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Oxidative Stress Regulation by DJ-1 in the Retinal Pigment Epithelium

Vera L. Bonilha,

Department of Ophthalmology, Cleveland Clinic Lerner College of Medicine, The Cole Eye Institute, i31, 9500 Euclid Avenue, Cleveland, OH 44195, USA

Mary E. Rayborn,

Department of Ophthalmology, Cleveland Clinic Lerner College of Medicine, The Cole Eye Institute, i31, 9500 Euclid Avenue, Cleveland, OH 44195, USA

Xiaoping Yang,

Department of Ophthalmology, Cleveland Clinic Lerner College of Medicine, The Cole Eye Institute, i31, 9500 Euclid Avenue, Cleveland, OH 44195, USA

Chengsong Xie, and

Laboratory of Neurogenetics, National Institute of Aging, NIH, Bethesda, MD 20892, USA

Huaibin Cai

Laboratory of Neurogenetics, National Institute of Aging, NIH, Bethesda, MD 20892, USA

Abstract

DJ-1 is a protein expressed in many tissues including the brain where it has been extensively studied due to its association with Parkinson's Disease (PD). DJ-1 was reported to function as an antioxidant, redox-sensitive molecular chaperone, and transcription regulator, which protected cells from oxidative stress by modifying signaling pathways that regulate cell survival. Here we discuss our progress toward characterization of the DJ-1 function in the protection of RPE to oxidative stress.

Keywords

Retinal pigment epithelium; Oxidative stress; Reactive oxygen species; DJ-1; Histology

81.1 Introduction

DJ-1 gene encodes a highly conserved protein with 189 amino acids and a molecular weight of \sim 20 kDa, which belongs to the ThiJ/PfpI protein superfamily [1]. The function of ThiJ is not fully known, but it may be related to the biosynthesis of thiamins while PfpI is an intracellular protease, responsive to growth conditions and present in most bacteria and archea [2, 3, 4]. Therefore, both DJ-1 sequence and structure suggested that this protein would be involved in cellular viability.

Correspondence to: Vera L. Bonilha.

Initially, DJ-1 was reported as a novel oncogene showing a transforming activity when expressed together with H-ras [5]. Later, it was reported that the gene for human DJ-1 (PARK7) is mutated in rare forms of recessively inherited PD [6].

Recently, we have identified DJ-1 peptides in both young and aged retinal pigment epithelium (RPE) lysates starting with a proteomic approach. We also reported immunolocalization of DJ-1 protein in both young and aged rat RPE cells [7]. Here we extend our initial observations of DJ-1 expression and further discuss DJ-1 function in the RPE cells.

81.2 DJ-1 Function and Oxidative Stress Protection

At the subcellular level, under basal conditions, DJ-1 is found mostly in the cytoplasm and to a lesser extent in the mitochondria [6, 8, 9] and nucleus [9]. Under conditions of oxidative stress, more DJ-1 redistributes to mitochondria and later to the nucleus, and this cellular redistribution correlates with the ability of DJ-1 to confer neuroprotection [8, 10, 11]. As previously described in several cell types, under baseline conditions, DJ-1 displays a diffuse cytoplasmic and nuclear staining of all the RPE cell lines analyzed (Fig. 81.1a). Oxidative stress induced by incubation of RPE cultures with H_2O_2 leads to a visible increase in immunocytochemical staining for DJ-1 (Fig. 81.1b). In addition, these RPE cultures displayed an induced intracellular redistribution of DJ-1 to a perinuclear localization (mitochondria, data not shown). Confirmation of the increased DJ-1 protein levels in RPE cells under oxidative stress was also demonstrated by Western blot analysis (data not shown).

Several reports suggest that DJ-1 robustly protects cells from oxidative stress [8, 12–18]. First, reports showed that flies and mice deficient in the gene encoding DJ-1 are indeed more susceptible to oxidative toxins [13, 15, 19–21]. Second, it was reported that DJ-1 can eliminate H_2O_2 in vitro by becoming oxidized itself and thus functioning as a scavenger of reactive oxygen species (ROS) [17, 22]. It was also established that cysteines at residues 46, 53, and 106 become oxidized upon oxidative stress, resulting in scavenging of ROS, enhancing DJ-1 association with mitochondria and allowing DJ-1 to protect cells from oxidative stress [8, 14, 23].

To explore the functional consequences of DJ-1 mutation, adenoviruses carrying both DJ-1 full length and DJ-1 with the three cysteine at residues 46, 53, and 106 mutated to serine were generated and used as agents to induce overexpression of exogenous DJ-1. Control and RPE cultures overexpressing both types of exogenous DJ-1 were exposed to oxidative stress through incubation with H_2O_2 followed by incubation with the fluorescent indicator used to assess ROS generation 5-(and-6)-carboxy-2',7'-difluorodihydrofluorescein diacetate (carboxy-DCFDA) and confocal microscopy (Fig. 81.2). Interestingly, no significant amount of ROS was detected in ARPE-19 over-expressing exogenous full-length DJ-1 and exposed to oxidative stress (Fig. 81.2c) and in ARPE cultures at basal conditions (Fig. 81.2a). In contrast, a significant generation of ROS was observed when ARPE-19 cultures were exposed to oxidative stress (Fig. 81.2b) and when RPE cultures overexpressing the mutated DJ-1 were exposed to oxidative stress (Fig. 81.2d). This data suggested that high levels of

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We have also analyzed histologically the eyes of young adult DJ-1 KO mice and noted that the RPE in these mice is characterized by patchy thinning (Fig. 81.3b, arrows) when compared to the control RPE (Fig. 81.3a). Our data suggested that normal RPE structure requires DJ-1 expression. To further relate RPE DJ-1 expression with oxidative stress regulation, we analyzed the expression of 7,8-dihydro-8-oxoguanine (8-oxoG), the most abundant oxidized base generated in vivo by various types of ROS, in control and DJ-1 KO RPE with antibodies specific to this DNA oxidation (Fig. 81.3c, d). Immunocytochemical staining of 8-oxoG, was significantly elevated in the DJ-1 KO RPE (Fig. 81.3d) and photoreceptor inner segments (Fig. 81.3d, arrowheads) when compared to control RPE (Fig. 81.3c). These data indicated that lack of DJ-1 expression leads to increased oxidative stress in RPE in vivo.

81.3 Conclusions

Age-related macular degeneration (AMD) is the most common cause of irreversible blindness in the elderly population in industrialized countries. Studies carried out in man, AMD animal models, and RPE cell lines point toward an important role of oxidative stress in the development of AMD. Due to the evidence presented here, connecting DJ-1 to protection against oxidative stress, it is conceivable that DJ-1 function may also be related to oxidative stress implicated in AMD pathology. The years to come will bring further definition of key proteins and pathways involved in the regulation of oxidative stress by DJ-1 in RPE cells as well as a better understanding of its function in AMD.

Acknowledgments

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Abbreviations

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Fig. 81.1.

Immunocytochemical analysis of intracellular distribution of DJ-1 on RPE monolayers exposed to oxidative stress. Immunocytochemical staining of ARPE-19 cells for DJ-1 demonstrates that at basal conditions **a** DJ-1 is diffused in the cytoplasm and nuclei (*) of the cells. With 1 h of exposure to 800 μ M H₂O₂, **b** a pronounced aggregated perinuclear staining (*arrows*) for DJ-1 is apparent. Bar = $20 \mu m$

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Fig. 81.2.

Staining of ROS generation on ARPE-19 monolayers expressing endogenous and exogenous DJ-1 and exposed to oxidative stress by exposure to 800 μ M H₂O₂ for 1 h. Incubation of RPE monolayers with carboxy-DCFDA indicates that no ROS is generated by the RPE cells at basal conditions (**a**). While upon oxidative stress incubation, appreciable ROS generation throughout the entire cell body of cells is observed (**b**). RPE cultures transduced with an adenovirus carrying the full-length hDJ-1 DNA did not display ROS generation when exposed to oxidative stress (**c**). However, RPE cultures transduced with an adenovirus carrying DJ-1 with the three cysteine at residues 46, 53, and 106 mutated to serine (**d**). Displayed appreciable ROS generation when cells were exposed to oxidative stress. Bar = 20 µm

Fig. 81.3.

Degeneration in the neural retina of young adult DJ-1 KO mouse. Plastic sections of both control (**a**) and DJ-1 KO (**b**) retinas stained with toluidine blue highlighted the RPE and photoreceptor cells. The RPE of the DJ-1 KO displayed thinning (**b**, arrows). Cryosections of retinas were also labeled with 8-oxoG (**c, d**) to detect DNA oxidation. Analysis showed that 8-oxoG immunoreactivity is significantly increased in the photoreceptor inner segments (arrowheads) and the RPE cells of DJ-1 KO (**d**) when compared to the control (**c**) retinas. ONL outer nuclear layer, OS photoreceptor outer segments, RPE retinal pigment epithelium, Bar $\mathbf{a}, \mathbf{b} = 25 \text{ }\mu\text{m}$ and $\mathbf{c}, \mathbf{d} = 40 \text{ }\mu\text{m}$