

Patterning artificial lipid bilayer on nanostructured surfaces

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Abstract: Artificial lipid bilayer on solid substrate plays an important role as an interface between nanotechnology and biology. In this study, grid structures were patterned on Au-Nb-glass substrate and artificial bilayer was prepared on these structures. The fluidity was checked using fluorescence recovery after photobleaching (FRAP), and neuronal adhesion was monitored on such structure using EphrinA5-tethered lipid bilayer. EphrinA5 is a ligand that binds to the Eph receptors of rat cortical neurons and influences cellular adhesion. Our result elucidated that influence of these nanopatterned protein-tethered lipid bilayer on cellular guidance and signaling can address many underlying mechanisms of cellular functioning and help us to understand and differentiate the signaling procedure in cancer and neurodegenerative diseases.

Keywords: Au-Nb, FRAP, EphrinA5, lipid bilayer, neurodegenerative diseases

Introduction

Artificial lipid bilayer on solid substrate plays an important role as an interface between nanotechnology and biology. This biomimetic membrane serves as an excellent model system to study cellular signaling and guidance. For signaling, cells confine the lipids and membrane proteins into specific domain, known as membrane domain.¹ The lateral diffusion of the membrane, due to its fluidic nature, is responsible for such spatial organization and compartmentalization of the signaling molecules. Artificially prepared lipid bilayer also retains this property.² By using barriers or constraints on the surfaces, this fluidity can be restricted, and both lipid and lipid-tethered proteins can be compartmentalized within these domains.³ In this study, grid structures were patterned on Au-Nb-glass substrate and artificial bilayer was prepared on these structures. The fluidity was checked using fluorescence recovery after photobleaching (FRAP), and neuronal adhesion was monitored on such structure using EphrinA5-tethered lipid bilayer.⁴ EphrinA5 is a ligand that binds to the Eph receptors of rat cortical neurons and influences cellular adhesion.⁵

Materials and methods

Fabrication of structures and characterization using scanning electron microscopy (SEM): 1–2 nm of Nb was deposited on glass surface prior to the coating of the photoresist, poly(methyl methacrylate) (PMMA). By using e-beam writer, the desired structures were transposed on the PMMA surface. PMMA was developed, and successively Au was deposited. The undesired Au was removed using the liftoff technique. Finally, the Nb was removed by reactive ion etching. SEM (FEI, Hillsboro, OR, USA) was performed to characterize the structures.

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Bilayer preparation and FRAP

1-Palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC) was dried under vacuum on the wall of a glass vessel, 0.1 M phosphate-buffered saline (PBS) or EphrinA5 was dissolved in detergent, and 0.1 M PBS was added to this dried lipid, vortexed, and sonicated to prepare liposomes or proteoliposomes, finally extruded through 100 nm membrane by Avanti mini extruder (Avanti Polar Lipids, Inc) to get small unilamellar vesicles (SUVs). These SUVs were fused on the cleaned and plasma activated patterned surface to get lipid bilayer. For FRAP 5% Oregon Green 1,2-dihexadecanoyl-*sn*-glycero-3-phosphoethanolamine (Oregon Green 488 DHPE) doped POPC-SUVs were deposited on the patterned surface. Bleaching was done by using a 473-nm laser from Rapp OptoElectronics coupled into a Zeiss Axio Scope, and a series of images were made to track the recovery of fluorescence.

Neuronal adhesion on EphrinA5-tethered lipid bilayer

About 40,000/mL neurons were cultured on EphrinA5 incorporated lipid bilayer. Bright field images were captured

just after 4 hours of cell adhesion with and without EphrinA5-tethered lipid bilayer.

Results and discussion

Figure 1 shows a schematic representation of the Au grid pattern on the glass surface and SEM images of the pattern. Grid structures made of Au (wall of 10 nm in height and 0.2 μm in width) in 5 different ranges (2.5, 2, 1.5, 1, and 0.5 μm) were fabricated on a glass substrate with a spacing of 10 μm between them. All these grids were fabricated in arrays. The SEM images show different regions of the patterns after the fabrication step. The white lines in the images show the Au walls of the structures. Five percent Oregon Green doped POPC-SUVs were deposited on the patterned surface for bilayer preparation and fluorescence imaging for FRAP experiments. Bleaching was done using a laser and recovery was monitored by capturing the images of the same area at different time intervals. FRAP was done for each region of the grid structures, and it was found that the lipids are mobile within the grids (Figure 2). This could be accounted for two reasons. First, there could be nano gaps within the grids through which lipids can move from

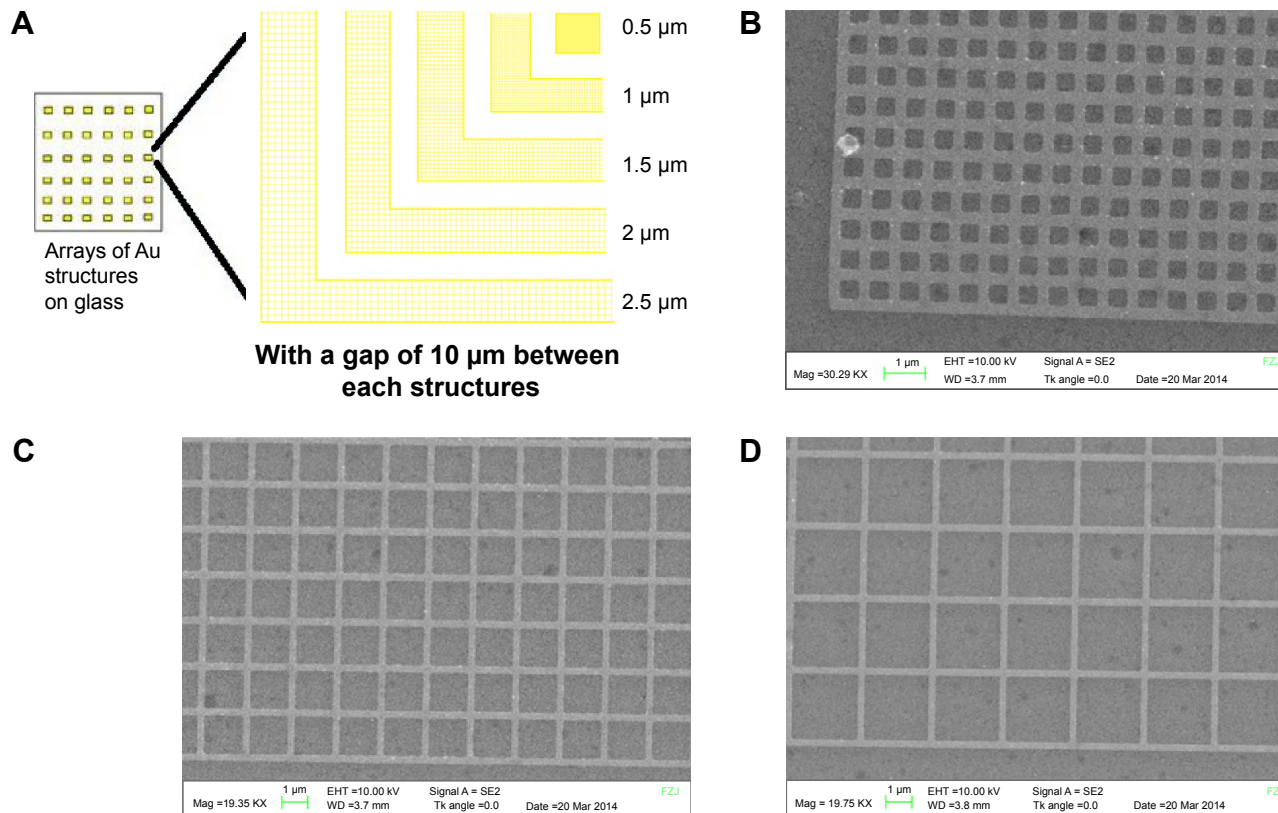


Figure 1 Schematics of the grid pattern (A) and the scanning electron microscope images of the patterns with 0.5, 1.5, and 2.5 μm grid (B–D, respectively).

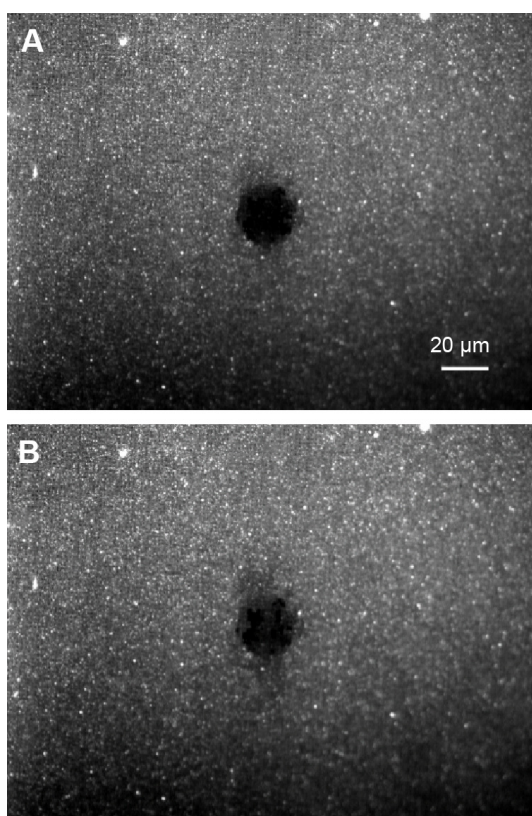


Figure 2 Fluorescence recovery after photobleaching of Oregon Green 1,2-dihexadecanoyl-*sn*-glycero-3-phosphoethanolamine doped lipid bilayer in the length/breadth of 1.5 μ m grid.

Notes: Images captured just after bleaching (A) and 421 seconds after bleaching (B).

one grid to another because there is always a possibility that during liftoff or cleaning process, chunks of Au are removed, which affect the homogeneity of the structure and create the nano gaps in the grid wall. However, 15-nm height could be too low for the lipids, and due to their lateral mobility, they could move from one compartment to another.

To check the distribution and diffusion in this guided geometry, a cell signaling protein EphrinA5 was incorporated into the model membrane. Rat cortical neurons were cultured on these EphrinA5-tethered lipid bilayer prepared on the patterned surface. Neuronal adhesion was monitored just after 4 hours of neuronal plating. Neuronal aggregation is observed on such patterned EphrinA5-tethered lipid bilayer (Figure 3).

Conclusion

In this report, a prepatterned glass substrate with Au structures in the range of nanometers is used for artificial lipid bilayer preparation. Repeated experiments of FRAP showed that the lipids are mobile and they can move within the structures. EphrinA5-tethered lipid bilayer can promote neuronal adhesion on

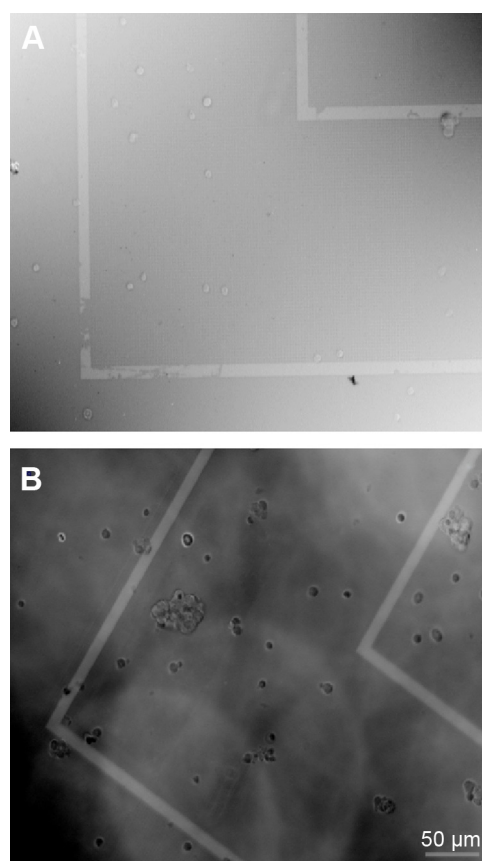


Figure 3 Neuronal culture on control substrate (A) and EphrinA5-tethered lipid bilayer (B).

these structures. The influence of these nanopatterned protein-tethered lipid bilayer on cellular guidance and signaling can address many underlying mechanisms of cellular functioning and help us to understand and differentiate the signaling procedure in cancer and neurodegenerative diseases.

Disclosure

The authors report no conflicts of interest in this work.

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