



# First Complete Genome Sequences of Human Sapovirus Strains Classified as GI.3, GI.4, GI.6, GI.7, and GII.7

Tomoichiro Oka,<sup>a</sup> Nobuhiro Iritani,<sup>b</sup> Mineyuki Okada,<sup>c</sup> Tomoko Ogawa,<sup>d</sup> Setsuko Iizuka,<sup>e</sup> Chika Tatsumi,<sup>e</sup> Seiya Harada,<sup>f</sup> Kei Haga,<sup>a\*</sup> Yen Hai Doan<sup>a</sup>

<sup>a</sup>Department of Virology II, National Institute of Infectious Diseases, Tokyo, Japan

<sup>b</sup>Division of Microbiology, Osaka Institute of Public Health, Osaka, Japan

<sup>c</sup>Chiba Prefecture Toso Meat Inspection Office, Chiba, Japan

<sup>d</sup>Division of Virology, Chiba Prefectural Institute of Public Health, Chiba, Japan

<sup>e</sup>Division of Virology, Shimane Prefectural Institute of Public Health and Environmental Science, Shimane, Japan

<sup>f</sup>Department of Microbiology, Kumamoto Prefectural Institute of Public Health and Environmental Science, Kumamoto, Japan

**ABSTRACT** We report here the first complete genome sequences of genotype GI.3, GI.4, GI.6, GI.7, and GII.7 sapovirus strains, detected from fecal samples of acute gastroenteritis patients. Complete or nearly complete genome sequences of all 18 genotypes of human sapoviruses are now available for phylogenetic analysis and primer design.

Genetically diverse sapovirus (SaV) strains have been detected in fecal specimens from patients with acute gastroenteritis (1–4). We have recently classified human SaV strains into 17 genotypes (i.e., GI.1 to GI.7, GII.1 to GII.7, GIV.1, GV.1, and GV.2) based on complete major structural protein (VP1) nucleotide sequence (5). Recently, an additional human SaV genotype (i.e., GII.8) has been proposed (6–8). Currently, complete or nearly complete genome sequences of 13 human SaV genotypes (i.e., GI.1, GI.2, GI.5, GII.1, GII.2, GII.3, GII.4, GII.5, GII.6, GII.8, GIV.1, GV.1, and GV.2) are available in public databases. In this study, we determined the first complete genome sequences of genotypes GI.3, GI.4, GI.6, GI.7, and GII.7.

Viral RNA was extracted from fecal suspensions using the High Pure viral RNA kit (Roche) or QIAamp viral RNA minikit (Qiagen). Library preparation from the purified RNA for sequencing on an Illumina MiSeq platform (Illumina) and *de novo* assembly of consensus SaV genome sequence were performed first, as described (9, 10). For the GII.7 SaV strain, we performed seminested long reverse transcriptase PCR (RT-PCR) (amplicon size, 5.7 kb) (10, 11) using the newly designed forward primer 5'-ATGGCTTCYAAGCCATTCTACC-3', which corresponds to the 22 nucleotides (nt) from the predicted start codon of the open reading frame (ORF) 1 of genotype GII.1 (GenBank accession no. AJ249939 and AY237419), GII.2 (AY237420), GII.3 (AY603425), GII.5 (LC190463), and GII.6 (AY646855) SaV genomes in combination with two primers designed in the previously determined VP1-encoding-region sequence (12). Furthermore, the 3' ends of SaV genomes of the GI.4, GI.7, and GII.7 SaV strains (~2.5 kb) were amplified by RT-PCR with a gene-specific forward primer and the reverse primer TX30SXN, which was complementary to the 3'-end poly(A) tail (9–11). The 5' terminal nucleotide sequence of the five SaV strains was further determined by seminested PCR-based 5'-RACE (rapid amplification of cDNA ends) (~0.5 kb) (9, 10). These PCR products were purified and sequenced directly and/or after cloning using the BigDye Terminator Cycle sequence kit v3.1 and the 3130 Genetic Analyzer capillary sequencer (Applied Biosystems), or library preparation (from the purified PCR product) and an

**Received** 9 February 2018 **Accepted** 23 February 2018 **Published** 22 March 2018

**Citation** Oka T, Iritani N, Okada M, Ogawa T, Iizuka S, Tatsumi C, Harada S, Haga K, Doan YH. 2018. First complete genome sequences of human sapovirus strains classified as GI.3, GI.4, GI.6, GI.7, and GII.7. *Genome Announc* 6:e00168-18. <https://doi.org/10.1128/genomeA.00168-18>.

**Copyright** © 2018 Oka et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Tomoichiro Oka, [oka-t@nih.go.jp](mailto:oka-t@nih.go.jp).

\* Present address: Kei Haga, Department of Molecular Virology and Microbiology, Baylor College of Medicine, Houston, Texas, USA.

Illumina MiSeq sequencer (10). The full-length genome sequences of the genotype GI.3, GI.4, GI.6, GI.7, and GI.7 SaV strains were assembled using the Sequencher program v4.10.1 (GeneCodes) and analyzed with Genetyx-Mac software v16.0.4 (Genetyx Corporation).

The genomes of GI.3 Hu/OH08021/2008/JP, GI.4 Hu/SV/Chiba/000496/2000, GI.6 Hu/SV/Chiba/000764/2000, GI.7 Hu/D1714-B/2008/JPN, and GI.7 Hu/20072248/2008/JP SaV strains consist of 7,442, 7,436, 7,443, 7,452, and 7,462 nt, respectively, excluding the poly(A) tail. All of these SaV genomes were predicted to encode two ORFs, a short 5' untranslated region (UTR) (12 or 13 nt long) and a 3'-UTR (78 to 112 nt long). The 5' terminal sequence was conserved as GTG, similarly to those of other SaVs (10, 11).

The new sequence data determined in this study will be useful in designing more broadly reactive primers and probes for human SaV detection PCR, as well as establishment of a nonstructural protein coding region typing scheme like that recently established for norovirus (13).

**Accession number(s).** The genome sequences of GI.3 Gu/OH08021/2008/JP, GI.4 Hu/SV/Chiba/000496/2000, GI.6 Hu/SV/Chiba/000764/2000, GI.7 Hu/D1714-B/2008/JPN, and GI.7 Hu/20072248/2008/JP SaV strains have been deposited in GenBank under the accession no. [AB623037](#), [AJ606693](#), [AJ606694](#), [AB522390](#), and [AB630067](#), respectively.

## ACKNOWLEDGMENTS

This work was partly supported by Grants-in-Aid for Scientific Research (C) from the Ministry of Education, Culture, Sports, Science, and Technology of Japan and by the Research Program on Emerging and Re-emerging Infectious Diseases Program from the Japan Agency for Medical Research and Development (AMED).

## REFERENCES

- Harada S, Okada M, Yahiro S, Nishimura K, Matsuo S, Miyasaka J, Nakashima R, Shimada Y, Ueno T, Ikezawa S, Shinozaki K, Katayama K, Wakita T, Takeda N, Oka T. 2009. Surveillance of pathogens in outpatients with gastroenteritis and characterization of sapovirus strains between 2002 and 2007 in Kumamoto Prefecture, Japan. *J Med Virol* 81:1117–1127. <https://doi.org/10.1002/jmv.21454>.
- Iizuka S, Oka T, Tabara K, Omura T, Katayama K, Takeda N, Noda M. 2010. Detection of sapoviruses and noroviruses in an outbreak of gastroenteritis linked genetically to shellfish. *J Med Virol* 82:1247–1254. <https://doi.org/10.1002/jmv.21791>.
- Okada M, Yamashita Y, Oseto M, Ogawa T, Kaiho I, Shinozaki K. 2006. Genetic variability in the sapovirus capsid protein. *Virus Genes* 33: 157–161. <https://doi.org/10.1007/s11262-005-0051-7>.
- Iritani N, Yamamoto SP, Abe N, Kubo H, Oka T, Kaida A. 2016. Epidemics of GI.2 sapovirus in gastroenteritis outbreaks during 2012–2013 in Osaka City, Japan. *J Med Virol* 88:1187–1193. <https://doi.org/10.1002/jmv.24451>.
- Oka T, Wang Q, Katayama K, Saif LJ. 2015. Comprehensive review of human sapoviruses. *Clin Microbiol Rev* 28:32–53. <https://doi.org/10.1128/CMR.00011-14>.
- Kagning Tsinda E, Malasao R, Furuse Y, Gilman RH, Liu X, Apaza S, Espetia S, Cama V, Oshitani H, Saito M. 2017. Complete coding genome sequences of uncommon GI.8 sapovirus strains identified in diarrhea samples collected from Peruvian children. *Genome Announc* 5:e01137–17. <https://doi.org/10.1128/genomeA.01137-17>.
- Liu X, Jahuir H, Gilman RH, Alva A, Cabrera L, Okamoto M, Xu H, Windle HJ, Kelleher D, Varela M, Verastegui M, Calderon M, Sanchez G, Sarabia V, Ballard SB, Bern C, Mayta H, Crabtree JE, Cama V, Saito M, Oshitani H. 2016. Etiological role and repeated infections of sapovirus among children aged less than 2 years in a cohort study in a peri-urban community of Peru. *J Clin Microbiol* 54:1598–1604. <https://doi.org/10.1128/JCM.03133-15>.
- Sanchez GJ, Mayta H, Pajuelo MJ, Neira K, Xiaofang L, Cabrera L, Ballard SB, Crabtree JE, Kelleher D, Cama V, Bern C, Oshitani H, Gilman RH, Saito M, Sapovirus Working Group. 2017. Epidemiology of sapovirus infections in a birth cohort in Peru. *Clin Infect Dis* <https://www.ncbi.nlm.nih.gov/pubmed/29309577>.
- Oka T, Doan YH, Haga K, Mori K, Ogawa T, Yamazaki A. 2017. Genetic characterization of rare genotype GI.5 sapovirus strain detected from a suspected food-borne gastroenteritis outbreak among adults in Japan in 2010. *Jpn J Infect Dis* 70:223–224. <https://doi.org/10.7883/yoken.JIID.2016.468>.
- Oka T, Doan YH, Shimoike T, Haga K, Takizawa T. 2017. First complete genome sequences of genogroup V, genotype 3 porcine sapoviruses: common 5'-terminal genomic feature of sapoviruses. *Virus Genes* 53: 848–855. <https://doi.org/10.1007/s11262-017-1481-8>.
- Oka T, Lu Z, Phan T, Delwart EL, Saif LJ, Wang Q. 2016. Genetic characterization and classification of human and animal sapoviruses. *PLoS One* 11:e0156373. <https://doi.org/10.1371/journal.pone.0156373>.
- Oka T, Mori K, Iritani N, Harada S, Ueki Y, Iizuka S, Mise K, Murakami K, Wakita T, Katayama K. 2012. Human sapovirus classification based on complete capsid nucleotide sequences. *Arch Virol* 157:349–352. <https://doi.org/10.1007/s00705-011-1161-2>.
- Kroneman A, Vega E, Vennema H, Vinjé J, White PA, Hansman G, Green K, Martella V, Katayama K, Koopmans M. 2013. Proposal for a unified norovirus nomenclature and genotyping. *Arch Virol* 158:2059–2068. <https://doi.org/10.1007/s00705-013-1708-5>.