REVIEW



FDA-approved drugs that are spermatotoxic in animals and the utility of animal testing for human risk prediction

Elizabeth R. Rayburn ¹ · Liang Gao ² · Jiayi Ding ³ · Hongxia Ding ⁴ · Jun Shao ³ · Haibo Li^{2,3}

Received: 25 August 2017 / Accepted: 5 October 2017 / Published online: 24 October 2017 © Springer Science+Business Media, LLC 2017

Abstract

Purpose This study reviews FDA-approved drugs that negatively impact spermatozoa in animals, as well as how these findings reflect on observations in human male gametes.

Methods The FDA drug warning labels included in the DailyMed database and the peer-reviewed literature in the PubMed database were searched for information to identify single-ingredient, FDA-approved prescription drugs with spermatotoxic effects.

Results A total of 235 unique, single-ingredient, FDA-approved drugs reported to be spermatotoxic in animals were identified in the drug labels. Forty-nine of these had documented negative effects on humans in either the drug label or literature, while 31 had no effect or a positive impact on human sperm. For the other 155 drugs that were spermatotoxic in animals, no human data was available.

Conclusion The current animal models are not very effective for predicting human spermatotoxicity, and there is limited information available about the impact of many drugs on human spermatozoa. New approaches should be designed that more accurately reflect the findings in men, including more studies on human sperm in vitro and studies using other

Elizabeth R. Rayburn and Liang Gao should be considered joint first authors.

- Haibo Li ntlihaibo2015@163.com
- CLIA Laboratory Director, Birmingham, AL 35242, USA
- Department of Clinical Laboratory Sciences, Nantong Maternity and Child Health Hospital, Nantong 226018, China
- Department of Reproductive Medicine, Nantong Maternity and Child Health Hospital, Nantong 226018, China
- ⁴ Pharmacodia (Beijing) Co., Ltd, Beijing 100085, China

systems (ex vivo tissue culture, xenograft models, in silico studies, etc.). In addition, the present data is often incomplete or reported in a manner that prevents interpretation of their clinical relevance. Changes should be made to the requirements for pre-clinical testing, drug surveillance, and the warning labels of drugs to ensure that the potential risks to human fertility are clearly indicated.

 $\textbf{Keywords} \ \, \text{Drug warning labels} \cdot \text{Infertility} \cdot \text{Reproductive toxicology} \cdot \text{Spermatotoxicity}$

Introduction

It has been estimated that approximately 5–10% of men in developed countries are affected by infertility due to abnormalities in spermatozoa (number, morphology, motility, etc.) or semen (volume, composition, viscosity, etc.) [1]. There are numerous causes, including genetics, hormone imbalances, dietary insufficiencies, exposure to environmental or occupational toxicants, or the use/abuse of drugs (recreational, overthe-counter, or prescription). Given that approximately half of the US population has taken a prescription drug in the last month [2], prescription drugs may represent a common source of spermatotoxicity.

The present review focuses on the spermatotoxic effects (from the generation of spermatogonia to fertilization) of prescription drugs in various experimental animals. This review was prompted by a previous publication that examined the impact of FDA-approved drugs on human spermatogenesis [3]. In that study, frequent discrepancies were noted in the drug warning labels between the adverse effects of drugs on human and animal spermatogenesis, as well as some inconsistencies between the published peer-reviewed literature and the DailyMed database (a federal repository of drug warning



labels). In addition to providing an overview of the spermatotoxicity of prescription drugs in animals, the review also provides a comparison to the reported human spermatotoxicity and addresses how the current data reflect the risk of these drugs to human sperm. This review also suggests ways to improve the current male reproductive toxicity testing and reporting of results.

Materials and methods

Database search and inclusion/exclusion criteria

The FDA indicates that any finding of toxicity should be reported on the drug labels in the "Warnings," "Adverse Reactions," and "Precautions" sections, as well as any other area where such a warning would be appropriate. The DailyMed database (http://dailymed.nlm.nih.gov/dailymed/index.cfm) is a federal database that contains approximately 85% of the US FDA-approved drug labels. This database represents the largest and most comprehensive database of the warning labels for FDA-approved drugs and is updated whenever a label changes.

Stemmed key words were used to search the full-text drug labels included in the DailyMed database (also available at: https://www.fda.gov/ScienceResearch/ BioinformaticsTools/ucm289739.htm) as described in a previous study [4]. These stemmed keywords (deferen, epidid, interstitial, Leydig, semen, semin, Sertoli, sperm, and testi) covered numerous terms related to spermatogenesis, pathological changes of the male reproductive organs (such as testicular tumors, testitis, and epididymitis), and direct effects on the spermatozoa. To ensure that the effects were due to the specific drugs, drugs with indirect effects on sperm (i.e., drugs with teratogenic effects impacting the fertility of male offspring) were excluded. Only FDA-approved prescription drugs developed for administration to humans were examined. Over-the-counter (OTC) drugs were excluded because the information available on their labels was generally insufficient and most have not been subjected to rigorous reproductive toxicity testing. The drugs with multiple active ingredients were also excluded to reduce the complexity of the analysis. When there were redundant labels for drugs produced by different companies or drugs with more than one label that had been updated at different times, only the most recent label was analyzed. Other labels for drugs intended for veterinary use, medicinal foods, medical devices, dietary supplements, cosmetics, etc., were also excluded. When the final stemmed keyword search was performed on August 8, 2017, there were 95,873 labels included in the DailyMed database. After restricting the information to single-ingredient drugs as described above, a

total of 251 unique prescription drug labels were identified that showed a negative impact on sperm, which were further narrowed down to 235 drugs because some of them were the same drug in different formulations that led to the same adverse effects (i.e., aripiprazole/aripiprazole lauroxil).

Data extraction and analysis

Two authors independently searched the DailyMed database and extracted relevant data. When there were differences in the descriptions extracted between these authors, they discussed the findings with additional authors, and the group's consensus regarding the findings was used for the analysis.

A PubMed search was also performed to determine whether animal or human toxicity had been reported for the 235 drugs identified to impact animal spermatogenesis based on the DailyMed labels. The PubMed database was searched using the terms ([generic drug name] AND sperm*, fertil*, semen, semin*, or testi*).

Results

The general findings of the database searches

A total of 235 FDA-approved drugs intended for human use and containing a single active ingredient were found to impact spermatogenesis in at least one type of animal. All of these drugs could result in a decrease in the sperm count/concentration or a decrease in sperm quality, viability, or fertilization capacity.

Rats were the model most often reported to be affected by the drugs (specifically reported for 184 of the 235 drugs). This was expected, because rats are the most commonly used animal model for reproductive toxicity testing. Dogs are often used to confirm the findings in rats, and were thus also frequently reported to experience changes in spermatogenesis due to the exposure to prescription drugs. There were many instances where only rats were listed as having changes in spermatogenesis following drug exposure, either because there was no toxicity in other animals or because confirmation testing was not performed. Interestingly, in about half of the cases, spermatotoxicity was only reported for dogs. This suggests that dogs may be particularly sensitive to spermatotoxicants. Only 34 of the 235 drugs were reported to affect primate spermatogenesis. However, it is likely that many of the drugs were not tested in primates given the ethical concerns and high expenses associated with testing in primates. In addition, although spermatotoxicity was reported for 11 other drugs in DailyMed, the type of animal(s) affected was not specified in the drug label.



The FDA-approved drugs reported to adversely affect animal spermatogenesis

Unsurprisingly, antineoplastic agents were the category of drugs most often reported to have effects on animal spermatogenesis. Antineoplastic agents can cause temporary or permanent damage to the spermatozoa, germ cells, or support cells. Hormones and hormone antagonists were also expected to have adverse effects on spermatogenesis, so it is not surprising that they make up the other category of drugs most frequently noted to be spermatotoxic in this study. Both classes of drugs are well-recognized as having a potential negative impact on male fertility, and protocols are in place in most hospitals and clinics to counsel patients about these effects and the means to preserve spermatozoa for future use.

On the other hand, there were numerous other drugs with unexpected spermatotoxicity, where the intended target/ mechanism of action is not generally considered to impact sperm. These drugs were from various classes, including adrenergic agents (icatibant, silodosin), analgesics (carbamazepine, morphine), antihypertensive agents (tamsulosin, reserpine), anti-infectious agents (metronidazole, miconazole), cardiovascular agents (bosentan, spironolactone), and immunosuppressants (sirolimus, azathioprine). Such agents are of greater concern because the adverse effects of these agents on fertility are not well-recognized and may not be addressed when the drugs are prescribed.

The types of spermatotoxic effects reported for animals

The most common adverse effect of the drugs was a reduced sperm count (due to reduced spermatogenesis or the death of developed and developing sperm), followed by reduced sperm motility, abnormal sperm morphology, testicular atrophy, and tumor formation (most frequently benign Leydig cell tumors in rodents). Similar observations of reduced sperm counts, reduced motility, and abnormal sperm morphology have been noted in humans for some of these drugs (see Table 1). In contrast to animal studies, where drug-related tumor formation was relatively common, drug-induced testicular tumors seem to be rare in humans. While some agents, such as anabolic steroids, are considered to cause testicular atrophy in men [44], the histological findings are generally not available, except for rare cases of injury or malignancy that require surgery. This makes it difficult to assess the full extent of the changes in humans.

Of note, most of the reports of adverse effects did not specify how many (what percentage) of the animals were affected. Therefore, it is unclear whether the observed adverse effect was common or relatively rare. The reporting was similarly vague for human effects, with many labels failing to indicate what percentage of the population was affected by the specific type of toxicity.

PubMed findings of human effects for these drugs

Only about 11% (26/235) of the drug warning labels that indicated that the agent had a negative impact on animal sperm (any aspect of spermatogenesis or toxicity to the sperm themselves) also had information in the drug label about a negative effect in humans. There were reports of adverse effects on human sperm for another 36 of these drugs in the PubMed database, although some of these publications were of in vitro studies, case studies, or studies of drug combinations that included the drug of interest (Table 1). Thus, between the drug labels in DailyMed and the reports included in PubMed, less than 30% of the drugs found to adversely affect animal sperm have been found to adversely affect human sperm.

For about half of the drugs that cited a negative impact on animal sperm, there were no data in DailyMed or PubMed about their impact on human sperm (Table 2). Eleven of the drugs without any publications in PubMed indicated that they had effects in humans in the DailyMed labels, although specific details were generally lacking regarding the number of patients affected. Interestingly, there were 33 drugs indicated to have negative effects in animals that were shown to have no effect or a positive impact on human sperm in the published literature (Table 3).

Discussion

The status of spermatotoxicity testing and relevance of the current testing

Testing for and reporting spermatotoxicity

The guidelines for industry issued by the FDA for new drug development stipulate that "...reproductive or developmental toxicity, whether in valid reproductive/developmental or other relevant nonclinical studies, should be evaluated to estimate the likelihood or other reproductive or developmental risk for humans". In addition, the "Females and Males of Reproductive Potential" section requires the inclusion of information when there are human and/or animal data suggesting drug-associated effects on fertility and/or preimplantation loss effects (§201.57(c)(9)(iii))" [102]. The FDA, as well as the US Environmental Protection Agency, Occupational Safety and Health Administration, and other national and international organizations, have made efforts to both streamline testing and make the testing as accurate as possible, specifically taking into consideration the route, dose, frequency, and duration of exposure, as well as species-associated differences. The guidelines also emphasize the need to understand the potential impact of drugs on fertility via both pre-clinical studies in animals and subsequent studies in patients, with an



 Table 1
 FDA-approved drugs with negative effects on sperm in both animals and humans

Main category	Drug name (generic)	Effect(s) in animals	Type of animal(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Support in PubMed	Support Reported effect(s) in PubMed
5-Alpha-reductase inhibitor	Dutasteride	Leydig cell adenomas, reduced cauda epiddymal sperm counts, reduced weights of the enididymis protate and seminal vesicles	Rats	Yes	Decreased sperm count, motility, and semen	[5]	Reversible decrease in the number and motility of sperm
Adrenergic agent	Silodosin	Decreased sperm viability and counts, changes in the testes and emididamides	Rats	No		[9]	Prevents sperm emission, but no
Alpha-blocker	Tamsulosin		Rats	No		[7]	Decreased sperm count
Analgesic	nydrocnlonde Acetaminophen	Increased abnormal sperm, reduced	Mice	No		[8]	Reduced motility, DNA
	Burrenormhine	spermatogenesis Lavdia call tumore	Pote	S. Y.		[0]	fragmentation
	Carbamazepine	Exyaig cen unions Benign interstitial cell adenomas in the testes, testicular atrophy, asnermatogenesis	Rats	N o		[10]	Decreased sperm motility and vitality
	Morphine sulfate	Changes in hormone levels (incl. testosterone and luteinizing hormone)	Rats	No		[11]	Decreased sperm motility
Anticonvulsant	Divalproex sodium	Reduced spermatogenesis, testicular atrophy	Rats and dogs	No		[12]	Decreased counts and motility, increased abnormal morphology, decreased resticular volume
	Oxcarbazepine	Benign testicular interstitial cell tumors	Rats	No		[13]	Increased abnormal morphology,
	Valproate sodium/	Reduced spermatogenesis and testicular atrophy	Rats and dogs	No		[12]	See divalproex above
Antidepressant	Varprore acid Clomipramine hvdrochloride	Phospholipidosis and testicular changes	Not specified	Yes	Testicular tumors	[14]	Reduced motility and abnormal morphology
	Paroxetine	Spermatogenic arrest, atrophic changes in the seminiferous tubules	Rats	Yes	Decreased sperm quality, enididymitis	[15]	Increases DNA fragmentation
	Vasopressin ^b	Decrease in the function and fertilizing ability of spermatozoa	Mice	No	chines in the	[16]	Decreased sperm count and motility
Antifungal	Ketoconazole	Increased abnormal sperm, decreased sperm mobility	Rats	No		[17]	Oligospermia, azoopermia
Antihyperlipidemic	Rosuvastatin calcium ^b	Spermatidic giant cells	Dogs and monkeys	No		[18]	Reversible azoospermia
Anti-infective agent	Miconazole	Chromosomal aberrations in spemnatocytes, morphological abnormalities in spem	Mice	No		[19]	Reduced vitality
	Lindane Quinine sulfate	Reduced spermatids Atrophy or degeneration of the seminiferous nibules decreased snern count and morility	Rats Mice	No No		[20, 21] [22]	Decreased quality, AR Decreased motility, altered semen volume
Antineoplastic agent	Busulfan°	Decreased spermatogenesis	Rats	Yes	Damaged sperm and testicular testicular tassue, testicular	[23]	Decreased spermatogenesis
	Carmustine° Cyclophosphamide	Testicular degeneration Decreased weights, atrophy, changes in spermatogenesis	Rats Mice and rats	No Yes	Interference with spermatogenesis, testicular atrophy,	[24] [25]	Gem/Leydig cell failure Azoospermia/oligospermia, decreased motility, abnormal morphology



(continued)
Table 1

Main category	Drug name (generic)	Effect(s) in animals	Type of animal(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Support in PubMed	Support Reported effect(s) in PubMed
	Daunorubicin	Testicular atrophy, aplasia of spermatocytes,	Dogs	No	azoospermia, oligospermia	[26]	Decreased sperm count, motility
	nydrochloride Doxorubicin hydrochloride ^{a, c}	complete aspermatogenesis Decreased testicular weights and hypospermia, diffuse degeneration of the seminiferous tibules decreased engangement	Mice	No		[26, 27]	and vitality Damages DNA, decreased sperm count, motility and vitality
	Etoposide°	Testicular atrophy	Rats	No		[28]	Decrease in sperm count
	Fludarabine phosphate ^b	Decreased testicular weights, degeneration and necrosis of spermatogenic epithelium of the	Mice, rats and dogs	Yes	Damaged sperm and testicular tissue	[29]	(possini) tevetsine) DNA damage
	Hydroxyurea	resticular atrophy, decreased spermatogenesis	Rats	No O		[30]	Adverse effects on sperm
	Ifosfamide (alphabetize)	Testicular atrophy with degeneration of the seminiferous tubular epithelium, decreased	Dogs, rats	No		[31]	Sub-fertility, increased FSH and decreased inhibin-B
	Imatinib mesylate ^b	spermanogenesis Decreased testicular and epididymal weights and nervent motile snerm	Rats	No		[32]	Oligozoospermia
	Leuprolide acetate	Testicular interstitial cell adenomas	Rats	Yes	Suppressed testicular steroidogenesis, testicular atrouby	[33]	Decreased sperm count
	Temozolomide ^b	Testicular atrophy, syncytial cells/immature	Rats and dogs	No	find an object	[34]	Decreased motility, abnormal
	Vincristine sulfate ^c	Spettii tottilation Testicular degeneration and atrophy, epididymal	Rats	No		[35]	morphology Increased infertility
Antipsychotic Antiulcer agent	Risperidone ^b Cimetidine	asperma Decreased sperm motility and concentration Benign Leydig cell tumors	Beagle dogs Rats	No No		[36] [37]	Retrograde ejaculation Reduced sperm count, increased
Antiviral agent	Ribavirin	Abnormalities in sperm	Mice	No		[38]	Decreased density, motility and
Cardiovascular agent Nitroglycerin	Nitroglycerin	Testicular tumors, increased interstitial cell	Rats	No		[39]	Reversible decrease in motility
Hormones, hormone substitutes, and	Finasteride	Testicular Leydig cell adenomas, decreased weights of seminal vesicles and prostate	Mice, rabbits	Yes	Reversible decrease in ejaculate volume and	[5]	Reversible decrease in number and motility
antagonists	Oxandrolone ^{b, c}	Reduction of spermatogenesis and decreased weights of the testes, prostate, and seminal vesicles	Rats	Yes	Suppressed special per ejacularion Suppressed special	[40]	Reversible azoospermia
	Testosterone	Suppressed spermatogenesis	Rats	Yes	oligozoospermia Suppressed spermatogenesis,	[41]	Decreased concentration
Hypoglycemic agent	Hypoglycemic agent Atorvastatin calcium		Rats	No	oligozoospermia	[42]	



Table 1 (continued)							
Main category	Drug name (generic)	Effect(s) in animals	Type of animal(s) affected	Human toxicity in DailyMed?	Human Reported effect(s) in toxicity in humans (DailyMed)	Support in PubMed	Support Reported effect(s) in PubMed
Immunosuppressive Sirolimus agent	Sirolimus	Aplasia and aspermia in the epididymis, decreased sperm motility and concentration, and increased abnormal sperm Atrophy of the testes, epididymides, prostate, and seminiferous tubules and/or reduction in sperm counts	Rats	Yes	Reversible azoospermia	[43]	At least one abnormal parameter in 35% of men (number, viability, motility, morphology, etc.) Decreased concentration and motility

^a PubMed data based on in vitro studies

^b PubMed data based on small studies involving 10 or fewer patients, including case reports

PubMed data based on studies of combination therapy, no data exist for the agent administered alone in humans

understanding that there are some inherent disadvantages associated with testing in animals [103].

Despite the various considerations made to improve the testing process and provide the best predictions of toxicity, there have been numerous studies that reported a lack of efficacy for toxicity testing, including reproductive toxicity testing [104-107]. In a previous study, a total of 65 labels for single-ingredient FDA-approved drugs were found in DailyMed that indicated that the drugs had a negative impact on human spermatogenesis [3]. In agreement with our present findings suggesting that animal studies have limited predictive power for human spermatotoxicity, fewer than half of the drug labels (26 of 65) indicated that they had effects on animal spermatogenesis. There was support in the literature for an additional six drugs affecting animal spermatogenesis, but this means that approximately half of the prescription drugs that negatively affect human spermatogenesis did not have an animal correlate reported in either the drug label or the peerreviewed literature. This may reflect the inadequacy of the animal model(s) tested, insufficient testing, and/or ineffective reporting of the findings of animal studies.

The relationship between the reported effects in animals and the effects in humans

In the present study, approximately 17% of the single-ingredient, FDA-approved, clinically prescribed drugs included in the DailyMed database (235/1318) had an adverse effect on spermatogenesis in at least one animal. Less than 30% of these drugs have been reported to adversely affect human sperm (in either the drug warning label or the published literature included in PubMed). However, this finding does not indicate that the "false positive" rate of animal toxicity testing is > 70%, because there were no data in either database about the impact of more than half of the drugs on human sperm (see Table 2). Therefore, the "false positive" rate for animal testing of the drugs with reported findings was about 35% (33/93).

Interestingly, of the 33 drugs indicated to have negative effects in animals that were shown to have no effect or a positive impact on human sperm in the literature (Table 3), four had data in their FDA labels (included in the DailyMed database) indicating that they had a negative impact on human sperm. Since the DailyMed database is supposed to contain up-to-date information, it is possible that new data had been generated since the PubMed studies were published demonstrating the negative effect reported in the label.

It is also possible that there were differences in the experimental conditions (different ages of subjects, subjects of different ethnicities, subjects with different background diseases or stages of disease, or different timing of sample collection after administration, etc.), and these could have led to the different results. Because details were not present in the FDA labels included in the DailyMed database, it is unknown



 Table 2
 Drugs without clear information about the impact on men

,		1					
Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
Analgesic	Capsaicin	Reduction in the number and percent of motile Rats	Rats	No		May affect motility by binding	[45]
Anesthetic	Lidocaine hydrochloride monohydrate ^a	Sperm, reduced sperm counts Decreased homogenization-resistant sperm head count, daily sperm production, and snermatogenic efficiency	Rats	S O		Increases the spermicidal effects of other compounds	[46]
Antibacterial agent	Dapsone	Reduced sperm motility	Rats	Yes	Orchitis,	No studies identified	
	Moxifloxacin/moxifloxacin	Adverse effects on sperm morphology	Rats	No	merunty	No studies identified	
	Telavancin hydrochloride	Altered sperm parameters	Rats	No		No studies identified	
	Cefotetan disodium	Reduced testicular weight and seminiferous tubule degeneration	Rats	No		No studies identified	
	Trovafloxacin mesylate	Testicular degeneration	Rats and dogs	No		No studies identified	
Anticholinergic	Glycopyrrolate	Diminished seminal secretions	Dogs	No		No studies identified	
agent Anticonvulsant	Clobazam	Increased abnormal sperm	Rats	No		No studies identified	
Antidepressant	Fluoxetine ^a	Testicular and epididymal microscopic lesions	Rats	No		In vitro spermicidal effects	[47]
	Flivoxamine maleate ^a	and decreased sperm concentrations Decreased snerm count decreased enididymal	Rats	Yes	Hematospermia	In vitro snermicidal effects	[47]
		weight		3			
	Goserelin acetate	Atrophic histological changes in the testes, epididymis, seminal vesicles and prostate	Rats	No		No studies identified	
	Imipramine pamoate ^a	gland, suppressed spermatogenesis Atrophy of the seminiferous tubules,	Dogs	No		Decreased viability and motility	[48]
		spermatogenic arrest					
	Methyldopa/methyldopate hydrochloride	Decreased sperm count, sperm motility, number of late spermatids	Rats	No		No studies identified	
	Nimodipine ^a	Leydig cell adenomas	Rats	No		Blocks the AR	[49]
	Selegiline	Epididymal and testicular hypoplasia, decreased sperm count and concentration	Dogs, rats	No		No studies identified	
	Trimipramine maleate	Degeneration of seminiferous tubules	Not specified	No		No studies identified	
	Nebivolol hydrochloride	Testicular Leydig cell hyperplasia and adenomas, effects on spermatogenesis	Mice and rats	No		No studies identified	
Antidote for ethylene glycol	Fomepizole	Decreased testicular mass	Rats	No		No studies identified	
Antiemetic	Dronabinol ^a	Decreases in spermatogenesis, number of developing germ cells, and number of	Rats	No		Decreased motility and AR	[50]
Antiepileptic	Felbamate	Benign interstitial cell tumors of the testes	Rats	No		A case report suggested that it was [51]	[51]
	Rufinamide	Decreased sperm count and motility	Rats	No		No studies identified	



ਲ੍ਹ
tinu
Ē
3
7
<u>[</u>
ab
=

Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human Reported effect(s) toxicity in in humans DailyMed? (DailyMed)	s) Notes based on the PubMed search	PubMed reference(s)
Antihypertensive agent	Hydralazine hydrochloride Isradipine Eplerenone Reserpine ^a	Benign interstitial cell tumors Benign Leydig cell tumors and testicular hyperplasia Decreased weights of seminal vesicles and epididymides Malignant tumors of the seminal vesicles	Rats Rats Rats Mice	° 2 ° 2	No studies identified No studies identified No studies identified Reduced motility and fertilization	[52]
Anti-infective agent	Metronidazole ^a Trimetrexate glucuronate Linezolid		Rats Mice and rats Rats, dogs	2 ° ° 2	Spermicidal No studies identified No studies identified	[53]
	Metloquine hydrochloride Micafungin sodium Luliconazole Miltefosine	Histopathological lesions in the epididymides Vacuolation of epididymal ductal epithelial cells and reduced sperm count, seminiferous tubular atrophy and decreased epididymal sperm Decreased sperm counts Testicular Leydig cell adenoma, testicular atrophy, reduced numbers of viable sperm	Kats, dogs Rats, dags Rats	° 2 ° 2	No studies identified No studies identified No studies identified No studies identified	
Anti-inflammatory agent Antimigraine agents	Suprofen Celecoxib Zileuton Eletriotan hydrobromide	Testicular atrophy/hypoplasia Epididymal hypospermia, slight dilation of the Juvenile rats seminiferous tubules Benign Leydig cell tumors Testicular interstitial cell adenomas Rats	Rats Juvenile rats Rats Rats	% % % % % % %	No studies identified No studies identified No studies identified No studies identified	
Antineoplastic agent	Abiraterone acetate Ado-trastuzumab emtansine	Testicular atrophy, aspermia/hypospermia, reduced sperm counts and motility, altered morphology Degeneration of seminiferous tubules with hemorrhage in the testes, decreased weights of the epididymides, prostate, and seminal vesicles	Rats and monkeys	g Q Q	Decreased testosterone and decreased weights of androgen-responsive organs in a xenograft fetal testis model No studies identified	[54]
	Afatinib Alemtuzumab	zoospermia, increased apoptosis in s and atrophy in the seminal and prostate fects on sperm parameters, d sperm count and motility	Rats Transgenic mice	° 2	No studies identified The target of the antibody (CD52) may have homology with gp20 on the sperm membrane, but the clinical impact is unknown	[55]



Table 2 (continued)

Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
	Altretamine	Testicular atrophy, decreased spermatogenesis, atrophy of testes, seminal vesicles and ventral prostate	Rats	No		No studies identified	
	Asparaginase	Decreased sperm count and motility	Rats	No		No studies identified	
	Axitinib	Testicular atrophy, decreased numbers of germinal cells, hypospermia or abnormal sperm forms, reduced sperm density and	Rats	No		No studies identified	
	Belinostat	Reduced organ weights of the testes/epididymides, delay in testicular maturation	Dogs	No		No studies identified	
	Bexarotene	Testicular degeneration	Dogs	No		No studies identified	
	Bortezomib	Degenerative changes in the testes	Rats	No		No studies identified	
	Brentuximab vedotin	Seminiferous tubule degeneration, Sertoli cell vacuolation, reduced spermatogenesis or	Rats	No		No studies identified	
	Cabazitaxel	aspennia Degeneration of seminal vesicles and seminiferous tubule atrophy, minimal arithalial cincle call nearests in antididumic	Rats, dogs	No O		No studies identified	
	Cabozantinib	Decreased sperm counts and reproductive organ weights, testicular degeneration and decreased spermatocytes and snermatids	Rats, mice	No		No studies identified	
	Chlorpromazine ^a	Chromosomal aberrations in spermatocytes and abnormal sperm	Rodents	No		Decreased motility	[99]
	Cladribine	Testicular degeneration	Monkeys	No		No studies identified	
	Clofarabine	Seminiferous tubule and testicular degeneration, atrophy of interstitial cells	Mice, rats and dogs	No		No studies identified	
	Cobimetinib	Testicular degeneration	Dogs	No		No studies identified	
	Crizotinib	Testicular pachytene spermatocyte degeneration	Rats	No		No studies identified	
	Dabrafenib	Testicular degeneration/depletion	Rats and dogs	Yes	Impaired spermatogenes- is, decreased sperm count	No studies identified	
	Decitabine	Abnormal histology, decreased sperm number	Mice	No	•	No studies identified	
	Docetaxel	Testicular atrophy or degeneration	Dogs	No		No studies identified	
	Enzalutamide	Atrophy of the prostate and seminal vesicles	Rats	No		No studies identified	
	Eribulin mesylate	Testicular toxicity (hypocellularity of seminiferous epithelium with hypospermia/aspermia)	Rats and dogs	No V		No studies identified	
	Everolimus	, Jones Jone	Rats	Yes			[57]



G
inue
cont
٣
e 7
Ī
La

Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
	Decreased sperm motility, sperm count, and plasma testosterone levels			Azoospermia or oligozoospermia (< 1% of patients)	Increases testosterone, but clinical impact unclear	
Fulvestrant	Testicular Leydig cell tumors, loss of spermatozoa from seminiferous tubules, seminiferous tubular atrophy, and deconstruction changes in the savididumidae.	Rats	No		No studies identified	
Gemcitabine hydrochloride	Hypospermatogenesis	Mice	No		No studies identified	
Idarubicin	Testicular atrophy, inhibition of spermatogenesis and sperm maturation with few or no mature snerm	Dogs	No		No studies identified	
Idelalisib	Decreased epididymidal and testicular weights, reduced sperm concentration	Rats	No		No studies identified	
Irinotecan hydrochloride	Atrophy of male reproductive organs	Rodents	No		No studies identified	
Ixabepilone	Testicular atrophy or degeneration	Dogs	No		No studies identified	
Lenvatinib	Testicular hypocellularity of the seminiferous epithelium and desquamated seminiferous epithelial cells in the epididymides	Dogs	No O		No studies identified	
Nilutamide	Benign Leydig cell tumors	Rats	Yes	Testicular atrophy	No studies identified	
Omacetaxine mepesuccinate	Degeneration of the seminiferous tubular epithelium, hypospermia/aspermia in the epididymides	Mice	No O		No studies identified	
Oxaliplatin	Testicular degeneration, hypoplasia, and atrophy	Dogs	No		Reduced inhibin-B (predictive of poor spermatogenesis and infertility)	[58]
Palbociclib	Decreased organ weight, atrophy or degeneration, hypospermia, intratubular cellular debris, lower sperm motility and density.	Rats and dogs	No No		No studies identified	
Panobinostat	Prostate atrophy accompanied by reduced secretory granules, testicular degeneration, oligospermia, and increased epididymal debris	Dogs	No No		No studies identified	
Pazopanib hydrochloride	Reduced sperm production and testicular sperm concentrations, epididymal sperm concentrations and sperm motility, atrophy and degeneration of the testes, aspermia, hypospermia, and cribiform changes in the exididucing	Rats	N O		No studies identified	
Pemetrexed disodium	lity, hypospermia, and testicular	Mice	No		No studies identified	



Drug category

Table 2 (continued)

Table 2 (Collinaca)							
Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
	Pentostatin	Seminiferous tubular degeneration	Dogs	No		No studies identified	
	Ponatinib	Degeneration of the epithelium of the testes	Rats and monkeys	No		No studies identified	
	Porfimer sodium	Discoloration and hypertrophy of the testes	Rats	No		No studies identified	
	Regorafenib	Tubular atrophy and degeneration in the testes, atrophy in the seminal vesicle, and cellular debris and oligospermia in the epididymides	Rats	No V		No studies identified	
	Romidepsin	Testicular degeneration	Rats	No		No studies identified	
	Sorafenib	Testicular atrophy or degeneration; degeneration of the epididymis, prostate, and seminal vesicles; oligospermia	Rats, dogs	No		No studies identified	
	Topotecan	Multinucleated spermatogonial giant cells	Dogs	No		No studies identified	
	Trabectedin	Histopathological signs of hemorrhage and degeneration in the testes	Rats	No		No studies identified	
	Trifluoperazine ^a	Chromosomal aberrations in spermatocytes and abnormal sperm	Rodents	No		Decreased motility	[49]
	Valrubicin	Testicular degeneration, germ cell depletion, spermatid giant cells and karyomegaly	Dogs	No		No studies identified	
	Venetoclax	Testicular toxicity (germ cell loss)	Dogs	No		No studies identified	
	Vinorelbine	Decreased spermatogenesis and	Rats	Yes	Damage to	No studies identified	
	Vismodegib	prostate/seminal vestere secretions Decreased % motile sperm, hypospermia,	Rats, dogs	No	spermatozoa	Present in semen, but	[65]
	Ziv-aflibercept	germ cell degeneration Decreased sperm motility, alterations in sperm	Monkeys	No		impact unclear No studies identified	
Anti-osteoporosis	Risedronate sodium	morphology Testicular and epididymal atrophy	Rats	No		No studies identified	
Anti-Parkinson agent	Apomorphine hydrochloride	Leydig cell tumors	Rats	No		No studies identified	
	Ropinirole hydrochloride	Testicular Leydig cell adenomas	Rats	No		No studies identified	
	Rotigotine	Leydig cell tumors, decreased epididymal	Rats	No		No studies identified	
	Pimavanserin tartrate	Decreased density and motility of sperm, cytoplasmic vacuolation in the epididymis	Rats	No		No studies identified	
Antipsychotic agent	Aripiprazole/aripiprazole	Disturbances in spermatogenesis	Rats	No		No studies identified	
	Paliperidone/paliperidone palmitate	Decreased sperm motility and concentration	Beagle dogs	No		No studies identified	
	Prochlorperazine edisylate/- prochlorperazine maleate	Chromosomal aberrations in spermatocytes and abnormal sperm	Rodents	No No		No studies identified	



ф (р
inue
onti
၁
7
le
Q
$\mathbf{I}_{\mathbf{a}}$

,							
Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
Antituberculosis agent/antiinfectious	Rifabutin	Testicular atrophy	Baboons and rats	No		No studies identified	
Antiulcer	Dexlansoprazole	Testicular interstitial cell adenomas	Rats	No		No studies identified	
	Lansoprazole	Testicular interstitial cell adenomas, proliferative changes in the Leydig cells, including benion neoplasia	Rats	No		No studies identified	
Antiviral agent	Boceprevir	Testicular degeneration	Rats	No		No studies identified	
	Cidofovir	Inhibition of spermatogenesis	Rats and monkeys	No		No studies identified	
	Daclatasvir	Reduced prostate/seminal vesicle weights,	Rats	No		No studies identified	
	Entecavir	minimally increased dysmorphic sperm Seminiferous tubular degeneration	Rodents and dogs	No		No studies identified	
	Famciclovir	Atrophy of the seminiferous tubules,	Rats, mice and	No		No studies identified	
		reduction in sperm count, and/or increased incidence of sperm with abnormal morphology or reduced motility	sgop				
	Ganciclovir	Hynosnermatogenesis	Mice doos	Yes	Testionlar	No studies identified	
					hypotrophy, aspermatogen- esis (dose- dependent)		
	Penciclovir	Atrophy of the seminiferous tubules,	Rats and dogs	No		No studies identified	
		increased sperm with abnormal morphology, reduced motility					
	Trifluridine	Testicular atrophy	Mice	No		No studies identified	
	Valganciclovir	Hypospermatogenesis	Mice, rats and dogs	Yes	Inhibited spermatogene-sis	No studies identified	
Attention deficient disorder agent	Atomoxetine hydrochloride	Decreased epididymal weight and sperm number	Rats	No		No studies identified	
Bone density conservation agent	Ibandronate sodium	Decreased sperm production and altered sperm morphology	Rats	o N		No studies identified	
Cardiovascular	Dexrazoxane	Testicular atrophy	Rats	No		No studies identified	
agent	Naratriptan hydrochloride	Testicular/epididymal atrophy accompanied by spermatozoa depletion	Rats	No		No studies identified	
	Propafenone hydrochloride	Decreased spermatogenesis	Rabbits, dogs, and monkeys	No		No studies identified	
	Ambrisentan	Testicular tubular degeneration, effects on the sperm count and morphology	\mathbb{R}^{3}	No		No studies identified	



5	3
Ξ	3
÷	
Ş	ξ
٢	۰
•	1
٥	۵
	4
4	

Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
	Bosentan	Testicular tubular atrophy	Rodents	Yes	Decreased sperm	No studies identified	
	Dofetilide	Testicular atrophy and epididymal oligospermia	Rats, mice and dogs	No	11100	No studies identified	
	Spironolactone	Testicular interstitial cell tumors	Rats	No		2/9 men showed decreased sperm density, but overall, no significant difference	[09]
	Macitentan	Testicular tubular atrophy	Rats	No		No studies identified	
Central nervous	Carisoprodol	Reduced testes weight and sperm motility	Mice	No		No studies identified	
system agent	Eszopiclone	Decreased sperm number and motility, increase in morphologically abnormal sperm (mid and high doses)	Rats	No		No studies identified	
	Fentanyl citrate ^a	Decreased percent mobile sperm, sperm concentrations, increased abnormal sperm	Rats	No		Modest inhibition of motility (based on a CASA)	[61]
	Oxazepam	Testicular interstitial cell adenomas	Rats	No		,	
	Caffeine citrate	Spermatogenic cell degeneration	Rats	No		Different studies have shown	[62, 63]
						conflicting (potentially dose-related) effects	
Contrast medium	Gadobenate dimeglumine	Abnormal spermatogenic cells	Rats	No		No studies identified	
	Gadopentetate dimeglumine	Decreased testes and epididymis weights	Rats	No		No studies identified	
	Gadoversetamide	Reduction and degeneration of spermatocytes, degeneration of the germinal epithelium of the testes, presence of germ cells in the epididymides, reduced sperm count	Rats	No O		No studies identified	
Dermatological agent	Dimethyl fumarate	Leydig cell adenomas, increased nonmotile sperm, testicular atrophy, hypospermia, testicular hyperplasia	Mice, rats, and dogs	N _o		No studies identified	
	Tretinoin	Decreased sperm count and motility	Rats	No		Other retinoids had no effect on sperm	[64]
Dispersing agent	Hyaluronidase, ovine	Testicular degeneration	Not specified	No		The hyaluronidase level is associated with the fertilization rate, but the clinical impact of systemic or nonreproductive treatment is unclear	[65]
Enzyme inhibitor	Eliglustat	Adverse effects on sperm morphology, testes (germ cell necrosis), and sloughed cells in the epididymis	Rats	N _o		No studies identified	
	Tofacitinib	Benign Leydig cell tumors	Rats	No		No studies identified	
Epithelial growth	Palifermin	Decreased epididymal sperm counts	Rats	No		No studies identified	
10001	Desloratadine	Decreased sperm numbers and motility	Rats	No		No studies identified	



continued)
Table 2

Drug category	Generic name	Effect(s) on animal spermatogenesis	Animal type(s) affected	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Notes based on the PubMed search	PubMed reference(s)
Histamine antagonist Hormones, hormone substitutes, and hormone antagonists	Conjugated estrogens, estradiol estradiol acetate, estradiol cypionate	Increased frequency of carcinomas of testis	Not specified	°Z		Estrogen decreases the acrosome reaction in vitro, but is required for normal sperm production. Therefore, effects are likely concentration-dependent	[66, 67]
	Estropipate Flutamide	Increased frequency of carcinomas of testis Testicular interstitial cell adenomas of the testes	Not specified Rats	No Yes	Interference with testosterone, decreased	No studies identified Unclear, suggested to have minimal transient effects on sperm	[88]
	Ospemifene Histrelin acetate	Atrophy of the prostate and seminal vesicles	Rats Rats	No Ves	Tectionlar attortive	No studies identified	
Hypnotic agent	Doxepin hydrochloride	Increased percentages of abnormal sperm and decreased sperm motility	Rats	No No	responding and pro-	No studies identified	
	Ramelteon Rozerem	Benign Leydig cell tumors Leydig cell tumors	Rats Rats	o N N		No studies identified No studies identified	
Hypoglycemic agent	Acarbose	Benign Leydig cell tumors	Rats	No		No studies identified	
	Canagliflozin	Testicular Leydig cell tumors, decreased sperm velocity, increased number of abnormal sperm	Rats	No		No studies identified	
	Chlorpropamide	Suppression of spermatogenesis	Rats	No		No studies identified	
	Fenofibrate/fenofibric acid	Benign testicular interstitial cell tumors	Rats	No		No studies identified	
	Fluvastatin sodium	Tubular degeneration and aspermatogenesis in testes, vesiculitis of seminal vesicles	Hamsters, rats	No		No studies identified	
	Mifepristone ^a	Reduced testicular size	Rats	No		Decreased AR	[69]
Immunosuppressive agent	Azathioprine sodium	Reduced spermatogenesis, sperm viability and sperm count	Mice	No.		Conflicting findings, may be age-dependent or related to the disease status	[70, 71]
Immunological factor	Interferon gamma-1b	Decreased spermatogenesis and sperm counts, increased abnormal sperm (very high doses)	Mice and monkeys	N _o		Effect of local or systemic treatment unclear, but no effect on sperm treated in vitro	[72]
	Pimecrolimus	Decreased testicular and epididymal weights, testicular sperm counts, motile sperm	Rats	oN :		Other mTOR inhibitors had adverse effects	[43]
	Teriflunomide	Reduced epididymal sperm count	Rats	No			
	Thalidomide	Testicular pathological and histopathological effects	Rabbits	Yes	Orchitis	Present in semen, but impact unclear	[73]
Muscle relaxant agent	Dantrolene sodium	Testicular tumors	Rats	No		No studies identified	



reference(s) PubMed Reported effect(s) Notes based on the PubMed No studies identified (DailyMed) in humans DailyMed? toxicity in Human ₂ ₂ ŝ Š ž Animal type(s) Monkeys Rats Rats Rats Rats Tubular atrophy, degeneration in the testes and Reduced sperm motility, increased percentage Changes in sperm morphology and motility of abnormal sperm (broken sperm with Interstitial cell adenoma of the testes Effect(s) on animal spermatogenesis spermiogenic granuloma in the Leydig cell adenomas detached heads) epididymides Naloxegol oxalate Generic name Lubiprostone Roflumilast Aflibercept Avanafil Fable 2 (continued) Ophthalmic drug DE4 inhibitor Purgative agent DE5 inhibitor Drug category

PubMed data based on in vitro studies

why there were conflicting data. These findings illustrate the difficulties associated with reproductive toxicity testing. Moreover, they demonstrate the need for better record-keeping and transparency in terms of the data generated, because it is very difficult to interpret the findings in the drug labels based on the data available.

Difficulties associated with animal testing for spermatotoxicity

The current reproductive toxicity testing should be able to provide useful results for clinical extrapolation. However, as noted above, our current study and previous studies about spermatotoxicity [3, 105, 106] have shown that studies in animals have relatively low predictive value for human toxicity. In some cases, the lack of useful findings is due to the high doses of the compounds used in the toxicity studies. Nevertheless, even if the dose is proper in terms of the allometric scaling for animals based on body surface area (~ 12× the human dose (mg/kg b.w.) for a mouse, $\sim 2 \times$ for a dog, etc. [108]), the plethora of differences between humans and animals reduces the utility of the findings. For example, the pharmacokinetics and pharmacodynamics of drugs may not be similar between the animal model and humans, there may be organ/tissue-related differences that make it impossible to adequately predict or extrapolate the effects, and there may be other fundamental differences between species that abrogate the utility of testing [106]. This is particularly relevant for spermatotoxicity studies, where there are many important differences between rodents and humans, including the time required for spermatogenesis, the development of Leydig cell tumors, and the relative number of sperm produced [109-112]. Additional "uncertainty" factors have been used when extrapolating human toxicity from animal findings for occupational and environmental toxicants (i.e., when determining the benchmark dose or the acceptable daily intake based on the no observed adverse effects level (NOAEL) [113], but these types of studies are not usually employed as part of therapeutic drug testing and would be subject to the same issues described above.

Changes that should be made to the testing and reporting of spermatotoxicity

The most important findings of the present study include the following: only 27% (63/235) of the drugs that had a negative effect in animals reported in the drug label were found to have a documented negative impact on human sperm (in the drug labels or literature). Further, a previous study [3] showed that fewer than half of the drugs with labels indicating that they adversely affected human sperm had similar findings in animals. In addition, for more than half of the drugs found to negatively affect animal spermatogenesis or spermatozoa,



 Table 3
 Drugs where the animal findings were refuted by PubMed publications

Main indication	Drug name (generic)	Effect(s) in animals	Type of animal(s) affected/tested	Human toxicity in h DailyMed?	Reported effect(s) in humans (DailyMed)	Effect(s) in humans reported in PubMed	PubMed reference
Adrenergic agent	Icatibant acetate ^a	Testicular atrophy/degeneration, reduced sperm	Rats and dogs	No		No effect	[74]
Antibacterial agent	Doxycycline/doxycycline hyclate	Reduced motility, velocity and concentration, abnormal morphology	Rats	No		May improve sperm parameters in patients with infections	[75]
	Clarithromycin	Testicular atrophy	Rats and monkeys	No		Improves motility	[92]
Antiepileptic	Pregabalin	Decreased sperm counts and sperm motility, increased sperm abnormalities	Rats	Yes E	Epidiymitis	No effect	[77]
Antigout agent	Colchicine/colchicinum	Abnormal sperm morphology and reduced sperm counts in males	Not specified	Yes	Azoospermia, oligospermia	No effect	[78]
Antihyperlipidemic agent	Pravastatin sodium	Sperm abnormalities	Rats	No	•	No effect	[62]
Antihypertensive agent	Metoprolol	Reversible adverse effects of spermatogenesis (in some studies)	Rats	No o		Minimally inhibits motility	[80]
0	Pindolol	Testicular atrophy and/or decreased spermatogenesis	Rats	No		No effect	[80]
	Prazosin ^a	Testicular changes consisting of atrophy and necrosis	Rats and dogs	No		No effect	[81]
Anti-infective agent	Griseofulvin ^b	Suppression of spermatogenesis	Rats	No		No effect	[82]
	Minocycline	Reduced sperm cells and motile sperm, increased	Rats	No		Improves semen parameters	[83]
	hydrochloride	abnormal sperm, including absent heads, misshapen heads and abnormal flagella				in men with infections	
	Tinidazole	Testicular degeneration, spermatogenic effects	Rats	No		Improves sperm parameters	[92]
						as part of treatment for Π . pylori infections	
Anti-inflammatory agent	Ketoprofen	Abnormal spermatogenesis or inhibition of spermatogenesis	Rats and dogs	No		Treatment may improve male infertility	[84]
)	Oxaprozin	Testicular degeneration	Beagle dogs	No		No effect	[85]
Antineoplastic	Bicalutamide	Testicular benign interstitial (Leydig) cell tumors	Rats	No		No effect	[98]
agent	Dasatinib	Reduced size and secretion of seminal vesicles, immature prostate, seminal vesicle, and testis	Rats	No		No effect	[87]
	Epirubicin hydrochloride°	Atrophy of the testes and epididymis, reduced spermatogenesis	Mice and rats	No		No effect	[88]
	Fluorouracil ^a	Kills differentiated spermatogonia and spermatocytes	Mice	No		No effect	[68]
	Letrozole	he seminiferous tubular epithelium	Rats	N _o		Improves sperm parameters as part of treatment for <i>H. pylori</i> infections	[06]
	Paclitaxel ^c	Testicular atrophy/degeneration	Rodents	No		No effect	[91]
	Toremifene citrate	Testicular tumors	Mice	No		Improves sperm parameters and hormone levels	[92]
Antiviral agent	Acyclovir	Testicular atrophy and aspermatogenesis	Rats and dogs	No		No effect	[63]
	Miglustat		Rats	No		No effect	[94]



Table 3 (continued)

Main indication	Drug name (generic)	Effect(s) in animals	Type of animal(s) affected/tested	Human toxicity in DailyMed?	Reported effect(s) in humans (DailyMed)	Effect(s) in humans reported in PubMed	PubMed
		Decreased spermatogenesis with altered sperm morphology and motility, decreased fertility, and testicular interstitial cell adenomas					
	Tenofovir	None	Rats	No		Decreased progressive motility	[66]
Bone density conservation	Raloxifene hydrochloride	Raloxifene hydrochloride Testicular interstitial cell tumors	Mice			Improves sperm concentration and	[96]
agent						morphology and increased testosterone in oligospermic med	
Cholinergic agonist Pilocarpine	Pilocarpine	Decreased sperm motility, and morphologic evidence Rats of abnormal sperm	Rats	No		No effect	[26]
Dermatological agent	Clobetasol propionate	Increased weights of the seminal vesicles	Rats	N _o		Treatment of scrotal dermatitis improved the sperm count and motility	[86]
	Isotretinoin	Depression of spermatogenesis	Dogs	No		No effect	[64]
Hypoglycemic agent	Simvastatin	Seminiferous tubule degeneration (necrosis and loss of spermatogenic epithelium, testicular atrophy, decreased spermatogenesis, spermatocytic degeneration and giant cell	Rats Dogs	No		No effect	[79]
	Lovastatin	Testicular atrophy, decreased spermatogenesis, spermatocytic degeneration, giant cell formation	Dogs	No		No effect	[66]
Opioid antagonist	Naltrexone ^a	Increased testicular mesotheliomas	Rats	No		Increased motility (opposite the effects of morphine)	[100]
	Tadalafil	Degeneration and atrophy of the seminiferous tubular epithelium, decrease in spermatogenesis	Beagle dogs	Yes	Decreased sperm concentration	Improves concentration and motility	[101]

 a Based on in vitro findings b Based on small studies of < 10 patients, including case studies c Based on studies of combination treatment

the impact in humans is unclear either because no testing or surveillance has been performed, or because the results have not been reported. Even in cases where there was clear reporting of the type of human spermatotoxicity and the number of individuals affected, the data were frequently unclear regarding whether the effects were reversible and how long it took to recover potency. These findings suggest that the current system is not performing well and there is a need for better models, different types of pre-clinical testing, different endpoints, and a better system (and/or different regulations) for reporting data.

It should be noted that most of the general methods used to assess reproductive toxicity were often developed more than 50 years ago, although there have been updates in terms of automation and some of the details (i.e., sub-types of sperm motility) of the analyses. Our data add to the already extensive body of evidence that animal testing (certainly for spermatotoxicity, but also for many other types of toxicity) is not adequate to predict human risk. However, completely retiring the rat is not possible, at least at present. There are currently no other models that provide better predictive value for the various steps of reproduction. Direct testing on human sperm samples is useful and should be implemented whenever possible, but cannot identify changes in hormone levels, reproductive tissues, or germ or support cells. These parameters can only be evaluated in vivo.

While animal testing cannot be avoided at present, there are various steps that should be taken immediately to improve the efficacy and clinical relevance of the testing and reporting of data. First, the general protocol for testing should be reconsidered. The usual one-size-fits-all approach to toxicity testing should be scrapped in favor of integrative studies taking into account the differences in pharmacokinetics/ pharmacodynamics of the drug (if known), the intended target population (the age and disease background of the patients may be relevant to the toxicity), the effects of structurally and mechanistically similar drugs, and any information that can be obtained from in silico studies (high-throughput testing of the compound's predicted structure for interactions with characterized receptors and biomacromolecules). It may also be necessary to change the endpoints of studies. Instead of assessing sperm at a single time point (at a pre-specified time after a single dose, or at the end of a chronic study), it would likely be better to collect sperm at several different time points. Collection of multiple samples (including collection on more than 1 day prior to starting treatment) is particularly important for clinical trials given the high variability in human sperm production [111, 114]. Additional testing to confirm recovery would provide helpful information for counseling patients on the long-term adverse effects.

Perhaps the most important changes would be those made to the reporting of the findings. Although the FDA receives the full results of the pre-clinical toxicity studies and clinical safety studies, including the numbers of animals used for preclinical testing (or patients included in a clinical trial), this information is frequently lacking from the drug labels in DailyMed. A clear statement of the number of individuals evaluated and the number or percentage of these affected by each adverse effect would help to interpret the findings. It would also be useful for this information to specify the extent of the adverse impact, such as the exact percentages of sperm with abnormal morphology, rather than a blanket statement indicating that there were morphological changes. This information would not need to appear on the main drug label included with the packaging of each drug, but should be linked online (with the link included in the printed information) so that the full details of the study are readily available from DailyMed.

Another point that is commonly lacking in both the DailyMed labels and the data in journal articles is the reversibility of the spermatotoxicity and time to recovery (if recovery is possible). This information is frequently missing for pre-clinical studies, largely because sperm are usually collected at a single time point, sometimes at necropsy. Although clinical trials are more likely to assess the long-term effects of a drug, the long-term reproductive outcomes are often not evaluated. This is partly because such endpoints are difficult to add to standard surveillance protocols because the collection of semen is more invasive and potentially associated with ethical concerns, compared to physical examinations and blood collection.

Finally, computing power has been rapidly increasing during the past few decades, and research studies are helping to illuminate the molecular basis of various physiological processes. Therefore, efforts should be made to better understand the signaling and cellular interactions involved in normal fertility to permit in silico modeling of possible interference by investigational drugs. This would inform confirmatory studies to be performed in vitro on human sperm or iPSC-derived or other sources of germ cells, could suggest biomarkers to allow for more sensitive detection of effects in vivo (in animals and/ or humans), and would predict the toxicity likely to be encountered. As in silico modeling continues to improve, it may become possible to evaluate the interactions of various agents and various pathways simultaneously and to combine such findings with those from in vitro and limited in vivo studies for an integrated approach, more accurately predicting the effects in humans [115, 116].

Until toxicity testing is optimized, it is necessary to consider that there is a potential for human risk whenever there is a positive finding of spermatotoxicity in non-clinical or pre-clinical studies for one or more endpoints. The details of the findings and the level of evidence should be included to help physicians and/or pharmacists inform their patients of the risks of specific treatments to their fertility.



Conclusion

Our present findings illustrate the status of male reproductive toxicity testing and provide a useful reference about the FDAapproved drugs known to affect animal spermatogenesis and how these findings correlate with the human clinical setting. These findings emphasize the need for a better understanding of basic sperm biology to allow for the prediction of spermatotoxicity based on a drug's effects (both targeted and nonspecific). A greater understanding of the normal variations in the sperm of men of various ages and different health status would also help to understand what should be considered abnormal during clinical trials and long-term surveillance. In addition, since the predictive value of animal testing is so low, there is a need for a more comprehensive evaluation of the effects on human sperm in vitro and continued development of in silico and alternative in vitro studies so that potential changes can be estimated before the initiation of human clinical trials. Such an understanding could be used to determine whether the normal observations carried out during clinical trials should be increased or altered to better detect changes.

There are several limitations to this review that should be noted. First, only single-ingredient, FDA-approved drugs indicated for human use that were included in the DailyMed database were analyzed in this study. Although this database is the largest of its type, there are still a few drugs for which information is missing. In addition, all over-the-counter medications, including vitamins, supplements, and herbal/alternative medications, were excluded because the toxicity information on these agents is typically lacking. However, these excluded agents may be associated with spermatotoxicity, either when taken alone or by potentiating the effects of other drugs. The present investigation also focused on the direct effects of drugs on spermatozoa, which represents only a small aspect of male fertility. In addition, there may have been some reporting or publication bias for the databases. However, whenever possible, only large and well-designed clinical trials were included for the PubMed data, and it is noted whenever small-scale studies or case reports are cited in the tables. For the DailyMed database, the data sources are often unclear, so the accuracy of the data is also unclear.

Despite these limitations, the present data are useful in that they provide a comprehensive overview of the prescription drugs that affect animal spermatogenesis, as well as an illustration of the relatively poor performance of animal testing for predicting spermatotoxicity. Several suggestions have been made which might improve the testing and reporting of the data, hopefully leading to better patient care.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflicts of interest.

References

- Brezina PR, Yunus FN, Zhao Y. Effects of pharmaceutical medications on male fertility. J Reprod Infertil. 2012;13:3–11.
- Centers for Disease Control and Prevention. National Center for Health Statistics. Therapeutic drug use. In: Health, United States, 2015: with special feature on racial and ethnic health disparities. [Internet] Hyattsville, MD. 2016; [Cited August 8, 2017]. Available from: http://www.cdc.gov/nchs/fastats/drug-use-therapeutic.htm.
- Ding J, Shang X, Zhang Z, Jing H, Shao J, Fei Q, et al. FDAapproved medications that impair human spermatogenesis. Oncotarget. 2017;8:10714–25.
- Li H, Shi Q. Drugs and diseases interacting with cigarette smoking in US prescription drug labelling. Clin Pharmacokinet. 2015;54: 493–501.
- Smarr MM, Kannan K, Chen Z, Kim S, Buck Louis GM. Male urinary paracetamol and semen quality. Andrology. 2017; https:// doi.org/10.1111/andr.12413.
- Amory JK, Wang C, Swerdloff RS, Anawalt BD, Matsumoto AM, Bremner WJ, et al. The effect of 5alpha-reductase inhibition with dutasteride and finasteride on semen parameters and serum hormones in healthy men. J Clin Endocrinol Metab. 2007;92:1659

 –65.
- Kobayashi K, Masumori N, Hisasue S, Kato R, Hashimoto K, Itoh N, et al. Inhibition of seminal emission is the main cause of anejaculation induced by a new highly selective alpha1Ablocker in normal volunteers. J Sex Med. 2008;5:2185–90.
- Ramdurg S, Ambekar A, Lal R. Sexual dysfunction among male patients receiving buprenorphine and naltrexone maintenance therapy for opioid dependence. J Sex Med. 2012;9:3198–204.
- Reis RM, de Angelo AG, Sakamoto AC, Ferriani RA, Lara LA. Altered sexual and reproductive functions in epileptic men taking carbamazepine. J Sex Med. 2013;10:493–9.
- Agirregoitia E, Subiran N, Valdivia A, Gil J, Zubero J, Irazusta J. Regulation of human sperm motility by opioid receptors. Andrologia. 2012;44(Suppl 1):578–85.
- Hamed SA, Moussa EM, Tohamy AM, Mohamed KO, Mohamad ME, Sherif TM, et al. Seminal fluid analysis and testicular volume in adults with epilepsy receiving valproate. J Clin Neurosci. 2015;22:508–12.
- Isojärvi JI, Löfgren E, Juntunen KS, Pakarinen AJ, Päivänsalo M, Rautakorpi I, et al. Effect of epilepsy and antiepileptic drugs on male reproductive health. Neurology. 2004;62:247–53.
- Maier U, Koinig G. Andrological findings in young patients under long-term antidepressive therapy with clomipramine. Psychopharmacology. 1994;116:357–9.
- Tanrikut C, Feldman AS, Altemus M, Paduch DA, Schlegel PN. Adverse effect of paroxetine on sperm. Fertil Steril. 2010;94: 1021–6.
- Puri S, Puri VN. Urinary vasopressin in male infertility. Arch Androl. 1985;14:263–5.
- Pont A, Graybill JR, Craven PC, Galgiani JN, Dismukes WE, Reitz RE, et al. High-dose ketoconazole therapy and adrenal and testicular function in humans. Arch Intern Med. 1984;144:2150–3.
- Tada Y, Hayashi T, Iwaki Y, Karita M, Taguchi S, Funabiki M, et al. Transient azoospermia following rosuvastatin medication for hypercholesterolemia. Clin Exp Obstet Gynecol. 2015;42:545–6.
- Wang J, Zhao Y, Jiang SB, Xia QH, Wei CX, Wang MW, et al. Assessment of tamsulosin as a potential male contraceptive in healthy volunteers. Urology. 2012;80:614–7.
- Gulati A, Tiwary AK, Jain S, Moudgil P, Gupta A. Intrasperm Ca2+ modulation and human ejaculated sperm viability: influence of miconazole, clotrimazole and loperamide. J Pharm Pharmacol. 2006;58:1145–51.



- Pant N, Shukla M, Upadhyay AD, Chaturvedi PK, Saxena DK, Gupta YK. Association between environmental exposure to p, p'-DDE and lindane and semen quality. Environ Sci Pollut Res Int. 2014;21:11009–16.
- Silvestroni L, Fiorini R, Palleschi S. Partition of the organochlorine insecticide lindane into the human sperm surface induces membrane depolarization and Ca2+ influx. Biochem J. 1997;321:691–8.
- Yeung CH, Cooper TG. Effects of the ion-channel blocker quinine on human sperm volume, kinematics and mucus penetration, and the involvement of potassium channels. Mol Hum Reprod. 2001;7:819–28.
- Anserini P, Chiodi S, Spinelli S, Costa M, Conte N, Copello F, et al. Semen analysis following allogeneic bone marrow transplantation. Additional data for evidence-based counselling. Bone Marrow Transplant. 2002;30:447–51.
- Chatterjee R, Mills W, Katz M, McGarrigle HH, Goldstone AH. Germ cell failure and Leydig cell insufficiency in post-pubertal males after autologous bone marrow transplantation with BEAM for lymphoma. Bone Marrow Transplant. 1994;13:519–22.
- Fairley KF, Barrie JU, Johnson W. Sterility and testicular atrophy related to cyclophosphamide therapy. Lancet. 1972;1:568–9.
- Bujan L, Walschaerts M, Brugnon F, Daudin M, Berthaut I, Auger J, et al. Impact of lymphoma treatments on spermatogenesis and sperm deoxyribonucleic acid: a multicenter prospective study from the CECOS network. Fertil Steril. 2014;102:667–74.e3.
- Baumgartner A, Schmid TE, Cemeli E, Anderson D. Parallel evaluation of doxorubicin-induced genetic damage in human lymphocytes and sperm using the comet assay and spectral karyotyping. Mutagenesis. 2004;19:313–8.
- Suzuki K, Yumura Y, Ogawa T, Saito K, Kinoshita Y, Noguchi K. Regeneration of spermatogenesis after testicular cancer chemotherapy. Urol Int. 2013;91:445–50.
- Chatterjee R, Haines GA, Perera DM, Goldstone A, Morris ID. Testicular and sperm DNA damage after treatment with fludarabine for chronic lymphocytic leukaemia. Hum Reprod. 2000;15:762–6.
- National Toxicology Program. NTP-CERHR monograph on the potential human reproductive and developmental effects of hydroxyurea. NTP CERHR MON. 2008;21:vii–viii. v, ix-III1
- Williams D, Crofton PM, Levitt G. Does ifosfamide affect gonadal function? Pediatr Blood Cancer. 2008;50:347–51.
- Mariani S, Basciani S, Fabbri A, Agati L, Ulisse S, Lubrano C, et al. Severe oligozoospermia in a young man with chronic myeloid leukemia on long-term treatment with imatinib started before puberty. Fertil Steril. 2011;95:1120.e15–7.
- Swerdloff RS, Heber D, Bhasin S, Rajfer J. Effect of GnRH superactive analogs (alone and combined with androgen) on testicular function in man and experimental animals. J Steroid Biochem. 1983;19:491–7.
- Strowd RE, Blackwood R, Brown M, Harmon M, Lovato J, Yalcinkaya T, et al. Impact of temozolomide on gonadal function in patients with primary malignant brain tumors. J Oncol Pharm Pract. 2013;19:321–7.
- Sieniawski M, Reineke T, Josting A, Nogova L, Behringer K, Halbsguth T, et al. Assessment of male fertility in patients with Hodgkin's lymphoma treated in the German Hodgkin Study Group (GHSG) clinical trials. Ann Oncol. 2008;19:1795–801.
- Holtmann M, Gerstner S, Schmidt MH. Risperidone-associated ejaculatory and urinary dysfunction in male adolescents. J Child Adolesc Psychopharmacol. 2003;13:107–9.
- Wang C, Lai CL, Lam KC, Yeung KK. Effect of cimetidine on gonadal function in man. Br J Clin Pharmacol. 1982;13:791–4.
- Hofer H, Donnerer J, Sator K, Staufer K, Scherzer TM, Dejaco C, et al. Seminal fluid ribavirin level and functional semen

- parameters in patients with chronic hepatitis C on antiviral combination therapy. J Hepatol. 2010;52:812–6.
- Mckinney KA, Boyle P, Thompson W. Effect of glyceryl trinitrate on sperm motility and lipid peroxidation in normozoospermic men. Int J Androl. 1995;18:307–12.
- Peña JE, Thornton MH Jr, Sauer MV. Reversible azoospermia: anabolic steroids may profoundly affect human immunodeficiency virus-seropositive men undergoing assisted reproduction. Obstet Gynecol. 2003;101:1073–5.
- Kolettis PN, Purcell ML, Parker W, Poston T, Nangia AK. Medical testosterone: an iatrogenic cause of male infertility and a growing problem. Urology. 2015;85:1068–72.
- Pons-Rejraji H, Brugnon F, Sion B, Maqdasy S, Gouby G, Pereira B, et al. Evaluation of atorvastatin efficacy and toxicity on spermatozoa, accessory glands and gonadal hormones of healthy men: a pilot prospective clinical trial. Reprod Biol Endocrinol. 2014;12:65.
- 43. Zuber J, Anglicheau D, Elie C, Bererhi L, Timsit MO, Mamzer-Bruneel MF, et al. Sirolimus may reduce fertility in male renal transplant recipients. Am J Transplant. 2008;8:1471–9.
- Rahnema CD, Lipshultz LI, Crosnoe LE, Kovac JR, Kim ED. Anabolic steroid-induced hypogonadism: diagnosis and treatment. Fertil Steril. 2014;101:1271–9.
- De Toni L, Garolla A, Menegazzo M, Magagna S, Di Nisio A, Šabović I, et al. Heat sensing receptor TRPV1 is a mediator of thermotaxis in human spermatozoa. PLoS One. 2016;11: e0167622.
- Moudgil P, Gupta A, Sharma A, Gupta S, Tiwary AK. Potentiation of spermicidal activity of 2',4'-dichlorobenzamil by lidocaine. Indian J Exp Biol. 2002;40:1373–7.
- Kumar VS, Sharma VL, Tiwari P, Singh D, Maikhuri JP, Gupta G, et al. In vitro—the spermicidal and antitrichomonas activities of SSRI antidepressants. Bioorg Med Chem Lett. 2006;16:2509–12.
- Levin RM, Amsterdam JD, Winokur A, Wein AJ. Effects of psychotropic drugs on human sperm motility. Fertil Steril. 1981;36: 503–6.
- 49. José O, Hernández-Hernández O, Chirinos M, González-González ME, Larrea F, Almanza A, et al. In vitro-recombinant human ZP3-induced sperm acrosome reaction: evidence for the involvement of T- and L-type voltage-gated calcium channels. Biochem Biophys Res Commun. 2010;395:530–4.
- Whan LB, West MC, McClure N, Lewis SE. Effects of delta-9tetrahydrocannabinol, the primary psychoactive cannabinoid in marijuana, on human sperm function in vitro. Fertil Steril. 2006;85:653–60.
- 51. Yerby MS, McCoy GB. Male infertility: possible association with valproate exposure. Epilepsia. 1999;40:520–1.
- Chan SY, Tang LC. Effect of reserpine on fertilizing capacity of human spermatozoa. Contraception. 1984;30:363–9.
- El-Gizawy SA, Aglan NI. Formulation and evaluation of metronidazole acid gel for vaginal contraception. J Pharm Pharmacol. 2003;55:903–9.
- Spade DJ, Hall SJ, Saffarini CM, Huse SM, McDonnell EV, Boekelheide K. Differential response to abiraterone acetate and di-n-butyl phthalate in an androgen-sensitive human fetal testis xenograft bioassay. Toxicol Sci. 2014;138:148–60.
- 55. Focarelli R, Francavilla S, Francavilla F, Della Giovampaola C, Santucci A, Rosati F. A sialoglycoprotein, gp20, of the human capacitated sperm surface is a homologue of the leukocyte CD52 antigen: analysis of the effect of anti-CD52 monoclonal antibody (CAMPATH-1) on capacitated spermatozoa. Mol Hum Reprod. 1999;5:46–51.
- Hong CY, Chaput de Saintonge DM, Turner P. Effects of chlorpromazine and other drugs acting on the central nervous system on human sperm motility. Eur J Clin Pharmacol. 1982;22:413–6.
- Krämer BK, Neumayer HH, Stahl R, Pietrzyk M, Krüger B, Pfalzer B, et al. RADA2307 Study Group. Graft function,



- cardiovascular risk factors, and sex hormones in renal transplant recipients on an immunosuppressive regimen of everolimus, reduced dose of cyclosporine, and basiliximab. Transplant Proc. 2005;37:1601–4.
- Levi M, Shalgi R, Brenner B, Perl G, Purim O, Amit L, et al. The impact of oxaliplatin on the gonads: from bedside to the bench. Mol Hum Reprod. 2015;21:885–93.
- Grunewald S, Jank A. New systemic agents in dermatology with respect to fertility, pregnancy, and lactation. J Dtsch Dermatol Ges. 2015;13(4):277-89; quiz 290. https://doi.org/10.1111/ddg.12596.
- Caminos-Torres R, Ma L, Snyder PJ. Gynecomastia and semen abnormalities induced by spironolactone in normal men. J Clin Endocrinol Metab. 1977;45:255–60.
- Xu B, Wang ZP, Wang YJ, Lu PH, Wang LJ, Wang XH. The toxic effect of opioid analgesics on human sperm motility in vitro. Drug Chem Toxicol. 2013;36:205–8.
- Sobreiro BP, Lucon AM, Pasqualotto FF, Hallak J, Athayde KS, Arap S. Semen analysis in fertile patients undergoing vasectomy: reference values and variations according to age, length of sexual abstinence, seasonality, smoking habits and caffeine intake. Sao Paulo Med J. 2005;123:161–6.
- Moussa MM. Caffeine and sperm motility. Fertil Steril. 1983;39: 845–8.
- Török L, Kádár L, Kása M. Spermatological investigations in patients treated with etretinate and isotretinoin. Andrologia. 1987;19:629–33.
- Abdul-Aziz M, MacLusky NJ, Bhavnani BR, Casper RF. Hyaluronidase activity in human semen: correlation with fertilization in vitro. Fertil Steril. 1995;64:1147–53.
- Vigil P, Toro A, Godoy A. Physiological action of oestradiol on the acrosome reaction in human spermatozoa. Andrologia. 2008;40:146–51.
- Schlegel PN. Aromatase inhibitors for male infertility. Fertil Steril. 2012;98:1359–62.
- Neumann F, Diallo FA, Hasan SH, Schenck B, Traore I. The influence of pharmaceutical compounds on male fertility. Andrologia. 1976;8:203–35.
- Ko JK, Huang VW, Li RH, Yeung WS, Ho PC, Chiu PC. An in vitro study of the effect of mifepristone and ulipristal acetate on human sperm functions. Andrology. 2014;2:868–74.
- Koyun M, Baysal YE, Usta MF, Akman S, Güven AG. Evaluation of reproductive functions in male adolescents following renal transplantation. Pediatr Transplant. 2009;13:697–700.
- Dejaco C, Mittermaier C, Reinisch W, Gasche C, Waldhoer T, Strohmer H, et al. Azathioprine treatment and male fertility in inflammatory bowel disease. Gastroenterology. 2001;121:1048–53.
- Estrada LS, Champion HC, Wang R, Rajasekaran M, Hellstrom WJ, Aggarwal B, et al. Effect of tumour necrosis factor-alpha (TNF-alpha) and interferon-gamma (IFN-gamma) on human sperm motility, viability and motion parameters. Int J Androl. 1997;20:237–42.
- Teo SK, Harden JL, Burke AB, Noormohamed FH, Youle M, Johnson MA, et al. Thalidomide is distributed into human semen after oral dosing. Drug Metab Dispos. 2001;29:1355–7.
- Miska W, Schill WB. Influence of bradykinin antagonists on the motility of human spermatozoa enhanced by bradykinin. Arch Androl. 1994;33:1–5.
- Vicari E. Effectiveness and limits of antimicrobial treatment on seminal leukocyte concentration and related reactive oxygen species production in patients with male accessory gland infection. Hum Reprod. 2000;15:2536–44.
- El-Garem Y, El-Sawy M, Mostafa T. Seminal Helicobacter pylori treatment improves sperm motility in infertile asthenozoospermic men. Urology. 2014;84:1347–50.
- Sikka SC, Chen C, Almas M, Dula E, Knapp LE, Hellstrom WJ. Pregabalin does not affect sperm production in healthy volunteers:

- a randomized, double-blind, placebo-controlled, noninferiority study. Pain Pract. 2015;15:150–8.
- Haimov-Kochman R, Ben-Chetrit E. No or variable effect—the effect of colchicine treatment on sperm production and function: a review. Hum Reprod. 1998;13:360–2.
- Dobs AS, Miller S, Neri G, Weiss S, Tate AC, Shapiro DR, et al. Effects of simvastatin and pravastatin on gonadal function in male hypercholesterolemic patients. Metabolism. 2000;49:115–21.
- Curtis-Prior PB, Gadd AL. Beta-adrenoceptor antagonists and human sperm motility. J Pharm Pharmacol. 1990;42:220–2.
- Hellstrom WJ, Wang R, Peterson CA, Varady JC, Gesundheit N, Sikka SC. Effects of alprostadil and prazosin on motility, viability and membrane integrity of human sperm. J Urol. 1998;159:1559–62.
- Ko EM, Lowry RB, Martin RH. Analysis of sperm karyotypes in a patient treated with griseofulvin. Arch Androl. 2007;53:157–60.
- 83. Malallah YA, Zissis NP. Effect of minocycline on the sperm count and activity in infertile men with high pus cell count in their seminal fluid. J Chemother. 1992;4:286–9.
- Barkay J, Harpaz-Kerpel S, Ben-Ezra S, Gordon S, Zuckerman H.
 The prostaglandin inhibitor effect of antiinflammatory drugs in the therapy of male infertility. Fertil Steril. 1984;42:406–11.
- 85. Knuth UA, Kühne J, Crosby J, Bals-Pratsch M, Kelly RW, Nieschlag E. Indomethacin and oxaprozin lower seminal prostaglandin levels but do not influence sperm motion characteristics and serum hormones of young healthy men in a placebocontrolled double-blind trial. J Androl. 1989;10:108–19.
- Morgante E, Gradini R, Realacci M, Sale P, D'Eramo G, Perrone GA, et al. Effects of long-term treatment with the anti-androgen bicalutamide on human testis: an ultrastructural and morphometric study. Histopathology. 2001;38:195–201.
- Tauer JT, Ulmer A, Glauche I, Jung R, Suttorp M. Long-term imatinib treatment does not cause testicular toxicity in male adolescents with chronic myeloid leukemia and in a juvenile rat model. Klin Padiatr. 2014;226:169–74.
- Hill M, Milan S, Cunningham D, Mansi J, Smith I, Catovsky D, et al. Evaluation of the efficacy of the VEEP regimen in adult Hodgkin's disease with assessment of gonadal and cardiac toxicity. J Clin Oncol. 1995;13:387–95.
- Hong CY, Chiang BN, Ku J, Wu P. Screening the in vitro spermimmobilizing effect of some anticancer drugs. Hum Toxicol. 1985;4:461–4.
- Ribeiro MA, Gameiro LF, Scarano WR, Briton-Jones C, Kapoor A, Rosa MB, et al. Aromatase inhibitors in the treatment of oligozoospermic or azoospermic men: a systematic review of randomized controlled trials. JBRA Assist Reprod. 2016;20:82–8.
- Pectasides D, Pectasides E, Papaxoinis G, Skondra M, Gerostathou M, Karageorgopoulou S, et al. Testicular function in poor-risk nonseminomatous germ cell tumors treated with methotrexate, paclitaxel, ifosfamide, and cisplatin combination chemotherapy. J Androl. 2009;30:280–6.
- Farmakiotis D, Farmakis C, Rousso D, Kourtis A, Katsikis I, Panidis D. The beneficial effects of toremifene administration on the hypothalamic-pituitary-testicular axis and sperm parameters in men with idiopathic oligozoospermia. Fertil Steril. 2007;88:847–53.
- Douglas JM Jr, David LG, Remington ML, Paulsen CA, Perrin EB, Goodman P, et al. A double-blind, placebo-controlled trial of the effect of chronically administered oral acyclovir on sperm production in men with frequently recurrent genital herpes. J Infect Dis. 1988;157:588–93.
- Amory JK, Muller CH, Page ST, Leifke E, Pagel ER, Bhandari A, et al. Miglustat has no apparent effect on spermatogenesis in normal men. Hum Reprod. 2007;22:702–7.
- van Leeuwen E, Wit FW, Repping S, Eeftinck Schattenkerk JK, Reiss P, van der Veen F, et al. Effects of antiretroviral therapy on semen quality. AIDS. 2008;22:637–42.



- Tsourdi E, Kourtis A, Farmakiotis D, Katsikis I, Salmas M, Panidis D. The effect of selective estrogen receptor modulator administration on the hypothalamic-pituitary-testicular axis in men with idiopathic oligozoospermia. Fertil Steril. 2009;91: 1427–30
- Dwivedi C, Long NJ. Effect of cholinergic agents on human spermatozoa motility. Biochem Med Metab Biol. 1989;42:66–70.
- Hendry WF, Munro DD. "Wash leather scrotum" (scrotal dermatitis): a treatable cause of male infertility. Fertil Steril. 1990;53: 379–81.
- Farnsworth WH, Hoeg JM, Maher M, Brittain EH, Sherins RJ, Brewer HB Jr. Testicular function in type II hyperlipoproteinemic patients treated with lovastatin (mevinolin) or neomycin. J Clin Endocrinol Metab. 1987;65:546–50.
- Agirregoitia E, Valdivia A, Carracedo A, Casis L, Gil J, Subiran N, et al. Expression and localization of delta-, kappa-, and mu-opioid receptors in human spermatozoa and implications for sperm motility. J Clin Endocrinol Metab. 2006;91:4969–75.
- Corvasce A, Albino G, Leonetti T, Buonomo AF, Marucco EC.
 Once-a-day tadalafil administration improves the spermogram parameters in fertile patients. Arch Ital Urol Androl. 2015;87:210–3.
- 102. U.S. Food and Drug Administration. Guidance for industry: pregnancy, lactation, and reproductive potential: labeling for human prescription drug and biological products—content and format. 2014, Draft guidance. [Internet] [Cited 8 Aug 2017]. Available from: http://www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/UCM425398.pdf.
- 103. U.S. Food and Drug Administration. Guidance for industry: reproductive and developmental toxicities—integrating study results to assess concerns. [Internet] September 2011. [Cited 8 Aug 2017]. Available from: www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/ucm079240.pdf.
- Schapaugh AW, McFadden LG, Zorrilla LM, Geter DR, Stuchal LD, Sunger N, et al. Analysis of EPA's endocrine screening battery and recommendations for further review. Regul Toxicol Pharmacol. 2015;72:552–61.

- Mak IW, Evaniew N, Ghert M. Lost in translation: animal models and clinical trials in cancer treatment. Am J Transl Res. 2014;6: 114–8
- Odom DT, Dowell RD, Jacobsen ES, Gordon W, Danford TW, MacIsaac KD, et al. Tissue-specific transcriptional regulation has diverged significantly between human and mouse. Nat Genet. 2007;39:730–2.
- Shanks N, Greek R, Greek J. Are animal models predictive for humans? Philos Ethics Humanit Med. 2009;4:2.
- Nair AB, Jacob S. A simple practice guide for dose conversion between animals and human. J Basic Clin Pharm. 2016;7:27–31.
- Gray LE Jr, Wilson V, Noriega N, Lambright C, Furr J, Stoker TE, et al. Use of the laboratory rat as a model in endocrine disruptor screening and testing. ILAR J. 2004;45:425–37.
- Cook JC, Klinefelter GR, Hardisty JF, Sharpe RM, Foster PM. Rodent Leydig cell tumorigenesis: a review of the physiology, pathology, mechanisms, and relevance to humans. Crit Rev Toxicol. 1999;29:169–261.
- Johnson L, Petty CS, Neaves WB. A comparative study of daily sperm production and testicular composition in humans and rats. Biol Reprod. 1980;22:11233–43.
- Misell LM, Holochwost D, Boban D, Santi N, Shefi S, Hellerstein MK, et al. A stable isotope-mass spectrometric method for measuring human spermatogenesis kinetics in vivo. J Urol. 2006;175: 242–6.
- Gaylor D, Ryan L, Krewski D, Zhu Y. Procedures for calculating benchmark doses for health risk assessment. Regul Toxicol Pharmacol. 1998;28:150–64.
- Neaves WB, Johnson L, Porter JC, Parker CR Jr, Petty CS. Leydig cell numbers, daily sperm production, and serum gonadotropin levels in aging men. J Clin Endocrinol Metab. 1984;59:756–63.
- Oksel C, Ma CY, Liu JJ, Wilkins T, Wang XZ. Literature review of (Q)SAR modelling of nanomaterial toxicity. Adv Exp Med Biol. 2017;947:103–42.
- Ford KA. Refinement, reduction, and replacement of animal toxicity tests by computational methods. ILAR J. 2016;57:226–33.

