

Sequence analysis

A novel data structure to support ultra-fast taxonomic classification of metagenomic sequences with k -mer signatures

Xinan Liu^{1,†}, Ye Yu^{1,†}, Jinpeng Liu², Corrine F. Elliott¹, Chen Qian³ and Jinze Liu^{1,*}

¹Department of Computer Science and ²Biostatistics and Bioinformatics Shared Resource Facility, Markey Cancer Center, University of Kentucky, Lexington, KY, USA and ³Department of Computer Engineering, UC Santa Cruz, Santa Cruz, CA, USA

*To whom correspondence should be addressed.

[†]The authors wish it to be known that, in their opinion, the first two authors should be regarded as Joint First Authors.

Associate Editor: Cenk Sahinalp

Received on April 7, 2017; revised on June 16, 2017; editorial decision on June 29, 2017; accepted on July 3, 2017

Abstract

Motivation: Metagenomic read classification is a critical step in the identification and quantification of microbial species sampled by high-throughput sequencing. Although many algorithms have been developed to date, they suffer significant memory and/or computational costs. Due to the growing popularity of metagenomic data in both basic science and clinical applications, as well as the increasing volume of data being generated, efficient and accurate algorithms are in high demand.

Results: We introduce MetaOthello, a probabilistic hashing classifier for metagenomic sequencing reads. The algorithm employs a novel data structure, called l -Othello, to support efficient querying of a taxon using its k -mer signatures. MetaOthello is an order-of-magnitude faster than the current state-of-the-art algorithms Kraken and Clark, and requires only one-third of the RAM. In comparison to Kaiju, a metagenomic classification tool using protein sequences instead of genomic sequences, MetaOthello is three times faster and exhibits 20–30% higher classification sensitivity. We report comparative analyses of both scalability and accuracy using a number of simulated and empirical datasets.

Availability and implementation: MetaOthello is a stand-alone program implemented in C++. The current version (1.0) is accessible via <https://doi.org/10.5281/zenodo.808941>.

Contact: liuj@cs.uky.edu

Supplementary information: Supplementary data are available at *Bioinformatics* online.

1 Introduction

Metagenomics is the study of genomic content obtained in bulk from an environment of interest, such as the human body (Huttenhower and Human Microbiome Project Consortium, 2012), seawater (Venter *et al.*, 2004), or acidic mine drainage (Tyson *et al.*, 2004). Metagenomics studies often generate tens of millions of sequencing reads in order to capture the presence of microbial

organisms and quantify their relative abundances, rendering the classification and analysis of these data a logistical challenge.

One of the major computational challenges in the analysis of metagenomic data is the classification of each sequencing read into the most-specific biological taxon to which sequence conservation supports its assignment. Specifically, a read is classified as belonging to a taxon if it has high sequence similarity with the reference

genomes collected for that taxon, a process made possible by the large deposits of reference sequences collected in recent years for a variety of microbial species. In 2014 alone, more than 10 000 sequence records were newly added to the NCBI RefSeq database thanks to the accessibility of high-throughput sequencing technology.

Existing classification methods can be divided into two broad categories: alignment-based and alignment-free. The former approach, implemented most popularly as BLAST (Altschul *et al.*, 1990), assigns each read to the taxon that affords the best alignment with its reference genomes. Several methods, including MEGAN (Huson *et al.*, 2007), PhymmBL (Brady and Salzberg, 2009) and NBC (Rosen *et al.*, 2011), apply additional machine-learning techniques to BLAST results to increase classification accuracy. These methods are often slower than BLAST alone, rendering them computationally prohibitive for large-scale analysis of many millions of short reads. However, the recent development of Centrifuge (Kim *et al.*, 2016) has significantly improved the scalability of the alignment-based algorithm using FM-index. Besides using genomic sequences as reference, the recently published tool Kaiju (Menzel and Krogh, 2015) performs alignments towards protein sequences, achieving faster classification speed than existing tools.

The other line of work, including but not limited to LMAT (Ames *et al.*, 2013), Kraken (Wood and Salzberg, 2014) and Clark (Ounit *et al.*, 2015), classifies a read using exact k -mer matches between the read and the collection of reference sequences belonging to the target taxon, thereby avoiding inefficient base-by-base alignment while maintaining a sensitivity and specificity comparable to the alignment-based approach. This approach is generally faster than alignment-based methods and allows for greater flexibility in reference material because it requires only the collection of k -mers extracted from reference sequences belonging to each taxon. Thus k -mers extracted from DNA or RNA sequencing data can be included as reference material without being assembled, increasing the sensitivity of the algorithm in capturing natural variants that are often missed using reference genomes alone.

The above alignment-free algorithms rely on the use of indexing structures for k -mer matching. For example, Kraken indexes its lexicographically sorted k -mer database using a minimizer offset array, while Clark uses a hash table to store the mapping between a k -mer and its classification information. Both Kraken and Clark require computers with large memory to support the construction of their indexing structures (at least 120 GB RAM) and the classification of reads (at least 70 GB RAM) to achieve optimal classification accuracy. Although there exist variations on both algorithms requiring smaller memory footprints, they often afford significantly lower sensitivity or slower execution speed compared to the full version. For this reason, the ever-increasing amount of sequencing and reference-genome data call for tools with better scalability in both memory and computation.

In this paper, we present a new algorithm, dubbed MetaOthello, for taxonomic classification of sequencing reads. Our algorithm builds upon taxon-specific k -mer signatures to support direct assignment to any level in the taxonomy. It employs a novel data structure, l -Othello, to support ultra-fast k -mer classification, achieving at least an order-of-magnitude improvement in speed over the state-of-the-art methods, Kraken and Clark, and [runs] three times faster than Kaiju. In the meantime, MetaOthello also substantially reduces the memory footprint, typically requiring only one-third [the memory] of the aforementioned methods. This modest memory requirement allows our algorithm to run on typical lab servers with 32 GB RAM, rendering it more accessible to biological researchers than

those with memory requirements achievable only by supercomputers. Additionally, our algorithm is capable of conducting hierarchical top-down taxonomic classification and delivers performance competitive to, if not better than, other algorithms in both sensitivity and specificity as validated by benchmarking on a variety of datasets.

2 Algorithms

2.1 k -mer taxon signatures

A k -mer is a length k subsequence of genomic sequences; for any sequence of length L , there exist a maximum of $L - k + 1$ possible k -mers. Metagenomic reference material consists of one or more complete reference genomes belonging to an organism. Increasingly sophisticated sequencing techniques have permitted discovery of distinct reference genomes for a single species of organism, thereby capturing genomic variations that are often important to the functionality of the microbial species. The number of genomes (whether draft or complete) available as metagenomic reference material increases with each new discovery. If we consider each dataset as a collection of k -mers, a given taxon can be described by the set of k -mers present in the reference sequences belonging to its taxonomic subtree. The problem of classifying a metagenomic read thus simplifies to the identification of the taxon that best matches the set of k -mers associated with the target read. When k is sufficiently large (e.g. $k \geq 20$), the majority of k -mers are unique to the species carrying them. These species-specific k -mers may serve as signatures, directly implicating the appropriate taxonomic classification. However, a significant proportion of k -mers is present in multiple species, making them unique only to higher-ranking taxa. In this paper, we formalize the taxonomic specificity of a k -mer as the signature of a taxon: A k -mer is considered to be a **signature** of a taxon if (i) the k -mer does not appear in any genomic references belonging to ancestors or siblings of the target taxon, but only to sequences belonging to the taxon's subtree, and (ii) the k -mer is not a signature of any lower-ranked taxon in the subtree. Equivalently, the taxon evincing a k -mer signature is the lowest common ancestor (LCA) of all species in the taxonomy whose reference genomes contain that k -mer (Ames *et al.*, 2013; Tu *et al.*, 2014).

In this way, as illustrated in Figure 1A, the set of all k -mers present in the genomic references of a taxonomy can be divided into disjoint collections, each of which contains the set of signature k -mers belonging to a single node in the taxonomy tree. Formally, let S be the set of all k -mers present in genomic references annotated by the taxonomy and let $T = \{1, 2, \dots, |T|\}$ be the taxa (nodes) present in the taxonomy. Then S can be divided into $|T|$ disjoint sets, $S = \{S_0, S_1, \dots, S_t, \dots, S_{(|T|-1)}\}$, where for any node $t \in T$, S_t corresponds to the set of k -mer signatures belonging to taxon t . Thus, there exists a mapping, $g : S \rightarrow T$, such that $g(s) = t$ if the k -mer, $s \in S$, is a signature of the taxon, $t \in T$.

2.2 k -mer classification with l -Othello

The core data structure of MetaOthello is called l -Othello. l -Othello is essentially a hashing classifier, first designed for fast-forwarding information base queries (Yu *et al.*, 2016). It is capable of classifying a key to the appropriate member of a large collection of categories with high efficiency in memory and speed. In our particular application, l -Othello supports the mapping between a k -mer signature and the corresponding taxon.

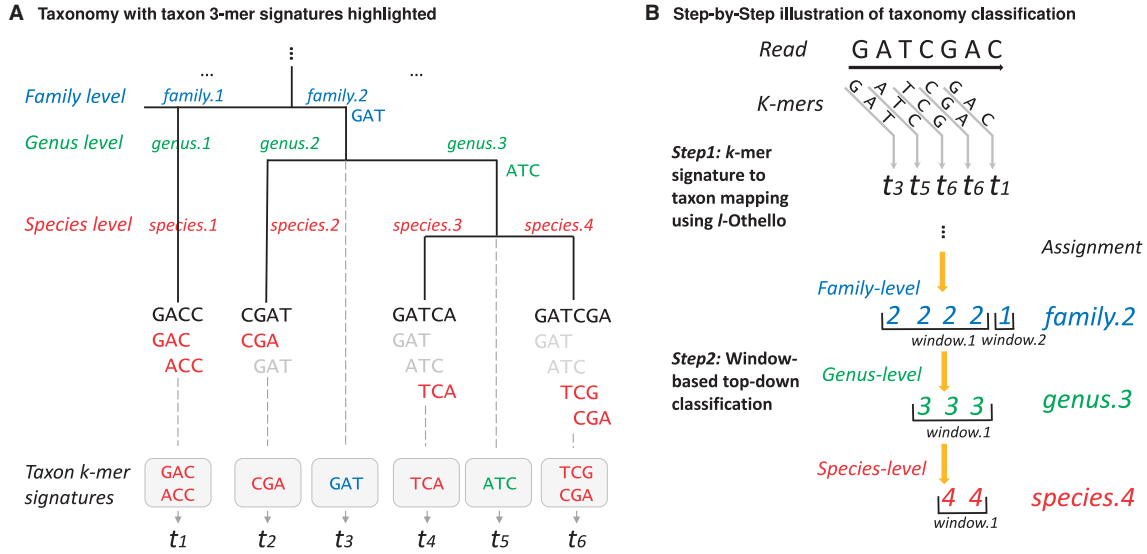


Fig. 1. Illustration of MetaOthello algorithm. (A) An example of taxonomy with reference sequences in the leaf nodes. 3-mers that are signatures to each taxon are highlighted in bold font with different shades. (B) A two-step approach to read classification (Color version of this figure is available at *Bioinformatics* online.)

2.2.1 Overview of the *l*-Othello data structure

l-Othello maintains a query function between any given k -mer and a taxon: $\tau: S_U \rightarrow \{0, 1, \dots, 2^l - 1\}$, where S_U is the universal set of k -mers (i.e. $S_U = \sigma^k$, $\sigma = \{A, C, G, T\}$). l is determined by the total number of taxa T , where $l = \lceil \log_2 |T| \rceil$. The algorithm satisfies the following properties: (i) *l*-Othello is always able to retrieve the correct taxon ID corresponding to a valid k -mer signature; that is, for any $s \in S$, $\tau(s) = t$, where t is the ID of the taxon to which s is specific. (ii) When provided an *alien* k -mer (i.e. a k -mer that does not appear in the reference sequences), *l*-Othello is able to recognize the alien with high success rate but may carry a slight risk of assigning it to a random taxon in the taxonomy.

Figure 2 shows an example *l*-Othello structure. An *l*-Othello data structure works by maintaining a pair of hash functions, $\langle h_a, h_b \rangle$ and two arrays of l -bit integers, A and B . $h_a: S_U \rightarrow \{0, 1, \dots, m_a - 1\}$ and $h_b: S_U \rightarrow \{0, 1, \dots, m_b - 1\}$, where m_a and m_b are integer values determined during *l*-Othello construction. The relationships among the elements of A and B can be viewed as a bipartite graph $G = (U, V, E)$, where nodes in U and V correspond to elements in A and B respectively. A query of k -mer s on the graph yields a node index $i = h_a(s)$ in U and a node index $j = h_b(s)$ in V . The classification of k -mer s is determined by the values at $A[h_a(s)]$ and $B[h_b(s)]$, via a \oplus (bitwise XOR) operation:

$$\tau(s) = A[h_a(s)] \oplus B[h_b(s)].$$

The bit-wise XOR operation of integers has the following property: For any l -bit integer x , $x \oplus x = 0$, $x \oplus 0 = x$.

When *l*-Othello is properly constructed, $\tau(s) = t$ for any k -mer specific to taxon t , i.e. $s \in S_t$. The success of *l*-Othello relies on assigning bitmap values on both sides of the bipartite graph, where the \oplus operation between two nodes can directly generate the class membership. Setting the bitmap of two nodes with no initial values is fairly simple and can be achieved in multiple ways. For example, when $l = 1$ and assuming the membership of a key is 1, we can assign the bit values of the two nodes u_i and v_j as either $A[i] = 0, B[j] = 1$ or $A[i] = 1, B[j] = 0$. However, if the value of one node has already been determined by another key, then only the remaining value is altered. For example, if $A[i]$ is already set as 0, given $A[i] \oplus B[j] = 1$,

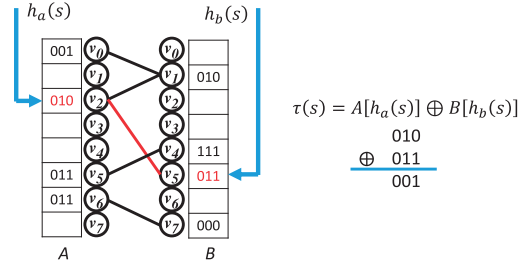


Fig. 2. An example of *l*-Othello with $l=3$ classifying $n=5$ k -mers. Left: Bipartite graph G and corresponding bitmaps A and B . Each edge in G represents a k -mer. Hash functions h_a and h_b map the k -mer s into corresponding locations in A and B . Right: Query of s returns $\tau(s) = (001)_2 = 1$

then $B[j] = 1$. In the worst-case scenario, both $A[i]$ and $B[j]$ have already been determined by their involvement with other keys. This situation creates a cycle in the graph when edge u_i and v_j is added, possibly resulting in a conflict and failed assignment of a bit value. When a conflict arises, we have two options: remove the k -mer or select a different hash function. Theorem 1 shows that for a randomly selected pair of hash functions $\langle h_a, h_b \rangle$, the probability of G being cyclic is extremely low. In our experiment, fewer than 100 k -mers among 6 billion were removed due to conflicts. Additionally, because multiple k -mers manifest in one read, losing one k -mer does not significantly affect the accuracy of the algorithm, so we may omit k -mers whose inclusion would cause a conflict.

THEOREM 1 Suppose h_a, h_b are randomly selected from a family of random hash functions such that $h_a: S \rightarrow \{0, 1, \dots, m_a - 1\}$ and $h_b: S \rightarrow \{0, 1, \dots, m_b - 1\}$. Given a set of k -mers $S = S_0 \cup S_1 \cup \dots \cup S_{2^l - 1}$, let $n = |S|$. Construct a bipartite graph $G = (U, V, E)$, where an edge $(u_i, v_j) \in E$ if and only if there is a k -mer $s \in S$ such that $h_a(s) = i$ and $h_b(s) = j$. Let χ be the number of cycles in G . Then χ converges to a Poisson distribution with parameter $\lambda = -\frac{1}{2} \ln(1 - c^2)$, where $c = \frac{n}{\sqrt{m_a m_b}}$.

The proof of Theorem 1 can be found in Supplementary Section S1. We recommend the values of m_a and m_b as follows: Let m_a and m_b

be powers of 2, where m_a is the smallest value such that $m_a \geq 1.33n$, and m_b is the smallest value such that $m_b \geq n$. As a result, we have $2.67n \leq m_a + m_b < 4n$ and $0.14 < \lambda < 0.41$. In practice, we can always expect the number of cycles in an l -Othello to be less than 2 with probability 97%, and smaller than 9 with probability higher than 99.999%.

2.2.2 Time and space complexity for l -Othello construction and query

Construction Complexity: The construction of the l -Othello data structure generally follows a depth-first traversal of the bipartite graph; thus the complexity is $O(n)$, where n is the total number of k -mers. The memory complexity is $O(ln)$, where l is the number of bits to encode the number of categories. We can further reduce the memory cost by dividing the set of k -mers into smaller groups based on prefixes, commonly of length 3, corresponding to $g = 64$ groups. Each group contains approximately $\frac{n}{g}$ k -mers. For each group, we build an l -Othello, using time $O\left(\frac{n}{g}\right)$ with memory cost $O\left(\frac{n}{g}\right)$. In total, constructing g l -Othello structures still takes $O(n)$ time.

Query Complexity: For each query $\tau(s)$, l -Othello computes two hash values $h_a(s)$ and $h_b(s)$, and accesses two memory locations $A[b_a(s)]$ and $B[b_b(s)]$. The time complexity is $O(1)$. This procedure only includes a few basic arithmetic operations, resulting in an extremely fast execution speed.

2.2.3 On alien k -mers

An alien k -mer is defined as a k -mer that is not included during the construction of an l -Othello. In the context of taxonomic classification, they are those k -mers that are not included in any of the reference materials. Alien k -mers often arise due to noise or genomic sequences belonging to a novel species sampled by a sequence read. We have designed two strategies to detect alien k -mers. First, we would like to increase the randomness of assignments in the case of alien k -mers; secondly, we may add another bit in the bitmap (i.e. let $l > \log [|T|]$), doubling the number of categories, so that an alien k -mer has a much higher chance of being assigned to alien categories (i.e., categories not used by existing taxon labels).

Here we show how we may leverage the randomness of alien assignment to predict an alien k -mer within the l -Othello itself. We first discuss the query result for $l = 1$, and then we extend it to l -Othello. When $l = 1$, Othello classifies k -mers ($s \in S = S_0 \cup S_1$) into S_0 and S_1 . Each element in A or B is a 1-bit value. For a query of an alien k -mer $s' \notin S$, l -Othello still returns a value $\tau(s') \in \{0, 1\}$. For alien keys, $\tau(s') = A[b_a(s')] \oplus B[b_b(s')]$. Let a_0 and a_1 be the fraction of 0s and 1s in the bitmaps A respectively, i.e. $a_0 = \frac{|t|A[t]=0|}{m_a}$, $a_1 = \frac{|t|A[t]=1|}{m_a}$. Similarly b_0 and b_1 are the fractions in B . Suppose h_a and h_b are uniformly distributed random hash functions, and s' is an arbitrary k -mer in the universal set; then $\tau(s')$ returns 1 with probability $p_1 = a_0b_1 + a_1b_0$. Similarly, $\tau(s')$ returns 0 with probability $p_0 = a_0b_0 + a_1b_1$.

For l -Othello, a similar property also holds. Let $p(t)$ be the probability that the query of an alien k -mer returns exactly t . $\tau(s') = t$ indicates $A[b_a(s')] \oplus B[b_b(s')] = t$. Note that h_a and h_b are uniform random hash functions and are not correlated. Hence,

$$p(t) = P(\tau(s') = t) = \sum_{x=0}^{2^l-1} a_x b_{x \oplus t}$$

where a_x is the fraction of elements has value x in a and $b_{x \oplus t}$ is the fraction of elements has value $x \oplus t$ in B .

Given a particular l -Othello, we can always compute $p(t)$ values for all $t = 0, 1, \dots, 2^l - 1$ using time $O(2^{2^l} + n)$. These $p(t)$ values are affected by the occurrence frequency of each l -bit integer, namely a_x and b_x for all $0 \leq x < 2^l$. In some cases, these values are not uniformly distributed, which may result in imbalance among $p(t)$. Under such circumstances, we can tune these values by flipping the bitmaps of a connected component in the bipartite graph without changing $\tau(s)$ for $s \in S$. In practice, we can always tune the values so that $p(t)$ is of the same order of magnitude for all t , and all of them are approximately 2^{-l} . Two tuning approaches are described in Supplementary Section S2.

We can also explicitly detect alien k -mers by increasing l , thus intentionally expanding the number of targeted categories, where the majority of them are dummy (alien). Due to the randomness of class assignment in the presence of an alien, many alien k -mers are likely to fall in these dummy categories, and are thus recognized as alien. Formally, if for some s^* , $\tau(s^*) \geq |T|$, s^* is an alien. Thus, alien k -mers are recognized in this stage with probability $\sum_{x=|T|}^{2^l-1} p(x) \sim \frac{2^l - |T|}{2^l}$.

2.3 Taxonomic classification of sequencing reads

As illustrated in Figure 1B, given any sequencing read, our algorithm iterates over each k -mer from the beginning of the read and, for each k -mer, retrieves the taxon to which it is specific using l -Othello. Taxonomic classification of the read is determined by assembling the taxa for all k -mers in the read. The classification is straightforward when all k -mers indicate the same taxon, but this is not often the case. Disparate taxa are considered to be consistent if they belong to the same path in the taxonomy, meaning that one assignment is the higher rank of the other. When these taxa belong to different branches, they represent conflicting information. The issue is further complicated by the possibility of false taxonomic information returned from querying alien k -mers, where the k -mer in the read does not appear in any of the reference sequences.

To tackle this challenge, we have designed a window-based classification approach. A window is defined as a sequence of consecutive k -mers that are assigned to the same taxon of a given level. The window-based approach guards against false-positive assignments due to alien k -mers. Assuming that the taxon ID returned by an alien k -mer is random, the chance of having two consecutive alien k -mers return the same taxon ID is

$$\sum_{t=0}^{2^l-1} \left(p(t) \right)^2 \sim 2^{-l}.$$

This value is very small, regardless of k . Additionally, each window corresponds to a maximum read subsequence that matches the reference sequences. Thus, the longer the window, the longer the subsequence match, and the less likely the match is random. In comparison, other algorithms such as Kraken and Clark count the total number of k -mer matches, regardless of their spatial distribution across the read.

If multiple taxon windows are available, MetaOthello scores each of them using the summed squares of window sizes as in the following formula; the taxon with the maximum score will be selected:

$$Score(t) = \sum (u_i^t)^2,$$

where u_i^t denotes the number of k -mers in the i_{th} window classified to taxon t .

A k -mer signature belonging to a taxon is also specific to its higher-ranking taxa, so at higher taxonomic ranks, there exist more k -mers to distinguish a taxon from its siblings. Thus, longer k -mer windows and more-accurate classifications are expected at higher taxonomic ranks. Under this assumption, a ‘top-down’ strategy is adopted during read classification. Given a read sequence, MetaOthello starts the classification at the top rank and continues the classification down the ranks until there does not exist a sufficiently large k -mer window supporting the level. Based on the k -mer distribution in each taxon, MetaOthello establishes a threshold on minimum window size when the classification on that taxon requires. Theorem 2 shows that the minimum window-size threshold can be precomputed for each taxon prior to read classification. The minimum window size required for a taxon is determined by the probability of an alien k -mer query on l -Othello returning a taxon rooted in t and the acceptable false-positive rate. The larger the size of the taxon subtree, the higher the probability that a random alien k -mer may match to t and thus the longer the window required for reliable classification. Additionally, a larger window size will be required in order to lower the false-positive rate.

THEOREM 2. Given a user-defined false-positive rate λ and the total read number M , the minimum window-size threshold required for a taxon t can be computed as $\log_{p(t)} \frac{\lambda}{(1-\lambda)M}$, where p_t denotes the probability that an alien k -mer query on l -Othello returns a value in the taxon subtree with root t .

The proof is presented in Supplementary Section S3. For example, when t is a genus-level node, supposing $l=12$, then $p(t) \sim (1+7)2^{-l} = \frac{1}{256}$. Given 10 million reads and suppose $\lambda = 0.001$, then $\log_{p(t)} \frac{\lambda}{(1-\lambda)M} = 3.42$, and only windows larger than three will be taken into consideration when determining the read assignment.

3 Experimentation and evaluation

3.1 Classification accuracy and the relative abundance of taxon-specific k -mers

Accurate classification of a read to a taxon is largely dependent upon the presence of k -mer signatures. Thus the abundance of these signature k -mers (i.e. the proportion of taxon-specific k -mers among all k -mers present in the reference sequences for the taxon) becomes an important indicator of the capability of our algorithm. Thus we first investigate the correlation between classification accuracy and the relative abundance of taxon-specific k -mers.

Classification accuracy is computed as the fraction of reads assigned correctly. Using a next-generation sequencing (NGS) read simulator called ART (Huang *et al.*, 2012), we simulated 10 000 reads for each of 2629 reference genomes in the NCBI RefSeq bacterial genome database, for a total of 26 290 000 reads. The database is available at ftp://ftp.ncbi.nih.gov/genomes/archive/old_refseq/Bacteria/. Each read is paired-end and of length 100 bp with a fragment size of 250 bp, generated using the default error profile for the HiSeq platform. Figure 3 shows the read-classification accuracy for each species as a function of species-specific k -mer proportion, where read assignments were generated by the MetaOthello algorithm using 20-mers or 31-mers, respectively. The scatter plot for either k -mer size demonstrates that the vast majority of all species manifest more than 50% 20-mers that are species-specific, and almost all species have 75% species-specific 31-mers. Although in general more species-specific k -mers afford better classification accuracy, these results suggest that the presence of 50% or more species-specific k -mers affords suitably high classification accuracy,

thereby demonstrating the utility of k -mer signatures in classifying metagenomic reads.

To investigate how the window-based approach improves MetaOthello’s performance over the widely adopted count-based approach as in Kraken and Clark, we implemented a count-based version of MetaOthello, and compared its performance against that of the window-based version executed on the same datasets. The results demonstrate the clear advantages of a window-based approach over count-based, especially on assignment precision. This suggests that the window-based approach is more effective at eliminating the false positives caused by alien k -mers. Detailed results are reported in Supplementary Section S5.

3.2 Comparison with state-of-the-art tools

We now assess the performance of MetaOthello in comparison to three state-of-the-art tools: Kraken (version 0.10.5 beta), Clark (version 1.2.3) and Kaiju (version 1.4.4). Besides the newly published tool Kaiju, Kraken and Clark were chosen based on the recommendation of a recent benchmarking paper (Lindgreen *et al.*, 2016), which evaluated 14 tools using six datasets and subsequently declared Kraken and Clark the best performers over Genometa (Davenport *et al.*, 2012), GOTCHA (Freitas *et al.*, 2015), LMAT (Ames *et al.*, 2013), MEGAN (Huson *et al.*, 2007, 2011), MG-RAST (Meyer *et al.*, 2008), the One Codex webserver, taxator-*tk* (Dröge *et al.*, 2015), MetaPhlAn (Segata *et al.*, 2012), MetaPhyler (Liu *et al.*, 2010), mOTU (Sunagawa *et al.*, 2013) and QIIME (Caporaso *et al.*, 2010). The comparison was benchmarked against three publicly available datasets: HiSeq, MiSeq and SimBA5. The same datasets have been used multiple times to evaluate a number of metagenomic classification tools, including Kraken in previous studies (Wood and Salzberg, 2014). All tools were executed using the same reference database (NCBI RefSeq as of October 1st, 2016), and all other parameters follow the default settings.

3.2.1 Classification accuracy

We first compare classification accuracy. Three different k -mer lengths (20-mer, 25-mer and 31-mer) are used to assess the performance relationship with k -mer size for Kraken, Clark and MetaOthello; Kaiju is not a k -mer-based algorithm. To facilitate the comparison and to mimic the sequencing data generated by current platforms, we discarded reads shorter than 36 bp and those whose taxon is not included in the reference taxonomy.

Reads were classified by each algorithm at three taxonomic levels: phylum, genus and species. MetaOthello, Kraken and Kaiju were able to classify reads at the three levels simultaneously, while Clark required three separate runs to conduct similar classifications. Thus for Clark, results from these runs were merged for the purpose of direct comparison. Precision and sensitivity were computed at each of the three classification levels. Precision is defined as the ratio between correctly assigned reads and the total number of reads in an assignment; sensitivity is calculated as the fraction of total reads assigned correctly. F-score (i.e. the harmonic mean of precision and sensitivity) was also calculated to quantify the balance between these two metrics. Results of the comparison are shown in Table 1.

In general, longer k -mers enhance the precision of read classification but decrease sensitivity, as observed in MetaOthello, Kraken and Clark. Within each dataset, the overall winner (in bold) was the one with highest F1-score when considering across all three k -mer sizes. In phylum-level classification, MetaOthello outperforms the other algorithms in all three datasets all using 20-mers. At both genus and species levels, MetaOthello exhibits the best performance

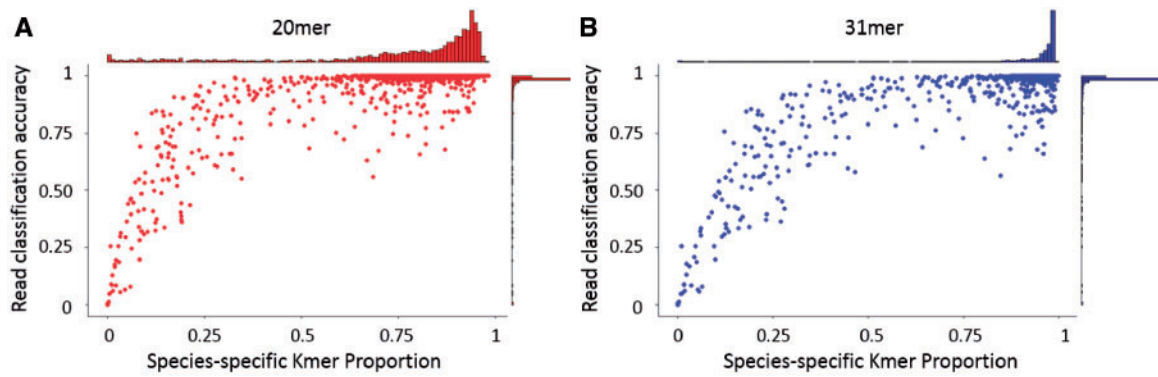


Fig. 3. Correlation between species-specific k -mer signatures and classification accuracy when $k=20$ (A) and $k=31$ (B). In each panel, the central figure depicts the correlation between species-specific k -mer proportion and read-classification accuracy for all species; the top histogram shows the distribution of species as a function of species-specific k -mer proportion, and the right histogram shows the distribution of classification accuracy for all species

Table 1. Accuracy of read taxonomic classification in terms of precision, sensitivity and F-score

			Phylum	Genus	Species	
			Prec/Sens/F-score	Prec/Sens/F-score	Prec/Sens/F-score	
HiSeq	MetaOthello	20mer	98.4/95.0/967	97.2/92.5/948	82.0/69.4/751	
		25mer	99.4/92.2/957	99.1/91.2/950	84.2/69.1/760	
		31mer	99.4/89.0/939	99.3/88.2/934	85.7/68.0/758	
	Kraken	20mer	97.8/94.8/963	96.1/92.0/940	80.2/69.2/743	
		25mer	99.7/92.3/959	99.1/91.4/951	83.7/69.4/759	
		31mer	99.7/88.3/937	99.3/87.6/931	85.4/67.6/745	
	Clark	20mer	97.7/95.5/966	95.1/92.6/939	76.4/69.5/728	
		25mer	99.7/92.1/958	99.1/91.2/950	83.5/69.2/757	
		31mer	99.7/88.8/940	99.3/88.1/934	85.4/68.0/757	
	MiSeq	Kaiju	20mer	99.4/68.7/812	98.6/65.1/785	89.2/34.7/499
			25mer	99.2/97.5/983	96.2/92.2/942	91.8/78.6/846
			31mer	99.6/95.4/975	97.4/91.4/943	93.0/78.3/850
MetaOthello		20mer	99.6/92.9/961	98.0/89.7/937	93.8/77.2/847	
		25mer	99.0/97.5/983	95.8/92.2/939	91.0/78.9/845	
		31mer	99.8/95.1/974	97.4/91.2/942	92.7/78.3/849	
Kraken		20mer	99.9/92.3/960	98.0/89.3/935	93.6/76.8/844	
		25mer	98.8/97.8/983	94.4/92.5/934	86.9/78.8/826	
		31mer	99.8/95.2/975	97.1/91.5/942	91.9/78.5/847	
Clark		20mer	99.9/92.7/962	98.0/89.8/937	93.4/77.3/846	
		25mer	99.5/75.7/860	98.5/68.0/805	95.2/40.6/570	
		31mer	99.9/99.7/998	99.6/95.8/977	99.3/84.2/911	
simBA5	MetaOthello	20mer	99.9/98.2/990	99.8/94.6/971	99.5/83.1/906	
		25mer	99.5/92.2/957	99.5/88.7/938	99.4/77.9/873	
		31mer	99.8/99.5/996	99.4/95.9/976	98.8/84.6/912	
	Kraken	20mer	99.9/98.5/992	99.8/95.0/974	99.5/83.8/909	
		25mer	99.9/94.2/970	99.9/90.9/952	99.7/80.0/887	
		31mer	99.8/99.6/997	98.5/95.8/971	94.4/84.2/890	
	Clark	20mer	99.9/98.4/992	99.8/94.8/973	99.4/83.4/907	
		25mer	99.9/93.5/966	99.9/90.2/948	99.7/79.2/883	
		31mer	99.6/75.6/860	97.9/65.9/788	96.5/46.7/630	

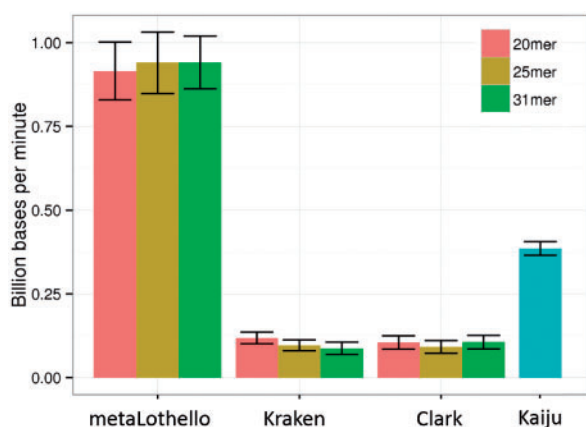
on two out of the three datasets using either 25-mers or 20-mers. Kraken performs the best in the remaining comparisons, followed closely by MetaOthello in both cases. In general, Kaiju delivered much lower (20–30%) sensitivity compared to the other three tools, due to its lack of capability in classifying reads from non-protein coding regions. We also ran Kaiju using two additional databases, *nr* and *proGenomes*, as recommended in Kaiju's manual. Although higher sensitivities were achieved in some datasets at some taxonomic levels, the overall accuracy is still considerably lower than the other algorithms. Detailed results of the two additional runs are reported in Supplementary Section S6.

3.2.2 Runtime and memory

Speed benchmarks were performed using the servers from Lipscomb High-Performance Computing at the University of Kentucky. The servers are equipped with Dell R820, Quad Intel E5-4640 8-core (Sandy Bridge) @ 2.4 GHz and 512 GB/node of 1600 Mhz RAM. Each algorithm was executed using eight threads and k -mer lengths as specified previously; all other parameters follow the default settings. The speed for each tool is presented in Figure 4. In general, MetaOthello achieved the highest processing speed, clocking roughly 1 billion bases per minute. This figure represents an order-of-magnitude improvement over Kraken and Clark, the two most-

Table 2. The proportion of reads classified into the top-five genera by each algorithm using different k -mer lengths

k -mer length	MetaOthello			Kraken			Clark			Kaiju
	20	25	31	20	25	31	20	25	31	
Streptococcus	14.32	13.24	12.12	14.58	13.29	12.18	14.25	13.15	12.02	10.05
Haemophilus	6.999	6.626	5.938	7.089	7.134	6.005	6.952	6.411	5.894	4.773
Prevotella	5.621	4.893	4.226	5.774	5.012	4.270	5.590	4.567	4.193	7.653
Veillonella	3.321	2.668	1.924	3.441	2.931	1.976	3.231	2.587	1.891	5.168
Neisseria	2.215	1.860	1.525	2.279	1.941	1.552	2.213	1.779	1.510	3.113

**Fig. 4.** Performance comparison in terms of billion bases processed per minute using different k -mer lengths

rapid state-of-the-art tools within the category of alignment-free classifiers. Impressively, the high speed does not entail a compromise in the memory requirement. MetaOthello only consumes about one-third (peak memory 27 GB) of the RAM required by Kraken and Clark (peak memory 73 GB).

The construction of the MetaOthello index from the NCBI RefSeq bacterial genome sequence database requires roughly 6 hours with peak memory usage up to 40 GB using 16 threads. In contrast, Kraken and Clark used 164 GB and 120 GB respectively for index construction, but both finished under 4 hours with 16 threads.

In summary, MetaOthello achieves a significant speedup with much smaller memory footprint in comparison with Kraken and Clark while delivering competitive or even superior performance in classification accuracy. While Kaiju is relatively scalable, it suffers from low sensitivity in classification.

3.3 Metagenomic classification of real datasets

3.3.1 Human Microbiome Project data

To assess the performance on real datasets of MetaOthello relative to Kraken, Clark and Kaiju, the three algorithms were run on sequencing data from three saliva samples (NCBI SRA accessions: SRS015055, SRS019120 and SRS014468) used in the Human Microbiome Project (Human and Project, 2012). We ran the three k -mer-based algorithms at each of three different k -mer length settings (20-mer, 25-mer and 31-mer) as with the simulated data. The three samples were analyzed separately, and the results were pooled together to assess the relative abundances of species. The top five most-abundant genera are presented in Table 2. The four tools reported the same five most-abundant genera: Streptococcus, Haemophilus, Prevotella, Veillonella and Neisseria, all of which are known to be associated with human saliva. Interestingly, although the absolute abundance (i.e. the fraction of total reads assigned to a

given genus) varies with k -mer size, the relative abundances remain stable except for Kaiju. The false-positive rate, however, cannot be assessed in this case.

4 Conclusion and discussion

In this paper, we present MetaOthello, a novel metagenomic sequencing read classifier. MetaOthello leverages a novel probabilistic hashing structure, l -Othello, to conduct taxonomic classification using taxon-specific k -mer signatures. The algorithm delivers ultra-fast and memory-efficient solutions to k -mer-based taxonomic classification. Within the set of alignment-free approaches, MetaOthello achieves an order-of-magnitude improvement in classification speed relative to the fastest algorithms, Kraken and Clark, while reducing the RAM requirement from 73 GB to 27 GB. MetaOthello exhibits high sensitivity and precision competitive with Kraken and Clark, and in most cases achieves a better balance between the sensitivity and specificity (as quantified by F-score). It is also three times faster than the protein alignment-based method Kaiju and delivers much higher classification accuracy.

Besides the application of metagenomics, we also expect that our data structure l -Othello may benefit other k -mer-based sequencing-matching methods with its advantages in memory and speed efficiency.

Acknowledgements

This work was previously submitted to and accepted by The Seventh RECOMB Satellite Workshop on Massively Parallel Sequencing (RECOMB-Seq 2017). We thank the reviewers for their valuable comments and suggestions.

Funding

This work was supported by National Science Foundation [CAREER award grant number 1054631 to J.L.; grant CNS-1701681 to C.Q.]; and the National Institutes of Health [grant number P30CA177558 and 5R01HG006272-03 to J.L.].

Conflict of Interest: none declared.

References

- Altschul, S.F. *et al.* (1990) Basic local alignment search tool. *J. Mol. Biol.*, **215**, 403–410.
- Ames, S.K. *et al.* (2013) Scalable metagenomic taxonomy classification using a reference genome database. *Bioinformatics (Oxford, England)*, **29**, 2253–2260.
- Brady, A. and Salzberg, S.L. (2009) Phymm and PhymmBL: metagenomic phylogenetic classification with interpolated Markov models. *Nat. Methods*, **6**, 673–676.
- Caporaso, J.G. *et al.* (2010) QIIME allows analysis of high-throughput community sequencing data. *Nat. Methods*, **7**, 335–336.

- Davenport, C.F. et al. (2012) Genometa – a fast and accurate classifier for short metagenomic shotgun reads. *PLoS ONE*, **7**, e41224.
- Dröge, J. et al. (2015) Taxator-tk: Precise taxonomic assignment of metagenomes by fast approximation of evolutionary neighborhoods. *Bioinformatics*, **31**, 817–824.
- Freitas, T.A.K. et al. (2015) Accurate read-based metagenome characterization using a hierarchical suite of unique signatures. *Nucleic Acids Res.*, gkv180.
- Huang, W. et al. (2012) ART: A next-generation sequencing read simulator. *Bioinformatics*, **28**, 593–594.
- Human, T. and Project, M. (2012) A framework for human microbiome research. *Nature*, **486**, 215–221.
- Huson, D.H. et al. (2007) MEGAN analysis of metagenomic data. *Genome Res.*, **17**, 377–386.
- Huson, D.H. et al. (2011) Integrative analysis of environmental sequences using MEGAN4. *Genome Res.*, **21**, 1552–1560.
- Huttenhower, C. and Human Microbiome Project Consortium (2012) Structure, function and diversity of the healthy human microbiome. *Nature*, **486**, 207–214.
- Kim, D. et al. (2016) Centrifuge: rapid and sensitive classification of metagenomic sequences. *bioRxiv*, (054965).
- Lindgreen, S. et al. (2016) An evaluation of the accuracy and speed of metagenome analysis tools. *Sci. Rep.*, **6**, 19233.
- Liu, B. et al. (2010) MetaPhyler: Taxonomic profiling for metagenomic sequences. In: *Proceedings – 2010 IEEE International Conference on Bioinformatics and Biomedicine, BIBM 2010*, pp. 95–100.
- Menzel, P. and Krogh, A. (2015) Kaiju: fast and sensitive taxonomic classification for metagenomics. *bioRxiv*, **7**, 1–9.
- Meyer, F. et al. (2008) The metagenomics RAST server a public resource for the automatic phylogenetic and functional analysis of metagenomes. *BMC Bioinformatics*, **9**, 386.
- Ounit, R. and Lonardi, S. (2016) Higher classification sensitivity of short metagenomic reads with CLARK-S. *Bioinformatics*, **32**, 3823–3825.
- Ounit, R. et al. (2015) CLARK: fast and accurate classification of metagenomic and genomic sequences using discriminative k-mers. *BMC Genomics*, **16**, 236.
- Rosen, G.L. et al. (2011) NBC: the Naive Bayes Classification tool webserver for taxonomic classification of metagenomic reads. *Bioinformatics (Oxford, England)*, **27**, 127–129.
- Segata, N. et al. (2012) Metagenomic microbial community profiling using unique clade-specific marker genes. *Nat. Methods*, **9**, 811–814.
- Sunagawa, S. et al. (2013) Metagenomic species profiling using universal phylogenetic marker genes. *Nat. Methods*, **10**, 1196–1199.
- Tu, Q. et al. (2014) Strain/species identification in metagenomes using genome-specific markers. *Nucleic Acids Res.*, **42**, e67–e67.
- Tyson, G.W. et al. (2004) Community structure and metabolism through reconstruction of microbial genomes from the environment. *Nature*, **428**, 37–43.
- Venter, J.C. et al. (2004) Environmental genome shotgun sequencing of the Sargasso Sea. *Science (New York, N.Y.)*, **304**, 66–74.
- Wood, D.E. and Salzberg, S.L. (2014) Kraken: ultrafast metagenomic sequence classification using exact alignments. *Genome Biol.*, **15**, R46.
- Yu, Y. et al. (2016) A Concise Forwarding Information Base for Scalable and Fast Flat Name Switching. *arXiv*, (1608.05699).