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Rescue of impaired sociability and anxiety-like behavior in adult *cacna1c*-deficient mice by pharmacologically targeting eIF2 α

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Abstract

CACNA1C, encoding the Ca_v1.2 subunit of L-type Ca²⁺ channels, has emerged as one of the most prominent and highly replicable susceptibility genes for several neuropsychiatric disorders. Ca_v1.2 channels play a crucial role in calcium-mediated processes involved in brain development and neuronal function. Within the *CACNA1C* gene, disease-associated single-nucleotide polymorphisms have been associated with impaired social and cognitive processing and altered prefrontal cortical (PFC) structure and activity. These findings suggest that aberrant Ca_v1.2 signaling may contribute to neuropsychiatric-related disease symptoms via impaired PFC function. Here, we show that mice harboring loss of *cacna1c* in excitatory glutamatergic neurons of the forebrain (fbKO) that we have previously reported to exhibit anxiety-like behavior, displayed a social behavioral deficit and impaired learning and memory. Furthermore, focal knockdown of *cacna1c* in the adult PFC recapitulated the social deficit and elevated anxiety-like behavior, but not the deficits in learning and memory. Electrophysiological and molecular studies in the PFC of *cacna1c* fbKO mice revealed higher E/I ratio in layer 5 pyramidal neurons and lower general protein synthesis. This was concurrent with reduced activity of mTORC1 and its downstream mRNA translation initiation factors eIF4B and 4EBP1, as well as elevated phosphorylation of eIF2 α , an inhibitor of mRNA translation. Remarkably, systemic treatment with ISRIB, a small molecule inhibitor that suppresses the effects of phosphorylated eIF2 α on mRNA translation, was sufficient to reverse the social deficit and elevated anxiety-like behavior in adult *cacna1c* fbKO mice. ISRIB additionally normalized the lower protein synthesis and higher E/I ratio in the PFC. Thus this study identifies a novel Ca_v1.2 mechanism in neuropsychiatric-related endophenotypes and a potential future therapeutic target to explore.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

ZDK and AMR contributed to the experimental design, data interpretation and wrote the paper. AC and NVDMG performed electrophysiological experiments and analyzed data. ZDK performed stereotaxic surgeries and analyzed data. ZDK and DKF conducted molecular and behavioral experiments. BR conducted molecular experiments. RCR and MB conducted behavioral experiments. MJG conducted electron microscopy experiments. All authors discussed and commented on the manuscript.

Supplementary Information accompanies the paper on the Molecular Psychiatry website (<http://www.nature.com/mp>)

INTRODUCTION

The *CACNA1C* gene, encoding the Ca_v1.2 subunit of L-type calcium channels (LTCCs), has emerged as a candidate susceptibility gene for multiple neuropsychiatric disorders¹⁻³ including bipolar disorder (BD),⁴ schizophrenia (SCZ),⁵ major depressive disorder (MDD),⁵ and autism spectrum disorder (ASD).⁶ *CACNA1C* is additionally present in biological pathways that are shared between SCZ, BD, and MDD.⁷ The Ca_v1.2 LTCCs are a key source of Ca²⁺ entry into cells and activate calcium-mediated signaling pathways⁸ that are critical for normal neuronal development, dendritic growth, and neuronal plasticity.⁸⁻¹⁰ The critical role of Ca_v1.2 channels in normal brain function is further underscored by human imaging and clinical studies that have reported structural and functional brain alterations and behavioral abnormalities in human carriers of *CACNA1C* disease-associated single-nucleotide polymorphisms (SNPs).¹¹

Neuropsychiatric disorders comprise a plethora of complex behavioral endophenotypes, many of which are shared across multiple disorders.^{11,12} While alterations in mood and emotion, including anxiety, form the core features of neuropsychiatric disorders, impairments in social and cognitive processing are prominent co-morbid symptoms that remain understudied and show limited improvement with current treatments.¹³ In humans, disease-associated SNPs in *CACNA1C* have been associated not only with heightened anxiety but also cognitive dysfunction and impaired facial emotional recognition,¹¹ a feature required for normal social behavior, suggesting a potential role for Ca_v1.2 LTCCs in mediating these functions.

The prefrontal cortex (PFC) is the central anatomical structure mediating higher-order social and cognitive function by having a top-down influence on several sub-cortical regions.^{14,15} Because of its wide range of outputs, the PFC can have different modulatory effects on distinct cognitive domains. In *CACNA1C* SNP carriers, neuroimaging studies have identified alterations in PFC structure¹⁶ and changes in cognitive processing-dependent PFC activity,¹¹ suggesting altered prefrontal cortical functioning. However despite the significant link between *CACNA1C* and neuropsychiatric disease, the underlying Ca_v1.2-mediated molecular mechanisms contributing to neuropsychiatric-related behaviors remain unknown.

One emerging hypothesis for the pathophysiological mechanisms mediating the cognitive impairments in diseases like SCZ and ASD is altered excitation to inhibition (E/I) balance, with higher E/I ratio reported in multiple mouse models with altered social and cognitive function.¹⁷⁻²⁰ Furthermore, higher E/I in the PFC via optogenetic manipulation was recently demonstrated to directly impair both social behavior and fear responses.²¹ Separately, impairments in cognitive functioning have also been reported in neuropsychiatric disease-associated mouse models that have decreased expression of factors involved in mRNA translation.²²⁻²⁵

While the precise mechanism by which the identified SNPs in *CACNA1C* confer risk for neuropsychiatric disorders is unknown, lower *CACNA1C* transcript levels have been reported in the brains of *CACNA1C* SNP-carrying SCZ²⁶ and BD patients,²⁷ suggesting that

decreased $Ca_v1.2$ signaling has the potential to impact brain function that can contribute to the disease symptoms observed in *CACNA1C*-associated disorders. Using a combination of genetic, behavioral, molecular and electrophysiological strategies, here we show that forebrain-specific *cacna1c* conditional knockout mice have lower levels of protein synthesis in the PFC, in parallel with lower levels of phospho-mTOR and higher levels of phospho-eIF2 α , a mRNA translation initiation repressor. Pharmacologically suppressing the downstream effects of elevated phospho-eIF2 α by the small molecule ISRIB, is sufficient to normalize the lower protein synthesis and higher E/I ratio in the PFC. Additionally, systemic treatment with ISRIB is able to rescue the deficit in sociability and elevated anxiety-like behavior. Together, our findings highlight a novel $Ca_v1.2$ -mediated mechanism via increased eIF2 α phosphorylation in the PFC that results in aberrant synaptic transmission and protein synthesis that may underlie neuropsychiatric-associated behavioral deficits.

MATERIALS AND METHODS

Detailed methods are provided in Supplementary Information.

Animals

In this study forebrain-specific *cacna1c* conditional knockout (fbKO) mice,²⁸ their wild-type (WT) littermates, and homozygous *cacna1c* floxed (*cacna1c*^{fl/fl}; Lee *et al.*²⁸) mice, all on a C57BL/6J background were utilized. All procedures were conducted in accordance with the Weill Cornell Medicine Institutional Animal Care and Use Committee rules.

Stereotaxic surgery

For focal knockdown of *cacna1c*, AAV2/2-Cre-GFP was stereotaxically delivered directly into the PFC of adult *cacna1c*^{fl/fl} mice.²⁸ Placement of surgical injections was confirmed using GFP immunohistochemistry.²⁸

Behavioral testing

All behavioral tests were performed in adult mice.

Three-chambered social approach—Social approach was performed using the three-chamber social interaction test.²⁹ 'Sociability' was measured as the amount of time the experimental animal spent with a stranger mouse compared to a novel object. Data is reported as time spent in the chambers as well as contact zones (a designated 1.5" area encircling the cup) containing the stranger mouse or novel object.

Fear conditioning—Context- and cue-associated fear conditioning³⁰ was performed, with some modifications.

Water-based Y-maze—The Y-maze was used to measure working memory as well as short- (1 h) and long-term (24 h and 7 days) memory retention. Working memory was measured over a 6 min period as percent of spontaneous alternations (defined as consecutive entry in the three different arms over total arm entries).³¹ For the memory tests, mice were trained over five trials to locate a submerged platform in one arm of the Y-shaped maze and

tested 1 h, 24 h and 7 days later for memory retention. Each test consisted of 5 trials (and reported as an average) with the latency and successes (calculated based on the errors made during each trial) to locate the submerged platform measured.

Morris water maze—Morris water maze (MWM) was performed as previously published³² with slight modifications.

Elevated plus maze—Anxiety was measured using elevated plus maze (EPM).²⁸

Electrophysiological methods

Whole-cell recordings—Postnatal day (P) 30-P45 male mice were anesthetized with isoflurane, decapitated and brains dissected. Coronal slices (400 μm) were obtained and whole-cell recordings were performed.³³

Molecular methods

All molecular experiments were performed on brain tissue from adult mice.

SUnSET—Protein synthesis was measured using a modified version of the SUnSET technique.^{22,34}

Subcellular fractionation and immunoblotting—Total protein lysates or synaptoneuroosomes³⁵ from the PFC were used for Western blot analysis. For details on antibodies used see Supplementary Table 1.

BDNF ELISA—Mature BDNF levels were measured using the ELISA protocol.³⁶

Drug treatment—A single dose of ISRIB (SML0843, Sigma, St. Louis, MO, USA) at 2.5 mg kg^{-1} (diluted in 6.25% DMSO and 6.25% PEG300) or vehicle was administered intraperitoneally 90 min prior to behavioral testing, protein analyses, or electrophysiological recordings.³⁷

Statistics

GraphPad Prism (La Jolla, CA, USA) was used for all statistical analyses. All statistical results and tests used are included in the figure legends. Sample size was chosen based on prior experience followed by power calculation. If power was greater than 0.7, additional animals were not added to the experiment. For most behavioral analyses a two-way analysis of variance was performed followed by a Bonferroni *post hoc* test if a main effect ($P < 0.05$) was observed. For independent sample sets, the D'Agostino and Pearson normality test was performed and for those that passed normality ($P > 0.05$), a two-tailed Student's *t*-test was performed with Welch's correction if variances were significantly different. For electrophysiological recordings that passed the normality test, a one-way analysis of variance was performed followed by a Bonferroni *post hoc* test if a main effect ($P < 0.05$) was observed. For sample sets that failed to pass normality ($P < 0.05$), a Kruskal–Wallis *H*-test was performed with a Dunn's uncorrected *post hoc* test if a main effect was observed.

RESULTS

***Cacna1c* forebrain knockout mice display decreased sociability and increased cue-associated fear responses**

Ca_v1.2 LTCCs are located primarily at the postsynaptic membrane³⁸ of excitatory neurons (Supplementary Figure 1) where they tightly regulate Ca²⁺-mediated gene expression and synaptic plasticity. Because of the high prevalence of these channels in excitatory neurons and their critical role in brain activity, we generated forebrain-specific *cacna1c* conditional knockout (fbKO) mice that have a loss of *cacna1c* in excitatory neurons of the forebrain. *Cacna1c* fbKO mice have elevated anxiety-like behavior,²⁸ a symptom common across all *CACNA1C*-associated neuropsychiatric disorders, and deficient postnatal hippocampal neurogenesis,³⁹ also associated with neuropsychiatric disorders⁴⁰ making this an ideal mouse model to examine mechanisms downstream of Ca_v1.2 (*cacna1c*) that underlie neuropsychiatric-related behavioral and pathophysiological phenotypes.

In most neuropsychiatric and neurodevelopmental disorders, including SCZ, BD, MDD and ASD, altered emotional regulation is often accompanied by impaired social behavior.¹² Furthermore, human and animal studies have shown that anxiety and social interaction are intimately linked with overlapping neural mechanisms.⁴¹ Thus, we evaluated the role of *cacna1c* on social behavior utilizing the three-chamber social approach test. This behavior probes for voluntary initiation of social interaction with a novel conspecific mouse (Figure 1a) and recruits components of social anxiety.^{42,43} In the social approach test *cacna1c* fbKO mice displayed reduced sociability. While WT controls spent significantly more time in the chamber and contact zone containing the con-specific stranger mouse compared to the novel object, fbKO mice spent similar amounts of time with the stranger mouse and novel object (Figures 1b and c). This abnormal social behavior was not a result of decreased locomotor activity, as WT and fbKO mice traveled comparable distances during the social task (Figure 1d).

Increased emotional reactivity to aversive stimuli is a prominent feature of many neuropsychiatric disorders and can be modeled in animals using the classical fear conditioning paradigm.⁴⁴ Furthermore, trait anxiety, referring to an individual's predisposition to anxiety,⁴¹ has been associated with increased fear responses in both humans⁴⁵ and rodents.⁴⁶ Therefore, we next examined the role of *cacna1c* on fear-related behavior. We conducted cue- and context-associated fear conditioning, wherein mice acquire a conditioned fear response (freezing) by pairing an unconditioned stimulus (foot shock) with a conditioned stimulus (tone or context). During training, WT controls and fbKO mice displayed similar levels of freezing to both the cue (tone; Figure 1e) and context (Figure 1f), demonstrating no deficit in acquisition of fear or in sensitivity to foot shock. Interestingly, twenty-four hours later fbKO mice displayed higher freezing to the cue (Figure 1e) but not the context (Figure 1f). This heightened cue-associated fear response is consistent with our previous finding of elevated anxiety in these animals.²⁸ Together, these findings demonstrated that loss of *cacna1c* in forebrain glutamatergic neurons reduces sociability and enhances cue-associated fear responses.

***Cacna1c* forebrain knockout mice exhibit selective deficits in learning and memory**

Cognitive impairments are a prominent feature of neuropsychiatric disorders with deficits in PFC-specific cognitive functions being a hallmark of neuropsychiatric patients.¹³ In addition, in humans, cognitive function can be severely impacted by anxiety.⁴⁷ Therefore, we next examined cognitive function in *cacna1c* fbKO mice utilizing the water-based Y-maze task (Figure 1g) that, in part, recruits the PFC and measures egocentric navigation (using internal cues) to locate a submerged platform. In this test, fbKO mice displayed a significant deficit in learning, with longer time (Figure 1h) and fewer successful attempts (Supplementary Figure 2a) in locating the submerged platform. Similarly, during the short- and long-term memory tests, fbKO mice exhibited a significant deficit in locating the platform (Figure 1i and Supplementary Figure 2b). These behavioral deficits were not a consequence of a generalized functional impairment in the PFC as fbKO mice showed no deficit in the Y-maze working memory-related task (Figure 1j, Supplementary Figures 2c–d), a PFC-dependent cognitive test that is often altered in SCZ patients.⁴⁸

Next, to examine hippocampal-dependent learning and memory in fbKO mice, we employed the Morris water maze task (Figure 1k) that utilizes allocentric navigation (distal spatial cues). In this test, fbKO mice displayed no deficits in locating the hidden platform during learning (Figure 1l) or the memory probe tests (Figures 1m–o). Collectively, these results demonstrated that loss of *cacna1c* in forebrain glutamatergic neurons negatively impacts selective PFC-dependent but not hippocampal-dependent learning and memory.

Decreased sociability in mice with focal knockdown of *cacna1c* in the adult PFC

As neuroimaging studies have identified altered PFC structure and activity in *CACNA1C* SNP carriers¹¹ and SCZ-associated SNPs result in decreased *CACNA1C* expression in the PFC,²⁶ we wondered if focal ablation of *cacna1c* in the PFC of adult mice may be capable of inducing the behavioral deficits observed in *cacna1c* fbKO mice. Adenoassociated viral (AAV) vector-expressing Cre recombinase or control AAV-GFP was stereotaxically delivered into the PFC of adult *cacna1c^{fl/fl}* mice (Figure 2b) and tested in the above-mentioned behaviors (Figure 2a). Interestingly, mice with a focal knockdown of *cacna1c* in the adult PFC (PFC-Cre), that have lower *cacna1c* mRNA levels (Supplementary Figure 3a), as well as Ca_v1.2 protein levels as previously revealed by immunohistochemistry,²⁸ displayed a deficit in sociability in the three-chamber social approach test similar to that observed in fbKO mice (Figure 1b). While control (PFC-GFP) mice showed a preference for the chamber and contact zone containing the stranger mouse over the novel object, the PFC-Cre mice displayed no preference (Figures 2c and d). No difference in locomotor activity was observed as PFC-Cre and PFC-GFP mice traveled similar distances during the social task (Figure 2e). In addition, in agreement with our previous report,²⁸ PFC-Cre mice displayed increased anxiety-like behavior spending significantly less time in the open arm of the elevated plus maze (EPM; Figure 2f). However, in contrast to the fbKO mice, knockdown of *cacna1c* in the adult PFC had no effect on cue fear conditioning (Figure 2g) or on learning and memory in the Y-maze (Figures 2i and j; Supplementary Figures 3b–c), suggesting a potential developmental role of Ca_v1.2 or involvement of additional brain regions for these tasks. Similar to fbKO mice, PFC-Cre mice displayed no deficits in working memory (Figure 2k; Supplementary Figures 3d–e) or contextual fear conditioning

(Figure 2h). Thus, knockdown of *cacna1c* in the adult PFC is sufficient to drive the deficit in sociability and elevate anxiety-like behavior as observed in *cacna1c* fbKO mice, but does not impact learning and memory or fear associations.

***Cacna1c* forebrain knockout mice have decreased initiation of mRNA translation in the PFC**

Alterations in protein synthesis, via dysregulated cap-dependent mRNA translation have been shown to underlie social deficits^{22,49} and anxiety.²⁴ Therefore, we measured the rate of protein synthesis in the brain of *cacna1c* fbKO mice utilizing the SUnSET technique.³⁴ Puromycin was injected into the lateral ventricle of WT and fbKO mice and levels of total puromycin-labeled proteins, indicative of active translation, were measured. We found significantly lower puromycin labeling in the PFC of fbKO mice (Figure 3a), with no differences in the somatosensory cortex (Figure 3b) or hippocampus (Figure 3c). Thus, loss of *cacna1c* in forebrain glutamatergic neurons results in lower *de novo* protein synthesis specifically in the PFC, consistent with the observed PFC- and not hippocampal-dependent behavioral deficits.

Alteration in the mTORC1 pathway-dependent translation has been associated with behavioral and synaptic deficits in multiple mouse models of neuropsychiatric disorders.⁵⁰ Furthermore, translation is tightly regulated at the level of initiation and elongation. Therefore, we quantified levels of several mRNA translation factors within the mTORC1 pathway (Figure 3d). In the PFC of *cacna1c* fbKO mice we found significantly lower levels of phosphorylated mTOR (P-mTOR) at serine (S) 2448, a marker of active mTORC1,⁵⁰ together with significantly decreased expression of the eukaryotic translation initiation factors, 4EBP1 and eIF4B and their active phosphorylated forms. We observed no difference in levels of the mTORC1-regulated elongation factor eEF2 (Figure 3e), or its downstream target brain-derived neurotrophic factor (BDNF; Figure 3f), an important mediator of neuropsychiatric-related behavioral endophenotypes.⁵¹ In addition, even though higher levels of total rpS6 protein were present in the PFC of fbKO mice (Figure 3e), we observed no difference in the active, phosphorylated ribosomal protein rpS6, involved in ribosome biogenesis, a step necessary for mRNA translation.

mRNA translation is also regulated by factors that are mTORC1-independent including eukaryotic initiation factor 2 α (eIF2 α ; Figure 3d), which when phosphorylated at S51 inhibits initiation of mRNA translation.²⁵ In the PFC of fbKO mice we found significantly higher levels of P-eIF2 α S51 (Figure 3e), consistent with the decreased protein synthesis in this region (Figure 3a). Collectively these data indicate that in the PFC, loss of *cacna1c* in forebrain glutamatergic neurons results in lower activity of mTORC1-dependent factors and eIF2 α , all of which are involved in initiation of cap-dependent mRNA translation. Systemic ISRIB normalizes the lower protein synthesis in the PFC. Because of the lack of a direct pharmacological activator of mTORC1 to correct the lower protein synthesis in the PFC of *cacna1c* fbKO mice, we tested whether a recently discovered small molecule ISRIB (integrated stress response inhibitor)³⁷ that potently blocks the inhibitory effects of higher P-eIF2 α S51 on mRNA translation³⁷ was sufficient to normalize the lower general protein synthesis in the PFC of fbKO mice and rescue the social deficit and anxiety-like behavior

(Figure 4a). Adult WT and fbKO mice were injected with puromycin in the lateral ventricle concurrent with an acute injection of either ISRIB (2.5 mg kg⁻¹, i.p.) or vehicle for a total of ninety minutes. While vehicle treated fbKO mice displayed lower puromycin labeling in the PFC, confirming lower protein synthesis (Figure 3a), ISRIB treatment significantly increased puromycin-labeled proteins to the level of WT controls (Figure 4b). These results demonstrated that acute treatment with ISRIB is sufficient to normalize the lower levels of general protein synthesis in the PFC of *cacna1c* fbKO mice.

Systemic ISRIB reverses the decreased sociability and increased anxiety in *cacna1c* forebrain knockout mice

To examine whether an acute treatment with ISRIB corrected the behavioral impairments, adult WT and fbKO mice were injected with either ISRIB (2.5 mg kg⁻¹, i.p.) or vehicle and ninety minutes later tested in social approach followed immediately by elevated plus maze (Figure 4c). Remarkably, ISRIB was sufficient to rescue the social deficit in *cacna1c* fbKO mice. While vehicle treated fbKO mice displayed a lack of social preference, ISRIB treated mice spent significantly more time with the stranger mouse over the novel object, looking similar to social behavior in WT animals (Figures 4d and f). In WT mice, ISRIB treatment had no effect on social preference (Figures 4d and f). Moreover, ISRIB relieved the anxiogenic phenotype displayed by *cacna1c* fbKO mice in EPM. Compared to vehicle-treated fbKO mice that spent equal amount of time in the open and closed arms, ISRIB-treated fbKO mice spent significantly greater time in the open arms compared to the closed arms, looking similar to the behavior displayed by vehicle treated WT mice (Figures 4g and i). Interestingly, in WT mice, ISRIB treatment appeared to marginally increase anxiety (Figures 4g and i). ISRIB had no effect on locomotor activity in either the social interaction (Figure 4e) or anxiety (Figure 4h) behavioral tests. These results demonstrated that an acute systemic treatment with ISRIB in adulthood was sufficient to normalize the social deficit and elevated anxiety in *cacna1c* fbKO mice.

Systemic ISRIB reverses the decreased sociability but not the increased anxiety in adult PFC-specific *cacna1c* knockdown mice To test whether suppressing the effects of phosphorylated eIF2 α with ISRIB can reverse the social deficit and anxiety-like behavior in adult PFC-specific *cacna1c* knockdown (PFC-Cre) mice, we also treated PFC-Cre mice with ISRIB and tested them in the social and anxiety behavioral tests (Figure 4c). While vehicle treated PFC-Cre mice displayed the sociability deficit, ISRIB treatment significantly increased the time spent in the chamber containing the stranger mouse over the novel object (Figures 4d and f). Surprisingly, in the EPM test, ISRIB was unable to normalize anxiety-like behavior. Both vehicle and ISRIB treated PFC-Cre mice spent more time in the closed arms of the maze (Figures 4g and i). ISRIB in PFC-Cre mice had no effect on locomotor activity during the social (Figure 4e) or anxiety behavioral tests (Figure 4h). These results suggest a convergent molecular mechanism underlying the social impairments in the developmental (fbKO) and adult (PFC-Cre) *cacna1c*-deficient mice but potentially divergent mechanisms mediating the anxiety-like behavior.

***Cacna1c* forebrain knockout mice exhibit higher excitatory-inhibitory balance in the PFC**

It has been hypothesized that excitatory/inhibitory (E/I) imbalance in the brain is one neuropathophysiological mechanism underlying the social deficits and anxiety-like behavior observed in patients with SCZ and ASD, as well as in rodent models.^{18,19} In particular, studies have identified altered excitatory neurotransmission⁵² and spine density⁵³ in layer 5 pyramidal neurons of the PFC in mouse models of ASD that display social impairments. Additionally, mouse models that have impaired protein synthesis and exhibit these behavioral deficits, have altered E/I balance in the PFC.^{22,54,55} Therefore, to further explore the cellular mechanism underlying the behavioral deficits in *cacna1c* fbKO mice, we performed whole-cell patch clamp recordings in acute brain slices from layer 5 pyramidal neurons of the PFC. In the PFC of fbKO mice there was significantly reduced amplitude of nifedipine-sensitive L-type calcium currents (Supplementary Figures 4a–b), demonstrating effective loss of function of Ca_v1.2 channels paralleling lower Ca_v1.2 protein levels in synaptoneurosomes from the PFC of fbKO mice (Supplementary Figure 4c). Examination of miniature excitatory synaptic currents (mEPSCs) revealed significantly higher amplitude and frequency of mEPSCs in fbKO mice compared to WT controls that was normalized with ISRIB (Figures 5a–c). Interestingly, fbKO mice displayed no difference in the amplitude of miniature inhibitory synaptic currents (mIPSCs; Figures 5d–e) but significantly higher frequency that was normalized to WT levels with ISRIB (Figures 5d and f). Resting membrane potentials (RMPs) were comparable between WT ($n = 7$; -69.2 ± 1.0 mV) and fbKO ($n = 9$; -69.1 ± 0.6 mV) mice. When measuring total synaptic charge transfer, reflecting the E/I ratio,²³ we found significantly higher charge transfer for mEPSCs but not mIPSCs in fbKO mice that was normalized to WT levels with ISRIB (Figures 5g–i).

We additionally examined protein levels of excitatory and inhibitory presynaptic markers, vesicular glutamate (VGLUT1) and GABA (VGAT) transporters in synaptoneurosomes isolated from the PFC and evaluated their ratio, a molecular measure previously used as a correlate of altered E/I balance.⁵⁶ In the PFC of fbKO mice, no difference in levels of VGLUT1 (Supplementary Figure 5b) but significantly lower VGAT (Supplementary Figure 5c) levels was observed. Furthermore, when comparing the VGLUT1/VGAT ratio, a molecular measure of E/I ratio, fbKO mice displayed a significantly higher ratio in the PFC (Figure 5j) consistent with the elevated E/I ratio. Remarkably, an acute treatment with ISRIB (Supplementary Figure 5a) was sufficient to normalize the higher VGLUT1/VGAT ratio in the PFC of fbKO (Figure 5j) that appeared to be a result of an increase in VGAT protein levels (Supplementary Figure 5c). These results demonstrate that loss of *cacna1c* in forebrain glutamatergic neurons results in higher E/I ratio in layer 5 pyramidal neurons that is normalized by ISRIB.

DISCUSSION

Here, we report that fbKO mice that have a loss of *cacna1c* in excitatory glutamatergic neurons of the forebrain and exhibit anxiety-like behavior,²⁸ display reduced sociability, enhanced cue-associated fear responses and demonstrate selective impairments in learning and memory. The social deficit and increased anxiety-like behavior is recapitulated in mice with a focal knockdown of *cacna1c* in the adult PFC, identifying *cacna1c* in this region as a

critical mediator of these behavioral deficits. Further exploration of the PFC of *cacna1c* fbKO mice identified higher E/I ratio, a physiological parameter that has been linked to neuropsychiatric-related phenotypes, as well as elevated VGLUT1/VGAT ratio, a molecular correlate of the higher E/I. Molecular studies in the PFC of *cacna1c* fbKO mice revealed decreased protein synthesis concurrent with downregulation of mTORC1-dependent mRNA translation initiation factors and upregulation of P-eIF2 α S51, an inhibitor of mRNA translation. Finally, acute systemic treatment in *cacna1c* fbKO mice with the small molecule ISRIB that suppresses the effect of P-eIF2 α S51 on mRNA translation was sufficient to normalize the lower protein synthesis and higher E/I balance in the PFC. Furthermore, ISRIB treatment was also able to correct the social deficit and elevated anxiety.

The precise functional contribution of *CACNA1C* SNPs to neuropsychiatric symptoms remains unknown. To date all disease-associated *CACNA1C* SNPs are intronic¹ with both an increase^{57,58} and decrease^{26,27,58,59} in *CACNA1C* expression and function reported in human samples. Irrespective, given that Ca_v1.2 channels play a crucial role in various neuronal functions, we hypothesize that higher or lower Ca²⁺ influx in neurons can alter neuronal and brain function and impact behavior in patients. This hypothesis is supported by the Ca_v1.2 Timothy syndrome (TS) mouse model that harbors a mutation that results in higher Ca²⁺ influx, displays a similar social deficit and persistent increase in cue-associated fear memory⁶⁰ as we report here in *cacna1c* fbKO mice. However, while the TS mouse displays increased freezing during the contextual fear memory test,⁶⁰ the *cacna1c* fbKO mice do not, demonstrating some differences between the two mouse models. However, it is possible that the lack of a phenotype in *cacna1c* fbKO mice is due to a ceiling effect in freezing levels, a question that will be addressed in future studies using alternative fear conditioning protocols. Our behavioral findings in *cacna1c* fbKO mice are comparable to the low extraversion, a personality trait that is characterized by reduced social interactions⁶¹ and poor learning performance⁶² reported in human *CACNA1C* SNP carriers. Interestingly, while the *cacna1c* fbKO mice displayed elevated anxiety,²⁸ the TS mouse did not,⁶⁰ suggesting that human *CACNA1C* SNPs that have been associated with increased anxiety^{63,64} could possibly result from decreased *cacna1c* levels in the PFC. Interestingly, the lack of a deficit in the working memory task in the *cacna1c* fbKO mice that, in part, recruits the PFC, suggests that *cacna1c* deficiency may not affect working memory. This is supported by a previous study that identified intact working memory in patients with the SCZ-associated *CACNA1C* SNPs.⁶⁵ However, as the *cacna1c* fbKO mice that we utilized lack *cacna1c* in excitatory, glutamatergic neurons, it is possible that this cell-type is not involved in working memory. This is supported by a recent finding of a role of the parvalbumin-positive interneurons of the PFC in working memory.⁶⁶ Together, these preclinical animal model findings support the growing idea and evidence that brain region and cell-type specific loss of *cacna1c* may contribute to varying neuropsychiatric-related endophenotypes.^{59,67}

The PFC has been identified as a key regulator of social cognition (reviewed in Bicks *et al.* 17) and an important modulator of anxiety.⁶⁸ Together with our previous findings,²⁸ here we show that *cacna1c* in the PFC regulates both sociability and anxiety. As ISRIB treatment is sufficient to correct both, the social deficit and the elevated anxiety in *cacna1c* fbKO mice, our findings suggest overlapping anatomical and molecular mechanisms underlying these

two endophenotypes. This is consistent with previous studies that have identified an overlap between anxiety and impaired social behavior.⁴¹ These include human studies that have reported increased social dysfunction with increased anxiety levels as well as animal studies that have identified increase in social interaction with anxiolytic treatments.⁴¹ Interestingly, ISRIB treatment in mice with knockdown of *cacna1c* in the adult PFC rescued only the social deficit and not the anxiety-like behavior highlighting the complexity of behavioral control at the circuit- and mechanistic level and suggests dissociability of anxiety and social behavior in certain conditions, such as that recapitulated by the adult PFC-specific *cacna1c* knockout model, that are possibly mediated by divergent neural mechanisms. It will be important in future studies to use such conditional and inducible knockout strategies to further examine *cacna1c* circuitry and mechanisms underlying complex behaviors such as anxiety and sociability. Future studies will also examine if ISRIB treatment directly in the PFC is sufficient to correct the social deficit and heightened anxiety.

The increase in E/I transmission and higher VGLUT1/VGAT ratio in the PFC of *cacna1c* fbKO mice, makes this a potential cellular perturbation mediating the behavioral impairments in these mice. This is supported by the observation that ISRIB in adult *cacna1c* fbKO mice normalizes the behavioral impairments, as well as the elevated E/I and VGLUT1/VGAT ratio. Our data in the *cacna1c* fbKO mouse model adds to the growing body of evidence that cortical E/I imbalance^{17–20} negatively impacts neuropsychiatric-related behavioral endophenotypes with higher prefrontal cortical E/I driving neuropsychiatric-related impairments in social behavior^{21,22,69} and anxiety.⁷⁰

Converging lines of evidence suggest that exaggerated mRNA translation is one mechanism underlying higher E/I and neuropsychiatric-related behavioral impairments.^{19,25,50} Our finding that lower mRNA translation can also result in higher E/I and produce similar behavioral deficits suggests that divergent downstream mRNA translation mechanisms can eventually result in comparable physiological and behavioral outcomes. We find altered activity of two primary mRNA translation pathways, mTORC1 that regulates global protein synthesis,⁵⁰ and eIF2 α that regulates both global protein synthesis and also mRNA-specific translation.²⁵ Although suppressing the effects of exaggerated phospho-eIF2 α by ISRIB is sufficient to normalize the lower levels of general protein synthesis in the PFC in *cacna1c* fbKO mice, our data does not rule out the possibility of a role of eIF2 α -specific mRNA translation of target genes (discussed below). Furthermore, ISRIB may initiate crosstalk between eIF2 α and mTORC1 downstream pathways⁷¹ and regulate similar mRNA targets, a future line of investigation.

mTORC1 has been extensively studied in neurodevelopmental and neuropsychiatric disorders⁵⁰ with most reporting an increase in mTORC1 activity (and thus increase in protein synthesis). In contrast, in *cacna1c* fbKO mice we find lower mTORC1 activity that parallels the lower protein synthesis. Interestingly lower mTORC1 has been reported in mice deficient in MeCP2,⁷² a downstream target of LTCCs⁷³ that also display social deficits and elevated anxiety⁷⁴ and have higher E/I.⁵⁵ Furthermore lower mTORC1 has been reported in patients with idiopathic autism⁷⁵ that exhibit core social behavioral impairments. Thus, lower mTORC1 activity can produce similar cellular and behavioral deficits as higher mTORC1 activity seen in several animal models of neuropsychiatric disease.

As compared to mTORC1, studies on the eIF2 α pathway are limited, although a role in synaptic plasticity and memory has been reported. Phospho-mutant eIF2 α (S51A) mice, with reduced levels of P-eIF2 α S51, require a lower threshold to elicit the late-phase of hippocampal LTP (L-LTP) and exhibit enhanced memory,⁷⁶ behavioral effects mimicked by ISRIB.³⁷ Similarly, knockout of GCN2, an upstream kinase of eIF2 α that has lower phospho-eIF2 α , results in similar L-LTP and short-term spatial memory enhancement⁷⁷ as seen in eIF2 α S51A phospho-mutant mice, supporting our findings that higher P-eIF2 α S51 can be detrimental to synaptic plasticity and behavior. Additionally, knockout of PERK, another upstream kinase of eIF2 α results in elevated metabotropic glutamate receptor (mGluR)-LTD⁷⁸ and enhanced fear memory.⁷⁹ These effects on synaptic plasticity and memory enhancement have been suggested to be a consequence of selective increase in eIF2 α -mediated translation of the transcription factor ATF4. ATF4 belongs to the CREB family of transcription factors⁸⁰ that are highly regulated by LTCCs,⁸¹ and is a repressor of CREB-mediated transcription,^{82,83} suggesting the possibility that the deficits in *cacna1c* fbKO mice may be mediated by ATF4. However, a recent study has demonstrated the necessity of ATF4 for regulation of spine number, morphology and synaptic plasticity,⁸⁴ suggesting that the effects of ATF4 may be finely tuned and also dependent on the disease model examined. Future studies will examine the contribution of the eIF2 α /ATF4 pathway to cellular and behavioral phenotypes we have identified.

Another question to be addressed in future studies is the mechanism by which eIF2 α contributes to the E/I imbalance, and how ISRIB corrects it. ATF4 has been shown to regulate EPSCs and maintain glutamatergic synapses.⁸⁴ Another potential route of regulation of E/I imbalance is via P-eIF2 α -selective translation of the oligophrenin-1 (OPHN1) mRNA, involved in regulation of cell surface levels of hippocampal glutamatergic AMPA receptors.^{85,86} Using VGLUT1 and VGAT as markers of excitatory and inhibitory synapses, we find lower levels of VGAT in the PFC of *cacna1c* fbKO mice that are restored to WT levels by ISRIB, suggesting that higher EPSPs could possibly result from decreased translation of VGAT or other markers of inhibitory transmission. A similar finding of elevated EPSP in parallel with increased P-eIF2 α has been reported and suggested to be a result of impaired synthesis of inhibitory elements.⁸⁷ Future studies will explore P-eIF2 α regulation of both excitatory and inhibitory pre- and postsynaptic proteins that contribute to the E/I imbalance observed in the PFC of *cacna1c* fbKO mice.

The mechanism by which deficient Ca_v1.2 signaling impairs eIF2 α and mTORC1 pathways remains unknown. One potential route is via activation of the PI3K/Akt pathway, implicated downstream of LTCCs,⁸⁸ that activates mTORC1 via phosphorylation and inhibition of the tuberous sclerosis complex TSC1/2,⁵⁰ a molecule linked to ASD-related symptoms. The Akt/PI3K pathway also negatively modulates eIF2 α phosphorylation via PKR-like ER kinase (PERK).⁸⁹ Thus, we hypothesize that loss of *cacna1c* resulting in deficient Ca_v1.2 signaling may inhibit Akt that subsequently (1) suppresses phosphorylation of TSC1/2 and increases its inhibitory effect on mTORC1 phosphorylation, and separately (2) activates PERK and induces eIF2 α phosphorylation.

Overall, the preclinical findings in this study provide evidence that the *cacna1c* forebrain knockout mouse model is a valuable animal model to study mechanisms downstream of

Ca_v1.2 (*cacna1c*) that contribute to neuropsychiatric pathophysiology and also highlights how *CACNA1C* can exert such pleiotropic effects on psychopathology related to multiple neuropsychiatric disorders. This study identifies a novel *in vivo* role of *cacna1c* in glutamatergic neurons, demonstrates causality between altered expression of this gene and development of neuropsychiatric-related behaviors in mice, and identifies a unique molecular mechanism in *cacna1c*-deficient mice that merits further exploration for understanding *cacna1c*-mediated behavioral endophenotypes.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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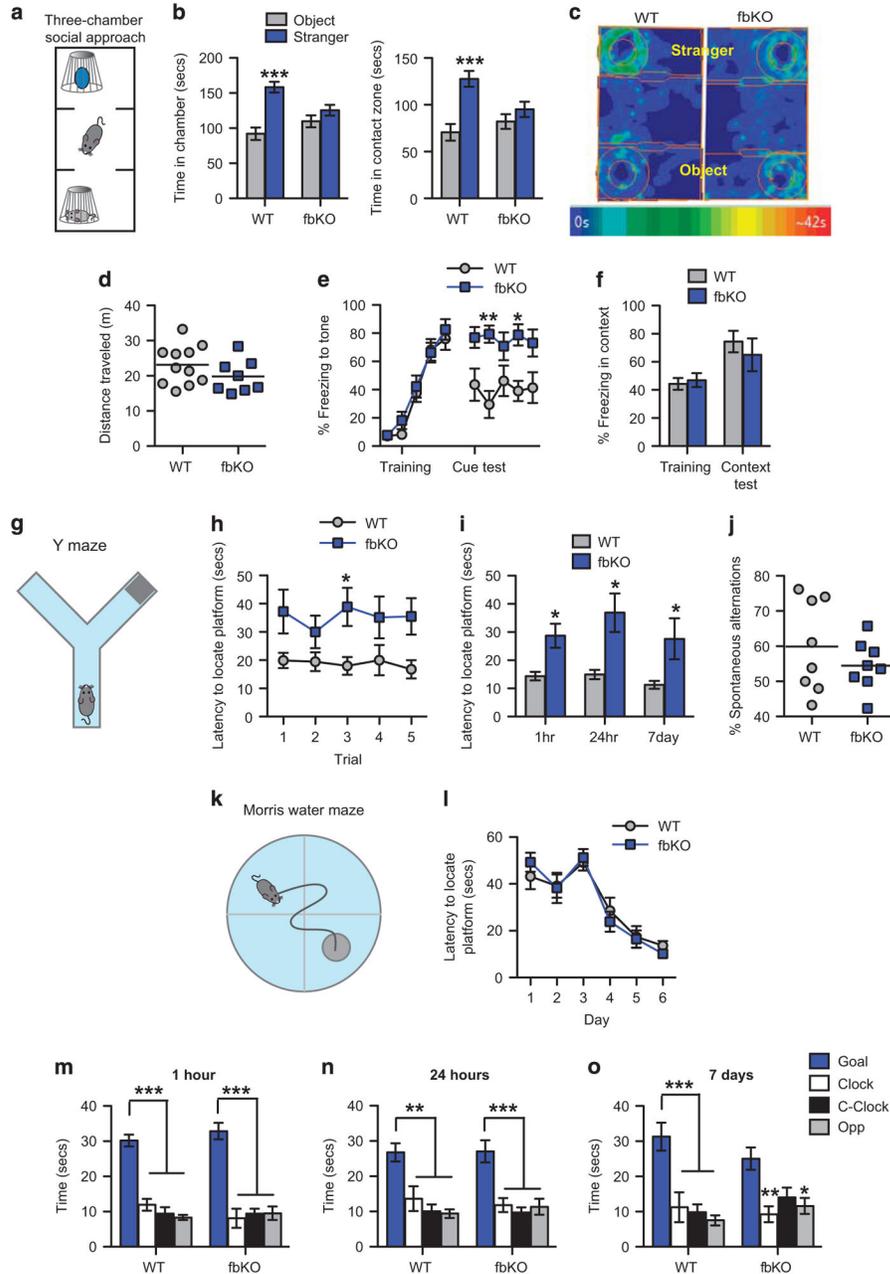


Figure 1. *Cacna1c* fbKO mice display a deficit in sociability and impaired learning and memory. (a) Schematic of the three-chamber social approach apparatus. (b) In the three-chamber social approach test, WT but not fbKO adult mice spent significantly more time in the chamber (left; two-way ANOVA, genotype \times chamber, $F_{1,34} = 8.865$, $P = 0.0053$) and contact zone (right; two-way ANOVA, genotype \times zone, $F_{1,34} = 6.6367$, $P = 0.0165$) containing the stranger mouse compared to the novel object (Bonferroni post hoc test, *** $P < 0.001$ vs object; WT $n = 11$, fbKO $n = 8$). (c) Representative heat maps showing time spent in the chambers (represented by rectangular compartments) and contact zones (represented by area

between outer circle and inner circle) containing the stranger mouse and novel object (represented by inner circle). **(d)** WT and fbKO mice traveled similar distances during the social approach test (WT $n=11$, fbKO $n=8$). **(e)** In cue fear conditioning, WT and fbKO mice displayed similar percent freezing to the tone during training (two-way ANOVA, main effect of tone, $F_{4,60} = 55.06$, $P < 0.0001$) but during the twenty-four hour cue fear test, fbKO mice exhibited significantly higher freezing compared to WT mice (Two-way ANOVA, main effect of genotype, $F_{1,60} = 34.27$, $P < 0.0001$; Bonferroni *post hoc* test, $*P < 0.05$, $**P < 0.01$ vs WT). Each data point represents freezing during a single tone presentation (WT $n=8$, fbKO $n=6$). **(f)** During contextual fear conditioning, WT and fbKO mice displayed similar percent freezing to the context during training and the twenty-four hour test (WT $n=6$, fbKO $n=6$). **(g)** Schematic of the water-based Y-maze apparatus. **(h)** In the Y-maze during training, fbKO mice displayed increased latency in locating the submerged platform compared to WT mice (Two-way ANOVA, main effect of genotype, $F_{1,75} = 24.1$, $P < 0.0001$; Bonferroni *post hoc* test, $*P < 0.05$ vs WT; WT $n=9$, fbKO $n=8$). **(i)** In the Y-maze memory tests, fbKO mice displayed significantly increased latency in locating the submerged platform at 1 h (Student's *t*-test with Welch's correction, $t_{(8)} = 3.188$, $P < 0.05$), 24 h (Student's *t*-test with Welch's correction, $t_{(7)} = 3.121$, $P < 0.05$) and 7 days (Mann-Whitney *U*-test, $U = 13.00$, $P < 0.05$) post-training ($*P < 0.05$, $**P < 0.01$ vs WT; WT $n=9$, fbKO $n=8$). **(j)** WT and fbKO mice exhibit similar percent of spontaneous alternations during the working memory Y-maze task (WT $n=8$, fbKO $n=8$). **(k)** Schematic of the Morris water maze (MWM) apparatus. **(l)** In the MWM during training, WT and fbKO mice displayed similar latency in locating the submerged platform (Two-way ANOVA, main effect of day, $F_{5,72} = 26.41$, $P < 0.0001$; $n = 7/\text{genotype}$). **(m–o)** In the MWM probe tests, WT and fbKO mice spent significantly more time in the goal quadrant relative to the other quadrants at 1 h (**m**; Two-way ANOVA, main effect of quadrant, $F_{3,48} = 70.41$, $P < 0.0001$), 24 h (**n**; Two-way ANOVA, main effect of quadrant, $F_{3,48} = 23.01$, $P < 0.0001$) and 7 days (**o**; Two-way ANOVA, main effect of quadrant, $F_{3,48} = 18.19$, $P < 0.0001$) post-training (Bonferroni *post hoc* test, $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ vs Goal; $n = 7/\text{genotype}$). Error bars represent mean \pm s.e.m. ANOVA, analysis of variance; fbKO, forebrain knockout; WT, wild type.

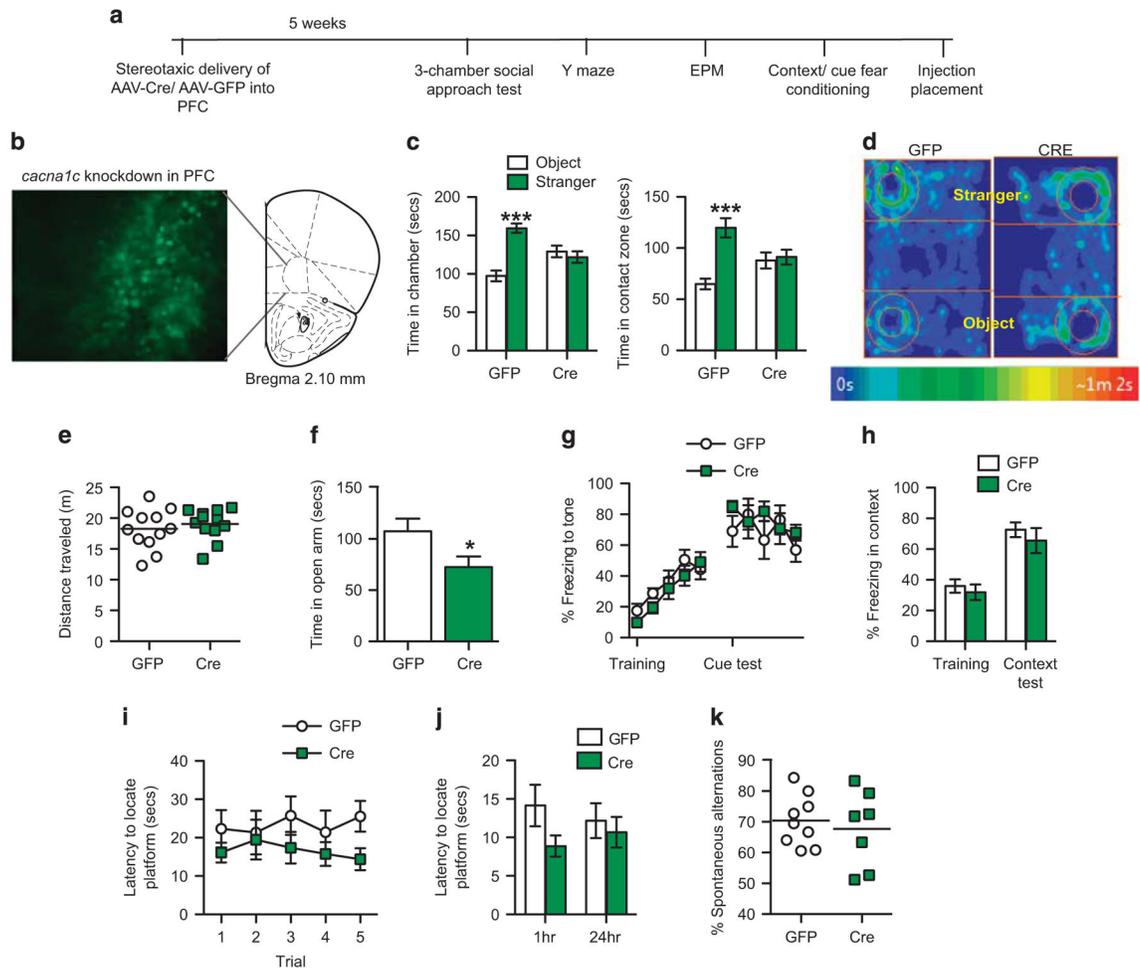


Figure 2.

Focal knockdown of *cacna1c* in the adult PFC recapitulates the social deficit and elevated anxiety-like behavior. **(a)** Outline of experimental design. AAV-Cre or AAV-GFP was stereotaxically injected into the PFC of adult *cacna1c* floxed mice and 5 weeks later tested in a battery of behavioral tests. **(b)** Representative image of GFP-positive cells expressed by AAV-Cre-GFP in the PFC of *cacna1c* floxed mice. **(c)** In the three-chamber social approach test, GFP but not Cre mice spent significantly more time in the chamber (left; Two-way ANOVA, genotype \times chamber, $F_{1,44} = 24.28$, $P < 0.0001$) and contact zone (right; two-way ANOVA, genotype \times zone, $F_{1,44} = 11.71$, $P = 0.0014$) containing the stranger mouse and novel object (Bonferroni *post hoc* test, *** $P < 0.001$ vs WT; GFP $n = 12$, Cre $n = 12$). **(d)** Representative heat maps showing time spent in the chambers and contact zones containing the stranger mouse and novel object during the social approach test. **(e)** GFP and Cre mice traveled similar distances during the social approach test (GFP $n = 12$, Cre $n = 12$). **(f)** Cre mice spent significantly less time in the open arm of the elevated plus maze compared to GFP mice (Student's *t*-test, $t_{13} = 2.156$, $P = 0.05$; * $P = 0.05$ vs GFP; GFP $n = 8$, Cre $n = 7$). **(g)** In cue fear conditioning, GFP and Cre mice displayed similar percent freezing to the tone during training (two-way ANOVA, main effect of tone, $F_{4, 70} = 12.7$, $P < 0.0001$) and the test. Each data point represents freezing during a single tone presentation (GFP $n = 8$,

Cre $n = 8$). **(h)** In contextual fear conditioning, GFP and Cre mice displayed similar percent freezing to the context during training and the test (GFP $n = 8$, Cre $n = 8$). **(i)** In the Y-maze, during training, Cre mice displayed slightly lower latency (main effect of treatment, $F_{1,75} = 5.374$, $P = 0.0232$) in locating the submerged platform compared to GFP mice (GFP $n = 9$, Cre $n = 8$). **(j)** In the Y-maze memory tests, GFP and Cre mice displayed similar latencies in locating the submerged platform at 1 h and 24 h post-training (GFP $n = 9$, Cre $n = 8$). **(k)** GFP and Cre mice exhibit similar percent of spontaneous alternations during the working memory Y-maze task (GFP $n = 8$, Cre $n = 8$). Error bars represent mean \pm s.e.m. ANOVA, analysis of variance; GFP, green fluorescent protein; PFC, prefrontal cortex.

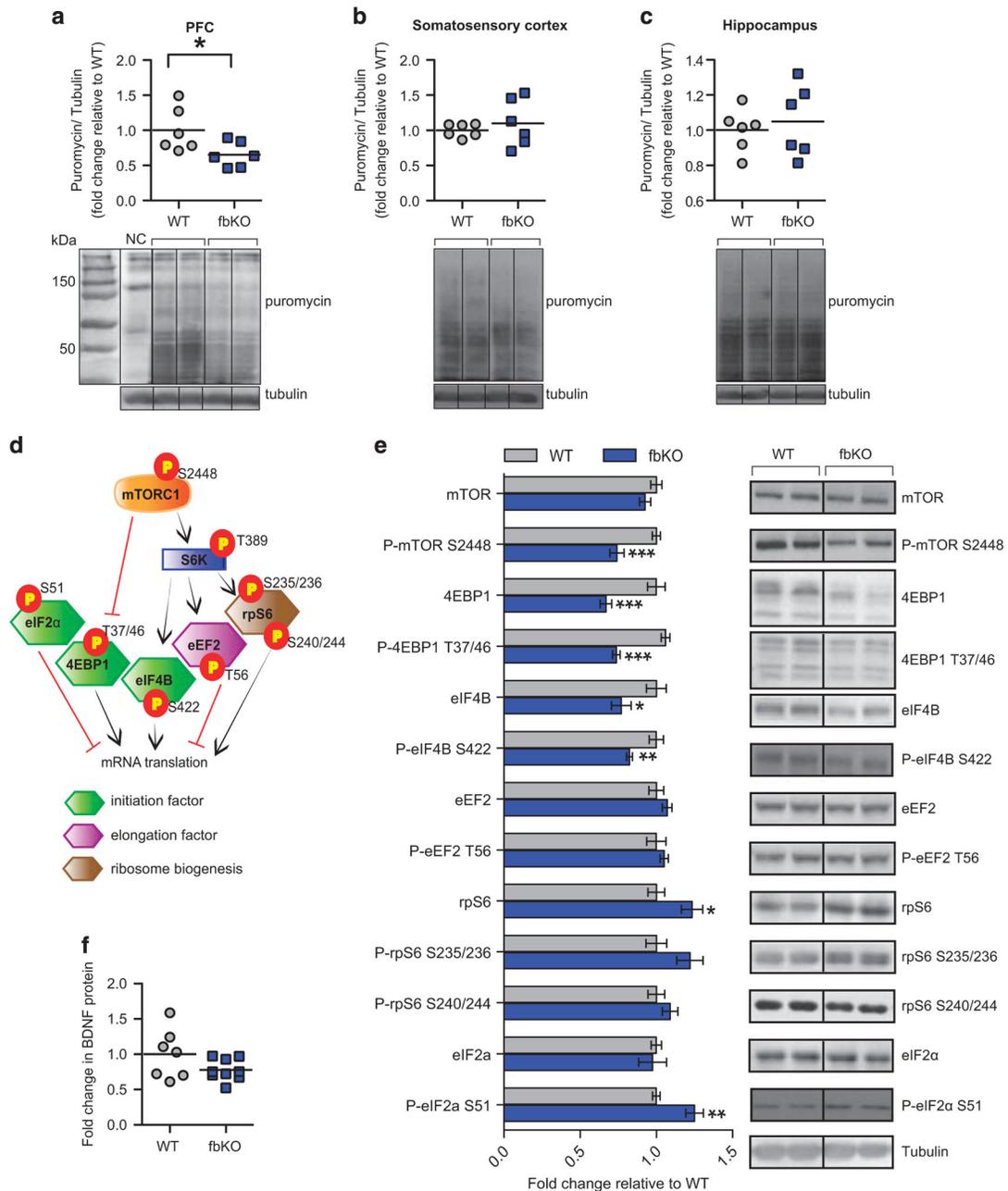


Figure 3. *Cacna1c* fbKO mice exhibit lower protein synthesis and lower mRNA translation initiation factors in the PFC. (a–c) Immunoblot analysis (top) and representative blots (bottom) of puromycin-labeled protein. Compared to WT mice, fbKO mice displayed significantly lower levels of puromycin-labeled proteins in the PFC (a; Student's *t*-test, $t_{(10)} = 2.330$, $P < 0.05$) but not the somatosensory cortex (b) or hippocampus (c; WT $n = 6$, fbKO $n = 6$). (d) Schematic representation indicating the relevant phosphorylation sites (indicative of their activity) of mTORC1, its substrates, and eIF2 α in their regulation of mRNA translation. (e) Immunoblot analysis (left) and representative blots of markers of mRNA translation in the

PFC from total protein lysates. Compared to WT mice, fbKO mice displayed significantly lower levels of P-mTOR S2448 (Student's *t*-test, $t_{(12)} = 4.410$, $P = 0.0009$), and its downstream substrates 4EBP1 (Student's *t*-test, $t_{(12)} = 4.739$, $P = 0.0005$), P-4EBP1 T37/46 (Student's *t*-test, $t_{(11)} = 8.627$, $P < 0.0001$), eIF4B (Student's *t*-test, $t_{(11)} = 2.494$, $P < 0.05$), and P-eIF4B S422 (Student's *t*-test, $t_{(11)} = 3.651$, $P = 0.0038$) as well as significantly higher levels of P-eIF2 α S51 (Student's *t*-test, $t_{(12)} = 3.972$, $P < 0.01$) in the PFC ($n = 6-7$ /genotype). (f) WT and fbKO mice display similar levels of mature BDNF protein levels in the PFC (WT $n = 7$, fbKO $n = 9$). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs WT. Error bars represent mean \pm s.e.m. fbKO, forebrain knockout; PFC, prefrontal cortex; WT, wild type.

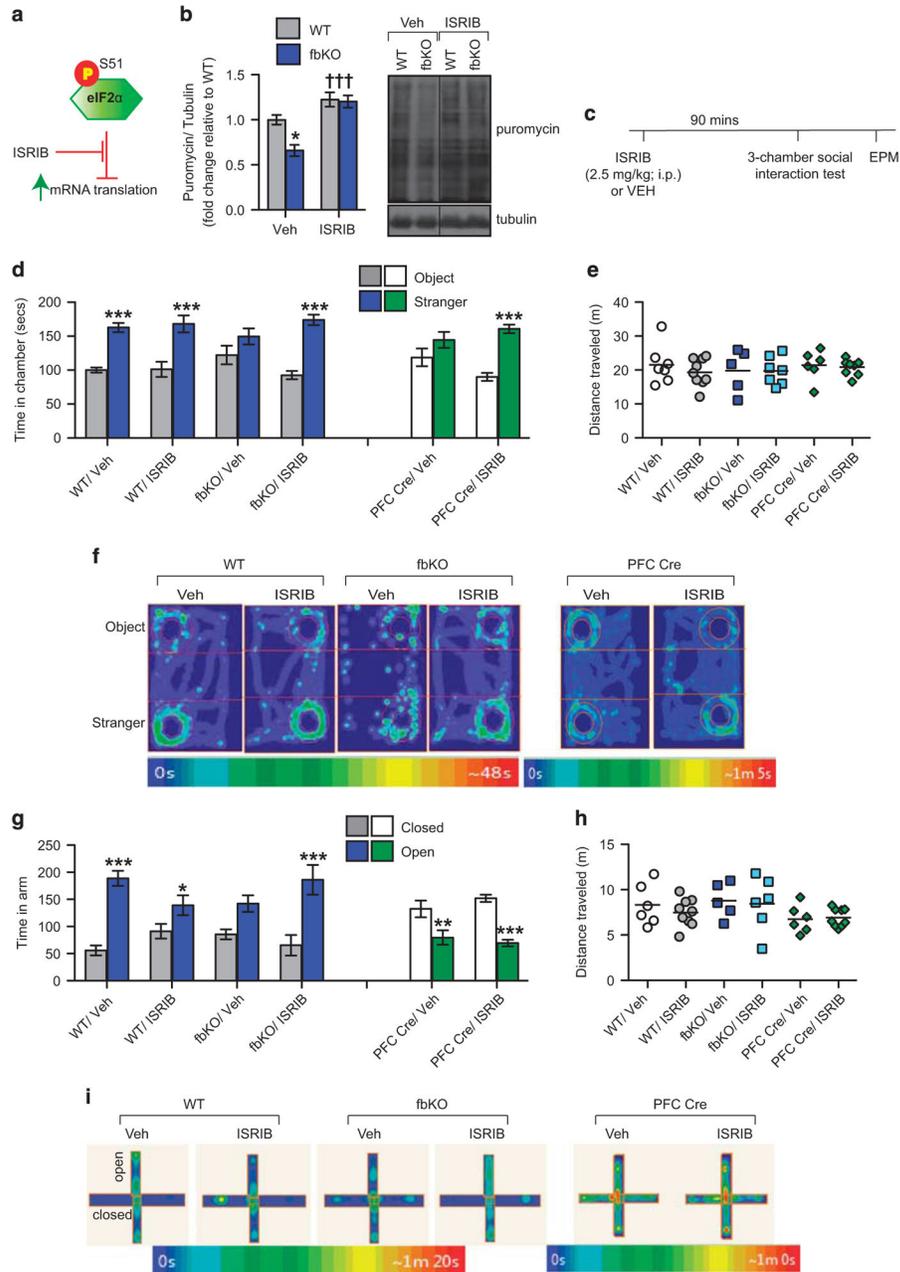


Figure 4.

ISRIB normalizes the lower protein synthesis in the PFC and rescues the social deficit and elevated anxiety in *cacna1c* fbKO mice. **(a)** Schematic depiction of ISRIB's inhibitory effect on the effect of P-eIF2 α S51 on general mRNA translation. **(b)** Immunoblot analysis (left) and representative blots (right) of puromycin-labeled protein in the PFC. Compared to WT mice, vehicle treated fbKO mice displayed significantly lower levels of puromycin-labeled proteins that was normalized to WT levels with ISRIB treatment (Two-way ANOVA, main effect of genotype, $F_{1,14} = 5.934$, $P = 0.0288$, main effect of treatment, $F_{1,14} = 26.78$, $P = 0.0001$; Bonferroni *post hoc* test, $*P < 0.05$ vs WT/Veh, $\dagger\dagger\dagger P < 0.001$ vs fbKO/Veh; WT/Veh

n = 3, fbKO/Veh *n* = 3, WT/ISRIB *n* = 5, fbKO/ISRIB *n* = 6) (c) Outline of experimental design. Adult WT, fbKO and PFC-Cre mice were given a single systemic injection of either ISRIB (2.5 mg/kg, i.p) or vehicle and ninety minutes later tested in social approach followed immediately by EPM. (d) In the three-chamber social approach test, vehicle and ISRIB treated WT mice (WT/Veh and WT/ISRIB, respectively) spent significantly more time in the chamber containing the stranger mouse compared to the novel object (Two-way ANOVA, main effect of chamber, $F_{1, 28} = 40.78$, $P < 0.0001$). In contrast, vehicle treated fbKO mice (fbKO/Veh) mice spent similar amounts of time in the chambers containing the stranger mouse and object while fbKO mice with ISRIB treatment (fbKO/ISRIB) displayed a normalization of this social deficit spending significantly more time with the stranger mouse compared to the object (Two-way ANOVA, treatment \times chamber, $F_{1, 20} = 8.12$, $P = 0.0099$), looking similar to WT/Veh mice. Similarly, vehicle treated PFC Cre mice (PFC Cre/Veh) spent similar amounts of time in the chamber containing the stranger mouse and object whereas PFC Cre mice following ISRIB treatment (PFC Cre/ISRIB) exhibited a normalization of this social deficit by spending significantly more time with the stranger mouse compared to the novel object (Two-way ANOVA, treatment \times chamber, $F_{1, 26} = 6.535$, $P = 0.0168$) looking similar to WT mice (Bonferroni *post hoc* test, $***P < 0.0001$ vs object; WT: Veh *n* = 7, ISRIB *n* = 9; fbKO: Veh *n* = 5, ISRIB *n* = 7; PFC-Cre: Veh *n* = 6, ISRIB *n* = 9). (e) All mice traveled similar distances during the social approach test (WT: Veh *n* = 7, ISRIB *n* = 9; fbKO: Veh *n* = 5, ISRIB *n* = 7; PFC Cre: Veh *n* = 6, ISRIB *n* = 9). (f) Representative heat maps showing time spent in the chambers containing the stranger mouse and novel object during the social approach test. (g) In the elevated plus maze, WT/Veh and WT/ISRIB mice spent significantly more time in the open arms relative to the closed arms of the maze, although there was a minor effect of ISRIB on anxiety-like behavior (Two-way ANOVA, treatment \times arm, $F_{1, 26} = 7.349$, $P = 0.0117$). fbKO/Veh mice spent similar amounts of time in the open and closed arms of the maze while fbKO/ISRIB mice displayed normalization of this behavior spending significantly more time in the open arm relative to the closed arm of the maze (Two-way ANOVA, main effect of arm, $F_{1, 18} = 19.64$, $P = 0.0003$), looking similar to WT/Veh mice. In contrast, both PFC Cre/Veh and PFC Cre/ISRIB mice spent significantly less time in the open arm of the maze compared to the closed arm (Two-way ANOVA, main effect of arm, $F_{1, 26} = 47.01$, $P < 0.0001$; Bonferroni *post hoc* test, $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ vs closed arm; WT: Veh *n* = 6, ISRIB *n* = 9; fbKO: Veh *n* = 5, ISRIB *n* = 6; PFC Cre: Veh *n* = 6, ISRIB *n* = 9). (h) All mice traveled similar distances in the elevated plus maze (WT: Veh *n* = 6, ISRIB *n* = 9; fbKO: Veh *n* = 5, ISRIB *n* = 6; PFC Cre: Veh *n* = 6, ISRIB *n* = 9). (i) Representative heat maps showing time spent in the open and closed arms of the elevated plus maze. Error bars represent mean \pm s.e.m. ANOVA, analysis of variance; EPM, elevated plus maze; fbKO, forebrain knockout; PFC, prefrontal cortex; WT, wild type.

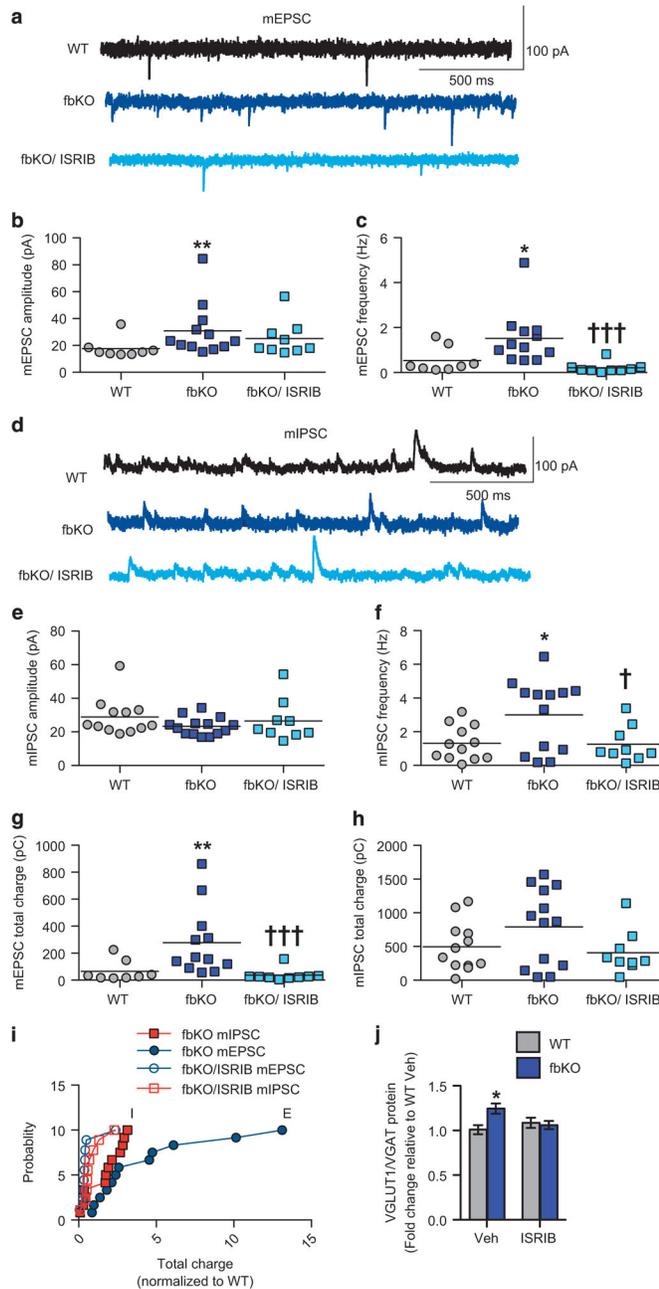


Figure 5.

Cacna1c fbKO mice exhibit higher excitatory/inhibitory ratio in layer 5 pyramidal neurons of the PFC that is normalized with ISRIB. (a) Representative traces of mEPSCs from layer 5 pyramidal neurons of the PFC. (b) fbKO mice displayed significantly higher amplitude of mEPSCs that remained unchanged with ISRIB (Kruskal–Wallis test, $H = 8.242$, $P = 0.0162$; Dunn’s uncorrected *post hoc* test, $**P < 0.01$ vs WT; WT $n = 8$, fbKO $n = 12$, fbKO/ISRIB $n = 9$ cells, from $n = 2–4$ mice per group). (c) fbKO mice displayed significantly higher frequency of mEPSCs that was normalized to WT levels with ISRIB (Kruskal–Wallis test, $H = 16.74$, $P = 0.0002$; Dunn’s uncorrected *post hoc* test, $*P < 0.05$ vs WT, $†††P < 0.0001$ vs

fbKO; WT $n=8$, fbKO $n=12$ cells, fbKO/ISRIB $n=9$ cells, from $n=2-4$ mice per group). (d) Representative traces of mIPSCs from layer 5 pyramidal neurons of the PFC. (e) fbKO mice displayed similar amplitude of mIPSCs compared to WT mice that was unchanged with ISRIB (WT $n=12$, fbKO $n=13$, fbKO/ISRIB $n=9$ cells, from $n=2-4$ mice per group). (f) fbKO mice displayed significantly higher frequency of mIPSCs that was normalized to WT levels with ISRIB (one-way ANOVA, $F_{2,33} = 4.909$, $P = 0.0141$; Bonferroni *post hoc* test, $*P < 0.05$ vs WT, $^{\dagger}P < 0.05$ vs fbKO; WT $n=12$, fbKO $n=13$, fbKO/ISRIB $n=9$ cells, from $n=4$ mice per group). (g) fbKO mice displayed significantly higher total charge transfer for mEPSCs that was normalized with ISRIB (Kruskal–Wallis test, $H = 14.2$, $P = 0.0008$; Dunn's uncorrected *post hoc* test, $**P < 0.01$ vs WT, $^{\dagger\dagger\dagger}P < 0.001$ vs fbKO; WT $n=8$, fbKO $n=12$, fbKO/ISRIB $n=9$ cells, from $n=2-4$ mice per group). (h) fbKO mice displayed similar total charge transfer for mIPSCs compared to WT mice that was unchanged with ISRIB (WT $n=12$, fbKO $n=13$, fbKO/ISRIB $n=9$ cells, from $n=2-4$ mice per group). (i) Higher relative change in mEPSC than mIPSC total charge transfer for each fbKO neuron normalized to the mean WT value that was corrected by ISRIB. (j) Compared to WT/Veh mice, fbKO/Veh mice had significantly higher levels of VGLUT1/VGAT ratio in the synaptoneurosome from the PFC while fbKO/ISRIB mice showed normalization of this elevated ratio (two-way ANOVA, treatment \times genotype, $F_{1,23} = 5.797$, $P = 0.0245$; Bonferroni *post hoc* test, $*P < 0.05$ vs WT; WT: Veh $n=6$, ISRIB $n=7$; fbKO: Veh $n=6$, ISRIB $n=7$). Error bars represent mean \pm s.e.m. ANOVA, analysis of variance; E, excitatory; fbKO, forebrain knockout, I, inhibitory; mEPSCs, miniature excitatory post synaptic currents; mIPSCs, miniature inhibitory post synaptic currents; WT, wild type.