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Discriminating MGMT Promoter Methylation Status in Patients with Glioblastoma Employing Amide Proton Transfer-Weighted MRI Metrics

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One of the authors (Dr. Fangyao Chen) has significant statistical expertise. No complex statistical methods were necessary for this paper.

Three study subjects have been previously reported in one of our previous papers, in which we evaluated the diagnostic values of APTw imaging in differentiate PCNSL and malignant gliomas, see Ref. [29].

Methodology: retrospective, diagnostic or prognostic study, performed at one institution.

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Abstract

Objectives—To explore the feasibility of using amide proton transfer-weighted (APTw) MRI metrics as surrogate biomarkers to identify the O6-methylguanine-DNA methyltransferase (MGMT) promoter methylation status in glioblastoma (GBM).

Methods—Eighteen newly diagnosed GBM patients, who were previously scanned at 3T and had a confirmed MGMT methylation status, were retrospectively analyzed. For each case, a histogram analysis in the tumor mass was performed to evaluate several quantitative APTw MRI metrics. The Mann-Whitney test was used to evaluate the difference in APTw parameters between MGMT methylated and unmethylated GBMs, and the receiver-operator-characteristic analysis was further used to assess the diagnostic performance.

Results—Ten GBMs were found to harbor a methylated MGMT promoter, and eight GBMs were unmethylated. The Mean, Variance, 50^{th} percentile, 90^{th} percentile, and Width_{10–90} APTw values were significantly higher in the MGMT unmethylated GBMs than in the MGMT methylated GBMs, with the areas under the receiver-operator-characteristic curves of 0.825, 0.837, 0.850, 0856, and 0.763, respectively, for the discrimination of MGMT promoter methylation status.

Conclusions—APTw signal metrics have the potential to serve as valuable imaging biomarkers for identifying MGMT methylation status in the GBM population.

Keywords

Glioblastoma; O6-methylguanine-DNA methyltransferase; Magnetic Resonance Imaging; Amide Proton Transfer-Weighted Imaging; Methylation

Introduction

Glioblastoma (GBM) is the most dismal type of primary malignant brain tumor in adults. Although tremendous effort has been made to optimize the treatment of GBM, most patients die within 15 months [1]. The miserable prognosis conferred by GBM is partly due to the tendency of GBMs to diffusely and extensively infiltrate into surrounding normal brain tissue [2]. In addition, the intrinsic or acquired resistance presents obstacles to successful chemoradiotherapy treatment [3, 4]. Recently, it has been demonstrated that some GBM cohorts, with inhibited DNA damage response mutations or pathways against chemotherapy, perceptibly modulated the tumor response [5–7]. The methylated O6-methylguanine-DNA methyltransferase (MGMT) promoter is one molecular marker that is indicative of longer survival in patients with GBM who receive alkylating agents, with both proved biochemical mechanisms and clinically verified evidence [8–10]. The postulated underlying mechanism showed that the methylated MGMT promoter could enhance chemosensitivity via silencing of the MGMT gene to eventually downregulate the expression of encoded MGMT protein, which functions as a key DNA repair enzyme that counteracts alkylate drugs [11].

Thus, the MGMT promoter methylation status test has become a routine screening test in patients with GBM as a personalized medicine strategy [12]. Unfortunately, current lab tests primarily require large tissue samples obtained from invasive surgical procedures, which are not only costly, but also pose risks. In recent years, a new hypothesis in cancer research has emerged that various genotypes or molecular alterations within GBM could lead to observable, altered imaging features [13, 14]. Hence, imaging-based methodologies have been implemented to provide insight into the MGMT methylation status in patients with GBM. Some studies have attempted to discover promising textural patterns derived from run-length textures [15] or the co-occurrence matrix [16] in MRI datasets via intricate algorithms. Several imaging textures were found to be feasible for the discrimination of GBMs with different MGMT promoter methylation statuses, despite the fact that it is timeconsuming and statistic overfitting [17]. Researchers also evaluated the efficacy of applying advanced MRI techniques to determine the status of MGMT methylation in GBMs, including diffusion imaging [18], diffusion tensor imaging [19, 20], and perfusion imaging [19-21]. Positron emission tomography (PET) was also studied to discover predictive markers for MGMT-expressed glioblastoma in vitro [22]. All these studies proposed that the MGMT molecular aspects of GBMs can be characterized quite convincingly by MRIderived tumor features or by a PET imaging agent. Yet, the results drawn from different research groups have been inconsistent, even when investigating the same advanced MRI imaging [18–21], or the sample sizes were still relatively limited with regard to compelling statistical conclusions.

Amide proton transfer (APT) imaging, the most developed branch of chemical exchangedependent saturation transfer (CEST) imaging [23, 24], is a novel molecular MRI technique that generates contrast based on endogenous cellular proteins in vivo [25, 26]. Several studies from different labs thus far have consistently demonstrated that APT-weighted (APTw) imaging has great potential for detecting malignant brain tumors [27–30] and other cancers [31, 32]. With regard to gliomas, APTw MRI has shown its unique clinical efficiency in defining the tumor burden [33], determining tumor grade [34-36], and monitoring treatment effects [37, 38]. Notably, it has been shown that APTw signal intensities have a positive correlation with cellularity and proliferation [35, 36]. The promoter of MGMT is often hypermethylated in many cancers, suggesting the decrease of its protein expression. It may affect other protein expressions downstream of MGMT [39]. Hence, protein-based APTw MRI may provide a new approach to non-invasively investigate the MGMT methylation status in GBMs. We hypothesize that GBMs with an unmethylated MGMT promoter will present higher APTw values than the MGMT methylated GBMs. The purpose of this retrospective study is to investigate whether the MGMT methylation status in GBMs can be stratified by APTw-MRI metrics.

Materials and methods

Subjects

This retrospective study was approved by the local Institutional Review Board, and the requirement for informed consent was waived. Patients with newly diagnosed GBM (grade IV astrocytoma), treated in Zhujiang hospital hospital between July 1, 2014 and August 31,

2016, were recruited. Enrollment criteria were: age 18 years; APTw and routine MRI scanning performed within 7 days preoperatively, including T_2 -weighted (T_2w), fluid-attenuated inversion recovery (FLAIR), T_1 -weighted (T_1w), and gadolinium-enhanced T_1 -weighted (Gd- T_1w) imaging; known MGMT methylation status; and no radiotherapy, chemotherapy, or surgery before imaging. Exclusion criteria included inferior image quality due to various reasons.

MRI data acquisition

MRI imaging was performed on a 3T MRI scanner (Achieva; Philips Medical Systems, Best, The Netherlands). A fat-suppressed, fast spin-echo pulse sequence was used to acquire APT image data, with the following parameters: radiofrequency saturation power, 2 μ T; saturation time, 0.8 sec; repetition time, 3 sec; matrix, 128 ×128 (reconstructed to be 400 × 400); field of view, 240×240 mm²; slice thickness, 6 mm; sensitivity-encoding factor, 2; and turbo-spinecho factor, 37. A single-slice, combined APTw imaging and Z-spectrum acquisition protocol was implemented on the maximum cross-sectional tumor slice (as determined by routine MRI images). As described before [33], this protocol had more acquisitions at and around ±3.5 ppm, facilitating a sufficient signal-to-noise ratio for APTw imaging (5 ml/s; 0.2 mL/kg body weight; Magnevist; Bayer Schering, Guangzhou, China) was acquired as the last sequence to avoid potential influence on the APTw signal intensity [40].

APTw image processing and analysis

All image data were processed using the interactive data language software (IDL, Version 7; Exelis Visual Information Solutions, Inc., Boulder, CO). To reduce possible motion artifacts during the scanning, the acquired APTw image or Z-spectrum series was registered to the saturated image at 3.5 ppm [41], which was further corrected for the B₀ inhomogeneity effect on a voxel-by-voxel basis. Then, Z-spectra (Z(offset), defined as water signal intensities with and without selective radio-frequency irradiation plotted as a function of saturation frequency offset, relative to water), magnetization transfer ratio asymmetry spectra (MTR_{asym}(offset) = Z(offset) – Z(-offset)), and MTR_{asym}(3.5ppm) images were calculated, as previously reported [25, 26]. Because of the contributions from the nuclear Overhauser enhancement effect at -3.5 ppm [42–46], and the asymmetry of the conventional semi-solid magnetization transfer effect [47], the calculated MTR_{asym}(3.5ppm) images are often called APTw images [48].

The acquired conventional MR images were co-registered to the corresponding saturated S_{sat} image at 3.5 ppm (which was co-registered with the APTw image) to fulfill quantitative APTw analyses [41]. Two radiologists (S.J. and X.W., with ten and seven years of experience in brain imaging, respectively) carefully drew the regions-of-interest (ROIs), in consensus. One large ROI contouring the whole area of Gd enhancement within the lesion on the Gd-T₁w image was drawn and defined as the tumor mass for the histogram ROI analysis. The contralateral normal-appearing white matter (CNAWM) was also analyzed for normalization, and relative APTw values were reported (ROI APTw - CNAWM APTw). Fig. 1 shows how the ROIs were drawn.

For each case, the Z-spectrum data and APTw value histogram data from the large wholetumor ROI were recorded. The histogram data were analyzed for Mean, Variance, Skewness, Kurtosis, 10^{th} percentile, 50^{th} percentile, 90^{th} percentile, and Mode values, as defined previously [49, 50]. We also evaluated the Width_{10–90} value (90^{th} percentile - 10^{th} percentile) in this histogram analysis.

The qualitative APTw signal features (ring-like, nodular, or patchy) of the MGMT unmethylated and methylated GBMs were further analyzed, using standard MRI sequences as a reference by the two above-mentioned radiologists with consensus. Then, invasive lobes of each lesion were recorded. The lesions with ventricles involved were specifically noted.

Histopathological evaluation

Operative tissue samples were re-evaluated by an experienced pathologist (Y.W.), blinded to the imaging findings, using the newest 2016 WHO classification of central nervous system tumors. The MGMT methylation status was assessed with a methylation-specific polymerase chain reaction, as described in the literature [51].

Statistical analysis

Mann-Whitney U-tests were used to analyze the statistical differences between quantitative APTw parameters for methylated and unmethylated GBMs after normality testing. Receiver operating characteristic (ROC) curves for the significant different APTw parameters were used to assess the diagnostic performance. Statistical analyses were performed using statistical software (SPSS, Version 23; Chicago, IL). P values < 0.05 were considered statistically significant.

Results

Patient characteristics

Eighteen patients (aged 20–67 years old), who fulfilled the eligibility criteria, based on their medical records, were retrospectively analyzed. Tumor tissue samples were available for MGMT analysis from all subjects by gross total resection (n = 15) or subtotal resection (n = 3). A methylated MGMT promoter was found in 10 cases (55.6%, 47.3 \pm 14.3 years), and an unmethylated MGMT promoter was proven in eight cases (44.4%, 51.1 \pm 12.4 years). In this retrospective study, three of the subjects were included in a previous paper [29].

MTRasym spectra for MGMT unmethylated and methylated GBMs

The average MTR_{asym} spectra for the two GBM groups—with a methylated MGMT promoter and an unmethylated MGMT promoter—were compared to explore the specific characteristics of the APT effect at an offset of ~3.5 ppm downfield from water (Fig. 2). In the offset range of 1–4.5 ppm, both GBM groups demonstrated stronger protein-based APT (at 3.5 ppm) and other CEST effects, compared to the CNAWM. For human studies at 3T, no obvious APT effect can be clearly observed at 3.5 ppm offset, due to the larger voxel size, the larger B₀ inhomogeneity, and the smaller absolute offset range. Notably, the CEST signal intensities in the offset range of 1–4.5 ppm were relatively higher in MGMT unmethylated GBMs than in MGMT methylated GBMs.

APTw image features for MGMT unmethylated and methylated GBMs

Of eight MGMT unmethylated GBMs, three lesions were limited to one single lobe, three lesions had infiltrated two lobes, and two lesions were mainly involved in the third ventricle. The APTw images often demonstrated these MGMT unmethylated GBMs as highly heterogeneous, ring-like or single nodular, hyperintense lesions, compared to the CNAWM. Figure 3A and B shows two typical examples of standard and APTw MR images for two MGMT unmethylated GBMs.

Of 10 patients with MGMT methylated GBM, seven cases were located in one single lobe, one case involved the right parieto-occipital lobes, and two involved the third ventricle. These MGMT methylated GBMs showed ring-like, scattered patchy, or nodular heterogeneous APTw hyperintensity, compared to the CNAWM. Figure 3C and D shows one example of a patient with an MGMT methylated GBM.

Quantitative APTw analyses

Based on the quantitative analysis and comparison, the APTw value metrics (Mean, Variance, 50^{th} percentile, 90^{th} percentile, and Width_{10–90}) were significantly different between the two groups (p < 0.05), and lesions that harbored an unmethylated MGMT promoter showed higher values than did MGMT methylated lesions, as listed in Table 1.

Figure 4A shows the whole-tumor APTw histograms obtained for GBMs with unmethylated and methylated MGMT promoters. The GBMs with an unmethylated MGMT promoter encompassed more voxels with APTw hyperintensity, compared to methylated GBMs. Histogram-based APTw metrics for two cohorts of GBMs are summarized in Table 1. With regards to the differences between the histogram-based APTw value metrics, the MGMT unmethylated GBMs had significantly higher Mean (2.54 ± 0.41 vs. 2.01 ± 0.42 ; P = 0.022), Variance (1.01 ± 0.34 vs. 0.59 ± 0.24 ; P = 0.011), 50th percentile (2.54 ± 0.36 vs. 1.99 ± 0.41 ; P = 0.012), 90th percentile (3.71 ± 0.45 vs. 2.93 ± 0.53 ; P = 0.006), and Width₁₀₋₉₀ (2.31 ± 0.42 vs. 1.87 ± 0.41 ; P = 0.049) values than the MGMT methylated GBMs. The 10th percentile and Mode values showed a higher trend in the MGMT unmethylated group, compared to the MGMT methylated (P = 0.186, 0.086, respectively). Skewness and Kurtosis showed no difference between the two groups (P = 0.963, 0.934, respectively), which indicates that the shapes of the histograms from the two groups are almost identical.

Prediction of MGMT promoter methylation status with APTw metrics

Mean, Variance, 50th percentile, 90th percentile, and Width_{10–90} APTw values differed significantly between the MGMT unmethylated and MGMT methylated groups. Based on ROC curve analyses (Fig. 4B), the 90th percentile values showed the highest area under the ROC curve (AUC = 0.856), and the Mean showed the highest accuracy (83.3%) in predicting the MGMT methylation status (Table 1). MGMT methylation status was, thus, predictable with APTw imaging, non-invasively.

Discussion

Recently, increasing evidence suggests that MGMT methylation status could be a strong predictive and prognostic factor in the GBM patient cohort who are undergoing chemotherapy with alkylating agents, both in newly diagnosed [8, 52–54] and recurrent [55, 56] patients. Thus, MGMT methylation status has been served as a stratification marker in randomized clinical trials. Predicting the MGMT methylation status before treatment, with methods such as MRI, is of paramount importance. Nevertheless, the early results published, to date, have presented some controversial conclusions with regard to the correlation between MRI parameters and MGMT methylation status [19, 57, 58]. When applying machine-learning and texture analysis, the texture features originating from T_2 w images were claimed to reach an AUC of 0.85 [16], or an accuracy of 71% [15]. The texture analysis reveals subtleties not seen by an observer to aid in determining the MGMT status, but at the cost of a huge amount of computational power and time-consuming registration.

Our study shows that GBMs with an unmethylated MGMT promoter are typically associated with relatively higher APTw signal intensity values, compared to methylated GBMs. Multiple APTw-MRI metrics are potentially capable of identifying the MGMT methylation status in GBMs. To the best of our knowledge, this is the first presentation to evaluate correlations between APTw imaging features and MGMT promoter methylation status in patients with GBM. It was anticipated that the potential to determine MGMT status noninvasively could improve treatment efficacy in patients with recurrent GBM, because the repeated surgery only for MGMT status determination is not the optimized regimen in clinical practice.

In this study, we implemented the whole-tumor-ROI-based histogram analysis for APTw value assessment. Strong and significant higher histogram-based Mean, Variance, 50^{th} percentile, 90^{th} percentile, and Width_{10–90} APTw values were identified in MGMT unmethylated GBMs, compared with methylated GBMs (Table 1). The corresponding AUCs were 0.813, 0.825, 0.837, 0.850, 0.856, and 0.763, respectively, for the determination of the MGMT methylation status (Fig. 4B). Among the aforementioned APTw parameters for the feasibility of the discrimination of the MGMT genotype of GBMs, the Mean APTw value showed the highest diagnostic accuracy (83.3%). Therefore, the APTw signals could be valuable imaging biomarkers with which to predict MGMT methylation status in GBMs.

Among the histogram parameters that were utilized in this study, the Skewness and Kurtosis, which depict the lack of symmetry and normal distribution of the data shape, as well as the Mode, which represents the value with highest frequency to be sampled, showed no difference (Fig. 4A). The most significantly different portion was found at the right tail in the histogram, where the 90th percentile was assigned. Thus, it could be reasonably extrapolated that the GBMs with an unmethylated MGMT promoter usually present a larger proportion of voxels with hyperintensity on APTw images than the MGMT methylated GBMs.

Generally, the stratification accuracy of our proposed APTw-MRI approach is a little higher than other methods reported, with quantitative imaging biomarkers extracted from some

advanced MRI sequences, and is comparable with the texture analysis when investigating big MRI datasets. The MGMT promoter methylation in cancer suggests a decrease in protein expression [59]. This may affect other protein expressions downstream of MGMT. Therefore, protein-based APTw-MRI is potentially a novel imaging biomarker for the prediction of MGMT methylation status. It should be kept in mind that the APTw-MRI signal is associated with a large group of cellular (mainly cytosolic) proteins, each contributing multiple amide groups. The exact quantitative explanation demands a further proteomics study, as shown in the previous work [60].

There are a few limitations that relate to our conclusions, which merit discussion. The first limitation was the relatively small sample size used in our study, necessitating a future large-scale study to obtain more conclusive results. The second was the semi-quantitative nature of the current APTw signal intensity, due to the contamination by the upfield nuclear Overhauser enhancement and other effects [43, 48]. Fortunately, several improved APTw imaging analysis or acquisition approaches [61–66] have been proposed to achieve a more pure APT effect. Notably, evidence has also shown that the APT effect would be the major contributor to the APTw image contrast obtained in our experimental setting [43, 67, 68]. Finally, APTw MRI is very sensitive to motion. Artifacts associated with the intraventricular CSF pulsation can often be found in ventricles on APTw images that could disturb the clinical diagnosis.

In conclusion, our early results show that multiple APTw-MRI metrics provide valuable information for the noninvasive discrimination of MGMT promoter methylation status in GBMs. The findings are of paramount importance for the management of alkylating agent chemotherapy and of particular benefit for ambiguous recurrent cases where biopsy or reoperation could potentially be avoided.

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Abbreviations and acronyms

APTw	amide proton transfer-weighted
MRI	magnetic resonance imaging
ROC	receiver operator characteristic curve
AUC	area under the curve
MGMT	O6-methylguanine-DNA methyltransferase
GBM	glioblastoma

CEST	chemical exchange-dependent saturation transfer
T ₂ w	T ₂ -weighted
T ₁ w	T ₁ -weighted
FLAIR	fluid-attenuated inversion recovery

gadolinium-enhanced T₁-weighted Gd-T₁w

ADC apparent diffusion coefficient

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Key Points

- APTw-MRI is applied to predict MGMT promoter methylation status in GBMs.
- GBMs with unmethylated MGMT promoter present higher APTw-MRI than methylated GBMs.
- Multiple APTw histogram metrics can identify MGMT methylation status.
- Mean APTw values showed the highest diagnostic accuracy (AUC = 0.825).



Fig. 1.

With reference to $T_{2}w(A)$, $T_{1}w(B)$, $Gd-T_{1}w(C)$, and APTw (**D**) images, an example of the placement of ROIs on the Gd- $T_{1}w$ image (**E**) and co-registered APTw image (**F**). One large ROI (black lines) contouring the whole area of Gd enhancement within the lesion on the Gd- $T_{1}w$ image was drawn and defined as the tumor mass. In addition, one ROI was placed in the CNAWM for normalization. The white arrow indicates artifacts associated with ventricles.

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A,B Comparison of the average MTR_{asym} spectra of the tumor mass and CNAWM for GBMs with an unmethylated MGMT promoter (n = 8, A) and a methylated MGMT promoter (n = 10, B). The CEST effects were clearly observed at multiple frequencies in the 1–4.5 ppm frequency offset range. C The average relative MTR_{asym} spectra of the tumor mass (MTR_{asym} spectra from tumor mass - MTR_{asym} spectra from CNAWM). The error bars in the figures are shown as standard errors.

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Fig. 3.

A, B Conventional and APTw MR images of two typical GBMs with an unmethylated MGMT promoter, illustrating the heterogeneous ring-enhancement characteristic of the disease on Gd-T₁w image. APTw image demonstrated the lesion with ring-like, strong hyperintensity. **C, D** Conventional and APTw MR images of two typical MGMT promotermethylated GBMs. Gd-T₁w image demonstrated a patchy Gd-enhancing tumor mass. APTw image showed the masses as a hyperintense nodule (C) or scattered patch (D).

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Fig. 4.

A Comparison of average tumor mass APTw histograms for all GBMs with an unmethylated (n = 8) and a methylated (n = 10) MGMT promoter. The GBMs with an unmethylated MGMT promoter show more voxels with APTw hyperintensity. **B** ROC analysis of APTw metrics as imaging biomarkers with which to distinguish GBMs with an unmethylated MGMT promoter from GBMs with a methylated MGMT promoter. The 90th Percentile APTw value demonstrated the highest AUC, at 0.856. AUC: area under the curve.

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Table 1

Quantitative APTw intensity values and the corresponding diagnostic performance for MGMT unmethylated and methylated GBMs.

	Unmethylated	Methylated	P value	AUC (95% Confidence interval)	Cut-off value	Sensiu vuy	Specificity	Accuracy
Mean	$2.54{\pm}0.41$	2.01 ± 0.42	0.022^{*}	0.825 (0.626–1.000)	2.26	87.5%	80%	83.3%
Variance	$1.01 {\pm} 0.34$	0.59 ± 0.24	$0.01 \ 1^{*}$	0.837 (0.649–1.000)	0.94	62.5%	%06	77.8%
Skewness	0.04 ± 0.52	0.06 ± 0.87	0.963					
Kurtosis	4.67 ± 1.93	4.80 ± 3.48	0.934					
10 th percentile	1.40 ± 0.53	1.06 ± 0.45	0.186					
50 th percentile	$2.54{\pm}0.36$	1.99 ± 0.41	0.012	0.850 (0.672–1.000)	2.25	75%	80%	77.8%
90 th percentile	$3.71 {\pm} 0.45$	2.93 ± 0.53	0.006^*	0.856 (0.674–1.000)	3.25	87.5%	40%	77.8%
Width ₁₀₋₉₀	2.31 ± 0.42	$1.87{\pm}0.41$	0.049^{*}	0.763 (0.537–0.988)	2.15	62.5%	80%	72.2%
Mode	2.45 ± 0.38	2.05 ± 0.47	0.086					