



# HHS Public Access

Author manuscript

*Behav Brain Res.* Author manuscript; available in PMC 2019 October 15.

Published in final edited form as:

*Behav Brain Res.* 2018 October 15; 352: 125–132. doi:10.1016/j.bbr.2017.10.005.

## Fetal Alcohol Spectrum Disorders: Zebrafish in the analysis of the milder and more prevalent form of the disease

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### Abstract

Fetal Alcohol Spectrum Disorders (FASD) represent a large unmet medical need. Exposure of the developing human embryo to alcohol can lead to life-long suffering. Despite the well documented deleterious effects of alcohol on the developing fetus, pregnant women continue to drink alcohol, and FASD remains the leading cause of preventable mental retardation and other behavioral abnormalities. Particularly prevalent are the milder forms of the disease cluster, representing children who do not show obvious physical signs and who may be undiagnosed or misdiagnosed. To develop treatment and diagnostic tools, researchers have turned to animal models. The zebrafish is becoming one of the leading biomedical research organisms that may facilitate discovery of the biological mechanisms underlying this disease and the identification of biomarkers that may be used for diagnosis. Here we review the latest advances of this field, mostly focussing on the discoveries made in our own laboratory and others with zebrafish employed to analyze the effects of moderate to low level of exposure to alcohol. We argue that the zebrafish represents unique advantages, and adding information obtained with this species to the mix of other animal models will significantly increase translational relevance of animal biomedical research for the analysis of human FASD.

### What is Fetal Alcohol Spectrum Disorder?

First introduced by Jones and colleagues, in 1973 [1], the term ‘fetal alcohol syndrome (FAS)’ was used to describe a pattern of physical abnormalities seen in children born to alcohol abusing mothers. Further investigation led to recognition of particular cranio-facial malformation, growth retardation, [1], as well as behavioural and cognitive deficits including social-emotional impairments as characteristic of fetal alcohol exposure induced abnormalities [2]. As knowledge and awareness of negative effects associated with fetal alcohol exposure grew, problems with the term “Fetal alcohol syndrome” and its diagnostic criteria came to light. Not all children born to alcohol abusing women displayed the severe symptoms previously described, i.e. cranio-facial malformation or growth retardation [3]. Furthermore, for some children, particularly those in adoptive or foster care, reports of maternal alcohol consumption were impossible to access [4]. The current diagnosis of Fetal Alcohol Spectrum Disorder (FASD) is now used for individuals with known or suspected

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fetal alcohol exposure and this diagnosis now includes cases with much less severe symptoms than those of FAS [3]. The modern diagnosis of FASD now encompasses FAS (with and without confirmation of maternal drinking), partial FAS, alcohol related neurological defects (ARND) and alcohol related birth defects (ARBD)[4]. Currently in the western world, FASD is the most common preventable form of developmental disability [5].

It has been confirmed that the most detrimental effects of fetal alcohol exposure are associated with periods of binge drinking [6]. Alvik and colleagues [7] reported that moderate to heavy drinking, entailing 5–7 drinks per drinking event, is less harmful than when a pregnant woman consumes 8+ alcohol beverages per drinking event. This latter pattern of fetal alcohol exposure was correlated with later problems in the child to a greater extent than other patterns of drinking [7]. May et al [8] found that women who ceased alcohol consumption after their first trimester decreased the risk of an FASD diagnosis in their offspring by up to 5 times, while alcohol consumption during the entire gestational period expectedly led to the worst behavioural outcomes in the child [8].

In Canada, the current diagnostic criteria begin with whether prenatal exposure to alcohol is confirmed or not, followed by the presence of 3 sentinel facial features (which include palpebral fissure length, philtrum and upper lip scores, as a small palpebral fissure, smooth philtrum and thin upper lip are markers for early alcohol exposure during the first trimester, then central nervous system (CNS) impairment [3]. This leads to the possible diagnosis of an individual to be: presenting with FASD, being at risk of FASD (this diagnosis is used with young children where CNS impairment may yet to be difficult to ascertain), and non-FASD individuals who require developmental care [3]. A recent collection of FASD data in Canada revealed that only 2.1% of individuals diagnosed with FASD met the criteria for FAS [9]. This same study also found that there were no disabilities that were specific to any subgroup of FASD individuals [9]. As the cost of FASD in Canada has been estimated from \$3.6 to 7.3 million per year [10], it is imperative to recognize that individuals which fall under the FASD diagnostic umbrella will display varied impairments and deficits. May et al. [11] have estimated the prevalence of FASD in the US and Western Europe to be 3–4 children per thousand, while a recent meta-analysis of international data estimates the world-wide prevalence of FAS to be 14.6 per 10 000 individuals [12]. That is, the latter study found that over 110 000 infants are born with the severe form of the disease, FAS, every year.

While research is being conducted to examine the mechanisms, e.g. key brain areas involved in fetal alcohol spectrum disorders [13,14,15], as well as the full spectrum of behavioral alterations resulting from embryonic alcohol exposure [16,17], most studies have focused on the analysis of the effects of high to moderate levels of alcohol exposure [18]. Information gathered from such studies is clearly needed, however, the findings may not extrapolate to patients exposed to only low levels of alcohol during their embryonic development. Briefly, the effects of exposure to low doses of alcohol remain understudied, a notable hiatus in research given that the milder FASD cases have been shown to be most prevalent [19,9].

## **Fetal alcohol exposure leads to lasting impairment in several domains of social behavior in humans**

Fetal alcohol exposure can result in impairments in a variety of neurodevelopmental domains, which include cognition, memory, attention, executive function, communication and social behavior [3]. Of central importance to the focus of this review are the effects of early alcohol exposure on affective and social behaviors.

Impaired social behavior following fetal alcohol exposure has been documented in individuals across their lifespan [20], from studies reporting weak sucking in nursing infants [21] and disorganized mother-child attachments [22] with babies with irritable temperaments [23] to adolescents and adults who have been found to display high rates of sexually inappropriate behavior [24] and higher rates of mental health disorders including depression [25,26,27] and increased unemployment [28]. Thomas et al [29] compared adaptive behaviors of children with FAS and IQ-matched control children and neurotypical control children. These authors measured social skills in various domains, and found that children with FAS displayed the most severe impairment in interpersonal relationship skills and concluded that the development of social abilities in FAS children was arrested [29]. More recent studies have found children with fetal alcohol exposure to score lower than controls in social skills [30], and to suffer from decreased ability in a social problem-solving task [31] and from difficulty interpreting the mental states and emotions of others [32]. Others found children with FASD to display poor social judgment and to have difficulty communicating in social contexts [33]. The latter study also found FASD children not to be able to learn from their past mistakes and to possess impaired or underdeveloped moral judgement.

Nash et al [17, 34] attempted to identify the defining behavioral characteristics of children with FASD with the goal of creating a screening tool to facilitate early intervention. The identified characteristics specific to FASD children and not common, for example, with ADHD (attention-deficit/hyperactivity disorder) [35] included display of cruelty, lack of guilt, acting young for the age, and the tendency to steal [17]. Children suffering from FASD have been shown to benefit from continued intervention especially when it is started early [36, 37]. Thus, improved diagnosis made as early as possible would have a major impact on the affected children and their families.

## **Mammalian models of human FASD**

A variety of animals has been used to study the effects of fetal alcohol exposure for decades [20]. Although the scientific literature on the effects of fetal alcohol exposure in humans is continually expanding, the measures of alcohol consumption are predominately based upon the mother's self report [38,39]. Furthermore, no human study can control the amount of alcohol the fetus is exposed to, the embryonic developmental time point at which the exposure occurs, or the frequency and duration of the exposure. Animal studies are necessary to systematically control these factors, and to understand the effects of parametrically changing every one of these factors. We know from the human literature that higher concentrations of alcohol exposure are most harmful to the developing fetus, and that periods of binge exposure result in the most impairment (cognitively and behaviorally) in

children [6,7]. Fetal alcohol effects have been studied in many species of animals focussing on the more severe end of the FASD spectrum, with findings mirroring some of what is seen in humans.

The facial and cranial malformations which are sentinel markers of high level of fetal alcohol exposure in infants and children [3] are also seen in macaques [40] and mice [41]. Furthermore, macaques exposed to prenatal alcohol were shown to display impairments in visual memory [42], as well as in sensory processing similar to children with FASD [43]. Prenatal alcohol exposed rats have been documented to avoid social interactions with conspecifics [44] and to display impaired social recognition of a known conspecific [45]. Both male and female rats exposed to fetal alcohol have been found to display sex-specific changes in play behavior with alcohol exposed males decreasing the frequency of play, while alcohol exposed females increasing the frequency of play compared to sex matched controls [46]. Increased aggression following prenatal exposure in rats has also been reported [47, 48], corroborating what has been reported in school aged children with FASD [49]. Although dominant in the literature, mammalian models are not the only attempts to investigate the effect of embryonic alcohol exposure in vertebrates. The zebrafish has been also proposed as an excellent model organism for this purpose [50, 51, 52].

### **General considerations of what makes an animal model acceptable with a focus on zebrafish**

Before discussing some of the specific advantages of zebrafish and the reasons for the increasing popularity of this species in behavioral neuroscience in general and FASD modeling in particular, we wish to consider a general question associated with the use of animals as models of human disorders. This question is particularly relevant in case of zebrafish, a species whose last common ancestor with humans lived approximately 400 million years ago. Briefly, the question concerns translational relevance. That is, can we use a species that is so distantly related to humans to draw any conclusions about human disorders? This single question is usually broken down to three distinct but related problems, known as the criteria of a proper animal model. First, the model should have face validity, i.e. the phenotypical alterations seen in the animal model should mimic at least some observable aspects of the human disorder. Second, the animal model should have construct validity, i.e. it should be mechanistically relevant for the human disorder. Third, it should have predictive validity, i.e. it should be able to detect efficacy of drugs (or in case of genetics, known effects of known mutations) previously shown to influence relevant phenotypes.

While in general the above three criteria are accepted broadly by the scientific community, what each criterion actually means may not be that clear. For example, deciding about face validity is not simple when one compares zebrafish and human behavioral responses. Central nervous system alterations induced by embryonic alcohol exposure affecting mechanisms underlying social behavior may manifest quite differently in zebrafish versus in humans. For example, how could one evaluate reduced empathy towards others in zebrafish? The evaluation of construct validity is also a complicated matter. The only level of the biological

organization at which this evaluation is clearly possible is the genome. The nucleotide sequence of human and homologous zebrafish genes has been found to be reaching and often exceeding 70% [53]. This level of sequence homology ascertains that if one identifies a zebrafish gene as being involved in FASD, for example, as a gene that encodes a protein that enhances or reduces the effects of embryonic alcohol exposure, one can be almost certain that a homologous gene involved in a similar function in humans will be identified. However, above the genetic level, mechanistic similarities between zebrafish and human may be more complicated to evaluate. Nevertheless, numerous evolutionarily conserved features, from neuroanatomy through neurotransmitter systems to behavior have been identified between zebrafish and humans [54], implying that zebrafish may be a useful model in biomedical research aimed at the analysis of human disorders. Last, we note that the requirement of predictive validity is often misunderstood. It is regularly used in pharmaceutical research whereby the model must show efficacy with previously developed mammalian drugs. The problem, however, with this requirement is that it may make the novel animal model miss novel compounds that act through previously undiscovered pathways or mechanisms.

### **Why should we use zebrafish in FASD research: General and specific advantages of this species?**

The zebrafish has become an important scientific tool in the study of the brain and behavior. Over the past 3 decades, it has gained in popularity as a test subject for a variety of reasons. Due to its diminutive size, large number of subjects can be easily and economically housed. Furthermore, its fast embryonic development and short period to reach sexual maturity facilitate breeding and cross-generational studies. But most importantly, from the perspective of FASD research it offers precision and complete control over when, how much and for how long the developing embryo is exposed to alcohol because eggs are externally fertilized and develop outside of their mother. Zebrafish eggs can be placed in an alcohol solution and the developing embryo inside the egg can absorb alcohol, an immersion-based alcohol delivery method that is non-invasive compared to most alcohol administration protocols employed with mammals.

The first pioneering studies that used zebrafish to investigate the behavioral and physical aspects of FASD employed high concentrations of alcohol and/or exposed the zebrafish embryos to alcohol for a prolonged period of time [55,56,57,58,59, 60]. These studies found that embryonic alcohol exposure led to increased mortality [56,61,59,], gross structural deformities [61;61,62], smaller body size [59;62], deformities to the inner ear and lateral line system [60]; and visual system defects[58], including cyclopia (eyes fuse together) [55,56, 59], microphthalmia (reduced sized eyes) [57,67]. High doses of embryonic alcohol and/or prolonged exposure periods have resulted in hypoactive larvae, with intermediate doses of alcohol leading to hyperactivity [63, 64]. These pioneering studies demonstrated that the zebrafish is a translationally relevant model for FASD, as high concentration and long alcohol exposure was found to lead to abnormalities that mimicked those found in the severe human forms of the FASD disease cluster, i.e. in FAS [57]. However, these zebrafish models of FAS did not recapitulate the symptoms of the less severe forms of FASD, the

symptoms of patients that exhibited behavioral, e.g. social behavioral, abnormalities without anatomical changes. Given that these less severe forms of the FASD cluster represent the greatest majority of cases, the need for a new animal model became obvious.

## A zebrafish model of the mild form of human FASD

In 2009, Fernandes & Gerlai exposed zebrafish embryos to low concentrations of alcohol and only for a brief 2-hour period at 24th hour post fertilization (hpf) [50]. This dosing procedure was chosen to mimic the milder and more prevalent forms of FASD, and to avoid the physical deformities found in severe human FAS. The exposed fish showed no gross malformations, increased mortality or morbidity and appeared to develop normally. At their adult stage, these zebrafish, which were exposed to alcohol concentrations ranging between 0.25 and 1.00% (vol/vol external bath), showed no apparent changes. However, thorough behavioural phenotyping revealed an interesting and fairly specific alteration. When presented with animated images of conspecifics, a computerized shoal stimulus at their adult stage of development, the fish that were exposed to alcohol during their embryonic development exhibited a significant and prior alcohol concentration dependent impairment. Fish exposed to lower concentrations of alcohol spent significantly reduced amount of time near the shoal stimulus compared control, alcohol unexposed fish. Whereas fish that were exposed to the highest concentration (1%) of alcohol during their embryonic development showed no shoal stimulus induced response at all, i.e. this highest dose of prior alcohol exposure abolished the shoaling response [50]. Importantly, the significant reduction of shoaling response found in the alcohol exposed fish was replicated using a different behavioral test and stimulus presentation method. Buske and Gerlai [68] used the same dosing procedure, but measured shoal cohesion in larval fish from post-fertilization day 7 through day 102 using live, freely moving shoals. Control, alcohol unexposed, fish were found to reduce inter-individual distance amongst shoal members as fish matured, resulting in a tighter, i.e. more cohesive shoal. Fish exposed to embryonic alcohol, on the other hand, exhibited significantly larger inter-individual distances, i.e., formed a much looser shoal, an impairment that became gradually more prevalent as the fish matured [68].

What could have led to the impaired ability of the alcohol exposed zebrafish to respond to social stimuli? It is possible that alcohol exposure impaired motor function and as a result, the exposed zebrafish could not swim to their shoal mates or maintain a proper distance from them. However, this possibility was excluded by finding no alterations in motor function. Swim path parameters, including total distance swam, turn angle, or the location of swimming were all statistically indistinguishable between control and alcohol exposed fish [50, 68]. Past studies which employed much longer exposure periods have also found locomotory responses in adult alcohol-exposed fish to be comparable to those of control animals [59,69]. Another possibility was that alcohol impaired the development of the visual system [56,57], and thus the alcohol exposed fish could not properly see their shoal mates or the animated shoal. However, this possibility was also excluded because the alcohol exposed fish were found to respond to the shoal images with reducing their activity just as much as control fish did [50] proving that they could see the animated images. Furthermore, others [68] have replicated these findings using freely moving shoals. The latter study found reduced shoal cohesion (increased inter-individual distance) among adult zebrafish exposed

to alcohol during their embryonic development as compared to control fish while detected no other changes in their activity or movement pattern. It is also notable that in the latter test, the freely moving experimental zebrafish could use both visual as well as lateral line cues (echolocation). In summary, these results suggested that the impaired responses of embryonic alcohol exposed fish to social stimuli were not due to alteration of simple performance features including motor function and perception. Another important discovery made more recently demonstrated that the impaired social response was long lasting, it could be demonstrated in two-year old zebrafish [70]. The latter result suggests that exposure to even low concentrations of alcohol and for a very short period of time during embryonic development of the zebrafish can essentially have life-long deleterious consequences, a notable warning for women inclined to consume alcohol during their pregnancy. What could be the behavioral mechanism underlying the impaired social behavior responses of the embryonic alcohol exposed zebrafish? Does finding altered responses to social stimuli with no changes in vision and motor function mean altered social behavior? Not necessarily.

The abnormal social behavioral responses documented by Fernandes and Gerlai [50] and Buske and Gerai [68] could be due to three possible reasons. The main adaptive function of shoaling has been shown to be predator avoidance [71]. Thus, altered fear or anxiety induced by embryonic alcohol exposure may also modify responses to shoaling stimuli, not because of altered mechanisms underlying social behavior per se, but because of altered mechanisms underlying fear and/or anxiety. Second, Al-Imari and Gerlai [72] showed that the sight of conspecifics is rewarding. A generalized impairment of motivation, e.g. altered mechanisms associated with reward pathways of the brain of zebrafish, may also lead to impaired responding to the social stimuli. Third, and finally, alterations more specific to mechanisms underlying social behavior itself, may be the reason.

### **Possible behavioral mechanisms underlying the impaired response to social stimuli in zebrafish exposed to low concentrations of embryonic alcohol**

To explore whether a change in fear or anxiety may be responsible for the reduction in social behavior previously documented [50,68], control fish and zebrafish treated with different concentrations of embryonic ethanol were exposed to a novel tank and subsequently to an animated image of a natural predator of the zebrafish, the clown knifefish (*Chitala ornata*) [73]. This same image was previously shown to reliably induce an avoidance response in control zebrafish bred and raised in the laboratory with no prior predator experience with this predator [74]. When exposed to a novel tank, zebrafish normally respond by remaining near the bottom portion of the tank, by displaying erratic movements (swimming with sharp and frequent turns), and by freezing (cessation of locomotion) [73]. Control and embryonic alcohol exposed fish were found statistically indistinguishable in the novel tank [75]. Similarly, control and embryonic alcohol treated fish were also statistically indistinguishable in their responses to the animated predator [75]. Anxiety is often defined in animal and human studies as a set of responses elicited by diffuse aversive stimuli that are continually present. Novelty is often used to elicit anxiety. Fear, on the other hand, is defined as a set of

behavioral responses induced by the appearance of a clearly defined aversive stimulus. Seguin et al [75] thus concluded that embryonic alcohol treatment as employed did not alter fear or anxiety in zebrafish. In summary, the behavioral deficits in shoaling responses induced by embryonic alcohol treatment is thus not likely due to altered fear or anxiety.

The second working hypothesis, i.e. whether a general change in motivation, e.g. a form of anhedonia, may underlie the reduced shoaling response, has not been systematically examined yet, although unpublished preliminary results (Fernandes and Gerlai) suggest that embryonic alcohol exposed fish respond normally to food reward (also see [76]). Thus, it is likely that the impaired response to social stimuli observed in zebrafish that were exposed to alcohol during embryonic development is indeed the result of abnormal social behavior.

What this abnormality entails, however, is not known. For example, the embryonic alcohol treated fish may not recognize conspecifics because they misinterpret social cues. Unfortunately, there is only limited amount of information available on what social cues zebrafish may pay attention to (e.g. [77]). It is possible that misinterpretation of social cues is specific to the shoaling context, but it is also possible that the impairment of the alcohol exposed fish extends to social behaviors other than shoaling, e.g. courtship or aggressive behaviors, an alteration that would mimic what has been found in human children suffering from FASD.

Luckily, the above questions may be easily addressed using zebrafish. For example, physical characteristics of the shoaling stimulus, e.g. the number of fish in the stimulus shoal, the sex composition of the shoal, as well as the visual appearance of the stimulus fish (color, shape, size, pattern), can be systematically manipulated, and the behavioral effects on the experimental fish studied. In a recent study, control, alcohol unexposed, zebrafish have been found to reliably distinguish numerically larger shoals from smaller ones as long as numerical ratios of 2:1 or larger were used [78]. This finding is particularly relevant given that quantity estimation abilities of animals are believed to be the evolutionary precursor of higher mathematical abilities found in humans [79,80] and FASD patients suffer from diminished mathematical reasoning [81–83]. It has also been shown that these mathematical impairments become more pronounced as maternal alcohol intake increases [84, 8].

Given that human FASD patients have often been found to misinterpret social cues and have aberrant sexual behavior, it is possible that exposure to embryonic alcohol disrupts some aspects of sexual behavior in zebrafish too. This could easily be analyzed in the context of reproductive behaviors or courtship behaviors, but could also be systematically investigated using animated images of shoals differing in sex composition. Such studies may reveal more finely grained differences between control and embryonic alcohol exposed zebrafish. They may also ascertain whether the social behavior impairing effects of the alcohol exposure are context specific. For example, Ruhl et al [85] reported that male zebrafish (not exposed to embryonic alcohol) prefer all female shoals to all male shoals, while female zebrafish prefer all male shoals to all female shoals. Notably, sex-specific effects of fetal alcohol exposure on social behaviors have been documented in mammals. Dahlgren et al. [86] reported that following alcohol exposure, adult male rats displayed female-typical sexual behavior when time spent near prospective mating partners was measured. This study was built upon



evidence showing that males displayed increased lordosis responses following prenatal alcohol exposure [87]. Neither studies found evidence of changes to sexual behaviors following alcohol exposure in female animals [86,87]. These findings are supported by more recent rodent studies which have found nearly half of the males exposed to prenatal alcohol not to be able to ejaculate [88]. Lugo et al [89] found that prenatal alcohol exposed male rats displayed decreased aggression and more female typical behaviors, such as hopping and darting, compared to unexposed control males.

Saverino & Gerlai [77] presented zebrafish with images of two moving animated shoals simultaneously. A specific feature of the presented stimulus fish was systematically altered, e.g., colour, pattern or shape, and the preference of the experimental zebrafish for the altered image versus the unaltered species typical image was measured. This pioneering study revealed numerous interesting and unexpected findings demonstrating that certain visible characteristics of the image enhance, others reduce, while yet others do not affect choice between or preference for the altered versus unaltered conspecific images [77]. In other words, experimental zebrafish were not indifferent to certain features of the presented images, an observation that may be utilized in the analysis of social behavior related abnormalities found in embryonic alcohol treated zebrafish too.

The question whether embryonic alcohol treatment altered aggression has also not been studied. Nevertheless, rodents exposed to alcohol during their embryonic development have been shown to exhibit increased aggression [90,91,48]. Aggression can also be reliably induced and quantified using zebrafish, for example, by pairing an experimental male with a size-matched unknown male, or by placing a mirror with a solitary male [92–94]. Recent pilot work in our lab has found fish exposed to 0.50% embryonic alcohol to display significantly more attacks, such as striking and chasing, towards a novel male compared to alcohol unexposed control fish (Seguin & Gerlai, unpublished).

Aside from social behaviors, there may be other behavioral impairments induced by alcohol exposure during embryonic development in zebrafish. For example, learning and memory have been found impaired in FASD patients [84, 95], in primates [96] as well as in rodents [45,97,98]. The severity of learning impairment has been found to positively correlate with the frequency of maternal binge drinking episodes [84]. Embryonic alcohol exposure has been found to particularly affect acquisition of memory [99]. Similar impairments in acquisition of memory has also been documented in rhesus monkeys [96]. In zebrafish, exposure to higher concentrations of alcohol administered for prolonged period of time has been found to induce learning deficits [51]. But immersion of the eggs into even as low as 0.25% (vol/vol) concentration of alcohol for as short a period as 2h during embryonic development has been found to lead to lasting impairment of associative learning in zebrafish [76]. However, notably, this learning deficit was induced when the embryos were exposed to alcohol at their 16<sup>th</sup> hour post-fertilization stage but not when they were exposed to the same dose of alcohol at their 24<sup>th</sup> hour post-fertilization stage (Fernandes & Gerlai personal observation), raising the intriguing, and yet unexplored, possibility that social deficits versus learning deficits may result from alcohol exposure at different stages of embryonic development. The above results demonstrate how little we know about the behavioral mechanisms underlying embryonic alcohol induced behavioral changes. Yet, they

also show that zebrafish may be a promising tool with which such changes can be studied. Similarly, little we know about the molecular or neurobiological mechanisms underlying embryonic alcohol exposure induced behavioral abnormalities in zebrafish. Nevertheless, the first pioneering studies have already revealed some notable changes.

## **Neurobiological correlates of altered social behavior in embryonic alcohol exposed zebrafish: The first steps toward discovering biological mechanisms**

Alcohol is a dirty drug, i.e. it is known to directly and indirectly interact with a large number of molecular targets and alter numerous biochemical, synaptic and other neurobiological processes. For example, a recent DNA microarray analysis studying the effects of chronic alcohol exposure, identified close to 2000 differentially expressed genes responding to alcohol treatment, 60% of which were functionally unknown [100]. Coupled with this complexity is the likely complication associated with a cascade of altered developmental processes that is triggered by all the changes alcohol induces in the embryonic brain at the time of exposure. Briefly, one may expect a large number of molecular and neurobiological mechanisms to be altered in the adult zebrafish brain that develops after embryonic alcohol exposure. To tackle such complexity, systematic and comprehensive analyses/phenotyping may be needed. For example, a comprehensive transcriptome and proteome analysis followed by bioinformatic study may reveal numerous mRNA and protein level changes clustered according to biological function or biochemical interactions. Alternatively, one may opt to employ a large-scale mutagenesis study in which mutations, and ultimately the genes harboring such mutations, that alter, exacerbate or diminish, embryonic alcohol induced changes may be identified. Similarly, one may decide to conduct a comprehensive drug screen to identify small molecules that enhance, alter, or negate the effects of embryonic alcohol exposure. None of these comprehensive, unbiased screens have been conducted yet, however.

Instead, only a hypothesis driven, proof of concept study has been performed. The impetus for this study came from the observation that the sight of conspecifics is rewarding. Al-Imari & Gerlai [72], found that the sight of conspecifics may be employed as a reinforcer in an associative learning task designed for the zebrafish. The authors used the sight of conspecifics (group of stimulus zebrafish placed in a small tank outside of a plus maze) as the unconditioned stimulus (US or reward), and paired it with a color cue (a red plastic cue card placed inside the maze adjacent to the stimulus fish) the conditioned stimulus or CS. They found experimental zebrafish to be able to associate the CS with the US, i.e. in a probe trial (during which no stimulus fish were present) found the experimental subjects to be able to show a significant preference for the previously neutral (not preferred) CS. Subsequently, Saif et al. [101] showed that the appearance of conspecific images triggers a robust increase of the levels of dopamine and DOPAC (dopamine's metabolite) in the brain of zebrafish without changes in other neurotransmitter levels. Furthermore, Scerbina et al. [102] showed that disruption of the dopaminergic system by administration of a dopamine D1-receptor antagonist results in a drug dose-dependent reduction of shoaling in zebrafish. The above results thus suggested that the sight of conspecifics is rewarding, a positive/attractive

stimulus, and that the dopaminergic system, known to mediate reward related processes, is involved in shoaling in zebrafish. Thus, we hypothesized that the reduced shoaling responses seen in embryonic alcohol exposed adult zebrafish may be the result of impaired dopaminergic function.

This working hypothesis turned out to be correct. Mahabir et al. [103] found embryonic alcohol exposure to significantly impair the development of the dopaminergic system. Using high precision liquid chromatography (HPLC), these authors found that the level of dopamine and of DOPAC increased relative to total brain protein weight as zebrafish matured, but this increase was dose dependently blunted, or abolished, by embryonic alcohol exposure, an effect that correlated with the previously observed [50] alcohol dose dependent impairment of shoaling responses. Furthermore, Fernandes et al. [70] revealed that this dopaminergic neurotransmitter system related impairment does not represent altered baseline responses. Zebrafish that were isolated for 24 hours, i.e. were not exposed to conspecifics or their sight for this period of time, exhibited no embryonic alcohol exposure related changes in dopamine or DOPAC levels. However, when these isolated fish were shown animated (moving) images of conspecifics, control fish responded with a robust and significant increase of dopamine and DOPAC levels, but 0.5% and 1.0% embryonic alcohol exposed fish did not. Again, the lack of conspecific sight induced dopamine and DOPAC increase in the embryonic alcohol exposed zebrafish correlated well with the impaired shoaling response also found in these fish. Recently, changes in dopamine receptors have been found following prolonged exposure to embryonic alcohol exposure [104]. It is also notable that the impairment may be fairly specific to the dopaminergic system. A systematic HPLC analysis of potential changes in the levels of neurotransmitters other than dopamine, has found an alcohol dose dependent reduction of serotonin, but no significant changes in the level of any other neurotransmitter, including glutamate, GABA, aspartate, glycine and taurine in the brain of adult zebrafish exposed to alcohol during their embryonic development (Mahabir et al., unpublished results). Of particular societal relevance is the increased risk of developing drug and alcohol abuse disorders in adulthood among patients who were exposed to alcohol during their embryonic development [105, 106]. Animal studies have also provided evidence of such increased risk [107], and suggested that changes in the dopamine system may underlie the behavioral abnormalities [108,109]. Recent studies have reported that early embryonic exposure to alcohol in zebrafish also results in increased addictive behaviors in the adult [104,110]. Using a low-concentration dosing procedure similar to the one employed by Fernandes & Gerlai [50] and Buske & Gerlai [68]. Sterling et al. [110] found that alcohol exposed fish increased their voluntary consumption of ethanol compared to controls. Similarly, Parker et al. reported increased conditioned place preference and habit formation in embryonic alcohol exposed zebrafish [104].

The above results suggest that the activity of the dopaminergic system of the adult zebrafish brain is significantly blunted by embryonic alcohol exposure, and this change does not represent a generalized, or overall, neuronal activity change, but instead it is fairly specific to this particular neurotransmitter system. But what could explain such abnormality? Although the number of dopaminergic neurons of the adult zebrafish brain is fairly small, and their neuroanatomical location is well described (see Schweitzer et al. and the references therein [111]) a detailed anatomical study characterizing potential changes induced by embryonic

alcohol exposure in this neurotransmitter system is lacking. Nevertheless, a working hypothesis formulated on the basis of the results presented in Fernandes et al. [70] is that dopaminergic neurons themselves may be intact. This hypothesis is supported by finding baseline (unstimulated) dopamine and DOPAC levels in embryonic alcohol exposed fish statistically indistinguishable from those found in control fish. The results imply that both dopamine production (levels of dopamine) as well as dopamine release and metabolism in the synaptic cleft (DOPAC levels) are unaltered. Yet, when stimulated using a natural stimulus (sight of conspecific images), the dopaminergic system remains unresponsive in the alcohol exposed fish. This result implies that afferent connections to the dopaminergic system may be altered during brain development by embryonic alcohol exposure, a working hypothesis whose validity will need to be ascertained in the future.

One known mechanism via which alcohol may induce abnormalities, including in the development of neuronal connections, is apoptotic cell death. Alcohol increases apoptotic cell death [112,113], a process that is otherwise crucial for neuronal pruning and thus normal brain development [114]. Studies employing, for example, higher alcohol concentrations or longer exposure periods have found developmental toxicity [115], and numerous developmental abnormalities [116], including decreased neuronal cell counts and abnormal patterns of neural branching [117, 61, 63], as well as upregulation of specific microRNAs [118]. We have started to investigate potential structural alterations and their source following low dose alcohol exposure, and have found a larger number of apoptotic neurons in the brain of zebrafish exposed to embryonic alcohol two hours after the exposure (Mahabir et al., unpublished results). We have also found enhanced expression of the pro-apoptotic protein Bax in the brain of embryonic alcohol exposed zebrafish (Mahabir et al., unpublished results). It is thus likely that neurons connecting to dopaminergic neurons and involved in the stimulation of such neurons may have developed abnormally in the brain of embryonic alcohol exposed fish, a hypothesis that will need to be tested by analyzing the adult connectome and the development of the connectome of embryonic alcohol exposed zebrafish.

## Conclusions

The zebrafish is a relative newcomer in behavioral brain research in general and in the analysis of the effects of embryonic alcohol exposure in particular. Nevertheless, despite its novel status, the increasing number of studies, some of which reviewed in this paper, suggests that this small and relatively simple laboratory organism may be useful in modeling and the analysis of alcohol related human disorders, including FASD. There are multiple reasons for this popularity mostly stemming from the realization that the zebrafish represents a reasonable compromise between system complexity and practical simplicity. Another argument for its use is that adding this species, which is evolutionarily distance from our own, to the list of laboratory study organisms, will enhance translational relevance through our increased ability to identify common features overlapping across these multiple study species. These overlapping features, arguably will allow us to identify evolutionarily most ancient and mechanistically most fundamental aspects of our own biology. The significant behavioral effects of alcohol demonstrated in zebrafish show that this fish, similarly to mammals, including humans, does respond to alcohol in a quantifiable manner. Although the

mechanisms underlying the behavioral changes induced by embryonic alcohol exposure are largely unknown, the first promising results have already been generated. Despite the approximately 400 million years of biological evolution separating the zebrafish from human, homologies, i.e. evolutionary conservation, have been identified at many levels of the biological organization of this fish. Although at the early stages of model development, zebrafish models of human alcohol related disorders show signs of face validity (similar behavioral effects across zebrafish and human), construct validity (similar mechanisms) as well as predictive validity (similar drug effects). It is therefore hoped that the mild (low concentration) embryonic alcohol exposure zebrafish model will advance our understanding of the mechanisms of human FASD, and will lead to both the development of treatments and the identification of diagnostic biomarkers.

## Acknowledgments

Supported by NSERC (311637) and NIH/NIAAA (R01 AA14357-01A2)

## References

1. Jones KL, Smith DW, Ulleland CN, Streissguth AP. Pattern of malformation in offspring of chronic alcoholic mothers. *Lancet*. 1973; 1(7815):1267–1271. [PubMed: 4126070]
2. Streissguth AP, Herman CS, Smith DW. Intelligence, behavior, and dysmorphogenesis in the fetal alcohol syndrome: a report on 20 patients. *J Pediatr*. 1978; 92:363–367. [PubMed: 632974]
3. Cook JL, Green CR, Lilley CM, Anderson SM, Baldwin ME, Chudley AE, ... Conry JL, et al. Fetal alcohol spectrum disorder: a guideline for diagnosis across the lifespan. *CMAJ*. 2016; 188(3):191–7. [PubMed: 26668194]
4. Landgraf MN, Nothacker M, Kop IB, Heinen F. The Diagnosis of Fetal Alcohol Syndrome. *Deutsche Arzteblatt International*. 2013; 110(42):703–710.
5. Clark ME, Gibbard WB. Overview of fetal alcohol spectrum disorders for mental health professionals. *Can Child Adolesc Psychiatr Rev*. 2013; 12(3):57–63.
6. Bailey BN, Delaney-Black V, Covington CY, Ager J, Janisse J, Hannigan JH, Sokol RJ. Prenatal exposure to binge drinking and cognitive and behavioral outcomes at age 7 years. *American Journal of Obstetrics and Gynecology*. 2004 191-1037-1043.
7. Alvik A, Aalen OO, Lindemann R. Early Fetal Binge Alcohol Exposure Predicts High Behavioural Symptom Scores in 5.5-Year-Old-Children. *Alcoholism: Clinical and Experimental Research*. 2013; 37(11):1954–1962.
8. May PA, Blankenship J, Marais A, Gossage JP, Kalberg WO, Joubert B, Cloete M, Barnard R, De Vries M, ... Seedat S. Maternal alcohol consumption producing fetal alcohol spectrum disorders (FASD): Quantity, frequency and timing of drinking. *Drug and alcohol dependence*. 2013; 133:502–512. [PubMed: 23932841]
9. Clarren S, Halliwell CI, Werk CM, Seabaldt RJ, Petrie A, Lilley C, Cook J. Using a Common Form for Consistent Collection and Reporting of FASD Data from Across Canada: A Feasibility Study. *J Popul Ther Clin Pharmacol*. 2015; 22(3):e211–228. [PubMed: 26567605]
10. Popova S, Lange S, Burd L, Chudley AE, Clarren SK, Rehm J. Cost of Fetal Alcohol Spectrum Disorder Diagnosis in Canada. *PLOS one*. 2013; 8(4)
11. May PA, Baete A, Russo J, Elliott AJ, Blankenship J, Kalberg WO, Buckley D, Brooks M, Hasken J, Abdul-Rahman O, Adam MP, Robinson LK, Manning M, Hoyme HE. Prevalence and characteristics of fetal alcohol spectrum disorders. *Pediatrics*. 2014; 134:855–66. [PubMed: 25349310]
12. Popova S, Lange S, Probst C, Gmel G, Rehm J. Estimation of national, regional and global prevalence of alcohol use during pregnancy and fetal alcohol syndrome: a systematic review and meta-analysis. *The Lancet Global Health*. 2017; 5(3):e290–299. [PubMed: 28089487]

13. Dudek J, Skocic J, Sheard E, Rovet J. Hippocampal abnormalities in Youth with Alcohol-Related Neurodevelopmental Disorder. *Journal of the International Neuropsychological Society*. 2014; 20:181–191. [PubMed: 24512673]
14. Rajaprakash M, Chakravarty MM, Lerch JP, Rovet J. Cortical morphology in children with alcohol-related neurodevelopmental disorder. *Brain and Behavior*. 2014; 4(1):41–50. [PubMed: 24653953]
15. Sowell ER, Mattson SN, Kan G, Thompson PM, Riley EP, Toga AW. Abnormal cortical thickness and brain-behavior correlation patterns in individuals with heavy prenatal alcohol exposure. *Cerebral Cortex*. 2008; 18:136–44. [PubMed: 17443018]
16. Koditwakku PW. Defining the Behavioral Phenotype in Children with Fetal Alcohol Syndrome Spectrum Disorders: A Review. *Neuroscience and Biobehavioral Review*. 2007; 31:192–201.
17. Nash K, Rovet J, Greenbaum R, Fantus E, Nulman I, Koren G. Identifying the behavioral phenotype in fetal alcohol spectrum disorder: sensitivity, specificity and screening potential. *Arch Women Ment Hlth*. 2006; 9:181–186.
18. Gray R, Mukherjee RAS, Rutter M. Alcohol consumption during pregnancy and its effects on neurodevelopment: what is known and what remains uncertain. *Addiction*. 2009; 104:1270–1273. [PubMed: 19215606]
19. Kilburn TR, Eriksen H-LF, Underbjerg M, Thorsen P, Mortensen EL, Landrø NI, et al. Low to Moderate Average Alcohol Consumption and Binge Drinking in Early Pregnancy: Effects on Choice Reaction Time and Information Processing Time in Five-Year-Old Children. *PLoS ONE*. 2015; 10(9):e0138611.doi: 10.1371/journal.pone.0138611 [PubMed: 26382068]
20. Kelly SJ, Day N, Streissguth AP. Effects of prenatal alcohol exposure on social behavior in humans and other species. *Neurotoxicology and Teratology*. 2000; 22:143–149. [PubMed: 10758343]
21. Martin DC, Martin JC, Streissguth AP. Sucking frequency and amplitude in newborns as a function of maternal drinking and smoking. *Currents in Alcoholism*. 1979; 5:359–366. [PubMed: 755637]
22. Coles CD, Platzman KA. Behavioral development in children prenatally exposed to drugs and alcohol. *Intl J Addict*. 1993; 28(13):1393–433.
23. O'Connor MJ, Sigman M, Kasari C. Attachment behavior of infants exposed prenatally to alcohol: Mediating effects of infant affect and mother-infant interaction. *Dev Psychopathol*. 1992; 4:243–256.
24. Streissguth AP, Barr HM, Kogan J, Bookstein FL. Final report to the Centres for Disease Control and Prevention. Seattle: University of Washington School of Medicine; 1996. Understanding the occurrence of secondary disabilities in clients with fetal alcohol syndrome (FAS) and fetal alcohol effects (FAE).
25. Famy C, Streissguth AP, Unis AS. Mental illness in adults with fetal alcohol syndrome or fetal alcohol effects. *Am J Psychiatry*. 1998; 155(4):552–554. [PubMed: 9546004]
26. O'Connor MJ, Shah B, Whaley S, Cronin P, Gunderson B, Graham J. Psychiatric illness in a clinical sample of children with prenatal alcohol exposure. *Am J Drug Alcohol Abuse*. 2002; 28:743–754. [PubMed: 12492268]
27. Fryer SL, McGee CL, Matt GE, Riley EP, Mattson SN. Evaluation of psychopathological conditions in children with heavy prenatal alcohol exposure. *Pediatrics*. 2007; 119(3):e733–741. [PubMed: 17332190]
28. Rangmar J, Hjern A, Vinnerljung B, Stronland K, Aronson M, Fahlke C. Psychosocial Outcomes of Fetal Alcohol Syndrome in Adulthood. *Pediatrics*. 2015; 135(1):e52–e58. [PubMed: 25535260]
29. Thomas SE, Kelly SJ, Mattson SN, Riley EP. Comparison of Social Abilities of Children with Fetal Alcohol Syndrome to Those of Children with Similar IQ Scores and Normal Controls. *Alcohol Clin Exp Res*. 1998; 22(2):528–533. [PubMed: 9581664]
30. Stevens S, Nash K, Koren G, Rovet J. Social Perspective Taking and Empathy in Children with Fetal Alcohol Spectrum Disorders. *J Int Neuropsych Soc*. 2015; 21:74–84.
31. Stevens SA, Major D, Rovet R, Koren G, Fantus E, Nulman I, Desrocher M. Social problem solving in children with fetal alcohol spectrum disorder. *J Popul Ther Clin Pharmacol*. 2012; 19:e99–110. [PubMed: 22535836]

32. Greenbaum RL, Stevens SA, Nash K, Koren G, Rovet J. Social cognitive and emotion processing abilities of children with fetal alcohol spectrum disorders: A comparison with Attention Deficit Hyperactivity Disorder. *Alcohol Clin Exp Res*. 2009; 33:1656–1670. [PubMed: 19624575]
33. Schonfeld AM, Paley B, Frankel F, O'Connor MJ. Executive functioning predicts social skills following prenatal alcohol exposure. *Child Neuropsychol*. 2006; 12(6):439–52. [PubMed: 16952889]
34. Nash K, Sheard E, Rovet J, Koren G. Understanding Fetal Alcohol Spectrum Disorders (FASDs): Toward identification of a behavioural phenotype. *Scientific World Journal*. 2008; 8:873–882. [PubMed: 18836653]
35. Stevens S, Nash K, Koren G, Rovet J. Autism characteristics in children with fetal alcohol spectrum disorders. *Child Neuropsychology*. 2013; 19(6):579–587. [PubMed: 23030694]
36. Rangmar J, Hjern A, Vinnerljung B, Stronland K, Aronson M, Fahlke C. Psychosocial Outcomes of Fetal Alcohol Syndrome in Adulthood. *Pediatrics*. 2015; 135(1):e52–e58. [PubMed: 25535260]
37. Nash K, Stevens S, Greenbaum R, Weiner J, Koren G, Rovet J. Improving executive functioning in children with fetal alcohol spectrum disorders. *Child Neuropsychology*. 2015; 21(2):191–209. [PubMed: 25010354]
38. Kaskutas LA, Graves K. Pre-pregnancy drinking: how drink size affects risk assessment. *Addiction*. 2001; 96(8):1199–209. [PubMed: 11487425]
39. Dukes K, Tripp T, Willinger M, Odendaal H, Elliott AJ, Kinney HC, ... Robinson F, et al. Drinking and smoking patterns during pregnancy: Development of group-based trajectories in the Safe Passage Study. *Alcohol*. 2017; 62:49–60. [PubMed: 28755751]
40. Clarren SK, Bowden DM. Fetal alcohol syndrome: a new primate model for binge-drinking and its relevance to human ethanol teratogenesis. *J Pediatr*. 1982; 101(5):819–24.
41. Lipinski RJ, Hammond P, O'Leary-Moore SK, Ament JJ, Pecevich SJ, et al. Ethanol-Induced Face-Brain Dymorphology Patterns Are Correlative and Exposure-Stage Dependent. *PLoS ONE*. 2012; 7(8):e43067.doi: 10.1371/journal.pone.0043067 [PubMed: 22937012]
42. Gunderson VM, Grant-Webster KS, Sackett GP. Deficits in visual recognition in low birth weight infant pigtailed monkeys (*Macaca nemestrina*). *Child Dev*. 1989; 100(1):119–127.
43. Schneider ML, Moore CF, Gajewski LL, Larson JA, Roberts AD, Converse AK, et al. Sensory processing disorders in a primate model: evidence from a longitudinal study of prenatal alcohol and prenatal stress effects. *Child Development*. 2008; 79(1):100–113. [PubMed: 18269511]
44. Varlinskaya EI, Mooney SM. Acute exposure to ethanol on gestational day 15 affects social motivation of female offspring. *Behav Brain Res*. 2004; 251:106–109.
45. Kelly SJ, Tran TD. Alcohol exposure during pregnancy alters social recognition and social communication in rats. *Neurotoxicology and Teratology*. 1997; 19(5):143–149.
46. Meyer LS, Riley EP. Social play in juvenile rats prenatally exposed to alcohol. *Teratology*. 1986; 34(1):1–7. [PubMed: 3764769]
47. Hamilton DA, Barto D, Rodriguez CI, Magcalas CM, Fink BC, Rice JP, Bird CW, Davie S, Savage DD. Effects of moderate prenatal ethanol exposure and age on social behavior, spatial response perseveration errors and motor behavior. *Behav Brain Res*. 2014; 269:44–54. [PubMed: 24769174]
48. Royalty J. Effects of prenatal ethanol exposure on juvenile play-fighting and post-pubertal aggression in rats. *Psychological Review*. 1990; 66(2):551–560.
49. Brown RT, Coles CD, Smith IE, Platzman KA, Silvestein J, Erickson S, Falek A. Effects of prenatal alcohol exposure at school age. II. Attention and behavior. *Neurotoxicol Teratol*. 1991; 13(4):369–376.
50. Fernandes Y, Gerlai R. Long-term behavioral changes in response to early developmental exposure to ethanol in Zebrafish. *Alcohol Clin Exp Res*. 2009; 33:601–609. [PubMed: 19183139]
51. Carvan MJ 3rd, Loucks E, Weber DN, Williams FE. Ethanol effects on the developing zebrafish: neurobehavior and skeletal morphogenesis. *Neurotoxicol Teratol*. 2004; 26(6):757–68.
52. Parker MO, Annan LV, Kanellopoulos AH, Brock AJ, Combe FJ, Baiamonte M, Teh MT, Brennan CH. The utility of zebrafish to study the mechanisms by which ethanol affects social behavior and anxiety during early brain development. *Progress in Neuro-Psychopharmacology & Biological Psychiatr*. 2014; 55:94–100.

53. Howe K, Clark MD, Torroja CF, Torrance J, Berthelot C, Muffato M, Collins J, et al. The zebrafish reference genome sequence and its relationship to the human genome. *Nature*. 2013; 496(7446): 498–503. [PubMed: 23594743]
54. Herculano AM, Maximino C. Serotonergic modulation of zebrafish behavior: towards a paradox. *Prog Neuropsychopharmacol Biol Psychiatry*. 2014; 55:50–66. [PubMed: 24681196]
55. Bilotta J, Bennett JA, Hancock L, Saszik S. Ethanol Exposure alter zebrafish development: A novel model of fetal alcohol syndrome. *Neurotoxicol Teratol*. 2004; 26:737–743. [PubMed: 15451038]
56. Arenzana FJ, Carvan MJ III, Aijon J, Sanchez-Gonzalez R, Arevalo R, Porteros A. Teratogenic effects of ethanol exposure on zebrafish visual system development. *Neurotoxicology and Teratology*. 2006; 28:342–348. [PubMed: 16574376]
57. Kashyap B, Frederickson LC, Stenkamp DL. Mechanisms for persistent microphthalmia following ethanol exposure during retinal neurogenesis in zebrafish embryos. *Visual Neuroscience*. 2007; 24(3):409–421. [PubMed: 17640445]
58. Matsui JI, Egana AL, Sponholtz TR, Adolph AR, Dowling JE. Effects of ethanol on photoreceptors and visual function in developing zebrafish. *Retinal Cell Biology*. 2006; 47(10):4589–4597.
59. Sylvain NJ1, Brewster DL, Ali DW. Zebrafish embryos exposed to alcohol undergo abnormal development of motor neurons and muscle fibers. *Neurotoxicol Teratol*. 2010 Jul-Aug;32(4):472–80. [PubMed: 20211721]
60. Zamora LY, Lu Z. Alcohol-induced morphological deficits in the development of octavolateral organs of the zebrafish (*Danio rerio*). *Zebrafish*. 2013 Mar; 10(1):52–61. [PubMed: 23461415]
61. Uribe PP, Asuncion JD, Matsui JI. Ethanol effects the development of sensory hair cells in larval zebrafish (*Danio rerio*). *PLOS one*. 2013; 8(12):1–9.
62. Shan SD, Boutin S, Ferdous J, Ali DW. Ethanol exposure during gastrulation alters neuronal morphology and behavior in zebrafish. *Neurotoxicol Teratol*. 2015; 48:18–27. [PubMed: 25599605]
63. Joya X, Garcia-Algar O, Vall O, Pujades C. Transient exposure to ethanol during zebrafish embryogenesis results in defects in neuronal differentiation: an alternative model system to study FASD. *PLoS One*. 2014 Nov 10.9(11):e112851. [PubMed: 25383948]
64. Lockwood B1, Bjerke S, Kobayashi K, Guo S. Acute effects of alcohol on larval zebrafish: a genetic system for large-scale screening. *Pharmacol Biochem Behav*. 2004; 77(3):647–54. [PubMed: 15006478]
65. Tal TL1, Franzosa JA, Tilton SC, Philbrick KA, Iwaniec UT, Turner RT, Waters KM, Tanguay RL. MicroRNAs control neurobehavioral development and function in zebrafish. *FASEB J*. 2012; 26(4):1452–61. [PubMed: 22253472]
66. Marris JA, Clendenon SG, Ratcliffe DR, Fielding SM, Liu Q, Bosron WF. Zebrafish fetal alcohol syndrome model: effects of ethanol are rescued by retinoic acid supplement. *Alcohol*. 2010; 44(7–8):707–715. [PubMed: 20036484]
67. Bailey JM, Oliveri AN, Zhang C, Frazier JM, Mackinnon S, Cole GJ, Levin ED. Long-Term Behavioral Impairment Following Acute Embryonic Ethanol Exposure in Zebrafish. *Neurotoxicol Teratol*. 2015; 48:1–8. [PubMed: 25599606]
68. Buske C, Gerlai R. Early embryonic ethanol exposure impairs shoaling and the dopaminergic and serotonergic systems in adult zebrafish. *Neurotoxicol Teratol*. 2011; 33:698–707. [PubMed: 21658445]
69. Baiamonte M, Brennan CH, Vinson GP. Sustained action of developmental ethanol exposure on the cortisol response to stress in zebrafish larvae and adults. *PLoS One*. 2015; 10(4):e0124488. eCollection 2015, Erratum in: *PLoS One*, 2015, 10(5), e0128050. doi: 10.1371/journal.pone.0124488 [PubMed: 25875496]
70. Fernandes Y, Rampersad M, Gerlai R. Embryonic alcohol exposure impairs the dopaminergic system and social behavioural responses in adult zebrafish. *Int J Neuropsychopharmacol*. 2015; 18:1–8.
71. Mikheev VN. Combined effects of predators and parasites on shoaling behavior of fishes. *Journal of Ichthyology*. 2009; 49(11):1032–1041.
72. Al-Imari L, Gerlai R. Sight of Conspecifics as reward in associative learning in zebrafish (*Danio rerio*). *Behavioral Brain Research*. 2008; 189:216–219.



73. Ahmed O, Seguin D, Gerlai R. An automated predator avoidance task in zebrafish. *Behav Brain Res.* 2011; 216:166–171. [PubMed: 20674614]
74. Ahmed T, Fernandes Y, Gerlai R. Effects of animated images of sympatric predators and abstract shapes on fear responses in zebrafish. *Behaviour.* 2012; 149:1125–1153.
75. Seguin D, Shams S, Gerlai R. Behavioral Responses to Novelty or to a Predator Stimulus Are Not Altered in Adult Zebrafish by Early Embryonic Alcohol Exposure. *Alcohol Clin Exp.* 2016; 40(12):2667–2675.
76. Fernandes Y, Tran S, Abraham E, Gerlai R. Embryonic alcohol exposure impairs associative learning performance in adult zebrafish. *Behav Brain Res.* 2014; 265:181–187. [PubMed: 24594368]
77. Saverino C, Gerlai R. The social zebrafish: behavioral responses to conspecific, heterospecific, and computer animated fish. *Behav Brain Res.* 2008; 191:77–87. [PubMed: 18423643]
78. Seguin D, Gerlai R. Zebrafish prefer larger to smaller shoals: analysis of quantity estimation in a genetically tractable model organism. *Anim Cognit.* 2017; 20(5):813–821. [PubMed: 28616841]
79. Halberda J, Mazocco MMM, Feigenson L. Individual differences in non-verbal number acuity correlate with maths achievement. *Nature.* 2008; 455(7213):655–U62.
80. Gilmore CK, McCarthy SE, Spelke ES. Symbolic arithmetic knowledge without instruction. *Nature.* 2007; 447(7144):589. [PubMed: 17538620]
81. Kopera-Frye K, Dehaene S, Streissguth AP. Impairments in number processing induced by prenatal alcohol exposure. *Neuropsychologia.* 1996; 34(12):1187–1196. [PubMed: 8951830]
82. Olson HC, Feldman JJ, Streissguth AP, Sampson PD, Bookstein FL. Neuropsychological deficits in adolescents with fetal alcohol syndrome: Clinical findings. *Alcohol Clin Exp Res.* 1998; 22(9):1998–2012. [PubMed: 9884144]
83. Crocker N, Riley EP, Mattson SN. Visual-spatial Abilities Relate to Mathematics Achievement in Children with Heavy Prenatal Alcohol Exposure. *Neuropsychology.* 2015; 29(1):108–116. [PubMed: 25000323]
84. Streissguth AP, Barr HM, Olson HC, Sampson PD, Bookstein FL, Burgess DM. Drinking during pregnancy decreases word attack and arithmetic scores on standardized tests- adolescent data from a population-based prospective-study. *Alcohol Clin Exp Res.* 1994; 18(2):248–254. [PubMed: 8048722]
85. Ruhl N, McRobert SP, Currie WJS. Shoaling preferences and the effects of sex ratio on spawning and aggression in small laboratory populations of zebrafish. *Lab Animal.* 2009; 38(8):264–269. [PubMed: 19626019]
86. Dahlgren IL, Matuszcyk JV, Hård E. Sexual orientation in male rats prenatally exposed to ethanol. *Neurotoxicol Teratol.* 1991; 13(3):267–269. [PubMed: 1886535]
87. Hård E, Dahlgren IL, Engel J, Larsson K, Liljequist S, Lindh AS, Musi B. Development of sexual behaviors in prenatally ethanol-exposed rats. *Drug Alcohol Depend.* 1984; 14(1):51–61. [PubMed: 6489152]
88. Ward IL, Ward OB, Winn RJ, Bielawski D. Male and female sexual behavior potentials of male rats prenatally exposed to the influence of alcohol, stress, or both. *Behav Neurosci.* 1994; 108:1188–1195. [PubMed: 7893411]
89. Lugo JN Jr, Marino MD, Cronise K, Kelly SJ. Effects of alcohol exposure during development on social behavior in rats. *Physiology & Behavior.* 2003; 78:185–194.
90. Kršiak M, Elis J, Pöschlová N, Mašek K. Increased aggressiveness and lower brain serotonin levels in offspring of mice given alcohol during gestation. *J Stud Alcohol.* 1977; 38:1696–1704. [PubMed: 562457]
91. Jacobson SW, Bihun JT, Chiodo LM. Effects of prenatal alcohol and cocaine exposure on infant cortisol level. *Dev Psychopathol.* 1999; 11:195–208. [PubMed: 16506530]
92. Oliveira RF, Silva JF, Simoes JM. Fighting Zebrafish: Characterization of Aggressive Behavior and Winner-Loser Effects. *Zebrafish.* 2011; 8(2):73–81. [PubMed: 21612540]
93. Teles MC, Dahlbom SJ, Winberg S, Oliveira RF. Social modulation of brain monoamine levels in zebrafish. *Behavioral Brain Research.* 2013; 253:17–24.

94. Gerlai R, Lahav M, Guo S, Rosenthal A. Drinks like a fish: Zebra fish (*Danio rerio*) as a behavior genetic model to study alcohol effects. *Pharmacol Biochem Behav.* 2000; 67:773–782. [PubMed: 11166068]
95. Spadani AD, Bazinet AD, Frye SL, Tapert SF, Mattson SN, Riley EP. BOLD Response During Spatial Working Memory in Youth with Heavy Prenatal Alcohol Exposure. *Alcohol Clin Exp Res.* 2009; 33(12):2067–2076. [PubMed: 19740135]
96. Schneider ML, Moore CF, Becker EF. Timing of moderate alcohol exposure during pregnancy and neonatal outcome in rhesus monkeys (*Macaca mulatta*). *Alcohol Clin Exp Res.* 2001; 25(8):1238–1245. [PubMed: 11505056]
97. Wagner JL, Zhou FC, Goodlett CR. Effects of one- and three-day binge alcohol exposure in neonatal C57BL/6 mice on spatial learning and memory in adolescence and adulthood. *Alcohol.* 2014; 48(2):99–111. [PubMed: 24507877]
98. Pei J, Job J, Kully-Marten K, Rasmussen C. Executive function and memory in children with Fetal Alcohol Spectrum Disorder. *Child Neuropsychology.* 2011; 17(3):290–309. [PubMed: 21718218]
99. Wilford JA, Richardson GA, Leech SL, Day NL. Verbal and visuospatial learning and memory function in children with moderate prenatal alcohol exposure. *Clinical and Experimental Research.* 2004; 28(3):497–507.
100. Pan Y, Mo K, Razak Z, Westwood JT, Gerlai R. Chronic Alcohol Exposure Induced Gene Expression Changes in the Zebrafish Brain. *Behav Brain Res.* 2011; 216:66–76. [PubMed: 20654657]
101. Saif M, Chatterjee D, Buske C, Gerlai R. Sight of conspecific images induces changes in neurochemistry in zebrafish. *Behav Brain Res.* 2013; 243:294–299. [PubMed: 23357085]
102. Scerbina T, Chatterjee D, Gerlai R. Dopamine receptor antagonism disrupts social preference in zebrafish, a strain comparison study. *Amino Acids.* 2012; 43:2059–2072. [PubMed: 22491827]
103. Mahabir S, Chatterjee D, Gerlai R. Strain dependent neurochemical changes induced by embryonic alcohol exposure in zebrafish. *Neurotox Terat.* 2013; 41:1–7.
104. Parker MO, Evans AM, Brock AJ, Combe FJ, Teh MT, Brennan CH. Moderate alcohol exposure during early brain development increases stimulus-response habits in adulthood. *Addict Biol.* 2016; 21(1):49–60. [PubMed: 25138642]
105. Alati R, Al Mamun A, Williams GM, O’Callaghan M, Najman JM, Bor W. In utero alcohol exposure and prediction of alcohol disorders in early adulthood: a birth cohort study. *Archives of General Psychiatry.* 2006; 63:1009–1016. [PubMed: 16953003]
106. Baer JS, Sampson PD, Barr HM, Connor PD, Streissguth AP. A 21-year longitudinal analysis of the effects of prenatal alcohol exposure on young adult drinking. *Archives of General Psychiatry.* 2003; 60:377–385. [PubMed: 12695315]
107. Chotro MG, Arias C, Laviola G. Increased ethanol intake after prenatal ethanol exposure: studies with animals. *Neurosci Biobehav Res.* 2007; 31:181–191.
108. Blanchard BA, Steindorf S, Wang S, LeFevre R, Mankes RF, Glick SD. Prenatal ethanol exposure alters ethanol-induced dopamine release in nucleus accumbens and striatum in male and female rats. *Alcohol Clin Exp Res.* 1993; 17:974–981. [PubMed: 8279684]
109. Wang J, Haj-Dahmane S, Shen RY. Effects of prenatal ethanol exposure on the excitability of ventral tegmental area of dopamine neurons in vitro. *J Pharmacol Exp Ther.* 2006; 319:857–863. [PubMed: 16905687]
110. Sterling ME, Chang GQ, Karatayev O, Chang SY, Leibowitz SF. Effects of embryonic ethanol exposure at low doses on neuronal development, voluntary ethanol consumption and related behaviors in larval and adult zebrafish: Role of hypothalamic orexigenic peptides. *Behav Brain Res.* 2016; 304:125–38. [PubMed: 26778786]
111. Schweitzer J, Löhr H, Filippi A, Driever W. Dopaminergic and noradrenergic circuit development in zebrafish. *Devel Neurobio.* 2012; 72:256–268.
112. Wang G, Bierberich E. Prenatal alcohol exposure triggers ceramide-induced apoptosis in neural crest-derived tissues concurrent with defective cranial development. *Cell Death and Disease.* 2010; 1:e46.doi: 10.1038/cddis.2010.22 [PubMed: 21364652]

113. Creeley CE, Olney JW. Drug-Induced Apoptosis: Mechanism by which Alcohol and Many Other Drugs an Disrupt Brain Development. *Brain Sciences*. 2013; 3(3):1153–1181. [PubMed: 24587895]
114. Jacobson MD, Weil M, Raff MC. Programmed Cell Death in Animal Development. *Cell*. 1997; 88:347–354. [PubMed: 9039261]
115. Tanguay RL, Reimers MJ. Analysis of ethanol developmental toxicity in zebrafish. *Methods Mol Biol*. 2008; 447:63–74. [PubMed: 18369111]
116. Cole GJ, Zhang C, Ojiaku P, Bell V, Devkota S, Mukhopadhyay S. Effects of ethanol exposure on nervous system development in zebrafish. *Int Rev Cell Mol Biol*. 2012; 299:255–315. [PubMed: 22959306]
117. Yin G, Yao F, Chen X, Wang N, Wang H, Chang HE, Yuan Z, Wu B. Ethanol reduces neural precursor cells and inhibits neuronal and glial differentiation in zebrafish embryos. *Nan Fang Yi Ke Da Xue Xue Bao*. 2014; 34(11):1555–1561. [PubMed: 25413049]
118. Soares AR, Pereira PM, Ferreira V, Reverendo M, Simões J, Bezerra AR, Moura GR, Santos MA. Ethanol exposure induces upregulation of specific microRNAs in zebrafish embryos. *Toxicol Sci*. 2012; 127:18–28. [PubMed: 22298809]