

VKORC1 and VKORC1L1 have distinctly different oral anticoagulant dose-response characteristics and binding sites

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Key Points

- VKORC1 is more sensitive than VKORC1L1 to OAC inhibition, whereby 4-hydroxycoumarin rodenticides are equally effective.
- In silico and in vitro analysis revealed OAC binding sites that are different for VKORC1 and VKORC1L1.

Vitamin K reduction is catalyzed by 2 enzymes in vitro: the vitamin K 2,3-epoxide reductase complex subunit 1 (VKORC1) and its isozyme VKORC1-like1 (VKORC1L1). In vivo, VKORC1 reduces vitamin K to sustain γ -carboxylation of vitamin K-dependent proteins, including coagulation factors. Inhibition of VKORC1 by oral anticoagulants (OACs) is clinically used in therapy and in prevention of thrombosis. However, OACs also inhibit VKORC1L1, which was previously shown to play a role in intracellular redox homeostasis in vitro. Here, we report data for the first time on specific inhibition of both VKOR enzymes for various OACs and rodenticides examined in a cell-based assay. Effects on endogenous VKORC1 and VKORC1L1 were independently investigated in genetically engineered HEK 293T cells that were knocked out for the respective genes by CRISPR/Cas9 technology. In general, dose-responses for 4-hydroxycoumarins and 1,3-indandiones were enzyme-dependent, with lower susceptibility for VKORC1L1 compared with VKORC1. In contrast, rodenticides exhibited nearly identical dose-responses for both enzymes. To explain the distinct inhibition pattern, we performed in silico modeling suggesting different warfarin binding sites for VKORC1 and VKORC1L1. We identified arginine residues at positions 38, 42, and 68 in the endoplasmic reticulum luminal loop of VKORC1L1 responsible for charge-stabilized warfarin binding, resulting in a binding pocket that is diametrically opposite to that of VKORC1. In conclusion, our findings provide insight into structural and molecular drug binding on VKORC1, and especially on VKORC1L1.

Introduction

In the 1940s, dicoumarol, a naturally occurring 4-hydroxycoumarin in fodder contaminated with spoiled clover, was identified as the cause of spontaneous hemorrhage in livestock.¹ Further homologs were synthesized and characterized by Link and colleagues for intended use as rodenticides and, later, as human oral anticoagulants (OACs).² Since the 1950s, OACs have been frequently prescribed to treat and prevent thromboembolism. Warfarin is the oral anticoagulant drug of choice in the United States, acenocoumarol in southern Europe and Asia, and phenprocoumon in Central Europe.³ Fluindione, a substituted 1,3-indandione, is used frequently in French-speaking countries.⁴ Dosing and efficacy of OACs depend on the specific compound and patient's pharmacogenetic and pharmacokinetic profile.⁵

4-Hydroxycoumarin-based OACs and rodenticides, as well as indandiones, directly inhibit vitamin K 2,3-epoxide reductase complex subunit 1 (VKORC1).⁶⁻⁸ VKORC1 catalyzes the reduction of vitamin K 2,3-epoxide (K>O) and vitamin K quinone (K) to vitamin K hydroquinone (KH₂), which is essential

to support γ -carboxylation, and thereby activation of vitamin K-dependent (VKD) proteins (eg, coagulation factor IX, FIX).⁹ Therefore, inhibition of VKORC1 limits the availability of KH₂, leading to reduced posttranslational modifications of VKD clotting factors.^{10,11} The isozyme of VKORC1, VKORC1-like 1 (VKORC1L1), possesses the same enzymatic reactions as VKORC1 in vitro.¹²⁻¹⁴ Although VKORC1 is essential for blood clotting, it was shown that VKORC1L1 cannot substitute for this function, and likely has other biological roles.¹⁴⁻¹⁶ Recent studies suggest that the unique function of VKORC1 may be a result of its specific tissue distribution and high expression in hepatocytes where VKD clotting factors are synthesized.^{12,17,18} Both enzymes were found to exhibit different affinities with regard to warfarin.^{14,17} However, detailed knowledge of the molecular mechanism and structural site of OAC action for VKORC1L1 are lacking.

In this study, we established *VKORC1* and *VKORC1L1* knockout (KO) HEK 293T cell lines to investigate specific inhibition of endogenously expressed VKORC1L1 and VKORC1, respectively, by various OACs and rodenticides. We used in silico modeling and docking to investigate OAC drug binding and verified putative interacting residues for VKORC1L1. Our findings demonstrate enzyme-specific differences in warfarin binding for VKORC1 and VKORC1L1 that are consistent with lower OAC sensitivity for VKORC1L1.

Material and methods

Generation of VKORC1 and VKORC1L1 single- and double-KO HEK 293T cell lines

HEK 293T cell lines with single and double gene KOs for VKORC1 and VKORC1L1 were generated using CRISPR/Cas9-based gene editing.¹⁹ Guide RNA target sequence was GCTCTACGCGCTGCACGTGAAGG for VKORC1 and CTTCTCCCGCTCCACGTGGTAGG for VKORC1L1. HEK 293T cells were maintained in Dulbecco's modified Eagle medium (Thermo Fisher Scientific, Dreieich, Germany) containing 1% vol/vol penicillin/streptomycin, 1 \times nonessential amino acids (Thermo Fisher Scientific) and 10% vol/vol fetal bovine serum (PAN, Aidenbach, Germany). Cells were seeded (25 000 cells per 96-well plate) and transfected with 200 ng plasmid containing mCherry Cas9 and a U6 promoter-driven guide RNA against VKORC1 or VKORC1L1 with Lipofectamine 2000 (Thermo Fisher Scientific). Three days after transfection, cells were seeded under limiting conditions to obtain single-cell clones. Single-cell clones were expanded for maintenance and analysis. For next-generation sequencing (Illumina, MiSeq, San Diego, CA), ~200 different single-cell clones were lysed in 100 μ L lysis buffer (0.2 mg/mL proteinase K, 1 mM CaCl₂, 3 mM MgCl₂, 1 mM EDTA, 1% Triton X-100, 10 mM Tris at pH 7.5) for genomic DNA preparation. Dual polymerase chain reaction barcoding and deep sequencing were performed as described by Schmidt et al.²⁰ Deep sequencing data were evaluated using the OutKnocker.org web tool, and positive clones were verified by next-generation sequencing deep sequencing, as described earlier.^{21,22}

Mammalian expression vectors

For solely human F9 expression, a pCMV6-XL4 vector was used (Origene, Rockville, MD). The pIRES vectors harboring

cDNA of human VKORC1 (hVKORC1) or human VKORC1L1 (hVKORC1L1), together with human F9, were generated as previously described.^{23,24} The VKORC1 loop swap construct was created by replacing amino acids Lys30 to Ser79 of VKORC1 with Glu37 to Pro86 of VKORC1L1, and vice versa. Variants of VKORC1L1 were generated by site-directed mutagenesis. For expression analysis, VKOR variants were cloned into pcDNA3.1/myc-His (Thermo Fisher Scientific).

Determination of vitamin K reductase and VKOR activity and IC₅₀ values

Wild-type (WT), VKORC1 KO and VKORC1L1 KO HEK 293T cells were transfected with F9 cDNA to assess endogenous vitamin K reductase (VKR) and vitamin K 2,3-epoxide reductase (VKOR) activity of VKORC1L1 and VKORC1. Double-KO (DKO) HEK 293T cells were transfected with pIRES vectors containing cDNAs of F9 together with VKOR variants (or cotransfected with F9 [pCMV6-XL4] and VKOR myc-tagged variants [pcDNA3.1/myc-His]). After 4 hours, transfection medium was replaced by expression medium (OptiMEM [Thermo Fisher Scientific], including 0.25% BSA [protease-free; Sigma-Aldrich, Munich, Germany] and 2.5 mM CaCl₂ [Sigma-Aldrich]) supplemented with 12 μ M vitamin K₁ quinone (K₁; in dimethyl sulfoxide, Sigma-Aldrich) or 12 μ M vitamin K₁ 2,3-epoxide (K₁>O; in ethanol, Sigma-Aldrich). For determination of half-maximum inhibitory concentrations (IC₅₀), various concentrations of 4-hydroxycoumarins or 1,3-indandiones were added. Stock solutions of acenocoumarol (Cayman, Ann Arbor, MI), warfarin, coumatetralyl, coumachlor, phenindione, coumarin, brodifacoum, bromadiolone (Sigma-Aldrich), dicoumarol (Merck, Darmstadt, Germany), flutidione (BOC Sciences, New York, NY), phenprocoumon (Roche Diagnostics, Mannheim, Germany), 4-hydroxycoumarin (Dr. Ehrenstorfer GmbH, Wesel, Germany), and ferulenol (Santa Cruz Biotechnology, Heidelberg, Germany) were prepared in ethanol. Seventy-two hours after transfection, medium was collected and concentrated (VivaSpin 6; 10-kDa cutoff; Sartorius, Göttingen, Germany), and FIX activity was assessed on the basis of an aPTT test, using the coagulation analyzer BCS XP (Siemens Healthcare, Erlangen, Germany). The FIX activity was normalized to negative control (ie, cells secreting FIX in the absence of OACs), and activated FIX (FIXa) values were plotted against OAC concentration. Statistical analyses were performed using GraphPad Prism 5.01 (GraphPad Software Inc., La Jolla, CA) to impose nonlinear regression dose-response curves with variable slope.

Determination of expression level of overexpressed VKOR variants

For quantification of overexpressed VKOR variants, HEK 293T cells were transfected with myc-tagged VKOR variants, using Lipofectamine 2000. Forty-eight hours after transfection, cells were harvested using lysis buffer (150 mM NaCl, 1% NP-40, 50 mM Tris-HCl at pH 8.0) containing protease inhibitor (cOmplete mini, EDTA-free, Roche Diagnostics). Expression was detected by western blot, using an anti-c-myc- antibody (M4439, Sigma-Aldrich), including a loading control (ERGiC-53, sc-271517, Santa Cruz Biotechnology, Heidelberg, Germany). The detection antibody was a goat anti-mouse HPR-conjugated antibody in both reactions (sc-2005, Santa Cruz Biotechnology).

Molecular modeling of human VKORC1 and VKORC1L1 with comparative dockings of OACs

We constructed hVKORC1L1 homology models based on the 3.6-Å resolution X-ray crystallographic structure of the prokaryotic (*Synechococcus* sp.) homolog of VKORC1 (PDB ID: 3KP9), using the I-TASSER automated modeling server (<https://zhanglab.cmb.med.umich.edu/I-TASSER/>).^{25,26} Of 5 generated models, the best one was chosen on the basis of the calculated C-score of -0.46 . Stereochemical quality of the model and template structures were assessed and compared, using the MOLPROBITY server (<http://molprobity.biochem.duke.edu/>). Comparison of backbone C $_{\alpha}$ alignment between the model and template structures was assessed using the Mustang function embedded in the YASARA molecular modeling software.²⁷ Visualization, graphical rendering, and structural analysis were performed using YASARA and Chimera software.^{27,28}

We used the automated AutoDock module (The Scripps Research Institute) with default settings for small-molecule docking embedded in YASARA (ver. 12.9.27; Elmar Krieger) to search for predicted warfarin binding poses on the hVKORC1L1 model.^{27,29} Structure data format files (.sdf) for the ligands: acenocoumarol (CID 54676537), bromadiolone (CID 547539), brodifacoum (CID 54680676), coumachlor (CID 54682651), coumatetralyl (CID 54678504), dicoumarol (CID 54676038), fluindione (CID 68942), phenindione (CID 4760), phenprocoumon (CID 54680692), and S-warfarin (CID 54688261) were downloaded from the Pubchem database (<https://pubchem.ncbi.nlm.nih.gov/search/>) and converted to PDB format (.pdb), using the AUTOSMILES function of YASARA. A course-screened rigid rolling ball global search was performed, allowing the energy-optimized, rigid warfarin DCO isomer configuration (deprotonated, open side-chain) to move relative to the fixed, rigid hVKORC1L1 model. In addition, 10 OAC structures were flexibly docked on the hVKORC1L1, as well as on a previously generated hVKORC1 model.²⁴ Representative docked poses on hVKORC1L1 were selected on the basis of lowest pseudo-energy function scores and the relative orientation with respect to predicted physiological positively charged arginine residues on hVKORC1L1. Docking poses on hVKORC1 for all inhibitors except warfarin were chosen on the basis of their similarity to a published warfarin docking pose identified for hVKORC1.²⁴ All selected docking poses were subjected to 100 ns membrane-embedded simulation with a membrane composition of 70% PC (phosphatidylcholine) and 30% (phosphatidylethanolamine). The docked complexes with the lowest energy in the simulation trajectory were chosen as the final docked structures and analyzed for identifying putative binding residues.

Measurement of cell viability

The influence of oxidative stress was assessed using CellTiter 96 AQueous One Solution Cell Proliferation Assay (Promega, Mannheim, Germany). Thereby, a tetrazolium compound is converted exclusively by viable cells into formazan, which is measured at 490 nm. WT, VKORC1 KO, and VKORC1L1 KO and DKO HEK 293T cells were seeded on 96-well plate (5×10^5 cells/mL). Cells were incubated with different concentrations of H₂O₂ (0-250 μ M) or 75 μ M H₂O₂ combined without K₁ or K₁>O (12 μ M). After 18 hours, 15 μ L of reagent was added and incubated for 2 to 4 hours, and absorbance was measured at 490 nm (Synergy 2; BioTek Germany, Bad Friedrichshall, Germany).

Results

Characterization of VKORC1 and VKORC1L1 single-KO and DKO HEK cell lines

Figure 1A summarizes the genetic profile of VKORC1 and VKORC1L1 single-KO and DKO cell lines. Both single-KO cell lines showed similar FIX activities (Figure 1B) compared with WT HEK 293T cells, whereas the DKO cell line exhibited no FIX activity when transfected with F9 cDNA solely and supplemented with 12 μ M K₁. Thus, single-KO cells are suitable to determine specific inhibition of endogenous VKORC1 (in VKORC1L1 KO cells) and endogenous VKORC1L1 activity (in VKORC1 KO cells). Alternatively, the DKO cell line allows investigation of any VKOR variant without confounding endogenous VKR and VKOR activity at a substrate level below 12 μ M.

Determination of IC₅₀ values of OACs and rodenticides for endogenous VKORC1 and VKORC1L1

IC₅₀ data for all compounds investigated in single VKOR KO cells and treated with K₁ are reported in Table 1, including the ratio of IC₅₀ values for endogenous VKORC1L1 with respect to endogenous VKORC1. Dose-response curves are shown in supplemental Figures 1 and 2. Four-parameter logistic curves could not fit to the coumarin and dicoumarol data. Dicoumarol dose-response for both VKOR enzymes were biphasic, and single-concentration data were used to estimate IC₅₀ concentration (VKORC1, 49.1% FIX activity at 5 nM; VKORC1L1, 48.2% FIX activity at 100 nM).

For all OACs VKORC1L1 uniformly required higher drug concentrations for inhibition (three- to \sim 100-fold greater; Table 1). Of the 4-hydroxycoumarin-based drugs, warfarin showed the highest difference in IC₅₀ concentrations between VKORC1 and VKORC1L1 (VKORC1L1 exhibits \sim 13-fold greater IC₅₀ than VKORC1). Acenocoumarol and phenprocoumon exhibited similar specificity (VKORC1L1 was three- to fourfold less sensitive than VKORC1). In contrast, the 1,3-indandiones showed markedly greater enzyme specificity. VKORC1 was inhibited at low nanomolar concentrations, whereas VKORC1L1 yielded up to 100-fold greater IC₅₀ values.

Notably, all rodenticides showed similar IC₅₀ values for both VKOR enzymes in low nanomolar range (Table 1; supplemental Figure 2).

To investigate the inhibitory potential on VKOR activity, warfarin, fluindione, and coumachlor were tested in the same assay setup, using 12 μ M K₁>O (instead of K₁) as substrate (supplemental Table 1; supplemental Figure 3). For warfarin and coumachlor, similar IC₅₀ values were obtained compared with K₁-treated cells. Interestingly, VKORC1L1 KO cells showed markedly lower susceptibility to fluindione under K₁>O supplementation.

In silico modeling of hVKORC1L1

The highest scored model from ITASSER for hVKORC1L1 showed a 4-transmembrane (4TM) structure, as previously described.¹³ Stereochemical parameters of this model agree favorably with the crystal structure represented by 3KP9. Structural alignment of the model with the 3KP9 structure showed a strong similarity of root-mean-square deviation <1 Å (ie, 0.86 Å) over 110 aligned residues. Similarly, alignment of the hVKORC1 and hVKORC1L1 model also showed strong structural similarity (ie, 0.96 Å over 146 aligned

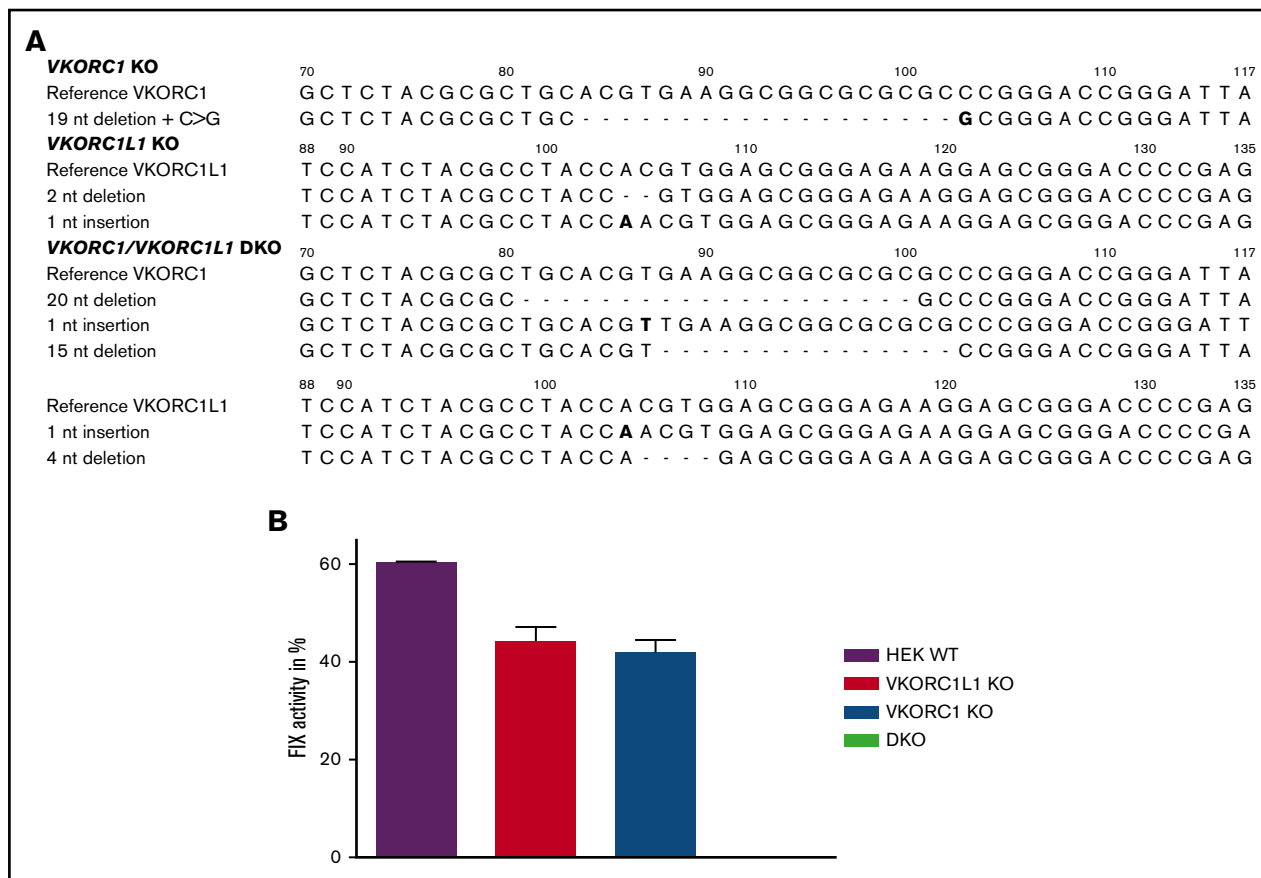


Figure 1. CRISPR/Cas9-mediated gene editing of *VKORC1* and *VKORC1L1* in HEK293T cells. (A) The wild-type sequence for both genes (c.70-116 of *VKORC1* and c.88-131 of *VKORC1L1*) is shown on the top, deletions are indicated by dashes, and insertion and substitution is shown in bold type. (B) Measurement of FIX activity after transfection of *human F9* cDNA and supplementation of 12 μ M vitamin K₁ in different genetically engineered cells. Columns represent measurements of endogenous *VKORC1* and *VKORC1L1* activity in HEK 293T WT, endogenous *VKORC1* activity (*VKORC1L1* KO), endogenous *VKORC1L1* activity (*VKORC1* KO), and in cells lacking both *VKOR* enzymes (*VKORC1/VKORC1L1* DKO cells). Columns show mean of triplicates \pm standard error of the mean.

residues).²⁴ Apart from the TM domains, the loops of both models appear similar in structure but very different to the thioredoxin-like domain on 3KP9 (Figure 2A-B).

Docking predictions for warfarin and other OACs on *VKORC1* and *VKORC1L1*

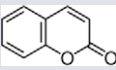
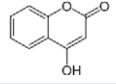
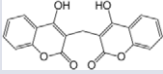
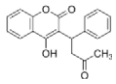
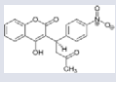
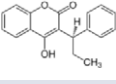
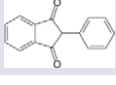
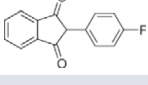
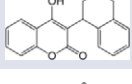
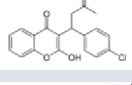
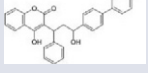
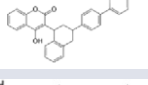
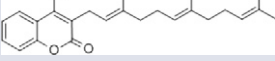
The dockings performed on the previously generated *VKORC1* model showed poses with pseudo-energy scores consistent for all OACs surveyed, and that are located similar to the published warfarin pose.^{24,30} The predicted pose on h*VKORC1L1* is located in the ER luminal loop, similar to the docking site on h*VKORC1*, but lies on the opposite site of the loop (Figure 2A-D). The ER luminal loops of h*VKORC1L1* and h*VKORC1* (Figure 2D) are similar in backbone structure, but differ greatly in their amino acid composition, especially with respect to the distribution of positively charged arginine residues. Because OACs bear a formal negative charge at physiological pH, the location of the predicted binding poses appears to be of great consequence with respect to possible charge interactions with the adjacent lying arginine residues that bear positive charges. Therefore, Arg38, Arg42, and Arg68 are likely to contribute to the OAC binding interactions with h*VKORC1L1* (supplemental Figure 4). Moreover, the predicted h*VKORC1L1* OAC binding pocket lacks any stacking

interactions and hydrogen bonds similar to those observed between warfarin and h*VKORC1*.³⁰

Mutation of specific loop residues of *VKORC1L1* to identify warfarin binding site

In a first approach, 2 variants were generated in which the loop of *VKORC1L1* was swapped into *VKORC1* cDNA and vice versa (supplemental Figure 6). In the DKO cells, IC₅₀ of *VKORC1*, *VKORC1L1*, and both loop swap variants were determined by overexpression of these variants together with *F9* (in a bicistronic vector or by coexpression of *VKOR* myc-tagged variants with *F9*). Western blot analysis of myc-tagged *VKOR* variants revealed similar protein expression (Figure 3C). Warfarin dose-response of *VKORC1* loop swap shifted to that for *VKORC1L1* WT, and *VKORC1L1* loop swap shifted to that of *VKORC1* WT (Figure 3A; supplemental Table 2). We further hypothesized that introducing the previously published *VKORC1* warfarin binding interface II (located in the loop) with Phe55 as major warfarin-binding residue in *VKORC1L1* leads to higher OAC susceptibility.^{24,30,31} Neither single (Leu62Phe) nor multi-*VKORC1L1* variants (Ala60Arg+Ala61Val+Leu62Phe) showed increased warfarin sensitivity. Indeed, these *VKORC1L1* variants required three- to fourfold higher warfarin IC₅₀ concentrations compared with *VKORC1L1* WT (Figure 3C; supplemental Table 2).

Table 1. IC₅₀ values of vitamin K antagonists

Compound class	Compound	Chemical structure	VKORC1 IC ₅₀ nM (95% CI)	VKORC1L1 IC ₅₀ nM (95% CI)	Ratio L1/C1
Inactive backbone	Coumarin		Not inhibitory*	Not inhibitory*	NA
Active 4HC backbone	4-hydroxycoumarin		1978 (1837-2130)	25110 (18530-34020)	12.7
Naturally occurring and therapeutic 4HC	Dicoumarol		5.0†	100.0†	20.0
Human therapeutic 4HC	Warfarin		1.9 (1.8-2.1)	25.2 (23.1-27.4)	13.4
Human therapeutic 4HC	Acenocoumarol		1.5 (1.4-1.7)	5.8 (5.4-6.2)	3.9
Human therapeutic 4HC	Phenprocoumon		3.6 (3.3-3.9)	10.5 (8.8-12.4)	2.9
Human therapeutic indandione	Phenindione		2.9 (2.6-3.3)	258.0 (213.4-312.0)	89.0
Human therapeutic indandione	Fluindione		4.8 (4.3-5.2)	268.5 (198.4-363.4)	55.9
First-generation rodenticide	Coumatetralyl		1.9 (1.8-2.1)	2.8 (2.7-3.0)	1.5
First-generation rodenticide	Coumachlor		0.7 (0.6-0.8)	1.0 (0.8-1.1)	1.4
Second-generation rodenticide	Bromadiolone		1.6 (1.5-1.7)	0.8 (0.8-0.9)	0.5
Second-generation rodenticide	Brodifacoum		0.5 (0.5-0.6)	0.4 (0.4-0.5)	0.8
Naturally occurring 4HC	Ferulenol		0.8 (0.8-0.9)	3.0 (2.4-3.8)	3.8

Overview of OACs investigated in the study, including their compound class as well as IC₅₀ assessed in VKORC1L1 KO and VKORC1 KO HEK 293T cells, using K₁ as substrate. Measurements were performed in triplicates.

4HC, 4-hydroxycoumarin; 95% CI, 95% confidence interval; C1, VKORC1; L1, VKORC1L1; NA, not applicable.

*VKR enzyme activities assessed up to 1 mM coumarin.

†Biphasic dose-response, no-curve fit with logistic function (VKORC1, 49.1%; VKORC1L1, 48.2% mean normalized activities with respect to 0 nM dicoumarol).

For verification of the in silico-predicted binding residues, positively charged arginine residues (Arg38, Arg42, Arg68) were mutated to negatively charged glutamate to maximize potential shifts in the respective dose-response curves. Warfarin dose-response data for single mutations Arg38Glu and Arg42Glu were shifted to greater values with respect to VKORC1L1 WT, whereas the values of Arg68Glu do not fall below 50% over the entire range of warfarin concentrations (Figure 3B). Arg38Glu+Arg42Glu revealed an eightfold higher IC₅₀, whereas the triple mutant Arg38Glu+Arg42Glu+Arg68Glu was completely resistant (Figure 3B).

Of note, the triple mutant showed reduced activities, yielding about 12% absolute FIX activity compared with ~25% of double arginine mutant and ~44% of VKORC1L1 WT. However, western blot analysis showed no difference in expression level for the VKORC1L1 variants (Figure 3D).

VKORC1L1 KO cells are most sensitive to oxidative stress

Assessment of cell viability revealed that all cell lines were sensitive toward H₂O₂ treatment, whereby VKORC1L1 KO cells were most

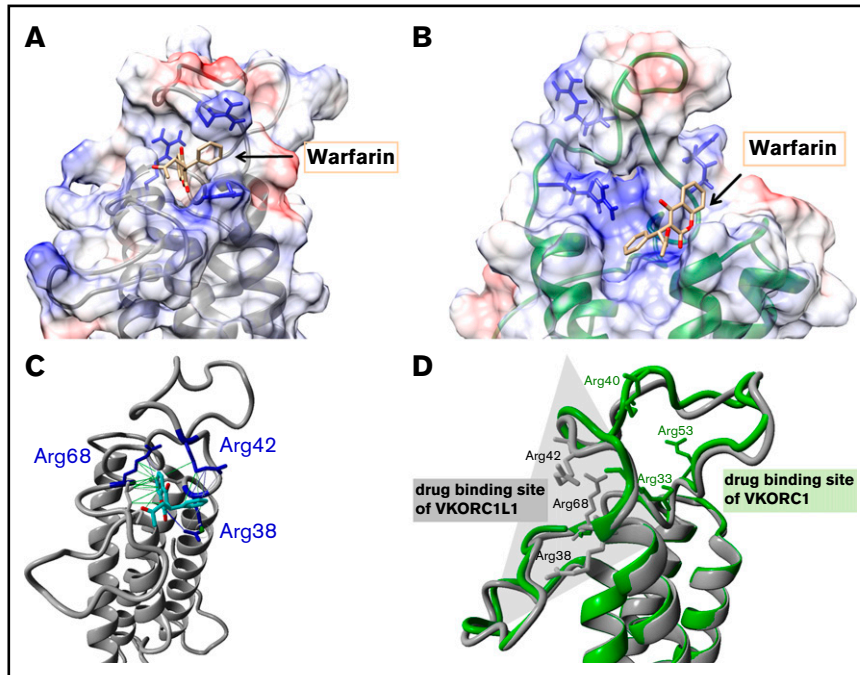


Figure 2. The binding poses for warfarin on VKORC1 and VKORC1L1. (A) Favored docking pose of warfarin on VKORC1L1 (gray-colored ribbon format). The electrostatic surface charge distribution is illustrated based on columbic charge distribution (blue, positive charge; red, negative charge). The clustered arginines on the endoplasmic reticulum (ER) luminal loop are depicted in blue and warfarin in beige-colored stick forms. (B) Favored docking pose of warfarin on VKORC1 (green-colored ribbon format). Charge distribution, arginines, and warfarin are colored as in panel A. (C) Interactions of the bound warfarin (beige stick form) with arginine residues (blue-colored stick forms) of VKORC1L1 (gray-colored ribbon format). Hydrophobic interactions between warfarin and VKORC1L1 residues are depicted as green lines, and charge-based (cation- π) interactions are depicted with blue lines of different intensities. (D) Structural alignment of the VKORC1 and VKORC1L1 models. The backbone of the VKORC1 and VKORC1L1 models are colored as in panels A and B. The arginine residues clustered around the favored warfarin binding site of each model are depicted in stick format. Opposite drug binding sites are highlighted with shaded triangles (gray for VKORC1L1, green for VKORC1).

sensitive. *VKORC1* KO and DKO cells exhibited similar cell viability as WT HEK cells (Figure 4A). K_1 supplementation seems to compensate cell stress for *VKORC1* KO cells, whereby $K_1 > O$ had no effect on cell viability. WT, DKO, and *VKORC1L1* KO cells showed no difference when supplemented with K_1 or $K_1 > O$ (Figure 4B).

Discussion

Here, we present a cell-based assay to independently study endogenous VKORC1 and VKORC1L1 inhibition by several compounds that affect vitamin K recycling. Previous studies have used different approaches to examine the capability of VKORC1 and VKORC1L1 to reduce K or $K > O$ by 2 different types of assays (Table 2). In the dithiothreitol (DTT) assay, direct conversion rates of $K > O$ to K are measured using HPLC.^{14,17,32,33} Alternatively, in the cell-based assays, a reporter protein that requires γ -carboxylation for biological activity is used as marker for VKR and VKOR activity.³⁴⁻³⁷ In several studies, WT FIX and chimeric FIX Gla domain-containing proteins were used as reporter for VKOR/VKR activity and were shown to reflect the in vivo situation properly.^{34,36-38} Until now, the majority of the studies could not discriminate between VKOR/VKR activities from endogenously expressed VKOR enzymes. Our assay analyzes endogenous inhibition of VKORC1 and VKORC1L1 independently by using genetically engineered *VKORC1L1* and *VKORC1* KO HEK 293T cell lines, respectively. Drug inhibition was not influenced by metabolic activity, as cytochrome P450

monooxygenases known to be involved in metabolism of vitamin K and warfarin are not expressed in HEK 293 cells.³⁷

VKORC1 and VKORC1L1 exhibit different inhibition characteristics for various OACs

A panel of 13 compounds with 4-hydroxycoumarin and 1,3-indandione backbones were analyzed for their inhibitory effect on VKORC1 and VKORC1L1 (Table 1). Consistent with in vivo data, naturally occurring coumarin showed no anticoagulatory effect in our in vitro assay.^{1,39} In contrast, VKORC1 and VKORC1L1 were both inhibited by 4-hydroxycoumarin at micromolar concentrations. The addition of 1 or more aromatic hydrocarbons of the variable 3-position side chains of 4-hydroxycoumarins correlates with decreased IC_{50} values, especially for VKORC1 (eg, 4-hydroxycoumarin [VKORC1 IC_{50} 2000 nM] and acenocoumarol [VKORC1 IC_{50} 1.6 nM]). It was recently shown that VKORC1 Phe55 participates in hydrophobic and stacking interactions to the 3-aromatic hydrocarbon of warfarin, thus explaining the increased affinities toward OACs with additional aromatic hydrocarbons.³⁰

In general, VKORC1 was more sensitive to therapeutic 4-hydroxycoumarin derivatives compared with VKORC1L1. The same inhibitory trend of coumarin derivatives regarding VKORC1 was shown by several in vitro and in vivo studies (Table 2).^{14,17,32-37} However, most studies relied on overexpressed VKOR enzymes. Tie et al were the

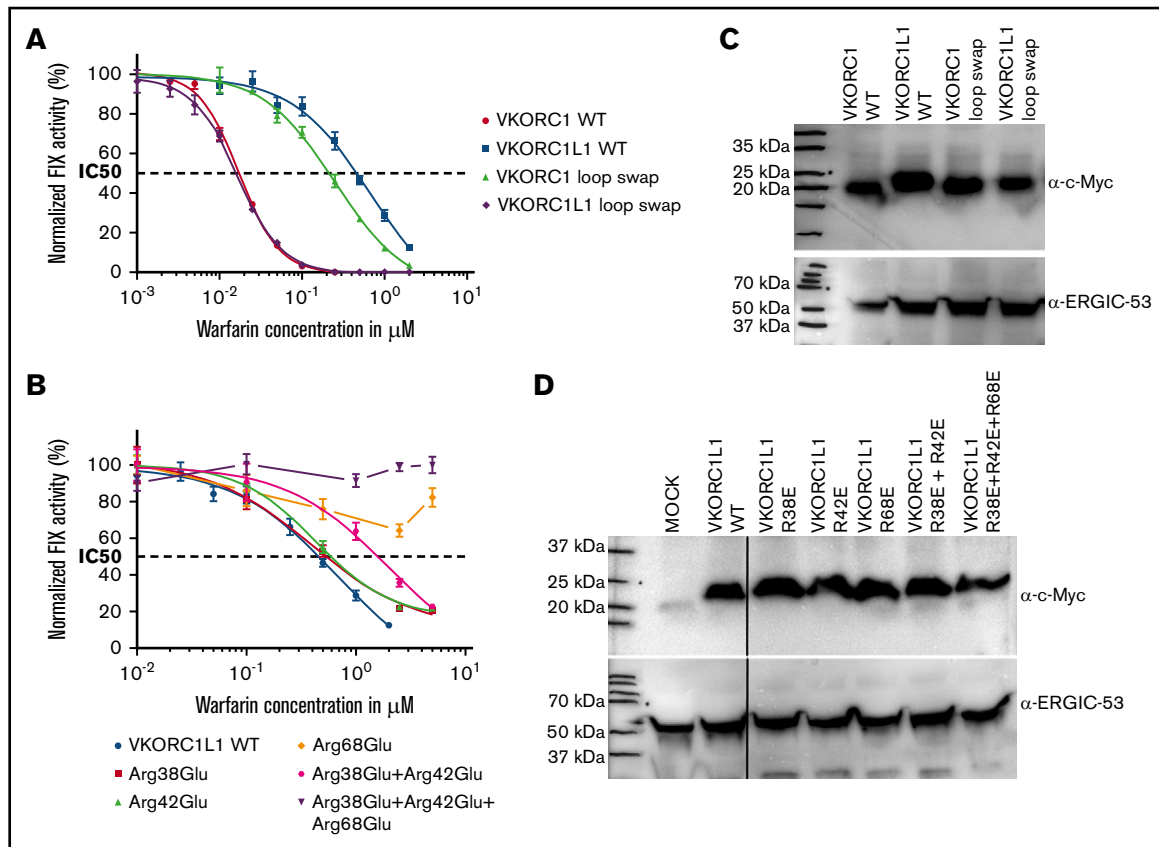


Figure 3. Inhibition curves for warfarin assessed in DKO cells. Exogenous VKR activity determined in DKO cells and assessed by overexpression of *VKOR* variants and *F9*. (A) Warfarin dose-response curves for VKORC1, VKORC1L1, and loop swap variants. (B) Warfarin dose-response curves for arginine variants of VKORC1L1. Measurements were performed in triplicates, values are shown as mean and error bars are represented as standard error of the mean. The coefficient of determination (R^2) calculated for each inhibition curve: (A) VKORC1, $R^2 = 0.9935$; VKORC1L1, $R^2 = 0.9628$; VKORC1 loop swap L1, $R^2 = 0.9826$. (B) VKORC1L1, $R^2 = 0.9628$; Arg38Glu, $R^2 = 0.9715$; Arg42Glu, $R^2 = 0.9875$; Arg38Glu + Arg42Glu, $R^2 = 0.9140$. (C-D) Expression of all VKOR variants detected by α -myc antibody (vertical lines have been inserted to indicate a repositioned gel lane). As loading control, α -ERGIC-53 was used.

first measuring endogenous VKORC1L1 inhibition in *VKORC1* KO HEK 293 cells.³⁵ However, they detected no difference in inhibition between WT and *VKORC1* KO HEK 293 cells, and *VKORC1L1* KO cells were not examined. Hammed et al studied inhibition of hVKORC1 and hVKORC1L1 by overexpression in *Pichia pastoris*, using the DTT assay that revealed ~30-fold higher susceptibility to warfarin for VKORC1 compared with VKORC1L1.¹⁷ In accordance with these results, we found a 30-fold difference in VKORC1 inhibition compared with VKORC1L1 when enzymes were overexpressed in DKO cells (Figure 3A; supplemental Table 1). Interestingly, measurement of endogenous VKOR inhibition by our assay revealed only 13-fold difference in warfarin sensitivity with lower IC_{50} values (Tables 1; supplemental Table 1). This might explain why studies using overexpressed VKOR enzymes, in general, reported higher IC_{50} values relative to the current study results.^{14,17,33,36} Various warfarin IC_{50} values, primarily for VKORC1, have been published and range between 25 nM and 24 μ M.^{13,32,33,35-37} Differences may arise from the type of assay used (DTT vs cell-based), the type of substrate (K_1 vs $K_1 > O$) and its concentration, and the relative protein expression levels (endogenously vs overexpressed), and source of cells (kidney- vs liver-derived) (Table 2). We observed that the type of substrate (K_1 or $K_1 > O$) did not affect warfarin dose responses when using the

same molarity (Tables 1; supplemental Table 1). Moreover, higher IC_{50} values detected for VKORC1 in other studies are probably a result of simultaneously measured endogenous VKORC1L1 activity, especially in cell-based assays. The low dose-response values and ability to discriminate between both VKOR enzymes indicate that our method using *VKORC1* and *VKORC1L1* KO cells is highly sensitive. Notably, polyploid endogenous VKORC1 expression may be similar in HEK 293T cells and native hepatocytes, the cells responsible for producing substantially all VKD coagulation factors.⁴⁰

Among the therapeutically used 4-hydroxycoumarins, acenocoumarol was most effective on VKORC1L1 inhibition, whereas warfarin required the greatest IC_{50} concentration. This suggests that at effective therapeutic dosages that lower VKORC1-mediated VKR activity and, hence, lower VKD coagulation factor activities, it is likely that VKORC1L1-mediated VKR activity is not appreciably affected in liver, and especially in nontarget tissues. However, the effect of VKORC1L1 inhibition in vivo at therapeutic OAC drug levels is not known yet. Although the cell-based assays are sensitive and reflect the in vivo situation, it is still unclear whether VKORC1L1 provides KH_2 for γ -carboxylation of VKD proteins in vivo, too. Results from mice studies indicate that VKORC1L1 is not able to

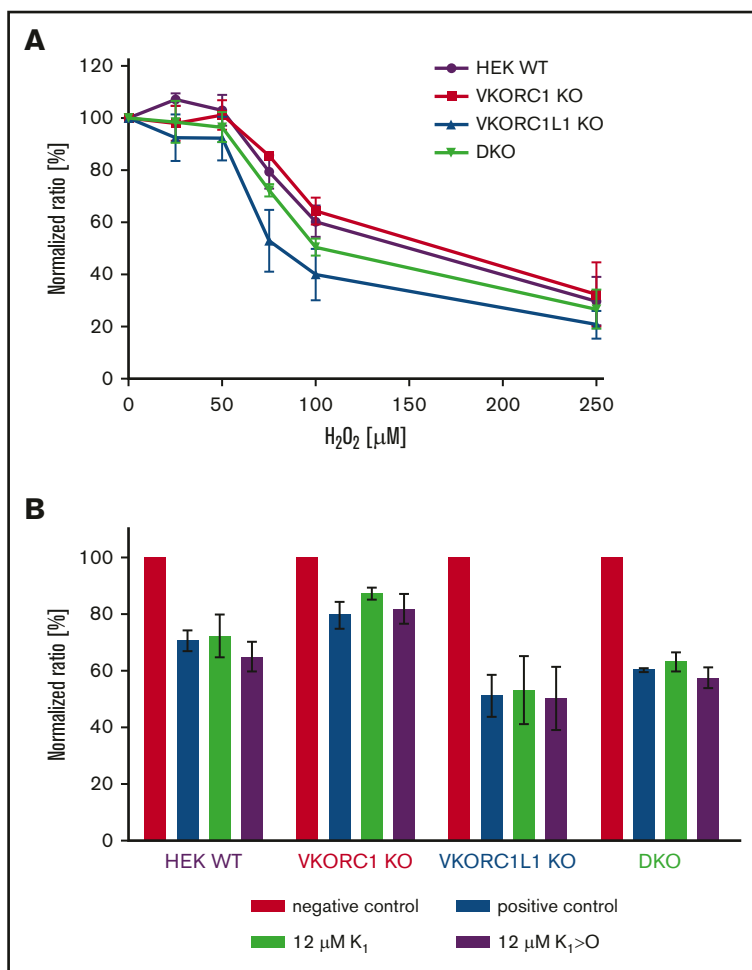


Figure 4. Effect of cell stress on VKOR KO cells. (A) Cell viability with regard to H₂O₂ treatment was assessed in WT (purple), VKORC1 KO (red), VKORC1L1 KO (blue), and DKO (green) HEK 293T. Curves showing normalized values for 3 replicates including error bars representing standard error of the mean. (B) Effect of K₁ and K₁>O treatment on cell viability. Negative control without H₂O₂; positive control with 75 μM H₂O₂. Cells were incubated with 75 μM H₂O₂ and K₁ or K₁>O (each 12 μM).

compensate lack of VKORC1 activity, as bleeding phenotype in constitutive *VKORC1*^{-/-} was not rescued.¹⁵ In liver, where coagulation factors are synthesized, VKORC1L1 expression is very low and is not upregulated in *VKORC1*^{-/-} mice.¹⁷ Assessment of VKOR activity in *VKORC1*^{-/-} liver microsomes showed markedly reduced activity (by 97%) compared with WT.¹⁷ However, *VKORC1L1*^{-/-} in osteoblasts revealed no difference in vitamin K-dependent osteocalcin levels compared with control mice.⁴¹ In an in vitro study, Westhofen et al postulated that VKORC1L1 plays a role in intracellular antioxidant.¹⁴ Therefore, we investigated whether our VKOR KO cell lines are sensitive toward oxidative stress, as well. Our results point into the same direction of Westhofen et al, with *VKORC1L1* KO cells being most sensitive toward H₂O₂ treatment (Figure 4A). Moreover, supplementation of K₁ slightly reversed H₂O₂ treatment in *VKORC1* KO cells, but not K₁>O. In contrast, DKO and *VKORC1L1* KO cells were not rescued by K₁ treatment, indicating that VKORC1L1 is the only enzyme responsible for this pathway. However, in WT HEK cells, endogenous VKORC1L1 did not improve cell viability under K₁ supplementation (Figure 4B). Both VKOR enzymes are present in those cells catalyzing K₁, which probably leads to lower substrate availability for VKORC1L1. Another possibility could be that VKORC1L1 is upregulated in *VKORC1* KO cells. More studies are needed to investigate the effects of VKOR enzymes on oxidative

stress. However, our results indicate that inhibition of VKORC1L1 may result in reduced antioxidative properties.¹⁴

For phenindione and flumidione, IC₅₀ values were almost similar to that of 4-hydroxycoumarin drugs regarding VKORC1 inhibition when determined in presence of K₁. In contrast, investigated indandiones required up to 50-fold greater concentrations compared with coumarin-based compounds for VKORC1L1 inhibition. Interestingly, VKORC1 exhibited greatly increased IC₅₀ values for flumidione in the presence of K₁>O (4.8 nM for K₁ vs 204 nM for K₁>O; supplemental Figure 6; Table 1; supplemental Table 1). Therefore, flumidione seems to have different binding characteristics in presence of K₁>O compared with K₁, as well as compared with 4-hydroxycoumarin-based drugs. We further conclude that K₁>O accumulation may affect the inhibitory potential of flumidione.

Inhibition of endogenous VKORC1 and VKORC1L1 by rodenticides and ferulenol

Rodenticides are long-lasting coumarin derivatives, whereas second-generation rodenticides emerged because of resistance against compounds of the first generation.⁴² The 3-position substitutions by long, phenyl side-chains yielded tremendous increase in potency and duration of action.⁴³ Our data show that rodenticides have increased affinities for both VKORC1 and

Table 2. Comparison of published VKOR inhibition data

	Gebauer ³²	Tie et al ³⁴	Westhofen et al ¹⁴	Tie et al ³⁵	Fregin et al ³⁶	Hammed et al ¹⁷	Bevans et al ³³	Haque et al ³⁷	Present study	Present study	Present study
Expression system/cell line	Rat liver microsomes	HEK 293	HEK 293T	HEK 293 WT and C1 KO	HEK 293T	<i>P. pastoris</i>	<i>P. pastoris</i>	HEK 293-C3	C1 KO and L1 KO HEK 293T	C1 KO and L1 KO HEK 293T	DKO HEK 293T
VKOR expression	Endogenous	Endogenous	Overexpression of both VKOR enzymes	Endogenous	Overexpression of VKORC1	Overexpression of both VKOR enzymes	Overexpression of VKORC1	Endogenous	Endogenous	Endogenous	Overexpression of both VKOR enzymes
Type of measurement	Direct	Indirect	Direct	Indirect	Indirect	Direct	Direct	Indirect	Indirect	Indirect	Indirect
Activity measurement	K>O conversion	FIXgla-PC	K>O conversion	FIXgla-PC	FIX	K>O conversion	K>O conversion	F9CH	FIX	FIX	FIX
Reporter expression	NA	Stable	NA	Stable	Transient	NA	NA	Inducible, stable	Transient	Transient	Transient
Substrate	25 mM K ₁ >O	5 μM K ₁ >O	50 μM K ₁ >O	5 μM K ₁ >O	10 μM K ₁	Up to 0.2 mM K ₁ >O	Up to 1 μM K ₁ >O	50 nM K ₁	12 μM K ₁	12 μM K ₁ >O	12 μM K ₁
Type of assay	DTT assay	ELISA	DTT assay	ELISA	Clotting assay	DTT assay	DTT assay	FACS	Clotting assay	Clotting assay	Clotting assay
Discrimination between C1 and L1	No	No	Yes (but remaining endogenous activity)	Yes (in KO cell line)	Yes (but remaining endogenous activity)	Yes	Yes	No	Yes	Yes	Yes
Warfarin	IC ₅₀ 2.2 μM	IC ₅₀ 100 nM	At 5 μM: C1: 53% inhibition L1: 29% inhibition	IC ₅₀ < 50 nM	IC ₅₀ 25 nM	K _i ratios: hL1/C1: 29× rL1/C1: 54×	IC ₅₀ 3-24 μM	IC ₅₀ 125 nM	IC ₅₀ : C1: 1.9 nM L1: 25.2 nM	IC ₅₀ : C1: 2.4 nM L1: 27 nM	IC ₅₀ : C1: 17.3 nM L1: 474 nM

Published data regarding inhibition of VKORC1 or VKORC1L1 by warfarin are summarized. K_i, inhibitory constant.

VKORC1L1 that may explain their potency. The difference between first- and second-generation rodenticides was not as great as shown by in vivo studies in rodents (Table 1; supplemental Figure 2).⁴³ Hence, studies with VKORC1 mutations should be performed to determine which rodenticide is most potent for inhibition of resistance phenotypes.

Moreover, ferulenol, a plant-derived 4-hydroxycoumarin, was reported to cause hemorrhagic symptoms in mammals.⁴⁴ Inhibitory potency of ferulenol was confirmed by our results, but values were not as high as described in literature.^{32,44} It is the most potent naturally occurring coumarin and is similarly effective as rodenticides and more effective than OACs on VKORC1.

VKORC1L1 is less OAC sensitive than VKORC1 because of different drug binding sites

To elucidate the basis for the different OAC susceptibilities of VKORC1 and VKORC1L1, the ER luminal loop of *VKORC1L1* was swapped in *VKORC1* cDNA and vice versa (Figure 3A; supplemental Figure 5). The loop swap variants are consistent with VKOR models comprising either 3 or 4 TM domains.^{45,46} Determination of IC₅₀ values for warfarin and fluroindione for VKORC1 loop swap yielded shifted doses responses toward VKORC1L1 (Figure 3A; supplemental Figure 6; supplemental Table 2). In accordance, VKORC1L1 loop swap turned out to be as sensitive as VKORC1 WT when using warfarin. These results strongly suggest that the loops of both VKOR enzymes play an important structural role in coumarin-based OAC binding. Interestingly, when using fluroindione, dose responses of VKORC1L1 loop swap did not completely shift to that observed for VKORC1 (supplemental Figure 6). We also detected differences in fluroindione IC₅₀ values for VKORC1 WT when supplemented with K₁>O compared with K₁, suggesting that fluroindione might have other drug binding characteristics on VKORC1 than 4-hydroxycoumarins (supplemental Figure 3; supplemental Table 1).

In a previously published warfarin binding pocket on VKORC1, 1 warfarin binding surface comprises residues Ser52 through Phe55 of the ER luminal loop, with Phe55 being the major amino acid binding warfarin.^{24,30,31} Therefore, we decided to swap this interface into VKORC1L1. The idea was to probe whether these warfarin interacting residues in VKORC1 contribute to higher binding affinity in VKORC1L1. Neither introducing the phenylalanine into the corresponding sequence of VKORC1L1 (VKORC1L1: Leu62Phe) nor complete swap of the binding surface (VKORC1L1: Ser59-Phe62) showed different susceptibility to warfarin, indicating that VKORC1 warfarin binding surface is different from VKORC1L1 (Figure 2D). Comparative visualization of the calculated charge distribution for the ER luminal loops of the VKORC1 and VKORC1L1 models suggests that the primary difference in binding sites might be attributed to the different clustering of positively charged arginines contributing to overall binding affinity of the anionic OAC drugs. In VKORC1, 3 arginine residues are clustered around the reported warfarin binding pocket and were found to result in resistance when mutated to glutamate residues (Arg33, Arg40, and Arg53), whereas other arginine residues in the loop of VKORC1 had no effect on

warfarin binding (Arg61, Arg35).³⁰ In contrast, in hVKORC1L1, 3 arginine residues at amino acid positions 38, 42, and 68 cluster around a binding pocket that is diametrically opposite to that one in VKORC1 (ie, on the other side of the short helix; Figure 2D). Substitution of these arginine residues by glutamate residues resulted in greater IC₅₀ values up to complete resistance. Interestingly, the corresponding arginine residues found in VKORC1L1 to affect warfarin binding have been shown to not affect OAC binding in VKORC1 (VKORC1L1: Arg68 = VKORC1:Arg61, VKORC1L1:Arg42 = VKORC1:Arg35; supplemental Figure 5).³⁰ These results, along with the loop-swapping data, are consistent with our prediction that this region is the binding site for warfarin in VKORC1L1 (Figures 2 and 3; supplemental Table 2). But why is OAC binding to VKORC1L1 weaker than to VKORC1? In VKORC1, where warfarin is stabilized by a combination of hydrogen bonds, aromatic stacking interactions, hydrophobic interactions, and charge-based interactions, unlike warfarin binding in VKORC1L1, appear to be stabilized only by charge-based and hydrophobic interactions. This might explain the poorer sensitivity of VKORC1L1 toward warfarin when compared with VKORC1. The distinct binding sites of VKORC1 and VKORC1L1 with comparably more amino acids being involved in OAC binding in VKORC1 may explain their different affinities to OACs. Furthermore, in silico docking predictions for other 4-hydroxycoumarins showed favorable docking poses in close apposition to all 3 experimentally determined arginines (supplemental Figure 4).

In conclusion, we have established KO HEK 293T cell lines that express either endogenously VKORC1 or VKORC1L1, and found that VKORC1 is more sensitive to OACs than VKORC1L1, whereas rodenticides apparently inhibit both VKOR enzymes equally. In silico analysis of VKORC1L1 and corresponding in vitro data strongly suggest that the warfarin binding sites for VKORC1 and VKORC1L1 are located in the loop but are different with respect to position and interacting residues.

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Authorship

Contribution: K.J.C. and K.L. provided experimental design; K.L. performed data collection, data analysis, and figure production; A.B. provided protein modeling; K.J.C., K.H., and V.H. provided CRISPR/Cas9 gene editing of HEK 293T cells; and K.J.C., K.L., K.H., V.H., M.W., and J.O. provided manuscript drafting and editing.

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