



Estimation of contemporary effective population size and population declines using RAD sequence data

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Abstract

Large genomic data sets generated with restriction site-associated DNA sequencing (RADseq), in combination with demographic inference methods, are improving our ability to gain insights into the population history of species. We used a simulation approach to examine the potential for RADseq data sets to accurately estimate effective population size (N_e) over the course of stable and declining population trends, and we compare the ability of two methods of analysis to accurately distinguish stable from steadily declining populations over a contemporary time scale (20 generations). Using a linkage disequilibrium-based analysis, individual sampling (i.e., $n \geq 30$) had the greatest effect on N_e estimation and the detection of population size declines, with declines reliably detected across scenarios ~10 generations after they began. Coalescent-based inference required fewer sampled individuals (i.e., $n = 15$), and instead was most influenced by the size of the SNP data set, with 25,000–50,000 SNPs required for accurate detection of population trends and at least 20 generations after decline began. The number of samples available and targeted number of RADseq loci are important criteria when choosing between these methods. Neither method suffered any apparent bias due to the effects of allele dropout typical of RAD data. With an understanding of the limitations and biases of these approaches, researchers can make more informed decisions when designing their sampling and analyses. Overall, our results reveal that demographic inference using RADseq data can be successfully applied to infer recent population size change and may be an important tool for population monitoring and conservation biology.

Introduction

One of the most important parameters in wildlife management and conservation biology is effective population size (N_e), with estimates providing insight into the demographic history and extinction risk of populations. Although N_e is informative about population viability and broadly applicable in ecology, conservation, and evolution, it is notoriously difficult to estimate (Luikart et al. 2010). Rarely is enough demographic information available from natural populations to directly estimate N_e , making indirect genetic estimates of considerable use, especially given their ease of

generation relative to direct demographic methods (Schwartz et al. 2007; Luikart et al. 2010; Dudgeon and Ovenden 2015; Andreotti et al. 2016). It is now possible to generate population genomic data for almost any species for the investigation of population and evolutionary history (Narum et al. 2013; Andrews et al. 2016; Nunziata et al. 2017). The increase in power and precision offered by a genomic approach is poised to greatly improve estimates of demographic history, including N_e , the timing of demographic events, and migration. Genomic-based demographic inference has yielded insight into invasion dynamics (Trucchi et al. 2016), climate-driven population shifts (Prates et al. 2016), and glacial refugium dynamics (Kopuchian et al. 2016) at historical timescales. However, as emphasized in a recent review, the application of genomic techniques in conservation studies has been rare (Shafer et al. 2015a). One obvious, but unanswered, question is whether genomic-based demographic inference methods have the ability to accurately characterize population history over a contemporary time scale (e.g., tens of generations),

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and whether there is a time lag between decline in census size and decline in N_e .

Previous work has begun to hint at the ability for genetic data to uncover recent population history. Simulation studies have suggested that microsatellite markers have the ability to detect bottlenecks and population size trends at a contemporary scale, but require sample sizes of 60 or more individuals and are not accurate with large (≥ 1000) population sizes (Tallmon et al. 2010; Antao et al. 2011). While increasing the number of microsatellite markers employed can increase the power to detect population size change (Hoban et al. 2013), in many cases researchers will not have access to, or resources to generate, ≥ 100 microsatellite markers. These studies either did not use single-nucleotide polymorphism (SNP) data which would be common in contemporary population genomic studies, or they simulated a small number (100–1000) of SNP markers (Antao et al. 2011; Hollenbeck et al. 2016). It is possible that the increased power offered by large genomic data sets can result in accurate estimates of population size trends over short timescales while using smaller numbers of sampled individuals.

Recent empirical studies have shown that coalescent-based demographic inference can accurately date documented introductions of populations occurring in the past few decades (McCoy et al. 2013; Fraser et al. 2015). Coalescence theory states that the probability of coalescence t generations ago is $(1 - (1/2N_e))^t \approx (1/2N_e)^t$, with the coalescent N_e estimated as the expected time of coalescence in generations, T , or $T = 2N_e$ (Nordborg and Krone 2002; Wakeley and Sargsyan 2009). Given these equations, when N_e is small enough, as is often the case in species of conservation concern, large sample sizes (individuals and/or loci) may be effective in estimating coalescent N_e at a contemporary scale as coalescent events will be clustered in the recent past. Consistent with this theory, a simulation study found that although large sample sizes are generally not needed for accurate demographic inference of ancient events, increased sampling of individuals increases accuracy of parameter estimates for more recent events (Robinson et al. 2014). Before these methods can be applied to real-world conservation biology, vigorous exploration is needed to estimate their accuracy with realistic sampling conditions to gain an understanding of implicit limitations and biases (Shafer et al. 2015a).

Restriction site-associated DNA sequencing (RADseq) is arguably the most popular method for generating genome-wide population genetic data from a reduced subset of the genome (Davey et al. 2011; Andrews et al. 2016). While RADseq can yield many thousands or tens of thousands of shared orthologous loci across individuals and populations, it also has inherent properties that lead to allele dropout, and consequently, missing data that may create biases in

population genetic results. Allele dropout via mutations in restriction cut sites and the shotgun nature of Illumina sequencing, which under-sequences loci or alleles can randomly lead to either missing genotypes for loci, or the misinterpretation of null alleles as homozygous at heterozygous loci. Both of these scenarios can result in skewed estimation of allele frequencies (Arnold et al. 2013), and a misrepresentation of the site frequency spectrum (Shafer et al. 2017). Simulation studies have highlighted the downstream effects of these biases in commonly estimated population genetic summary statistics (Gautier et al. 2013; Arnold et al. 2013) and in phylogenetic inferences (Huang and Knowles 2014). However, the effect of allele dropout in RADseq-based studies of N_e and contemporary population size trends has not been investigated.

Here we use an approach similar to Tallmon et al. (2010) and assess the ability of RADseq-generated SNP data and different N_e estimators to infer population abundance and population size trends (λ) over a contemporary time scale. We simulated ideal Wright–Fisher (W–F) populations over a range of known census sizes (N_C) and with either stable population size, or a steadily declining population. In ideal W–F populations $N_C = N_e$, so that estimates of N_e can be directly compared to the simulated N_C . Using both linkage disequilibrium-based analysis, and a coalescent-based analysis, we assess the estimation of N_e and population size trends. In doing so, we also evaluate the impacts of the various aspects of the population model (initial population size and the number of generations since λ began) on estimation, as well as the impacts of sampling, number of SNPs sampled, allele dropout, and data filtering.

Methods

Data simulation

We conducted simulations of RADseq data for populations with both stable and declining population sizes using the Python program simuPOP v1.1.4 (Peng and Kimmel 2005), a forward-time and individual-based population genetic modeling program. Prior to simuPOP simulations, initial haploid allele frequencies were generated with the coalescent simulator fastsimcoal2 v2.5.2.21 (fsc2; Excoffier et al. 2013) for 20,000 150 base pair (bp) loci using a diploid N_e of 1000. A mutation rate (μ) was randomly assigned to each locus from a log-normal distribution with a mean μ of 2.5E⁻⁸ and a log standard deviation of 1.3. This mutation rate has been robustly estimated in humans (Nachman and Crowell 2000) and similarly used in other RADseq simulation studies (Huang and Knowles 2014). We used this log-normal distribution of mutation rates among loci to account for variance in the mutation rate across the genome, and to

generate a large number of highly diverse loci. Our rationale was to generate a large number of SNPs typical of empirical RAD studies, while balancing computational demand of simulating even greater numbers of individual RAD loci variable in one or a few SNPs. This created a larger proportion of allele dropout than would be typical of empirical studies, but the loci retained to assess impacts of allele dropout should be comparable to those typical of empirical studies. Loci were generated as Arlequin-formatted files and were subsequently converted to Phylip format using the program PGDSpider v2.0.5.1 (Lischer and Excoffier 2012). Initial diploid genotypes for individuals in the simuPOP population were generated by pairing the fsc2-simulated alleles for each locus using random sampling with replacement, which approximated random mating and W–F populations. Diploid populations were constructed with initial population sizes of $n = 250, 500, \text{ and } 1000$, with 100 replicates constructed for each initial population size. Throughout the subsequent simulations, populations maintained an average sex ratio of 1 with random mating, non-overlapping generations, a fixed $\mu = 2.5E-8$ across all loci, and with no assignment to chromosomes. Under these conditions N_C should be approximately equal to N_e . All simulated populations went through an equilibrium phase of 10 generations to reach Hardy–Weinberg equilibrium (Waples 2006; Tallmon et al. 2010; Antao et al. 2011), after which each replicate diploid population evolved for one generation (t_{-1}) according to two separate deterministic growth rates that approximated a stable population ($\lambda = 1.0$) and a declining population ($\lambda = 0.9$). Data collection began at generation t_0 as the population evolved at the same λ for 20 generations as in Tallmon et al. (2010). In each simulation, genotypes from all loci were recorded after 0, 5, 10, 15, and 20 generations. Sample collection began with one generation after the initiation of the deterministic growth rate because inbreeding N_e estimates are reflective of the number of parents in the parental generation (Waples 2005). To assess the effect of the sample size of individuals, we sampled 15, 30, and 60 individuals from each of the specified generations.

In silico RADseq mutations and data filtering

Using custom Python scripts, we filtered RADseq loci from sampled individuals to mimic empirical RADseq data recovery and filtering conditions typically used in population genomic studies. To simulate allelic dropout as a result of a mutation in the restriction enzyme cutting site, all individual sequences were deleted containing a mutation in the first 8 bp, which represents our restriction cut site. To simulate missing data as a result of variation in sequencing coverage, we simulated the number of reads for each individual allele by drawing randomly from a Poisson

distribution with a mean of 10 (Huang and Knowles 2014). We imposed a sequencing coverage cutoff of 10, which is considered an efficient sequencing coverage cutoff for diploids. To be genotyped as heterozygous, individuals were required to have a coverage ≥ 5 reads per allele for a given locus. If one allele had a coverage ≥ 10 reads and the other had < 5 , the locus was recorded as homozygous for the higher-coverage allele due to allele dropout. Loci below these coverage cutoffs were recorded as missing data. All other sources of missing data and biases from sequencing errors, coverage cutoffs, and alignment errors were ignored here as they are not the focus of our study. These have been thoroughly reviewed in other studies, and are expected to cause general biases in all sequencing projects (Rokas and Abbot 2009, Pool et al. 2010, Huang and Knowles 2014).

We next filtered our simulated RADseq data using the criteria specific to the two analytical programs used in demographic estimation.

Linkage disequilibrium-based estimation

Linkage disequilibrium (LD) methods for N_e estimation assume unlinked loci. To remove the inclusion of linked sites within a RADseq locus, we used only the first SNP in a locus in all LD-based data sets. To examine whether the LD-based method produced unbiased N_e estimates with perfect detection of allele dropout, we analyzed data sets that removed all loci with missing data exclusively due to RADseq cut site mutations, hereafter referred to as the LD RAD mutation data set. We further examined how LD-based N_e estimation would be affected by the combined impacts of missing data from allele dropout due to RADseq cut site mutation and low sequencing coverage. For these analyses, we generated two filtered data sets that removed loci with $\geq 10\%$ and $\geq 50\%$ missing data; hereafter referred to as the 10% missing and 50% missing data sets, respectively.

Fastsimcoal2

In fsc2, the use of linked SNPs should not bias parameter estimation, so all data sets analyzed in this study used all SNPs in a locus. However, the inclusion of loci with missing data is expected to lead to a biased site frequency spectrum (SFS) and result in inaccurate parameter estimates (Excoffier et al. 2013). Therefore, we included only loci with no missing data across all sampled individuals. Only variable sites were included in the SFS. To examine the potential effects of allele dropout on N_e estimation in the program fsc2, we analyzed our simulated RADseq data under a range of filtering strategies that accounted for allele dropout due to mutations in restriction cut sites and insufficient sequencing coverage. First, we analyzed an unfiltered

data matrix with no allele dropout. Here the SFS was constructed using the complete 20,000 locus (3,000,000 bp) simulated data set, and is hereafter referred to as the fsc2 complete data set. Next, we examined the performance of N_e estimation in fsc2 when accounting for the perfect detection of allele dropout due to restriction cut site mutations. Here the SFS was constructed after removal of all loci with a restriction cut site mutation, hereafter referred to as the fsc2 RAD mutation data set. We examined the performance of N_e estimation in fsc2 when allowing for allele dropout due to both cut site mutation and low sequencing coverage, hereafter referred to as the fsc2 RAD mutation and coverage data set. Finally, to examine the impact of number of SNPs included in the joint SFS, we subsampled the fsc2 complete data set for 5000, 15,000, 25,000, 50,000, 100,000, and 150,000 SNPs.

N_e estimation and demographic inference

We used the program NeEstimator v2.01 (Do et al. 2014) to estimate N_e using the linkage disequilibrium method (Hill 1981). With finite population size and a limited number of parents, nonrandom associations of alleles at different genetic markers occur (i.e., linkage disequilibrium), even without any physical linkage on a chromosome (Waples and Do 2010). We estimated N_e from all sampled generations of our temporally simulated populations, employing all three LD-based data-filtering scenarios described above. In addition, we assessed the effect of excluding rare alleles using P_{crit} cutoffs, which is important in LD-based N_e estimation. For all data sets, we separately applied a P_{crit} of 0.01, 0.02, and 0.05. A P_{crit} of 0.02 has been recommended to balance precision and bias (Waples and Do 2010), although 0.05 is a common value used in SNP-based studies.

We used fsc2 to perform demographic inference using the joint SFS generated from serial samples taken at generations 0 (t_0) and 20 (t_{20}) in our temporally simulated populations. For all fsc2 analyses, we used a simple model of a single population with N_e at t_0 fixed at the known starting value and N_e in subsequent generations allowed to vary according to the model. Fixing N_e at t_0 allowed us to reduce the number of parameters estimated from the model, scale N_e estimation without a mutation rate, and ignore invariant sites in the SFS. Defined parameter ranges were uniformly distributed with N_e ranging from 1 to 10,000. A total of 100,000 simulations were performed to estimate the SFS, with a minimum and maximum of 10 and 100 loops (ECM cycles), respectively. The stopping criterion was defined as the minimum relative difference in parameters between two iterations, and was set to 0.001. A total of 50 replicate fsc2 runs were performed for each replicate simulation of a demographic scenario, and for each of the

three fsc2 filtering options described above. The overall maximum likelihood run across all 50 fsc2 replicates was retained as a point estimate for N_e^{t20} . Due to computational limitations, for each combination of initial population size and population growth rate, only the first 40 temporally simulated replicates (out of 100) were analyzed with fsc2.

Accuracy assessments

The performance of each N_e estimation method was evaluated for the overall accuracy of N_e estimates. To characterize the accuracy of N_e estimates across simulation replicates, we measured the root mean squared error (RMSE) calculated after removing infinitely large estimates by

$$\text{RMSE} = \sqrt{\frac{1}{m} \sum_{i=1}^m \left(\frac{1}{\hat{N}_{ei}} - \frac{1}{N_e} \right)^2},$$

where \hat{N}_{ei} is the estimated N_e in the i th ($i = 1-100$) replicate, and N_e is the simulated N_e . The RMSE was not calculated if over 50% of the estimates of \hat{N}_{ei} reached infinity.

Detection of population size change

To estimate population size trends, we calculated $\hat{\lambda}$ as the slope of a linear regression of the log transformation of N_e estimates from current and historical samples within a simulated replicate and we compared these to known λ . We performed these calculations for results generated from both NeEstimator and fsc2 using all simulated demographic scenarios, data-filtering scenarios, and P_{crit} levels. Following Tallmon et al. (2010), we recorded the proportion of times $\hat{\lambda} < 0.95$ when true $\lambda = 0.9$. This is a practical conservation scenario to identify populations that are declining by at least 5% per generation. We also assessed how often a stable population was incorrectly identified as declining as the proportion of times $\hat{\lambda} < 0.95$ when true $\lambda = 1.0$ (false positive rate).

Results

The number of SNPs generated in the simulation depended on the initial population size, imposed lambda, and the post-simulation filtering scenario used (LD-based data: Table 1; fsc2 data: Table S1). Consistent with theoretical expectations, in the LD-based SNP data sets, larger populations generally had more SNPs and lost genetic diversity less rapidly due to drift, and declining populations lost genetic diversity more rapidly than stable populations. The mean number of SNPs in the joint SFS was highly dependent on data-filtering method, with the number of shared SNPs

Table 1 Number of SNPs used for LD-based analysis resulting from simulations in simuPop

	λ	Initial variation at t_0			Final variation at t_{20}		
		RAD mutation # SNPs	10% # SNPs	50% # SNPs	RAD mutation # SNPs	10% # SNPs	50% # SNPs
N							
250	1.0	3660	3938	4527	3521	3695	4223
500	1.0	3718	4054	4657	3629	3876	4451
1000	1.0	3756	4130	4742	3703	4013	4611
250	0.9	3915	3941	4676	3099	2965	3457
500	0.9	3719	4053	4661	3392	3493	3975
1000	0.9	3756	4128	4739	3570	3773	4327

SNPs were calculated as average across the 100 replicates for each simulated scenario and are presented for the initial generation (t_0) after the 10-generation equilibrium phase and from the final generation (t_{20}). SNP levels are broken down across the different initial population sizes (N), population growth rates (λ), and data-filtering methods (LD RAD mutation, 10% missing, and 50% missing data sets).

between t_0 and t_{20} declining with allele dropout from both RADseq mutation and insufficient sequencing coverage. Although the number of SNPs will vary with study design, such as the number of individuals multiplexed in an Illumina sequencing lane, and coverage cutoffs, the number of SNPs we recovered in our simulations is comparable to empirical RADseq studies.

Stable population size estimation

LD-based estimation

Here we focus on results from estimation of \hat{N}_e at t_{20} under a $\lambda = 1.0$, where the accuracy of \hat{N}_e estimation was most influenced by the number of individuals sampled and the P_{crit} employed (Fig. 1; Fig. S1). Estimates of \hat{N}_e at time points t_0 through t_{15} were nearly identical to \hat{N}_e at t_{20} , and are not presented here. RMSE calculations yielding the lowest measures of error for all simulated demographic and filtering scenarios are presented in Table 2. The lowest individual sample size ($n = 15$) only produced meaningful results at a simulated population size of $n = 250$ and a $P_{crit} = 0.05$, with the majority of replicates at higher simulated population sizes and/or different filtering methods yielding either infinite \hat{N}_e or very wide ranges of parameter estimates. A full summary of the proportion of replicate estimates that reached infinity can be found in Tables S2–S4. In contrast, increased individual sampling ($n = 30$ and $n = 60$) produced more accurate estimates of \hat{N}_e over most demographic and data-filtering scenarios. Analyses of the LD RAD mutation data set generated \hat{N}_e estimates with the greatest accuracy and least variance; however, data sets with 10 and 50% missing data due to both cut site mutations and

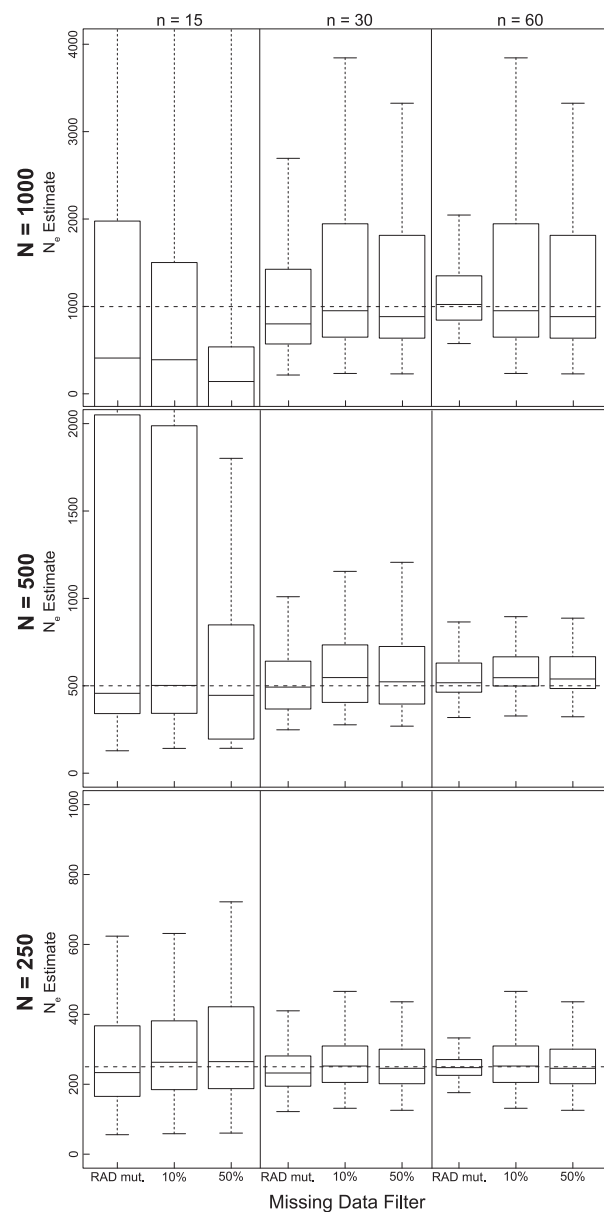


Fig. 1 Boxplots of the distribution of \hat{N}_e estimates from 100 replicate simulations for LD-based estimation at generation 20 from temporal simulations under stable population sizes ($\lambda = 1.0$) with a $P_{crit} = 0.05$. Dashed lines represent true N_e for the three population size models (1000, 500, and 250). Different missing data-filtering strategies are shown at the bottom of the figure. The number of individuals sampled is shown at the top

insufficient read coverage also generated similarly accurate \hat{N}_e estimates under many simulated population sizes and P_{crit} levels. The P_{crit} level yielding the most accurate results varied with the number of individuals sampled and simulated population size. Generally, including low frequency alleles with an $P_{crit} = 0.01$ appeared to have the largest effect by upwardly biasing \hat{N}_e and yielding the greatest variance (Fig. S1).

Table 2 RMSE values for all filtering scenarios for LD-based analysis in NeEstimator under a stable population ($\lambda = 1.0$)

		LD RAD mutation			10% missing data			50% missing data		
		(& P_{crit} level)			(& P_{crit} level)			(& P_{crit} level)		
		0.01	0.02	0.05	0.01	0.02	0.05	0.01	0.02	0.05
$N = 250$	$n = 15$	2.73E-03	2.73E-03	2.95E-03	3.41E-03	3.41E-03	2.73E-03	3.68E-03	3.68E-03	2.70E-03
	$n = 30$	1.41E-03	1.10E-03	1.25E-03	1.55E-03	1.11E-03	1.18E-03	1.532E-03	1.11E-03	1.23E-03
	$n = 60$	6.88E-04	6.17E-04	6.10E-04	1.55E-03	1.11E-03	1.18E-03	1.53E-03	1.11E-03	1.23E-03
$N = 500$	$n = 15$	Inf.	Inf.	1.60E-03	Inf.	Inf.	1.54E-03	Inf.	Inf.	1.62E-03
	$n = 30$	9.65E-04	7.98E-04	8.41E-04	1.16E-03	7.79E-04	8.06E-04	1.16E-03	7.79E-04	8.15E-04
	$n = 60$	4.59E-04	4.63E-04	4.40E-04	4.99E-04	4.56E-04	4.54E-04	5.02E-04	4.47E-04	4.43E-04
$N = 1000$	$n = 15$	Inf.	Inf.	1.99E-03	Inf.	Inf.	1.84E-03	Inf.	Inf.	1.87E-03
	$n = 30$	6.65E-04	7.76E-04	8.90E-04	8.03E-04	7.44E-04	7.87E-04	8.23E-04	7.32E-04	8.14E-04
	$n = 60$	3.01E-04	3.14E-04	3.04E-04	8.03E-04	7.44E-04	7.87E-04	8.23E-04	7.32E-04	8.14E-04

Bold values identify the lowest RMSE for a particular combination of population size (N), individual sampling level (n), and minor allele frequency cutoff (P_{crit}). Bold and italicized values identify the lowest RMSE for a particular population size. RMSE was not estimated if over 50% of the estimates of \hat{N}_e reached infinity for a particular parameter combination (Inf.)

Fastsimcoal2

Estimation of \hat{N}_e at t_{20} under a $\lambda = 1.0$ population model was most influenced by the number of SNPs included in the SFS (Fig. 2a, b, c) and, therefore, the allele dropout filtering scenario was used (Fig. 2S A–C). Overall, the fsc2 RAD mutation and fsc2 RAD mutation and coverage data sets yielded similar precision and accuracy compared to data sets using a similar number of randomly chosen SNPs from the fsc2 complete data set. Increased individual sampling had a slight improvement on accuracy and/or precision under all three population size models. However, analysis of 60-individual data sets in combination with lower numbers of SNPs (5000 and 10,000), including the fsc2 RAD mutation and coverage data sets, yielded a very wide range of estimates under all three population size models, with highly inaccurate and negatively biased estimates under a $n = 250$ model. In general, accuracy and precision in all scenarios proportionally decreased with the number of SNPs in the data set.

The 150,000 SNP data set yielded the lowest RMSE values for the $n = 250$ and $n = 500$ population models, and when sampling 60 individuals in the $n = 1000$ model (Table 3). Subsampled SNP data sets with 25,000 or more SNPs yielded only small decreases in RMSE with increasing numbers of SNPs. In the allele dropout data sets, the fsc2 complete data set yielded the lowest RMSE values for the $n = 250$ and $n = 500$ population models, and when sampling 60 individuals in the $n = 1000$ model (Table S5). Overall, fsc2 RAD mutation and fsc2 RAD mutation and coverage data sets had similar RMSE values.

Declining population size estimation

LD-based estimation

The number of generations since the beginning of a population decline was the biggest factor affecting the accuracy and precision of \hat{N}_e estimation (Fig. 3, Figs. S3–S5), with the variance in estimates decreasing over time as population size declined. Individual sampling also affected results, with an $n = 15$ yielding a greater estimation variance, particularly in earlier generations of the decline. Estimation using an $n = 30$ or 60 produced highly accurate estimates of \hat{N}_e in t_{10} through t_{20} . In general, \hat{N}_e estimation over time was only minimally affected by the initial population size, the missing data filter used, or the P_{crit} used. However, with individual samples size of $n = 15$ a P_{crit} of 0.05 lead to a greater proportion of finite \hat{N}_e (Table S8–S10).

Similarly, estimation of $\hat{\lambda}$ over different time intervals was most influenced by the number of generations passing between sampling events. The data filter used had minimal impact on the accuracy of $\hat{\lambda}$ estimation and we present results from analyses of the 10% missing data here (Table 4) with results from analysis of additional allele dropout data sets presented in Tables S6–S7. When sampling 30–60 individuals, the P_{crit} did not have a large impact on population trend detection, but with an individual samples size of 15, a P_{crit} of 0.05 improved population trend detection. For example, when sampling 15 individuals, population declines with an initial $n \leq 500$ were detected 67% of the time when at least ten generations passed, and increased to 85% of the time when 20 generations passed. With $n = 15$ and an initial $n = 1000$, at least 20 generations must pass for population declines to be detected 64% of the time. However, with $n = 15$ using a P_{crit} of 0.05 also increased the

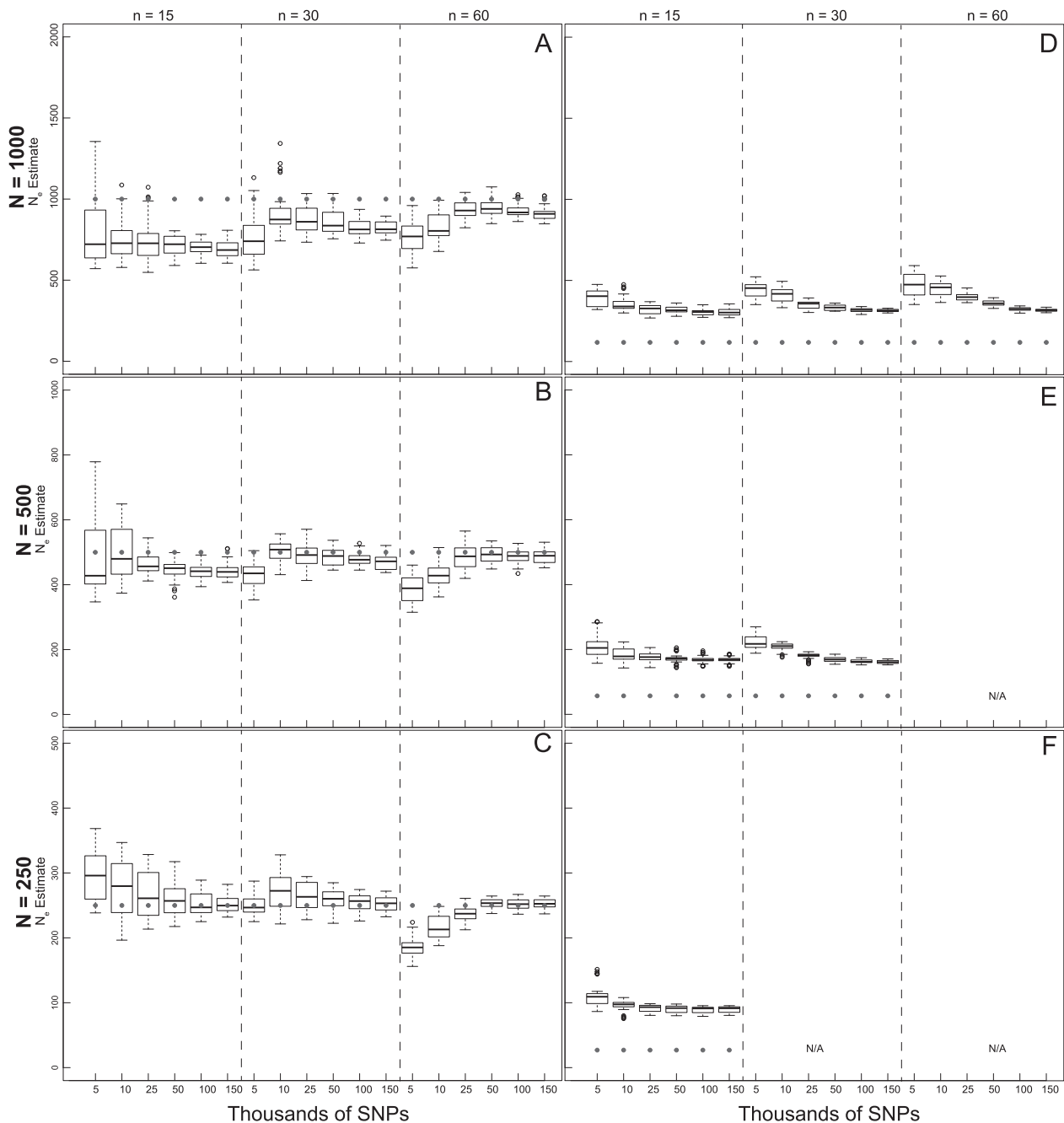


Fig. 2 Boxplots of the distribution of fastsimcoal2 estimates of \hat{N}_e at t_{20} from 40 replicate temporal simulations. **a–c** \hat{N}_e estimates under a stable population size ($\lambda = 1.0$) for population sizes of **(a)** 1000, **(b)** 500, and **(c)** 250. **d–f** \hat{N}_e estimates under declining population size ($\lambda = 0.9$), for initial population size of **(d)** 1000, **(e)** 500, and **(f)** 250. Red

dots represent the true N_e at t_{20} . Results are broken down across the number of individuals sampled (identified at the top of each panel) and the different numbers of SNPs used in analysis (identified at the bottom of each panel). For some parameter combinations, there were insufficient numbers of individuals for target n

false positive rate, where stable populations were incorrectly identified as declining with $\hat{\lambda}$ estimates of < 0.95 across many replicates (Table 5, Tables S11–S12). Increased individual sampling greatly improved the correct identification of a declining population. For example, under an $n = 1000$ model, sampling 60 individuals resulted in the correct identification of a population decline $> 95\%$ of the

time when 10 generations passed and correct identification $> 71\%$ of the time after just five generations.

Fastsimcoal2

The accuracy of \hat{N}_e at t_{20} was most influenced by the number of SNPs included in the joint SFS (Figs. 2d, e, f),

Table 3 RMSE values for increasing number of SNPs using fastsimcoal2 under a stable population ($\lambda = 1.0$)

		5000	10,000	25,000	50,000	100,000	150,000
$N = 250$	$n = 15$	0.000357	0.000357	0.000267	0.000201	0.000136	0.000101
	$n = 30$	0.000118	0.000234	0.000178	0.00014	0.000103	8.58E-05
	$n = 60$	0.000738	0.000367	0.000153	6.30E-05	6.14E-05	5.73E-05
$N = 500$	$n = 15$	0.000233	0.00017	0.000105	0.000153	0.000152	0.000147
	$n = 30$	0.00019	6.69E-05	8.29E-05	6.43E-05	6.10E-05	8.13E-05
	$n = 60$	0.000352	0.000191	8.78E-05	5.30E-05	5.33E-05	5.28E-05
$N = 1000$	$n = 15$	0.000217	0.000208	0.000217	0.000208	0.00022	0.000234
	$n = 30$	0.000205	8.59E-05	9.19E-05	9.73E-05	0.000115	0.000113
	$n = 60$	0.000188	1.18E-04	4.66E-05	4.04E-05	4.52E-05	5.50E-05

Bold values identify the lowest RMSE for a particular combination of population size (N) and individual sampling level (n). Bold and italicized values identify the lowest RMSE for a particular population size

and therefore also the allele dropout filter used (Fig. S2 D-F). Estimates of \hat{N}_e at t_{20} were positively biased across all data sets, with greater bias in data sets with fewer numbers of SNPs. Similarly, estimation of $\hat{\lambda}$ was most influenced by the number of SNPs included in the joint SFS. When sampling 5000–10,000 SNPs, population declines were detected < 50% of the time across most scenarios (Table S13). With samples of 50,000–150,000 SNPs, population declines were detected across most replicates for an initial N of 500 and 1000. Population declines were not reliably detected for an initial N of 250 for any sampling scenario. For the allele dropout data sets, population declines of $\hat{\lambda} < 0.95$ were detected across all 40 analyzed replicates using the fsc2 complete data set (Fig. S2 D–F). In contrast, none of the replicates for either fsc2 RAD mutation, or fsc2 RAD mutation and coverage data sets meet our criteria of $\hat{\lambda} < 0.95$, although most qualitatively indicated decline relative to N_e at t_0 . Stable populations were never identified as declining in any data set examined.

Discussion

Our results demonstrate that RADseq data have the potential to improve the inference of population demography and the detection of population declines on a very recent time scale. The linkage disequilibrium and coalescent methods we applied to estimate N_e use largely different sources of information from genomic data sets. The relative performance of these methods was influenced by different factors related to the study design, such as the number of individuals sampled (important for LD-based estimation) and the amount of variable data generated (important for coalescent estimation). Given that the accuracy and precision of N_e estimators hinge on aspects of the study design and the underlying population history, we further discuss these influences and provide guidelines for inferring N_e and

population size trends. While we compare and contrast the performance of both estimators, combining results from both methods in empirical studies may be the best approach to develop an encompassing view of overall population demographic history, as suggested by Waples (2016).

Performance of estimators

In our analysis of RADseq data, LD-based demographic inference generally outperformed coalescent-based inference for N_e estimation and the detection of population declines. However, there were limitations with LD-based inference, most notably with the number of sampled individuals required to provide both accurate and precise results. Sampling of 15 individuals led to large variance in estimates. This was most evident under a stable population size and in early generations of a population decline, particularly when population size was large (e.g., $N = 1000$). In contrast, increasing sampling to 30 individuals greatly increased the accuracy and precision of N_e estimates. This may be discouraging from the perspective of sampling, as many-population genetic studies sample far fewer than 30 individuals per population. However, in light of microsatellite-based simulations showing that 30 individuals resulted in largely biased N_e estimation (Tallmon et al. 2010), LD-based analysis of RADseq appears to provide new opportunities for accurate demographic inference.

In contrast, coalescent-based N_e estimation (using fsc2) was not greatly affected by the number of individuals sampled, with highly precise N_e estimates produced using as few as 15 individuals. This result is similar to those obtained with ABC estimates based on large genomic data sets (Robinson et al. 2014). The most significant limitation for the coalescent approach was the number of SNPs in the joint SFS, and therefore the data filter used. We found that sampling 25,000 SNPs, and in some cases as many as 50,000 SNPs, were required to obtain accurate estimates of

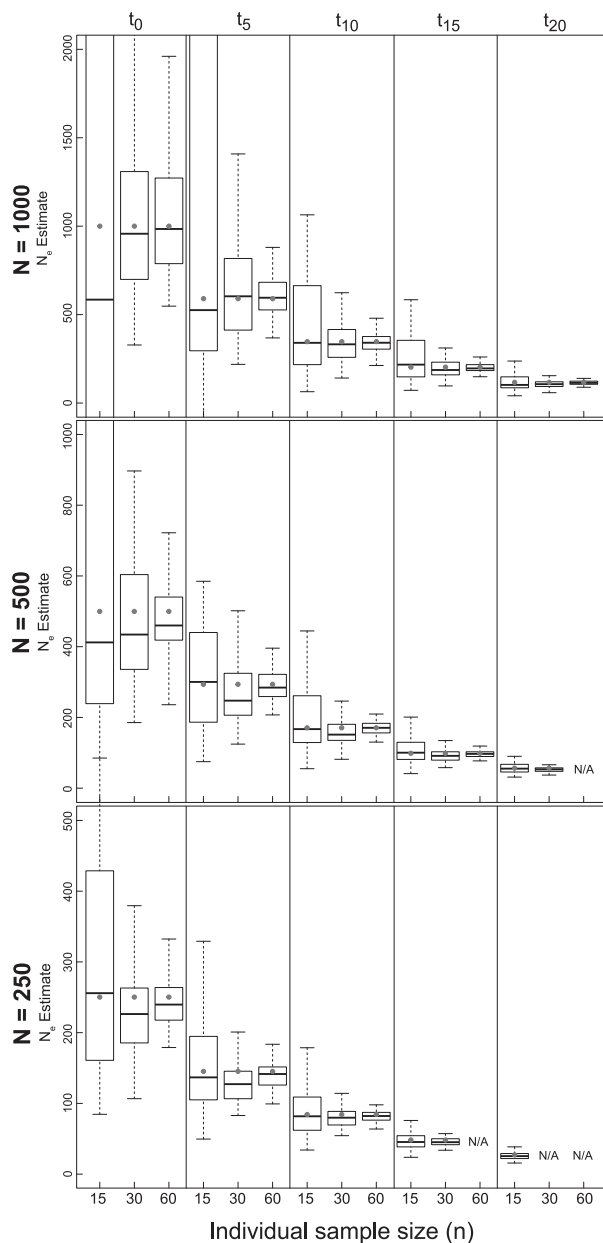


Fig. 3 Boxplots of the distribution of point estimates from 100 replicate simulations for LD-based N_e estimation from five temporal sampling points (t_0 – t_{20}) under declining population growth model ($\lambda = 0.9$) using the 10% missing data set and a $P_{\text{crit}} = 0.05$. Red dots represent true N_e over time, starting from an initial N of 1000 (top), 500 (middle), or 250 (bottom). Results are also broken down across different levels of individual sample size ($n = 15, 30$, or 60). For some parameter combinations, there were insufficient numbers of individuals for target n

N_e under a stable population model, with minimal increases in accuracy with greater number of SNPs. Previous simulation studies using coalescent-based ABC approaches found similar limitations with population size difficult to estimate even with 50,000 loci in some cases (Shafer et al. 2015b). All data sets yielded a consistent upward bias in N_e

estimation in the declining populations (Figs. 2d, e, f), and we are not sure what drives this estimation bias, but it was most pronounced in the data sets with fewer SNPs. Despite this positive bias, population declines were obvious using $\geq 50,000$ SNPs at 20 generations from initiating declines. Due to the intense computational needs inherent to *fsc2*, N_e was not estimated at earlier time points. Interestingly, detection of population declines were more difficult when initial population size was smaller (i.e., $N = 250$). While complete data sets similar to the ones used here are not attainable in empirical research, the positive correlation between numbers of SNPs and accurate coalescent-based N_e estimation is encouraging. Technological improvements and sequencing costs continue to increase our ability to generate more complete genome-wide SNP data, even when factoring in allele dropout. In contrast, increasing sample size, especially temporally, will remain difficult for many species. Our use of true N_e as a prior for one of our sampled years is also unlikely to be available in most study systems, which would further model complexity and add analytical time to an already computationally challenging set of analyses. Ultimately, coalescent-based demographic inference using a joint SFS-based method may be a great option for a more limited set of studies with access to large SNP data sets, and prior population information, as has been illustrated in a number of empirical studies (McCoy et al. 2013; Fraser et al. 2015; Nunziata et al. 2017).

Allele dropout and data filtering

Missing data via allele dropout in RADseq studies has been shown to affect a number of population genetic summary statistics, including measures of genetic diversity and population structure (Arnold et al. 2013; Gautier et al. 2013). Our results from parameter estimators for N_e are therefore encouraging, as increasing levels of missing data via allele dropout had little impact on LD-based N_e estimation and were generally comparable to the data set with no null alleles. Interestingly, while LD-based estimation was robust to the effects of allele dropout and missing data, the P_{crit} influenced \hat{N}_e accuracy and precision, particularly under a model of stable population size. These results are consistent with other studies (Waples and Do 2010), where the inclusion of low frequency alleles created a positive bias, while the exclusion of these alleles created a slightly negative bias, particularly at the lowest sample size (Waples and Do 2010). Also consistent with the guidelines outlined in Waples and Do (2010), when low individual samples sizes were used ($n = 15$) a P_{crit} of 0.05 yielded the most finite and accurate estimates, as it is the only P_{crit} that screened out singletons, which can bias \hat{N}_e .

In contrast to the LD-based analyses, the allele dropout filter used in the *fsc2* analyses did affect the results.

Table 4 Number of times that a declining population trend was correctly identified out of 100 replicate runs for LD-based analysis in NeEstimator under a declining population model ($\lambda = 0.9$)

		t_0-t_5			t_0-t_{10}			t_0-t_{15}			t_0-t_{20}		
		(& P_{crit} level)			(& P_{crit} level)			(& P_{crit} level)			(& P_{crit} level)		
		0.01	0.02	0.05	0.01	0.02	0.05	0.01	0.02	0.05	0.01	0.02	0.05
$N = 250$	$n = 15$	49	49	68	63	63	83	71	71	95	72	72	99
	$n = 30$	86	79	79	99	94	93	100	100	100	—	—	—
	$n = 60$	94	94	94	100	100	100	—	—	—	—	—	—
$N = 500$	$n = 15$	8	8	48	18	18	67	25	25	79	29	29	85
	$n = 30$	79	73	72	94	84	86	100	99	98	100	99	99
	$n = 60$	80	81	84	97	98	98	100	100	100	—	—	—
$N = 1000$	$n = 15$	0	0	35	0	0	40	2	2	53	2	2	64
	$n = 30$	57	65	66	70	80	79	76	91	91	78	97	97
	$n = 60$	71	73	74	96	95	96	100	100	100	100	100	100

Results are presented for increasing intervals of time and by the combination of population size (N), individual sampling level (n), and minor allele frequency cutoff (P_{crit}). Results are based on the analysis of data sets with 10% missing data.

For some parameter combinations, there were insufficient numbers of individuals for target n (—)

Table 5 Number of times that a population trend was incorrectly identified as declining out of 100 replicate runs for LD-based analysis in NeEstimator under a stable population model ($\lambda = 1.0$)

		t_0-t_5			t_0-t_{10}			t_0-t_{15}			t_0-t_{20}		
		(& P_{crit} level)			(& P_{crit} level)			(& P_{crit} level)			(& P_{crit} level)		
		0.01	0.02	0.05	0.01	0.02	0.05	0.01	0.02	0.05	0.01	0.02	0.05
$N = 250$	$n = 15$	26	26	38	24	24	29	24	24	24	20	20	10
	$n = 30$	29	27	25	20	13	12	15	9	8	3	1	1
	$n = 60$	8	9	8	1	1	1	1	1	1	0	0	0
$N = 500$	$n = 15$	1	1	38	0	0	32	2	2	29	3	3	23
	$n = 30$	47	41	43	36	21	22	29	16	15	20	8	8
	$n = 60$	26	24	27	6	6	6	1	1	1	0	0	0
$N = 1000$	$n = 15$	1	1	20	0	0	12	0	0	10	0	0	9
	$n = 30$	16	36	37	11	30	28	16	27	27	7	18	18
	$n = 60$	21	18	25	14	13	12	4	4	4	1	1	0

Results are presented for increasing intervals of time and by combination of population size (N), individual sampling level (n), and minor allele frequency cutoff (P_{crit}). Results are based on the analysis of data sets with 10% missing data.

However, allele dropout data sets did not appear to create any systematic bias compared to data sets using a similar number of randomly chosen SNPs from the fsc2 complete data set. Because these analyses preclude the use of loci with missing data, the direct impact of filtering loci by allele dropout was a major reduction of the number of SNPs included in the joint SFS. Contemporary population declines purge rare alleles, creating a predictable signature in the SFS (Nei et al. 1975; Gattepaille et al. 2013), with the likelihood of detecting this signature increasing with the number of SNPs included in the data set. We found that N_e estimation was accurate, and declines were reliably detected, using our data set containing $\geq 50,000$ SNPs. The generation of empirical data sets robust enough to detect

population declines may, therefore, require increased sequencing efforts to offset the effects of allele dropout by increasing the number of loci sampled and their coverage. Maybe counter intuitively, increased individual sampling does not solve this problem as adding individuals increases the probability of allele dropout through a cut site mutation or insufficient sequencing coverage, creating a smaller SNP matrix and decreasing precision in \hat{N}_e (Fig. 2S). Potentially, this result can be overcome by subsampling individuals for non-missing data (e.g., Papadopoulou and Knowles 2015).

Allele dropout often goes undetected in many studies, and our preliminary exploration suggests that the underlying population history of either stable or declining populations were recovered and point estimates were almost

always within an order of magnitude of real N_e . Previous simulation work has revealed that non-equilibrium demography, such as a population decline, can cause low N_e and result in fewer loci with missing data and more accurate allele frequency estimation (Arnold et al. 2013). Therefore, our findings should not be interpreted as applicable across systems, since we may have modeled scenarios (i.e., low N_e , steadily declining) that create evident signatures in the SFS at a contemporary time scale.

Practical considerations

Many additional factors influence N_e that we have not modeled here, including selection, migration, and overlapping generations (Slatkin 2008). In real populations, N_e rarely equals N_C , and changes in N_e could track any number of demographic changes, not exclusively N_C (Palstra and Ruzzante 2008). Further simulations are needed under more realistic scenarios to determine the application of evaluated methods across systems. One factor that must be considered with RADseq data sets and the LD-based approach is that although pairwise r^2 values (correlation of genes within individuals) increase with number of loci, SNPs on the same chromosome are not independent and will reduce the precision of \hat{N}_e because LD will be the result of physical linkage and not drift (Waples et al. 2016). The use of linked SNPs could be corrected for by using known genomic architecture (Waples et al. 2016); and is an important consideration in the application of LD-based N_e estimation to RADseq data.

Both LD and coalescent methods produced a time lag between census size declines and corresponding decline in N_e . The LD-based method has potential for accurate detection of population declines, generally after only 10 generations from initiation of a decline. However, if working with long-lived species with long-generation times, these 10 generations could equate to several decades within which populations could decline rapidly toward extinction with a little change in N_e . Given these findings, we emphasize that genomic monitoring is not a replacement for traditional census size monitoring in many cases, but may serve as an informative complement.

When inferring $\hat{\lambda}$ from \hat{N}_e for conservation purposes, false positives can lead to a waste of management resources when stable populations are misidentified as declining (Schwartz et al. 2007). The absence of any false positives in the fsc2-based λ estimation, and the lower number of individuals required, is promising for its application in conservation studies. However, the failure to detect declines in most replicates with < 25,000 SNPs highlights the need for very large SNP data sets, as well as temporal sampling, especially if quick detection of population declines is a goal. False positives for LD-based λ estimates were also low,

although this typically required larger sample sizes of at least 30 individuals. With large resources available to researchers, the application of both methods for demographic inference will be the ideal approach to take, but given constraints on sampling or sequencing, the results here can be useful for guiding decisions about how to design a conservation genetic study aimed at detecting recent population declines. Finally, even when temporal sampling is unavailable, N_e is itself an important indicator of population viability and evolutionary potential and RADseq data can serve as a valuable source of information for this parameter.

Data archiving

All simulation scripts Data available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.6d925>.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Andreotti S, Rutzen M, Van der Walt S, von der Heyden S, Henriques R, Meyer M, Oosthuizen H, Matthee CA (2016) An integrated mark-recapture and genetic approach to estimate the population size of white shark in South Africa. *Mar Ecol Progress Ser* 552:241–253
- Andrews KR, Good JM, Miller MR, Luikart G, Hohenlohe PA (2016) Harnessing the power of RADseq for ecological and evolutionary genomics. *Nat Rev Genet* 17:81–92
- Antao T, Pérez-Figueroa A, Luikart G (2011) Early detection of population declines: high power of genetic monitoring using effective population size estimators. *Evol Appl* 4:144–154
- Arnold B, Corbet-Detig RB, Hartl D, Bomblies K (2013) RAD-seq underestimates diversity and introduces genealogical biases due to nonrandom haplotype sampling. *Mol Ecol* 22:3179–3190
- Davey JW, Hohenlohe PA, Etter PD, Boone JQ, Catchen JM, Blaxter ML (2011) Genome-wide genetic marker discovery and genotyping using next-generation sequencing. *Nat Rev Genet* 12:499–510
- Do C, Waples RS, Peel D et al. (2014) NeEstimator V2: reimplementation of software for the estimation of contemporary effective population size (Ne) from genetic data. *Mol Ecol Resour* 14:209–214
- Dudgeon CL, Ovenden JR (2015) The relationship between abundance and genetic effective population size in elasmobranchs: an example from the globally threatened zebra shark *Stegostoma*

- fasciatum within its protected range. *Conserv Genet* 16:1443–1454
- Excoffier L, Dupanloup I, Huerta-Sanchez E, Sousa VC, Foll M (2013) Robust demographic inference from genomic and SNP data. *PLoS Genet* 9:e1003905
- Fraser BA, Kunstner A, Reznick DN, Dreyer C, Weigel D (2015) Population genomics of natural and experimental populations of guppies (*Poecilia reticulata*). *Mol Ecol* 24:389–408
- Gautier M, Gharbi K, Cezard T et al. (2013) The effect of RAD allele dropout on the estimation of genetic variation within and between populations. *Mol Ecol* 22:3165–3178
- Gattepaille LM, Jakobsson M, Blum MGB (2013) Inferring population size changes with sequence and SNP data: lessons from human bottlenecks. *Heredity* 110:409–419
- Gutenkunst RN, Hernandez RD, Williamson SH, Bustamante CD (2009) Inferring the joint demographic history of multiple populations from multidimensional SNP frequency data. *PLoS Genet* 5:1–11
- Hill WG (1981) Estimation of effective population size from data on linkage disequilibrium. *Genet Res* 38:209–216
- Hoban SM, Gaggiotti OE, Bertorelle G (2013) The number of markers and samples needed for detecting bottlenecks under realistic scenarios, with and without recovery: a simulation-based study. *Mol Ecol* 22:3444–3450
- Hollenbeck CM, Portnoy DS, Gold JR (2016) A method for detecting recent changes in contemporary effective population size from linkage disequilibrium at linked and unlinked loci. *Heredity* 117:207–216
- Huang H, Knowles LL (2014) Unforeseen consequences of excluding missing data from next-generation sequences: simulation Study of RAD Sequences. *Syst Biol* 65:357–365
- Kopuchian C, Campagna L, Di Giacomo AS et al. (2016) Demographic history inferred from genome-wide data reveals two lineages of sheldgeese endemic to a glacial refugium in the southern Atlantic. *J Biogeogr* 43:1979–1989
- Lischer HEL, Excoffier L (2012) PGDSpider: an automated data conversion tool for connecting population genetics and genomics programs. *Bioinformatics* 28:298–299
- Luikart G, Ryman N, Tallmon DA, Schwartz MK, Allendorf FW (2010) Estimation of census and effective population sizes: the increasing usefulness of DNA-based approaches. *Conserv Genet* 11:255–373
- McCoy RC, Garud NR, Kelley JL, Boggs CL, Petrov D (2013) Genomic inference accurately predicts the timing and severity of a recent bottleneck in a nonmodel insect population. *Mol Ecol* 23:136–150
- Nachman MW, Crowell SL (2000) Estimate of the mutation rate per nucleotide in humans. *Genetics* 156:297–304
- Narum SR, Buerkle CA, Davey JW, Miller MR, Hohenlohe PA (2013) Genotyping-by-sequencing in ecological and conservation genomics. *Mol Ecol* 22:2841–2847
- Nei M, Maruyama T, Chakraborty R (1975) The bottleneck effect and genetic variability in populations. *Evolution* 29:1–10
- Nordborg M, Krone SM (2002) Separation of time scales and convergence to the coalescent in structured populations. In: Slatkin M, Veuille M (eds) *Modern developments in theoretical population genetics*. Oxford University Press, Oxford, p 130–164
- Nunziata SO, Lance SL, Scott DE, Lemmon EM, Weisrock DW (2017) Genomic data detect corresponding signatures of population size change on an ecological time scale in two salamander species. *Mol Ecol* 26:1060–1074
- Palstra FP, Ruzzante DE (2008) Genetic estimates of contemporary effective population size: what can they tell us about the importance of genetic stochasticity for wild population persistence? *Mol Ecol* 17:3428–3447
- Papadopoulou A, Knowles LL (2015) Genomic tests of the species-pump hypothesis: recent island connectivity cycles drive population divergence but not speciation in Caribbean crickets across the Virgin Islands. *Evolution* 69:1501–1517
- Peng B, Kimmel M (2005) simuPOP: a forward-time population genetics simulation environment. *Bioinformatics* 21:3686–3687
- Pool JE, Hellmann I, Jensen JD, Nielsen R (2010) Population genetic inference from genomic sequence variation. *Genome Res* 20:291–300
- Prates I, Xue AT, Brown JL, Alvarado-Serrano DF, Rodrigues MT, Hickerson MJ, Carnaval AC (2016) Inferring responses to climate dynamics from historical demography in neotropical forest lizards. *Proc Natl Acad Sci USA* 113:7978–7985
- Robinson JD, Coffman AJ, Hickerson MJ, Gutenkunst RN (2014) Sampling strategies for frequency spectrum-based population genomic inference. *BMC Evol Biol* 14:254
- Rokas A, Abbot P (2009) Harnessing genomics for evolutionary insights. *Trends Ecol Evol* 24:192–200
- Schwartz MK, Luikart G, Waples RS (2007) Genetic monitoring as a promising tool for conservation and management. *Trends Ecol Evol* 22:25–33
- Shafer ABA, Wolf JBW, Alves PC et al. (2015a) Genomics and the challenging translation into conservation practice. *Trends Ecol Evol* 30:78–87
- Shafer ABA, Gattepaille LM, Stewart RE, Wolf JB (2015b) Demographic inferences using short-read genomic data in an approximate Bayesian computation framework: in silico evaluation of power, biases and proof of concept in Atlantic walrus. *Mol Ecol* 24:328–345
- Shafer ABA, Peart CR, Tusso S, Maayan I, Brelsford A, Wheat CW, Wolf JBW (2017) Bioinformatic processing of RAD-seq data dramatically impacts downstream population genetic inference. *Methods Ecol Evol* 8:907–917
- Slatkin M (2008) Linkage disequilibrium – understanding the evolutionary past and mapping the medical future. *Nat Rev Genet* 9:477–485
- Tallmon DA, Gregovich D, Waples RS et al. (2010) When are genetic methods useful for estimating contemporary abundance and detecting population trends? *Mol Ecol Resour* 10:684–692
- Trucchi E, Facon B, Gratton P, Mori E, Stenseth NC, Jentoft S (2016) Long live the alien: is high genetic diversity a pivotal aspect of crested porcupine (*Hystrix cristata*) long-lasting and successful invasion? *Mol Ecol* 25:3527–3539
- Wakeley J, Sargsyan O (2009) Extensions of the coalescent effective population size. *Genetics* 181:341–345
- Waples RS (2005) Genetic estimates of contemporary effective population size: to what time periods to estimates apply? *Mol Ecol* 14:3335–3352
- Waples RS (2006) A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. *Conserv Genet* 7:167–184
- Waples RS, Do C (2010) Linkage disequilibrium estimates of contemporary Ne using highly variable genetic markers: a largely untapped resource for applied conservation and evolution. *Evol Appl* 3:244–262
- Waples RK, Larson WA, Waples RS (2016) Estimating contemporary effective population size in non-model species using linkage disequilibrium across thousands of loci. *Heredity* 117:233–240
- Waples RS (2016) Making sense of genetic estimates of effective population size. *Mol Ecol* 25:4689–4691