this disorder, and allow for the development of therapeutic treatments to better the vision quality in BBS patients.

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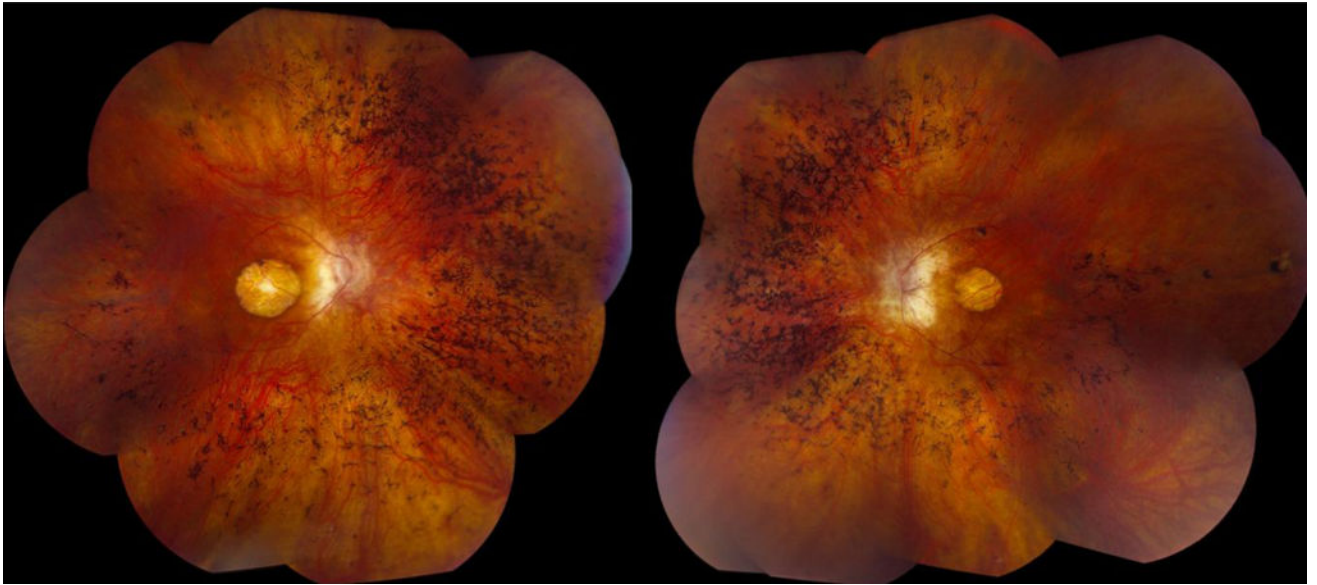
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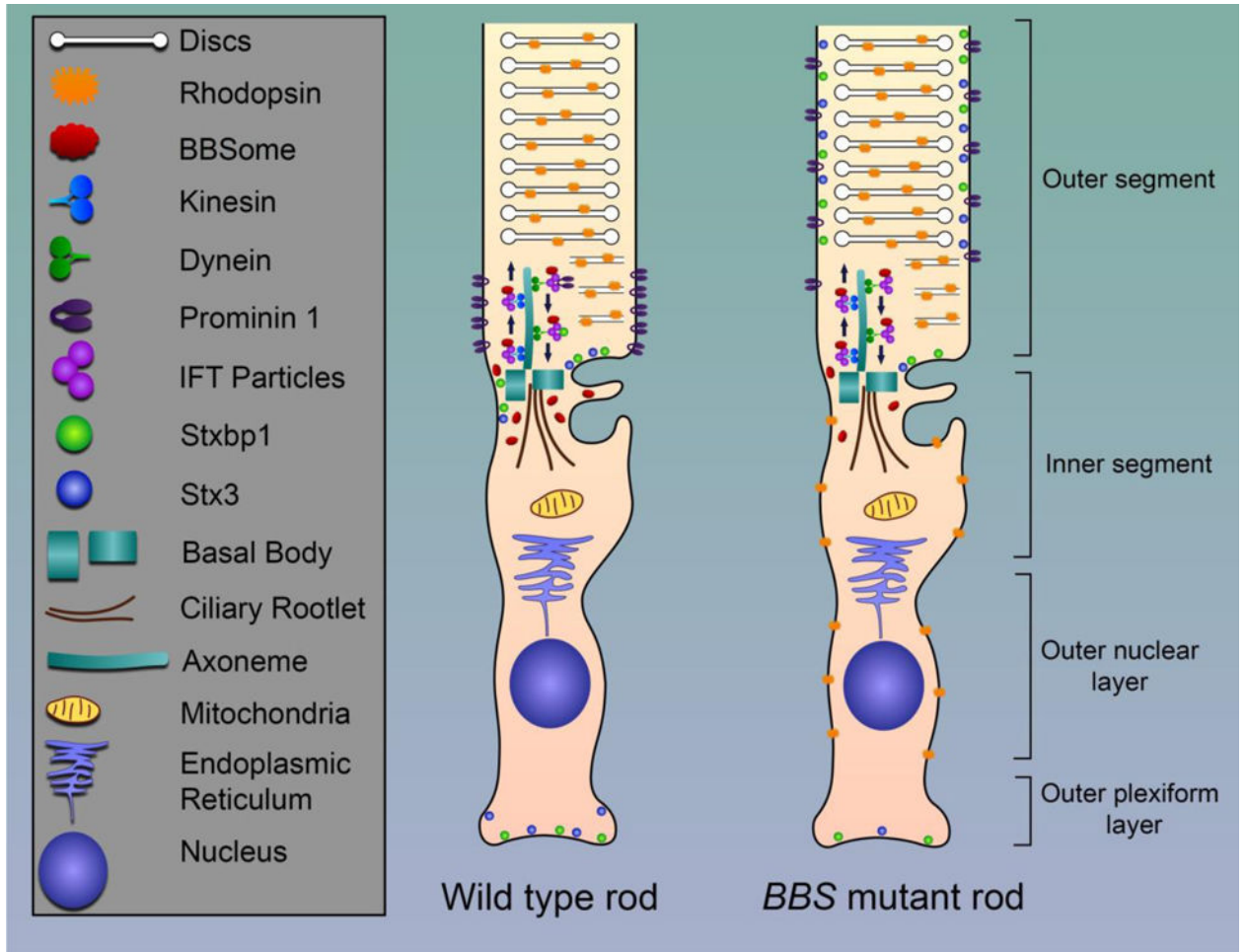
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**Figure 1.** Right (20/100 best corrected visual acuity (BCVA)) and left (20/80 BCVA) fundus photographs of a 47-year-old male with BBS caused by a homozygous M390R mutation in BBS1 showing the characteristic macular atrophy and peripheral bone spicule pigmentation. This patient has a history of postaxial polydactyly of the lower extremities and first noted visual symptoms at age 14.



**Figure 2.**

Schematic illustration of the rod photoreceptor for wild type mice and BBS mutant mice. The BBSome is involved in protein trafficking along the axoneme. In *Bbs8* and *Bbs17* mutant mice, proteins are mislocalized. In a wild type mouse, STX3 and STXBP1 are located at the synapse and base of the cilium. Rhodopsin is localized to the discs. In addition, Prominin 1 is located at the base of the outer segment. In BBS mutant mice, STX3 and STXBP1 are found throughout the outer segment. Rhodopsin is found at the inner segment and outer nuclear layer. In addition, Prominin 1 is found throughout the outer segment.

**Table 1**

Summary of BBS gene HGNC names, classification, and models of study.

<b>BBS Gene</b>	<b>HGNC</b>	<b>Classification</b>	<b>Support (Ref # are PMID)</b>
<i>BBS1</i>	<i>BBS1</i>	BBSome	Mouse (15322545, 18032602, 23160237) Zebrafish (24069149) Cell Culture (22500027)
<i>BBS2</i>	<i>BBS2</i>	BBSome	Mouse (15539463) Cell Culture (22500027)
<i>BBS3</i>	<i>ARL6</i>	BBSome Associated	Mouse (22139371) Zebrafish (21282186) Cell Culture (22139371)
<i>BBS4</i>	<i>BBS4</i>	BBSome	Mouse (16794820, 15173597) Zebrafish (24681783) Cell Culture (22500027, 24681783)
<i>BBS5</i>	<i>BBS5</i>	BBSome	Mouse (25849460) Zebrafish (16399798) Cell Culture (22500027)
<i>BBS6</i>	<i>MKKS</i>	BBSome Chaperonin	Mouse (15772095) Cell Culture (20080638)
<i>BBS7</i>	<i>BBS7</i>	BBSome	Mouse (23572516) Zebrafish (24938409) Cell Culture (22500027)
<i>BBS8</i>	<i>TTC8</i>	BBSome	Mouse (21646512) Zebrafish (20643117) Cell Culture (22500027)
<i>BBS9</i>	<i>PTHB1</i>	BBSome	Zebrafish (22479622) Cell Culture (22500027)
<i>BBS10</i>	<i>BBS10</i>	BBSome Chaperonin	Mouse (26273430) Zebrafish (16582908) Cell Culture (20080638)
<i>BBS11</i>	<i>TRIM32</i>	Non-BBSome	Mouse (19155210, 21775502) Zebrafish (16606853) Cell Culture (28498859)
<i>BBS12</i>	<i>BBS12</i>	BBSome Chaperonin	Mouse (22958920) Zebrafish (17160889) Cell Culture (20080638)
<i>BBS13</i>	<i>MKS1</i>	Non-BBSome	Mouse (19776033, 21045211) Zebrafish (18327255)
<i>BBS14</i>	<i>CEP290</i>	BBSome Associated	Mouse: (16632484, 21623382) Zebrafish (18327255) Cell Culture (25552655)
<i>BBS15</i>	<i>WDPCP</i>	Non-BBSome	Mouse (24302887) Zebrafish (24302887) Cell Culture (20671153)
<i>BBS16</i>	<i>SDCCAG8</i>	Non-BBSome	Mouse (24722439, 24302887) Zebrafish (20835237) Cell Culture (20835237, 24722439)
<i>BBS17</i>	<i>LZTFL1</i>	BBSome Associated	Mouse (27312011, 26216965) Cell Culture (22510444, 26216965)
<i>BBS18</i>	<i>BBIP1</i>	BBSome	Mouse (24316073) Cell Culture (24316073)
<i>BBS19</i>	<i>IFT27</i>	Non-BBSome	Mouse (25446516) Cell Culture (25446516)
<i>BBS20</i>	<i>IFT74</i>	Non-BBSome	Zebrafish (27486776)

<b>BBS Gene</b>	<b>HGNC</b>	<b>Classification</b>	<b>Support (Ref # are PMID)</b>
<i>BBS21</i>	<i>C8ORF37</i>	Non-BBSome	Mouse (32) Zebrafish (27008867)

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**Table 2**

Summary of BBS genes, references in which they were classified as causative genes of BBS, and BBS genes that cause other ciliopathies (Ref # are PMID).

Gene	Bardet-Biedl Syndrome	Non-Syndromic Retinitis Pigmentosa	Leber Congenital Amaurosis	Senior-Løken Syndrome	Joubert Syndrome	Meckel-Gruber Syndrome
<i>BBS1</i>	12118255	23143442				
<i>BBS2</i>	11285252	25541840				
<i>ARL6 (BBS3)</i>	15314642	19956407				
<i>BBS4</i>	11381270					
<i>BBS5</i>	15137946					
<i>MKKS (BBS6)</i>	10973238					
<i>BBS7</i>	12567324					
<i>TTC8 (BBS8)</i>	14520415	20451172				
<i>PTHH1 (BBS9)</i>	16380913					
<i>BBS10</i>	20805367					
<i>TRIM32 (BBS11)</i>	16606853					
<i>BBS12</i>	17160889					
<i>MKSI (BBS13)</i>	18327255				24886560	16415886
<i>CEP290 (BBS14)</i>	18327255		16909394	16682973	16682973	17564974
<i>WDRCP (BBS15)</i>	20671153					
<i>SDCCAG8 (BBS16)</i>	20835237			20835237		
<i>LZTFL1 (BBS17)</i>	22510444					
<i>BBIP1 (BBS18)</i>	24026985					
<i>IFT27 (BBS19)</i>	24488770					
<i>IFT74 (BBS20)</i>	27486776					
<i>C8ORF37 (BBS21)</i>	27008867	22177090				