

ANTIMICROBIAL RESISTANCE: George M. Eliopoulos, Section Editor

The Use of Noncarbapenem β -Lactams for the Treatment of Extended-Spectrum β -Lactamase Infections

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The continued rise in infections caused by extended-spectrum β -lactamase (ESBL)-producing pathogens is recognized globally as one of the most pressing concerns facing the healthcare community. Carbapenems are widely regarded as the antibiotics of choice for the treatment of ESBL-producing infections, even when in vitro activity to other β -lactams has been demonstrated. However, indiscriminant carbapenem use is not without consequence, and carbapenem overuse has contributed to the emergence of carbapenem-resistant Enterobacteriaceae. The use of non-carbapenem β -lactams for the treatment of ESBL infections has yielded conflicting results. In this review, we discuss the available data for the use of cephamycins, cefepime, piperacillin-tazobactam, ceftolozane-tazobactam, and ceftazidime-avibactam for the treatment of ESBL infections.

Keywords. ESBLs; carbapenems; cephamycins; cefepime; piperacillin-tazobactam.

Since their description in the 1980s, extended-spectrum β -lactamase (ESBL)-producing organisms have been recognized as a global threat [1–4]. Through the years, these enzymes have undergone substantial biochemical alterations resulting in the ability to more efficiently hydrolyze β -lactam antibiotics [5].

ESBLs have been detected worldwide in several gram-negative genus and species; however, they are most prevalent among *Escherichia coli* and *Klebsiella* species.

Carbapenems are widely regarded as the antibiotics of choice for the treatment of ESBL-producing infections, even when in vitro activity to other β -lactams is demonstrated [6, 7]. They are stable to ESBL hydrolytic activity and numerous studies demonstrate their efficacy [6]. However, carbapenem overutilization stimulates various resistance pathways including outer membrane protein (OMP) mutations and the selection of β -lactamases capable of hydrolyzing carbapenems [8]. Whenever equally efficacious alternate agents exist, efforts should be made to limit the use of carbapenem antibiotics.

The increased hydrolytic activity of ESBLs against third-generation cephalosporins, particularly ceftriaxone and cefotaxime, and their associated poor outcomes, usually disqualifies them from consideration for the treatment of ESBL-associated infections [9]. However, there are a number of scenarios that

give us pause as to whether other noncarbapenem β -lactams can be prescribed in place of carbapenems (Table 1).

In this review, we will summarize the data available on the use of noncarbapenem β -lactams compared to carbapenems for the treatment of ESBL infections. Our review is limited to antibiotics available in the United States, with the most in-depth discussion focused on piperacillin-tazobactam, as this agent appears to arouse the greatest debate. To date, there have been no adequately powered randomized controlled trials evaluating the use of β -lactam agents for ESBL infections. Therefore, we are left with weighing the pros and cons of available observational studies (Table 2), while accounting for the inherent limitations of this study design. Although we will not formally discuss non- β -lactam options in this review, several non- β -lactam options deserve consideration for the treatment of ESBL infections when susceptible in vitro, with some stipulations based on the source of infection and severity of illness.

CEPHAMYCINS

Cephamycins demonstrate consistent in vitro activity against ESBL-producing Enterobacteriaceae isolates, distinguishing them from AmpC cephalosporinases [10]. Concerns with administering cephamycins for the treatment of ESBL infections stem from reports demonstrating acquisition of OMP mutations and/or plasmids encoding AmpC cephalosporinases during exposure to these agents [11–14]. In at least 1 published case, a patient with a *K. pneumoniae* ESBL isolate became resistant to both flomoxef and carbapenem therapy, after exposure to flomoxef, due to an OMPk36 mutation, in combination with acquisition of the plasmid-mediated AmpC cephalosporinase gene *bla*_{DHA-1} [14]. It is unclear how frequently such mutations

Received 8 October 2016; editorial decision 24 December 2016; accepted 14 January 2017; published online January 25, 2017.

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Clinical Infectious Diseases® 2017;64(7):972–80

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Table 1. Potentially Favorable Circumstances for Noncarbapenem- β -Lactams in the Treatment of Extended-Spectrum β -Lactamase Infections

- What if noncarbapenem β -lactam minimum inhibitory concentrations are low?
- What if high-dose, frequent-interval β -lactams or cefepime is administered?
- What if extended-infusion noncarbapenem β -lactams are administered?
- If carbapenem antibiotics are administered when the burden of bacteria is highest, can therapy be transitioned to a noncarbapenem after a short period of time?
- If a β -lactamase inhibitor is administered, does the type of β -lactamase inhibitor matter (eg, tazobactam, sulbactam, clavulanic acid, or avibactam)?
- Does it matter if the ESBL resistance mechanism is a bla_{TEM} type, bla_{CTX-M} type, or bla_{SHV} type?
- Does the genus or species of the ESBL producer matter?
- Does the source of bacteremia and if source control measures were taken matter?
- Should the severity of illness determine if a carbapenem or noncarbapenem agent is administered?

Abbreviations: β -lactam, β -lactam- β -lactamase inhibitor; ESBL, extended-spectrum β -lactamase.

and gene acquisitions occur and what the predisposing host and environmental factors are.

A number of cephamycins are commercially available including cefoxitin, cefotetan, cefmetazole, flomoxef, and moxalactam. Unfortunately, clinical data evaluating cephamycins for ESBL infections are scarce, and data comparing the relative efficacy of the various cephamycins are virtually nonexistent. Existing observational studies comparing cephamycins and carbapenems are plagued by the inherent limitations of observational studies—most notably confounding by indication—as well as small sample sizes (Table 2) [15–20]. Only 1 of these studies showed improved outcomes among ESBL-infected patients

Table 2. Select Limitations of Existing Observational Studies Comparing Noncarbapenem β -Lactam Antibiotics and Carbapenems for the Treatment of Extended-Spectrum β -Lactamase Infections

- Inconsistent criteria for extended-spectrum β -lactamase production
- Confounding by indication (ie, ill-appearing patients more likely to receive the more “aggressive” therapy, ie, carbapenems)
- Differences in outcomes definitions
- Delays in initiating appropriate antibiotic therapy
- Classification issues for patients initially receiving empiric noncarbapenem β -lactam therapy, then transitioned to carbapenem therapy
- Large proportions of patients receiving combination antibiotic therapy
- Often single-center experiences
- Sample sizes limit sufficient power to detect differences between treatment approaches, if such differences exist
- Insufficient subgroups for analysis (eg, proportion of *Escherichia coli* vs *Klebsiella pneumoniae*, proportion of bla_{CTX-M} vs bla_{SHV})
- Disproportionate numbers of patients with low-inoculum and high-inoculum infections
- Differences in antibiotic susceptibility criteria utilized
- Differences in local epidemiology of in vitro activity of noncarbapenem β -lactams
- Insufficient data on dosing regimens
- Insufficient data on clinical outcomes with extended-infusion β -lactam therapy

treated with carbapenems compared with cephamycins, while the others could not detect differences [18] (Table 3). Optimal dosing regimens and achievable target attainment for various cephamycin minimum inhibitory concentrations (MICs) need to be explored [21].

We believe that cephamycins may be useful agents in the treatment of nonsevere ESBL-producing infections from urinary sources. Given the limited data on nonurinary sources and severe infections, it is the opinion of the authors that the use of cephamycins for severe ESBL infections and those originating outside of the urinary tract should be avoided until more data are available.

CEFEPIME

Cefepime is an oxyimino-cephalosporin agent with enhanced stability against degradation by β -lactamases. The current European Committee on Antimicrobial Susceptibility Testing (EUCAST) and Clinical and Laboratory Standards Institute (CLSI) susceptibility breakpoints for cefepime are 1 μ g/mL and 8 μ g/mL (accounting for drug dosing), respectively [22, 23]. There is concern that the current CLSI cefepime breakpoint leaves some ESBL enzymes in the susceptible range (ie, “hidden resistance”) [24].

Concerns about the diminished efficacy of cefepime for the treatment of ESBL infections with higher bacterial inoculums (eg, intra-abdominal infections, pneumonia, osteoarticular infections) have tempered enthusiasm for the use of cefepime for this indication. This so called “inoculum effect,” in which drug MICs increase dramatically in the presence of an increased bacterial load despite apparent initial susceptibility, has been observed in both in vitro and animal studies [25–31]. Highlighting one experience, Burgess and colleagues evaluated cefepime and meropenem against standard- and high-inoculum ESBL *Klebsiella pneumoniae* infections [32]. Each of these antibiotics maintained bactericidal activity against standard inoculums but at high inoculums, in contrast to meropenem, cefepime was unable to sustain bactericidal activity against ESBL producers. In this same series of experiments, both agents maintained >99% killing of high-inoculum non-ESBL infections over a 24-hour observation period. The contribution of the inoculum effect toward treatment failures has not been the subject of comprehensive clinical review.

Apart from the inoculum effect, an alternative explanation for poor outcomes associated with cefepime therapy relates to failure to meet necessary pharmacodynamic targets due to inadequate dosing and/or interval schedules [33]. A target of 50% free cephalosporin drug concentration greater than the drug MIC ($fT > MIC$) has been identified as the quantitative exposure necessary for effective cephalosporin bactericidal activity [34]. Monte Carlo simulation is a mathematical tool that may be applied to integrating multiple pharmacokinetic/pharmacodynamic (PK/PD) variables to estimate the probability of target attainment for specific

Table 3. Observational Studies Evaluating Clinical Outcomes of Patients with Extended-Spectrum β -Lactamase Infections Comparing Treatment with Cephamycins versus Carbapenems

Study	Cephamycin	Carbapenem	Organism(s)	ESBL Criteria	Sites and Sources of Infection	Severity of Illness at Infection Onset	Clinical Outcomes (Cephamycins vs Carbapenems)	Select Limitations ^a
Lee et al [15]	Flomoxef (n = 7)	n = 20	<i>Klebsiella pneumoniae</i> (100%)	Molecular confirmation	Site: blood-stream (100%) Sources: pneumonia (56%), intra-abdominal (19%), urinary (11%), SSTI (4%)	52% admitted to ICU	Mortality at 14 d: 29% vs 25% (ns ^b)	More severely ill patients in carbapenem arm
Doi et al [17]	Cefmetazole (n = 10)	n = 12	<i>Escherichia coli</i> (95%), <i>K. pneumoniae</i> (5%)	Disk diffusion	Site: urine (100%)	Not provided (but likely low)	Clinical cure at 4 weeks: 90% vs 100% (ns)	More patients in carbapenem group with bacteremia or complicated UTI; 90% of patients in cephamycin group received alternative agents initially
Yang et al [18]	Flomoxef (n = 29)	n = 28	<i>K. pneumoniae</i> (100%)	Disk diffusion	Site: blood-stream (100%) Source: fistula, graft, catheter (100%)	51% admitted to ICU	Mortality at 14 days: 55% vs 39% ($P < .05$)	Unclear if removal of infected hardware occurred at similar percentages across the 2 treatment groups
Pilmis et al [19]	Cefoxitin ^c (n = 8)	n = 31	<i>E. coli</i> (32%), <i>K. pneumoniae</i> (32%), <i>Enterobacter cloacae</i> (36%)	Not described	Site: urine (75%), bloodstream (25%)	Not provided	Clinical or microbiological relapse at 30 d: 13% vs 23% (ns)	Patients in carbapenem group more likely to be immunocompromised
Matsumura et al [20]	Empiric: cefmetazole or flomoxef (n = 26) Definitive: cefmetazole or flomoxef (n = 59)	Empiric: n = 45 Definitive: n = 54	<i>E. coli</i> (100%)	Disk diffusion	Site: blood-stream (100%) Source: urinary (45%), intra-abdominal (32%)	41% with severe sepsis	Mortality at 30 d in empiric group: 8% vs 9% (ns) Mortality at 30 d in definitive therapy group: 5% vs 9% (ns)	Patients in carbapenem group more ill and more likely to be immunocompromised

Abbreviations: ESBL, extended-spectrum β -lactamase; ICU, intensive care unit; ns, not significant; SSTI, skin and soft tissue infection; UTI, urinary tract infection.

^aSmall sample size, residual confounding, and confounding by indication are limitations for all included studies.

^bNot statistically significant using a P value $\leq .05$.

^cExcluding patients who initially received carbapenems and then converted to cephamycins.

$fT > MIC$ targets. There are a wide range of dosing regimens for cefepime, which may dramatically alter exposure and outcomes associated with treatment. Reese and colleagues showed that 2 g of cefepime every 12 hours was unable to achieve an adequate $fT > MIC$ in a medically complex population in which the cefepime MIC_{50} and MIC_{90} values were 8 $\mu\text{g/mL}$ and 16 $\mu\text{g/mL}$, respectively [35]. Others have shown similar results [36], prompting the CLSI to implement a susceptible dose-dependent category encouraging cefepime doses of 1 g every 8 hours or 2 g every 8 hours for organisms with MICs of 4 or 8 $\mu\text{g/mL}$, respectively [22].

The relative contribution of ESBL production and drug MIC towards cefepime efficacy remains controversial. Andes and Craig studied the impact of ESBL production on the activity of cefepime in a neutropenic murine thigh model of infection [37]. ESBL production in 5 isolates had no impact upon the $fT > MIC$ necessary for in vivo cefepime efficacy. Rather, they concluded, cefepime efficacy is predicted exclusively by the contribution

of the MIC in relation to the magnitude of drug exposure. However, cefepime failures for ESBL-producing infections have been observed with cefepime MICs as low as 1 $\mu\text{g/mL}$ [38], suggesting that both the presence of ESBLs and higher MICs likely contribute to cefepime failures.

Results of observational studies comparing the activity of cefepime and carbapenems for invasive ESBL infections have been conflicting with some studies showing no difference [39, 40] and others suggesting cefepime therapy is inferior [38, 41, 42] (Table 4). Lee and colleagues conducted an observational study including 17 patients with ESBL bacteremia receiving cefepime therapy and 161 patients receiving carbapenem therapy [42]. Patients receiving carbapenems were over 7 times more likely to survive than patients receiving cefepime. Mortality rates were lower with cefepime MICs ≤ 1 $\mu\text{g/mL}$ (17%) compared with MICs of 2–8 $\mu\text{g/mL}$ (46%). No patients received cefepime 2 g every 8 hours or continuous infusion cefepime to evaluate how

Table 4. Observational Studies Evaluating Clinical Outcomes of Patients with Extended-Spectrum β -Lactamase Infections Comparing Treatment with Cefepime versus Carbapenems

Study	Cefepime (Dosing if Available)	Carbapenem	Organism(s)	ESBL Criteria	Sites and Sources of Infection	Severity of Illness at Infection Onset	Clinical Outcomes (Cefepime vs Carbapenems)	Select Limitations ^a
Zanetti et al [41]	n = 13; 2 g every 8 h	n = 10	<i>Klebsiella pneu- moniae</i> (96%), <i>Enterobacter aerogenes</i> (4%)	Not provided	Site: pneumonia (100%)	100% admitted to ICU	Clinical response: 69% vs 100% ($P < .05$)	Cefepime MIC distributions not provided; baseline comparisons specifically for ESBL producers not provided
Goethaert et al [39]	n = 21; 2 g every 8 h	n = 23	<i>E. aerogenes</i> (100%)	Disk diffusion and molecular confirmation	Sites: pneumonia (64%), bloodstream (16%), urine (5%), intra-abdominal (14%), bone (0.3%)	100% admitted to ICU	Mortality at 30 d: 33% vs 26% (ns)	~80% of patients (equally in both groups) received combination antibiotic therapy; bacteremia more likely in carbapenem group; analysis limited to isolates con- taining bla _{TEM24} gene
Chopra et al [40]	Empiric: cefepime monotherapy n = 43 Definitive: cefepime monotherapy n = 9	Empiric: carbapenem monotherapy n = 14 Definitive: carbapenem monotherapy n = 33	<i>K. pneumo- niae</i> (83%), <i>Escherichia coli</i> (17%)	Disk diffusion	Site: bloodstream (100%) Sources: catheter (75%)	41% admitted to ICU	In-hospital mortality for empirical group: 40% vs 36% (ns) In-hospital mortality for definitive group: 33% vs 36% (ns)	Cefepime dosing not described; baseline comparisons of patients receiving cefepime vs carbapenems not provided
Lee et al [42]	Empiric: n = 21 Definitive: n = 17 1 g every 8 h (41%) and 2 g every 12 h (47%)	Empiric: n = 91 Definitive: n = 161	<i>Enterobacter cloacae</i> (55%), <i>E. coli</i> (24%), <i>K. pneumoniae</i> (21%)	ESBL Etest strip and molecular confirmation	Site: bloodstream (100%) Sources: pneumonia (24%), catheter (21%), intra-ab- dominal (16%), skin and soft tissue (6%), urinary (22%)	67% had Pitt bacte- remia score ≥ 4	Mortality at 30 d in definitive therapy group: 59% vs 17% ($P < .01$)	21% of patients receiving cefepime had cefepime MICs > 8 $\mu\text{g/mL}$; propensity score analysis used but no description of variables included in generating propen- sity score, making residual confounding possible
Wang et al [38]	n = 17; 2 g every 8 h (71%) and 1 g every 8 h (29%)	n = 51	<i>E. coli</i> (32%), <i>Klebsiella</i> spp (63%), <i>Proteus mirabilis</i> (3%)	ESBL Etest strip	Site: bloodstream (100%) Sources: catheter (44%), urinary (31%), biliary (9%), pneumonia 15%, intra-abdominal (13%), skin and soft tissue (3%)	29% admitted to ICU	Mortality at 14 d: 41% vs 20% ($P = .08$)	No patients received continuous-infusion cefepime

Abbreviations: ESBL, extended-spectrum β -lactamase; ICU, intensive care unit; ns, not significant; MIC, minimum inhibitory concentration.

^aSmall sample size, residual confounding, and confounding by indication are limitations for all included studies.

optimizing PK/PD could have impacted outcomes. However, in a study where all ESBL-infected patients received cefepime at 8-hour intervals and 70% of patients received 2 g every 8 hours, inferior outcomes were still observed in the cefepime group [38].

We believe cefepime can be considered for nonsevere ESBL infections where the agent can achieve high concentrations to ensure pharmacodynamic targets are met (eg, urinary tract infections (UTIs) with cefepime MICs ≤ 2 $\mu\text{g}/\text{mL}$). We do not favor the use of cefepime for serious ESBL infections. If cefepime is administered for nonsevere ESBL-producing infections with MICs of 4–8 $\mu\text{g}/\text{mL}$ based on susceptibility criteria, we recommend administering 2 g every 8 hours, possibly as a continuous infusion.

PIPERACILLIN-TAZOBACTAM

Current breakpoints for piperacillin-tazobactam (PTZ) according to the CLSI [22] and EUCAST [23] are ≤ 16 $\mu\text{g}/\text{mL}$ and ≤ 8 $\mu\text{g}/\text{mL}$, respectively. Despite a considerable proportion of ESBL isolates demonstrating susceptibility to PTZ [43], the role of this compound for patients infected with ESBL-producing pathogens remains unclear. Although ESBLs are generally inhibited by β -lactamase inhibitors, occasionally organisms produce multiple ESBLs simultaneously or have additional resistance mechanisms (eg, AmpC β -lactamases, OMP mutations), providing a complex background that may reduce the effectiveness of these agents [5]. Similar to cefepime, an “inoculum effect” has been proposed. This is supported by in vitro, animal data, and case reports [26, 30, 44–46]. In the time-kill experiments by Burgess and colleagues described above, PTZ maintained $>99\%$ killing against high inoculum non-ESBL *K. pneumoniae* isolates over a 24-hour period, but this effect was unsustainable against high-inoculum ESBL infections, where regrowth was observed at 8 hours [32].

Existing observational data have indicated contradictory results between outcomes of patients receiving PTZ and carbapenems for the treatment of ESBL infections (Table 5) [8, 47, 48–54]. Rodríguez-Baño and colleagues provided some of the earliest robust data evaluating β -lactam- β -lactamase inhibitors (β L- β LI) and carbapenems for the treatment of ESBL bacteremia by conducting a post hoc observational study of Spanish patients from 6 cohorts [47]. To overcome limitations with antibiotic regimen changes between empiric therapy and definitive therapy regimens, 2 nonmutually exclusive cohorts were constructed and independently analyzed. Thirty-day mortality was 10% and 19% in the empiric cohort and 9% and 17% in the definitive cohort for β L- β LI and carbapenems, respectively. Although these differences did not reach statistical significance, the absolute difference in mortality being almost twice as high in the carbapenem group gives us pause as to whether some selection bias remained that was not unaccounted for. There were some aspects to this study potentially compromising its generalizability. First, only *E. coli* isolates that typically contained *bla*_{CTX-M} genes were included. It is

unclear if similar outcomes would be observed with *K. pneumoniae* isolates containing *bla*_{SHV}-type genes. Second, approximately 70% of bloodstream isolates were from urinary and biliary sources (ie, “low-inoculum” infections). It remains undetermined if similar findings would have been observed if the majority of patients had bloodstream infections due to pneumonia or intra-abdominal infections. Third, approximately 13% of patients required intensive care unit (ICU) admission, signifying that most patients were not critically ill. Additionally, the median piperacillin MIC was only 2 $\mu\text{g}/\text{mL}$. Importantly, mortality was 4.5% when the MIC was ≤ 4 $\mu\text{g}/\text{mL}$ vs 23% for an MIC of ≥ 8 $\mu\text{g}/\text{mL}$. Finally, $>90\%$ of patients receiving PTZ were administered 4.5 g every 6 hours. Simulation models have shown a 99% probability of attaining PK/PD targets against ESBL producers with PTZ administered at 4.5 g every 6 hours when MICs are ≤ 8 $\mu\text{g}/\text{mL}$, compared with a probability of only 57% with piperacillin MICs of 16 $\mu\text{g}/\text{mL}$ [55].

A meta-analysis by Vardakas et al compared carbapenem and β L- β LI for ESBL bacteremia for both empiric use (273 β L- β LI vs 317 carbapenems) and definitive use (118 β L- β LI and 398 carbapenems) [6]. There was no difference in all-cause mortality between the empiric and definitive therapies. The meta-analysis was limited by considerable heterogeneity of the studies and because more severely ill patients tended to be prescribed carbapenems (Table 2). Unfortunately, adjustment for potential confounders using patient-level data was not feasible.

As the majority of patients had a urinary source for their bacteremia in the Rodríguez-Baño study, Ofer-Friedman and colleagues compared the efficacy of β L- β LI and carbapenems for the treatment of ESBL bacteremia, excluding urinary sources [8]. Thirty-day mortality was 60% for the PTZ group and 34% for the carbapenem group. These differences did not attain statistical significance ($P = .10$), although it is plausible that this is at least partly due to the restricted sample size. This limitation notwithstanding, this study suggests that for critically ill patients with ESBL bacteremia from nonurinary sources, PTZ therapy may lead to less desirable outcomes than carbapenem therapy.

Tamma et al compared 14-day mortality of patients receiving PTZ and carbapenems as empiric therapy in a cohort of patients with ESBL bacteremia who all received definitive carbapenem therapy [51]. The study population resembled the Ofer-Friedman et al study where about one-third of the isolates were *E. coli*, one-third of patients required ICU care, the majority of patients had “high-inoculum” infections, and most ESBL isolates had elevated PTZ MICs. In fact, 99% of organisms had piperacillin MICs of ≥ 4 $\mu\text{g}/\text{mL}$, with a median MIC of 8 $\mu\text{g}/\text{mL}$. Thirty-day mortality was higher in the PTZ group (Table 5). It should be noted that in both this study and the Ofer-Friedman et al study, the minority of patients received PTZ dosed at 4.5 g every 6 hours.

More recently, Gutiérrez-Gutiérrez et al conducted a multinational, observational study investigating this question in a design similar to the Rodríguez-Baño study [52]. In fact, some of the same

Table 5. Observational Studies Evaluating Clinical Outcomes of Patients with Extended-Spectrum β -Lactamase Bacteremia Comparing Treatment with β -Lactam- β -Lactamase Inhibitors versus Carbapenems

Study	β -Lactam	Carbapenem	Organism(s)	ESBL Criteria and β -Lactamase MIC Distribution, μ g/mL	Sources of Bacteremia	ICU Admission at Infection Onset	Clinical Outcomes (β -Lactam vs carbapenems)	Select Limitations ^a
Kang et al [49]	n = 36	n = 78	<i>Escherichia coli</i> (68%), <i>Klebsiella pneumoniae</i> (32%)	Not provided	Sources: not provided	Not provided	Mortality at 30 d: 22% vs 27% (ns)	Baseline comparisons not provided
Rodriguez-Baño et al [47]	Empiric cohort: n = 72 Definitive cohort: n = 54	Empiric cohort: n = 31 Definitive cohort: n = 120	<i>E. coli</i> (100%)	Molecular detection MICs: \leq 1 (29%), 2 (23%), 4 (11%), 8 (17%), 16 (20%)	Source: urinary or biliary (70%)	13%	Mortality at 30 d in empiric cohort: 10% vs 19% (ns) Mortality at 30 d in definitive cohort: 9% vs 17% (ns)	Generalizability to patients infected with ESBL bloodstream infections from high-inoculum sources, elevated piperacillin MICs, and severe infections not clear
Harris et al [50]	n = 24 (100% received 4.5 g per dose PTZ)	n = 23	<i>E. coli</i> (86%), <i>K. pneumoniae</i> (14%)	Cefotaxime nonsusceptible MICs: \leq 4 (71%) and 8 (29%)	Sources: urinary (47%), biliary (9%)	15%	Mortality at 30 d: 8% vs 17% (ns)	More immunocompromised patients in carbapenem group, generalizability to patients infected with ESBL bloodstream infections from high-inoculum sources, elevated piperacillin MICs, and severe infections not clear
Ofer-Friedman et al [8]	n = 10 (dosing regimens not described)	n = 69	<i>E. coli</i> (53%), <i>K. pneumoniae</i> (28%), <i>Proteus mirabilis</i> (19%)	Disk diffusion MICs: median 8	Sources: pneumonia (34%), skin and soft tissue (28%), biliary (17%), intra-abdominal (9%)	>50%	Mortality at 30 d: 60% vs 34% ($P = .10$) Mortality at 90 d: 80% vs 48% ($P = .03$)	Endpoint of 90-d mortality may not be representative of mortality due to poor antibiotic treatment choices; dosing not described
Tamma et al [51]	n = 103 (40% received 4.5 g per dose PTZ)	n = 110	<i>K. pneumoniae</i> (68%), <i>E. coli</i> (31%), <i>P. mirabilis</i> (1%)	Disk diffusion MICs: 2 (1%), 4 (39%), 8 (46%), 16 (14%)	Sources: catheter (46%), urinary (21%), intra-abdominal (17%), pneumonia (9%)	34%	Mortality at 14 d: 17% vs 8% ($P < .05$) Mortality at 30 d: 26% vs 11% ($P < .01$)	Only ~40% received 4.5 g every 6 h; no patients received extended-infusion therapy
Ng et al [53]	Empiric cohort: n = 97 (~100% received 4.5 g)	Empiric cohort: n = 57	<i>E. coli</i> (67%), <i>K. pneumoniae</i> (33%)	Resistance to third-generation cephalosporins MICs: not provided	Sources: catheter (4%), urinary (59%), biliary (9%), pneumonia (9%), intra-abdominal (5%)	9%	Mortality at 30 d: 31% vs 30% (ns)	PTZ MIC distribution not provided; unclear what proportion of infections were due to ESBL producers
Gutiérrez-Gutiérrez et al [52]	Empiric cohort: n = 170 (65% received 4.5 g per dose PTZ) Definitive cohort: n = 92 (83% received 4.5 g per dose PTZ)	Empiric cohort: n = 195 Definitive cohort: n = 509	<i>E. coli</i> (73%), <i>K. pneumoniae</i> (19%)	Elevated cephalosporin MICs with molecular confirmation of ~30% of cohort MICs: not provided	Sources: urinary (45%), biliary (12%)	11%	Mortality at 30 d in empiric cohort: 18% vs 20% (ns) Mortality at 30 d in definitive cohort: 10% vs 14% (ns)	Generalizability to patients infected with ESBL bloodstream infections from high-inoculum sources, elevated piperacillin MICs, and severe infections not clear

Abbreviations: β -Lactam, β -lactam- β -lactamase inhibitor; ESBL, extended-spectrum β -lactamase; ICU, intensive care unit; MIC, minimum inhibitory concentration; ns, not significant; PTZ, piperacillin-tazobactam.

^aSmall sample size, residual confounding, and confounding by indication are limitations for all included studies.

patients were included in both cohorts. This has been the largest study to date comparing the effectiveness of β L- β LI compared with carbapenems for the treatment of ESBL bloodstream infections, including 365 patients in the empiric therapy group and 601 patients in the targeted therapy group. This study was expanded to include patients infected with *K. pneumoniae* isolates, comprising 19% of cases. However, similar to the original study, only 11% of patients required ICU-level care and the majority of isolates were from urinary or biliary sources. Similar to the Rodriguez-Baño study, the vast majority of patients (83%) received 4.5-g PTZ dosing. Mortality was comparable between the study groups in both the empiric and definitive cohorts. The investigators attempted to overcome some of the differences with the Ofer-Friedman et al and Tamma et al studies by conducting a series of subgroup analyses comparing outcomes for patients treated with β L- β LI and carbapenems (eg, *E. coli* vs *K. pneumoniae* isolates, severe sepsis vs non-severely ill, urinary sources vs other sources). In subgroup analysis, the investigators were able to demonstrate that the point estimates and confidence intervals resembled estimates from the entire cohort. Although the investigators found no differences within the subgroups, it is unknown if this is an artifact of the small sample size within any individual subgroup. An important observation was that patients with ESBL-producing *K. pneumoniae* bloodstream infections had almost twice the odds of dying within 30 days compared to patients with ESBL-producing *E. coli* bacteremia.

Finally, Ng and colleagues evaluated 30-day mortality comparing empiric PTZ and carbapenem in 151 patients with presumed ESBL bloodstream infections. Thirty-day mortality was no different between the groups [53]. As with previous studies that resulted in similar outcomes between the treatment groups, a minority of patients were in the ICU (<10%), the majority of patients were infected with *E. coli* bacteremia, and almost 70% of patients had urinary or biliary sources of bacteremia. Patients in the PTZ group received 4.5-g dosing. Importantly, confirmatory ESBL testing was not conducted, so it is unclear what proportion of ceftriaxone-resistant isolates were indeed ESBL producers.

An important lingering question is as follows: Are the poorer outcomes in the Ofer-Friedman et al and Tamma et al studies related to the suboptimal performance of β L- β LI for critically ill patients with more aggressive infections (high inoculum, higher median PTZ MICs, greater proportion of *K. pneumoniae* isolates), or are they related to the underdosing of β L- β LI in both of these studies? Unfortunately, the answer to this question remains unknown. It is not clear if patients receiving β L- β LI would have had more favorable outcomes had target PK/PD exposures been achieved. Perhaps the discrepancies between these studies will be resolved with the MERINO trial [54], a multicenter, randomized noninferiority trial comparing meropenem 1 g every 8 hours and PTZ 4.5 g every 6 hours for ceftriaxone-nonsusceptible *E. coli* and *Klebsiella* species bloodstream infections (ClinicalTrials.gov identifier NCT02176122).

Unfortunately, the question of whether β L- β LI and carbapenems lead to equivalent outcomes when prescribed for ESBL infections remains unclear. Based on the experiences of the Rodriguez-Baño, Gutiérrez-Gutiérrez, and Ng studies, β L- β LI appear to be very reasonable options for low- to moderate-severity infections, infections resulting from urinary or biliary sources, and infections with piperacillin MICs <4 μ g/mL. For critically ill patients, patients with higher inoculum infections, and elevated piperacillin MICs, we believe that it might be more appropriate to administer carbapenem therapy, at least initially, until more data are available. Regardless, if PTZ is administered to patients with invasive ESBL infections, we would recommend administering 4.5 g every 6 hours (or 4.5 g every 8 hours as extended infusion) [55].

NEWER B-LACTAM-B-LACTAMASE INHIBITORS

The US Food and Drug Administration recently approved 2 new β L- β LI, ceftolozane-tazobactam and ceftazidime-avibactam, which are active in vitro against ESBL-producing organisms. Ceftolozane demonstrates good activity against Enterobacteriaceae. Similar to other oxyimino-cephalosporins, its activity is limited against ESBLs. Tazobactam is a potent, irreversible inhibitor of most ESBLs. The MIC₅₀/MIC₉₀ of this agent for ESBL-producing *E. coli* and *K. pneumoniae* are 0.5/4 μ g/mL and 4/>32 μ g/mL, respectively [56, 57]. Differences in MIC distributions may be reflective of discrepancies in ESBL genes present. The *bla*_{CTX-M} genes predominate in *E. coli*, whereas there is often a preponderance of *bla*_{TEM/SHV} in *K. pneumoniae*, with variations in local epidemiology [5].

Ceftolozane-tazobactam (in combination with metronidazole) was compared to meropenem for the treatment of complicated intra-abdominal infections in phase 2 [58] and phase 3 [59] trials that included 4 and 50 people, respectively, with ESBL-producing Enterobacteriaceae. Although the limited number of ESBLs precluded a robust analysis, this compound performed similarly against ESBL-producing and non-ESBL producing isolates.

Ceftazidime-avibactam is usually more active in vitro against ESBL producers than ceftolozane-tazobactam. The MIC₅₀/MIC₉₀ of this agent for ESBL-producing *E. coli* and *K. pneumoniae* are 0.12/0.25 μ g/mL and 0.5/1 μ g/mL, respectively [60]. Similar to ceftolozane-tazobactam, phase 2 [61] and phase 3 studies [62] compared ceftazidime-avibactam (plus metronidazole) vs meropenem for intra-abdominal infections, but did not specifically compare outcomes of ESBL-confirmed pathogens. Data from a phase 3 study comparing ceftazidime-avibactam and doripenem in UTIs showed similar microbiological response for ceftazidime-resistant Enterobacteriaceae, most of which were ESBL producers [63]. Although evidence thus far suggests a potential role for these new cephalosporin/ β -lactamase inhibitor antibiotics against ESBL-producing organisms, clinical data remain limited. Additionally, the significant expense of utilizing these new cephalosporin/ β -lactamase inhibitor agents is a limiting factor when alternative, less costly options are available.

CONCLUSIONS

Utilizing noncarbapenem β -lactams for the treatment of ESBL-producing organisms is an effective strategy to reduce carbapenem utilization and the associated downstream effects of carbapenem overuse. Available data suggest that cephamycins, cefepime, and β L- β LI are potential alternatives for frequently encountered ESBL clinical scenarios such as patients with mild to moderate “low-inoculum” infections, with the most robust data available for β L- β LI. On the other hand, carbapenems are still preferred, at least initially, for critically ill patients, infections with a high bacterial load, or elevated β -lactam MICs until more definitive data become available.

Notes

Financial support. This work was supported by the National Institutes of Health (award number 1K23AI127935) and an inhealth Pilot Project Discovery Program, both awarded to P. D. T.

Potential conflicts of interest. Both authors: No potential conflicts. Both authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References

1. Paterson DL, Hujer KM, Hujer AM, et al; International Klebsiella Study Group. Extended-spectrum beta-lactamases in *Klebsiella pneumoniae* bloodstream isolates from seven countries: dominance and widespread prevalence of SHV- and CTX-M-type beta-lactamases. *Antimicrob Agents Chemother* **2003**; 47:3554–60.
2. Villegas MV, Pallares CJ, Escandón-Vargas K, et al. Characterization and clinical impact of bloodstream infection caused by carbapenemase-producing Enterobacteriaceae in seven Latin American countries. *PLoS One* **2016**; 11:e0154092.
3. Jean SS, Coombs G, Ling T, et al. Epidemiology and antimicrobial susceptibility profiles of pathogens causing urinary tract infections in the Asia-Pacific region: results from the study for monitoring antimicrobial resistance trends (SMART), 2010–2013. *Int J Antimicrob Agents* **2016**; 47:328–34.
4. Lob SH, Kazmierczak KM, Badal RE, et al. Trends in susceptibility of *Escherichia coli* from intra-abdominal infections to ertapenem and comparators in the United States according to data from the SMART program, 2009 to 2013. *Antimicrob Agents Chemother* **2015**; 59:3606–10.
5. Bush K. Proliferation and significance of clinically relevant β -lactamases. *Ann N Y Acad Sci* **2013**; 1277:84–90.
6. Vardakas KZ, Tansarli GS, Rafailidis PI, Falagas ME. Carbapenems versus alternative antibiotics for the treatment of bacteraemia due to Enterobacteriaceae producing extended-spectrum β -lactamases: a systematic review and meta-analysis. *J Antimicrob Chemother* **2012**; 67:2793–803.
7. Paterson DL, Ko WC, Von Gottberg A, et al. Antibiotic therapy for *Klebsiella pneumoniae* bacteremia: implications of production of extended-spectrum beta-lactamases. *Clin Infect Dis* **2004**; 39:31–7.
8. Ofer-Friedman H, Shefler C, Sharma S, et al. Carbapenems versus piperacillin-tazobactam for bloodstream infections of nonurinary source caused by extended-spectrum beta-lactamase-producing Enterobacteriaceae. *Infect Control Hosp Epidemiol* **2015**; 36:981–5.
9. Paterson DL, Ko WC, Von Gottberg A, et al. Outcome of cephalosporin treatment for serious infections due to apparently susceptible organisms producing extended-spectrum beta-lactamases: implications for the clinical microbiology laboratory. *J Clin Microbiol* **2001**; 39:2206–12.
10. Matsumura Y, Yamamoto M, Nagao M, Tanaka M, Takakura S, Ichijima S. In vitro activities and detection performances of cefmetazole and flomoxef for extended-spectrum β -lactamase and plasmid-mediated AmpC β -lactamase-producing Enterobacteriaceae. *Diagn Microbiol Infect Dis* **2016**; 84:322–7.
11. Pangon B, Bizet C, Buré A, et al. In vivo selection of a cephamycin-resistant, porin-deficient mutant of *Klebsiella pneumoniae* producing a TEM-3 beta-lactamase. *J Infect Dis* **1989**; 159:1005–6.
12. Bradford PA, Urban C, Mariano N, Projan SJ, Rahal JJ, Bush K. Imipenem resistance in *Klebsiella pneumoniae* is associated with the combination of ACT-1, a plasmid-mediated AmpC beta-lactamase, and the fss of an outer membrane protein. *Antimicrob Agents Chemother* **1997**; 41:563–9.
13. Lee CH, Chia JH, Chu C, Wu TL, Liu JW, Su LH. In vivo selection of OmpK35-deficient mutant after cefuroxime therapy for primary liver abscess caused by *Klebsiella pneumoniae*. *J Antimicrob Chemother* **2006**; 58:857–60.
14. Lee CH, Chu C, Liu JW, Chen YS, Chiu CJ, Su LH. Collateral damage of flomoxef therapy: in vivo development of porin deficiency and acquisition of blaDHA-1 leading to ertapenem resistance in a clinical isolate of *Klebsiella pneumoniae* producing CTX-M-3 and SHV-5 beta-lactamases. *J Antimicrob Chemother* **2007**; 60:410–3.
15. Lee CH, Su LH, Tang YF, Liu JW. Treatment of ESBL-producing *Klebsiella pneumoniae* bacteraemia with carbapenems or flomoxef: a retrospective study and laboratory analysis of the isolates. *J Antimicrob Chemother* **2006**; 58:1074–7.
16. Kernéis S, Valade S, Geri G, et al. Cefoxitin as a carbapenem-sparing antibiotic for infections caused by extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae*. *Infect Dis (Lond)* **2015**; 47:789–95.
17. Doi A, Shimada T, Harada S, Iwata K, Kamiya T. The efficacy of cefmetazole against pyelonephritis caused by extended-spectrum beta-lactamase-producing Enterobacteriaceae. *Int J Infect Dis* **2013**; 17:e159–63.
18. Yang CC, Li SH, Chuang FR, et al. Discrepancy between effects of carbapenems and flomoxef in treating nosocomial hemodialysis access-related bacteremia secondary to extended spectrum beta-lactamase producing *Klebsiella pneumoniae* in patients on maintenance hemodialysis. *BMC Infect Dis* **2012**; 12:206.
19. Pilmis B, Parize P, Zahar JR, Lortholary O. Alternatives to carbapenems for infections caused by ESBL-producing Enterobacteriaceae. *Eur J Clin Microbiol Infect Dis* **2014**; 33:1263–5.
20. Matsumura Y, Yamamoto M, Nagao M, et al. Multicenter retrospective study of cefmetazole and flomoxef for treatment of extended-spectrum- β -lactamase-producing *Escherichia coli* bacteremia. *Antimicrob Agents Chemother* **2015**; 59:5107–13.
21. Guet-Revillet H, Emirian A, Groh M, et al. Pharmacological study of cefoxitin as an alternative antibiotic therapy to carbapenems in treatment of urinary tract infections due to extended-spectrum- β -lactamase-producing *Escherichia coli*. *Antimicrob Agents Chemother* **2014**; 58:4899–901.
22. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. 26th ed. CLSI supplement M100S. Wayne, PA: CLSI, **2016**.
23. European Union Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 1.1. Available at: http://www.eucast.org/clinical_breakpoints/. Accessed February 15th, 2017.
24. Kohner PC, Robberts FJ, Cockerill FR 3rd, Patel R. Cephalosporin MIC distribution of extended-spectrum- β -lactamase- and pAmpC-producing *Escherichia coli* and *Klebsiella* species. *J Clin Microbiol* **2009**; 47:2419–25.
25. Jacoby GA, Carreras I. Activities of beta-lactam antibiotics against *Escherichia coli* strains producing extended-spectrum beta-lactamases. *Antimicrob Agents Chemother* **1990**; 34:858–62.
26. Thomson KS, Moland ES. Cefepime, piperacillin-tazobactam, and the inoculum effect in tests with extended-spectrum beta-lactamase-producing Enterobacteriaceae. *Antimicrob Agents Chemother* **2001**; 45:3548–54.
27. Bedenić B, Beader N, Zagar Z. Effect of inoculum size on the antibacterial activity of cefpirome and cefepime against *Klebsiella pneumoniae* strains producing SHV extended-spectrum beta-lactamases. *Clin Microbiol Infect* **2001**; 7:626–35.
28. Szabó D, Máthé A, Filetőth Z, Anderlik P, Rókusz L, Rozgonyi F. In vitro and in vivo activities of amikacin, cefepime, amikacin plus cefepime, and imipenem against an SHV-5 extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* strain. *Antimicrob Agents Chemother* **2001**; 45:1287–91.
29. Rice LB, Yao JD, Klimm K, Eliopoulos GM, Moellering RC Jr. Efficacy of different beta-lactams against an extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* strain in the rat intra-abdominal abscess model. *Antimicrob Agents Chemother* **1991**; 35:1243–4.
30. Thauvin-Eliopoulos C, Tripodi MF, Moellering RC Jr, Eliopoulos GM. Efficacies of piperacillin-tazobactam and cefepime in rats with experimental intra-abdominal abscesses due to an extended-spectrum beta-lactamase-producing strain of *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* **1997**; 41:1053–7.
31. Jett BD, Ritchie DJ, Reichley R, Bailey TC, Sahn DF. In vitro activities of various beta-lactam antimicrobial agents against clinical isolates of *Escherichia coli* and *Klebsiella* spp. resistant to oxyimino cephalosporins. *Antimicrob Agents Chemother* **1995**; 39:1187–90.
32. Burgess DS, Hall RG 2nd. In vitro killing of parenteral beta-lactams against standard and high inocula of extended-spectrum beta-lactamase and non-ESBL producing *Klebsiella pneumoniae*. *Diagn Microbiol Infect Dis* **2004**; 49:41–6.
33. Maglio D, Ong C, Banevicius MA, Geng Q, Nightingale CH, Nicolau DP. Determination of the in vivo pharmacodynamic profile of cefepime against extended-spectrum-beta-lactamase-producing *Escherichia coli* at various inocula. *Antimicrob Agents Chemother* **2004**; 48:1941–7.
34. Craig WA. Interrelationship between pharmacokinetics and pharmacodynamics in determining dosage regimens for broad-spectrum cephalosporins. *Diagn Microbiol Infect Dis* **1995**; 22:89–96.

35. Reese AM, Frei CR, Burgess DS. Pharmacodynamics of intermittent and continuous infusion piperacillin/tazobactam and cefepime against extended-spectrum beta-lactamase-producing organisms. *Int J Antimicrob Agents* **2005**; 26:114–9.
36. Roos JF, Bulitta J, Lipman J, Kirkpatrick CM. Pharmacokinetic-pharmacodynamic rationale for cefepime dosing regimens in intensive care units. *J Antimicrob Chemother* **2006**; 58:987–93.
37. Andes D, Craig A. Animal model pharmacokinetics and pharmacodynamics In: Interscience Conference on Antimicrobial Agents and Chemotherapy, Chicago, IL **2001**. Abstract A-1099.
38. Wang R, Cosgrove SE, Tschudin-Sutter S, et al. Cefepime therapy for cefepime-susceptible extended-spectrum β -lactamase-producing Enterobacteriaceae bacteremia. *Open Forum Infect Dis* **2016**; 3:ofw132.
39. Goethaert K, Van Looveren M, Lammens C, et al. High-dose cefepime as an alternative treatment for infections caused by TEM-24 ESBL-producing *Enterobacter aerogenes* in severely-ill patients. *Clin Microbiol Infect* **2006**; 12:56–62.
40. Chopra T, Marchaim D, Veltman J, et al. Impact of cefepime therapy on mortality among patients with bloodstream infections caused by extended-spectrum β -lactamase-producing *Klebsiella pneumoniae* and *Escherichia coli*. *Antimicrob Agents Chemother* **2012**; 56:3936–42.
41. Zanetti G, Bally F, Greub G, et al; Cefepime Study Group. Cefepime versus imipenem-cilastatin for treatment of nosocomial pneumonia in intensive care unit patients: a multicenter, evaluator-blind, prospective, randomized study. *Antimicrob Agents Chemother* **2003**; 47:3442–7.
42. Lee NY, Lee CC, Huang WH, Tsui KC, Hsueh PR, Ko WC. Cefepime therapy for monomicrobial bacteremia caused by cefepime-susceptible extended-spectrum beta-lactamase-producing Enterobacteriaceae: MIC matters. *Clin Infect Dis* **2013**; 56:488–95.
43. Marchaim D, Sunkara B, Lephart PR, et al. Extended-spectrum β -lactamase producers reported as susceptible to piperacillin-tazobactam, cefepime, and cefuroxime in the era of lowered breakpoints and no confirmatory tests. *Infect Control Hosp Epidemiol* **2012**; 33:853–5.
44. Rice LB, Carias LL, Shlaes DM. In vivo efficacies of beta-lactam-beta-lactamase inhibitor combinations against a TEM-26-producing strain of *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* **1994**; 38:2663–4.
45. López-Cerero L, Picón E, Morillo C, et al. Comparative assessment of inoculum effects on the antimicrobial activity of amoxicillin-clavulanate and piperacillin-tazobactam with extended-spectrum beta-lactamase-producing and extended-spectrum beta-lactamase-non-producing *Escherichia coli* isolates. *Clin Microbiol Infect* **2010**; 16:132–6.
46. Zimhony O, Chmelnitsky I, Bardenstein R, et al. Endocarditis caused by extended-spectrum-beta-lactamase-producing *Klebsiella pneumoniae*: emergence of resistance to ciprofloxacin and piperacillin-tazobactam during treatment despite initial susceptibility. *Antimicrob Agents Chemother* **2006**; 50:3179–82.
47. Rodríguez-Baño J, Navarro MD, Retamar P, Picón E, Pascual Á; Extended-Spectrum Beta-Lactamases—Red Española de Investigación en Patología Infecciosa/Grupo de Estudio de Infección Hospitalaria. β -Lactam/ β -lactam inhibitor combinations for the treatment of bacteremia due to extended-spectrum β -lactamase-producing *Escherichia coli*: a post hoc analysis of prospective cohorts. *Clin Infect Dis* **2012**; 54:167–74.
48. Gavin PJ, Suseno MT, Thomson RB Jr, et al. Clinical correlation of the CLSI susceptibility breakpoint for piperacillin-tazobactam against extended-spectrum-beta-lactamase-producing *Escherichia coli* and *Klebsiella* species. *Antimicrob Agents Chemother* **2006**; 50:2244–7.
49. Kang CI, Park SY, Chung DR, Peck KR, Song JH. Piperacillin-tazobactam as an initial empirical therapy of bacteremia caused by extended-spectrum β -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae*. *J Infect* **2012**; 64:533–4.
50. Harris PN, Yin M, Jureen R, et al. Comparable outcomes for β -lactam/ β -lactamase inhibitor combinations and carbapenems in definitive treatment of bloodstream infections caused by cefotaxime-resistant *Escherichia coli* or *Klebsiella pneumoniae*. *Antimicrob Resist Infect Control* **2015**; 4:14.
51. Tamma PD, Han JH, Rock C, et al; Antibacterial Resistance Leadership Group. Carbapenem therapy is associated with improved survival compared with piperacillin-tazobactam for patients with extended-spectrum β -lactamase bacteremia. *Clin Infect Dis* **2015**; 60:1319–25.
52. Gutiérrez-Gutiérrez B, Pérez-Galera S, Salamanca E, et al. A multinational, preregistered cohort study of β -lactam/ β -lactamase inhibitor combinations for treatment of bloodstream infections due to extended-spectrum- β -lactamase-producing Enterobacteriaceae. *Antimicrob Agents Chemother* **2016**; 60:4159–69.
53. Ng TM, Khong WX, Harris PN, et al. Empiric piperacillin-tazobactam versus carbapenems in the treatment of bacteraemia due to extended-spectrum beta-lactamase-producing Enterobacteriaceae. *PLoS One* **2016**; 11:e0153696.
54. Harris PN, Peleg AY, Iredell J, et al. Meropenem versus piperacillin-tazobactam for definitive treatment of bloodstream infections due to ceftriaxone non-susceptible *Escherichia coli* and *Klebsiella* spp (the MERINO trial): study protocol for a randomised controlled trial. *Trials* **2015**; 16:24.
55. Kim A, Sutherland CA, Kuti JL, Nicolau DP. Optimal dosing of piperacillin-tazobactam for the treatment of *Pseudomonas aeruginosa* infections: prolonged or continuous infusion? *Pharmacotherapy* **2007**; 27:1490–7.
56. Farrell DJ, Flamm RK, Sader HS, Jones RN. Antimicrobial activity of ceftolozane-tazobactam tested against Enterobacteriaceae and *Pseudomonas aeruginosa* with various resistance patterns isolated in U.S. hospitals (2011–2012). *Antimicrob Agents Chemother* **2013**; 57:6305–10.
57. Sader HS, Farrell DJ, Flamm RK, Jones RN. Ceftolozane/tazobactam activity tested against aerobic gram-negative organisms isolated from intra-abdominal and urinary tract infections in European and United States hospitals (2012). *J Infect* **2014**; 69:266–77.
58. Lucasti C, Hershberger E, Miller B, et al. Multicenter, double-blind, randomized, phase II trial to assess the safety and efficacy of ceftolozane-tazobactam plus metronidazole compared with meropenem in adult patients with complicated intra-abdominal infections. *Antimicrob Agents Chemother* **2014**; 58:5350–7.
59. Solomkin J, Hershberger E, Miller B, et al. Ceftolozane/tazobactam plus metronidazole for complicated intra-abdominal infections in an era of multidrug resistance: results from a randomized, double-blind, phase 3 trial (ASPECT-cIAI). *Clin Infect Dis* **2015**; 60:1462–71.
60. Sader HS, Castanheira M, Flamm RK, Farrell DJ, Jones RN. Antimicrobial activity of ceftazidime-avibactam against gram-negative organisms collected from U.S. medical centers in 2012. *Antimicrob Agents Chemother* **2014**; 58:1684–92.
61. Lucasti C, Popescu I, Ramesh MK, Lipka J, Sable C. Comparative study of the efficacy and safety of ceftazidime/avibactam plus metronidazole versus meropenem in the treatment of complicated intra-abdominal infections in hospitalized adults: results of a randomized, double-blind, phase II trial. *J Antimicrob Chemother* **2013**; 68:1183–92.
62. Mazuski JE, Gasink LB, Armstrong J, et al. Efficacy and safety of ceftazidime-avibactam plus metronidazole versus meropenem in the treatment of complicated intra-abdominal infection: results from a randomized, controlled, double-blind, phase 3 program. *Clin Infect Dis* **2016**; 62:1380–9.
63. Wagenlehner FM, Sobel JD, Newell P, et al. Ceftazidime-avibactam versus doripenem for the treatment of complicated urinary tract infections, including acute pyelonephritis: RECAPTURE, a phase 3 randomized trial program. *Clin Infect Dis* **2016**; 63:754–62.