

## Melatonin signaling in mitochondria extends beyond neurons and neuroprotection: Implications for angiogenesis and cardio/gastroprotection

Amrita Ahluwalia<sup>a,1</sup>, Iwona M. Brzozowska<sup>b,1</sup>, Neil Hoa<sup>a</sup>, Michael K. Jones<sup>a,c</sup>, and Andrzej S. Tarnawski<sup>a,c,2</sup>

We read with interest the paper of Suofu et al. (1), demonstrating that neuronal mitochondria produce melatonin, which upon binding to its melatonin type 1 (MT<sub>1</sub>) receptor on the mitochondrial membrane (MM) inhibits cytochrome c release, caspase activation, and apoptosis. We commend the authors on their thorough investigation but wish to point out some major questions: (*i*) Is the presence of the MT<sub>1</sub> receptor in mitochondria limited to the neuronal cells or may it also apply to other cells, for example, endothelial cells that are components of blood vessels and are critical for delivery of oxygen and nutrients to all tissues, vessels' regeneration, and angiogenesis? (*ii*) Is the MT<sub>2</sub> receptor expressed on mitochondria and what is its role?

Melatonin is abundantly expressed beyond the neural system; for example, in gastrointestinal tissues, including the stomach, where it has been shown to protect gastric mucosa against stress, ischemia, and nonsteroidal antiinflammatory drug-induced injury, and to accelerate ulcer healing (2-6). We recently demonstrated that both MT<sub>1</sub> and MT<sub>2</sub> are expressed on the MM of gastric endothelial cells (GECs); MT<sub>1</sub> was expressed in MM while MT<sub>2</sub> was expressed in the nucleus and MM (Fig. 1). However, in contrast to the study of Suofu et al. (1), we found that treatment of GECs with exogenous melatonin (10  $\mu$ M) increased by 1.7-fold (P < 0.001) MM potential, which drives ATP synthesis (7). Moreover, treatment of GECs with exogenous melatonin increased the MM expression of MT<sub>1</sub> and MT<sub>2</sub> by 2.5- and 1.6-fold, respectively, and increased in vitro angiogenesis (new blood vessel formation) by 1.4-fold (P < 0.001). It should be noted that the angiogenic response to melatonin was independent of either cell proliferation or apoptosis, leading us to speculate that, at least in GECs, melatonin

signaling via mitochondria extends beyond cellular protection. Although, we have not yet determined the relative contributions of MT<sub>1</sub> vs. MT<sub>2</sub> in the angiogenic response of GECs to melatonin signaling, our finding that exogenous melatonin increased the MM expression of both  $MT_2$  and  $MT_1$  is at least indicative that both receptors are involved. Suofu et al. (1) did not assess the relative MM expression levels of MT<sub>1</sub> vs. MT<sub>2</sub>, but rather used the MT<sub>2</sub> selective inhibitor, 4P-PDOT, as evidence that melatonin signaling in neuronal mitochondria is mediated exclusively through MM-expressed MT<sub>1</sub>. Nevertheless, luzindole, which did prevent melatonin from blocking Ca<sup>2+</sup>-mediated cytochrome c release, has a much greater affinity for  $MT_2$  vs.  $MT_1$ . Moreover, the possibility that  $MT_2$  plays a "decoy" role, akin to Flt1 in VEGF signaling, cannot be excluded. In such a scenario,  $MT_2$  may play a more subtle role in regulating (e.g., dampening) melatonin signaling. On the other hand, the role of MT<sub>2</sub> in mitochondrial signaling by melatonin may be restricted to certain cell types (e.g., endothelial cells but not to brain neurons). The work of Suofo et al. (1) demonstrates novel melatonin signaling in neuronal mitochondria. Our work uncovered that this mechanism operates in endothelial cells that are major components of all blood vessels. We further suggest that melatonin signaling in mitochondria may also be applicable to other physiological processes, such as angiogenesis, gastroprotection, cardioprotection, and aging (3-6, 8, 9).

## **Acknowledgments**

This work was supported by Merit Review Award I01 BX000626-05A2 from the US Department of Veterans Affairs Biomedical Laboratory Research and Development Service (to A.S.T.).

and A.A., I.M.B., N.H., M.K.J., and A.S.T. wrote the paper. The authors declare no conflict of interest

Published under the PNAS license.

LETTER

<sup>&</sup>lt;sup>a</sup>Medical and Research Services, Veterans Affairs Long Beach Healthcare System, Long Beach, CA 90822; <sup>b</sup>Department of Anatomy, Jagiellonian University Medical College, 31-008 Cracow, Poland; and <sup>c</sup>Department of Medicine, University of California, Irvine, CA 92617 Author contributions: A.A. and A.S.T. designed research; A.A. and A.S.T. performed research; A.A., I.M.B., N.H., M.K.J., and A.S.T. analyzed data;

<sup>&</sup>lt;sup>1</sup>A.A. and I.M.B. contributed equally to this work.

<sup>&</sup>lt;sup>2</sup>To whom correspondence should be addressed. Email: atarnawski@yahoo.com.

Published online February 9, 2018.



Fig. 1. Localization of melatonin receptors  $MT_1$  and  $MT_2$  in MM of gastric endothelial cells (GECs). (A) Staining of mitochondria using MitoTracker, a fluorescent dye that stains mitochondria (red) in a manner dependent on mitochondrial membrane potential. (B) Immunofluorescence staining for  $MT_1$  in GECs (green). (C) Overlay of MitoTracker and  $MT_1$  immunostaining images showing localization of  $MT_1$  in MM as yellow/orange staining (arrows). (D) Staining of mitochondria using MitoTracker (red). (E) Immunofluorescence staining for  $MT_2$  in GECs (green); please note strong MT2 expression in the nucleus (N). (F) Overlay of MitoTracker and  $MT_2$  immunostaining images showing localization of  $MT_2$  in MM as yellow/orange staining (arrows).

- 1 Suofu Y, et al. (2017) Dual role of mitochondria in producing melatonin and driving GPCR signaling to block cytochrome c release. Proc Natl Acad Sci USA 114:E7997–E8006.
- 2 Bubenik GA (2008) Thirty four years since the discovery of gastrointestinal melatonin. J Physiol Pharmacol 59:33-51.
- 3 Karasek M (2004) Melatonin, human aging, and age-related diseases. Exp Gerontol 39:1723–1729.
- 4 Ma Z, et al. (2017) Melatonin and mitochondrial function during ischemia/reperfusion injury. Cell Mol Life Sci 74:3989–3998.
- 5 Brzozowska I, et al. (2002) Role of prostaglandins, nitric oxide, sensory nerves and gastrin in acceleration of ulcer healing by melatonin and its precursor, L-tryptophan. J Pineal Res 32:149–162.
- 6 Brzozowska I, Strzalka M, Drozdowicz D, Konturek SJ, Brzozowski T (2014) Mechanisms of esophageal protection, gastroprotection and ulcer healing by melatonin. Implications for the therapeutic use of melatonin in gastroesophageal reflux disease (GERD) and peptic ulcer disease. Curr Pharm Des 20:4807–4815.
- 7 Klusch N, Murphy BJ, Mills DJ, Yildiz Ö, Kühlbrandt W (2017) Structural basis of proton translocation and force generation in mitochondrial ATP synthase. *eLife* 6:6.
  8 Tengattini S, et al. (2008) Cardiovascular diseases: Protective effects of melatonin. *J Pineal Res* 44:16–25.
- 9 Lochner A, Huisamen B, Nduhirabandi F (2013) Cardioprotective effect of melatonin against ischaemia/reperfusion damage. Front Biosci (Elite Ed) 5:305–315.