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Effect of immediate and prolonged GLP-1 receptor agonist administration on uric acid and its kidney clearance: *post-hoc* analyses of four clinical trials

Lennart Tonneijck^{1,*}, Marcel H.A. Muskiet^{1,*}, Mark M. Smits¹, Petter Bjornstad², Mark H.H. Kramer¹, Michaela Diamant^{†,1}, Ewout J. Hoorn³, Jaap A. Joles⁴, and Daniël H. van Raalte¹

¹Diabetes Center, Department of Internal Medicine, VU University Medical Center, Amsterdam, The Netherlands ²Department of Pediatric Endocrinology, University of Colorado School of Medicine, Aurora, CO, United States; and Barbara Davis Center for Diabetes, University of Colorado Denver, Aurora, CO, United States ³Department of Internal Medicine, Division of Nephrology and Transplantation, Erasmus Medical Center, Rotterdam, The Netherlands ⁴Department of Nephrology and Hypertension, University Medical Center, Utrecht, The Netherlands

Abstract

Aims—To determine effects of glucagon-like peptide (GLP)-1 receptor agonists (RA) on uric acid (UA)-levels and kidney UA-clearance.

Material and methods—*Post-hoc* analyses of four controlled clinical trials, which assessed actions of GLP-1RA-administration on kidney physiology. Immediate effects of GLP-1RA exenatide-infusion versus placebo was determined in 9 healthy overweight males (Study-A) and in 52 overweight T2DM-patients (Study-B). Effects were also examined of 12-week long-acting GLP-1RA liraglutide versus placebo in 36 overweight T2DM-patients (Study-C) and of 8-week short-acting GLP-1RA lixisenatide versus once-daily titrated insulin-glulisine in 35 overweight T2DM-patients (Study-D). Plasma-UA, fractional (inulin-corrected) and absolute urinary-excretion of UA (UE_{UA}) and sodium (UE_{Na}), and urine-pH was determined.

Results—Median baseline plasma-UA levels was 5.39 to 6.33 mg/dL across all studies (17-22% of subjects were hyperuricemic). In study-A, exenatide-infusion slightly increased plasma-UA (+0.07 \pm 0.02mg/dL, P=0.04), and raised absolute-UE_{UA} (+1.58 \pm 0.65mg/min/1.73m², P=0.02), but

Correspondence to: Lennart Tonneijck, MD, Diabetes Center, Department of Internal Medicine, VU University Medical Center Amsterdam, De Boelelaan 1117, 1081 HV Amsterdam, The Netherlands, l.tonneijck@vumc.nl. [†]Deceased

^{*}Both authors contributed equally to this work

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Author contributions

L.T. designed Study-D, performed the measurements, analysed and interpreted the data and drafted and completed the final manuscript. M.H.A.M. designed Study-D, performed measurements, was involved in statistical analyses, interpretation of the data, and contributed to the drafting and critical revision of the manuscript. M.M.S. designed Studies A to C, performed measurements, was involved in statistical analyses, and contributed to the critical revision of the manuscript. M.D. designed all studies and was initially involved in the discussion and supervision. P.B., M.H.H.K, E.J.H., J.A.J. and D.H.v.R. contributed to the interpretation of the data, discussion of the intellectual content and critical editing of the manuscript.

did not affect fractional-UE_{UA} compared to placebo. Fractional-UE_{UA} and absolute-UE_{UA} correlated with increases in urine-pH (r:0.86, P=0.003 and r:0.92, P<0.001, respectively). Fractional-UE_{UA} correlated with increased fractional-UE_{Na} (r:0.76, P=0.02). In study-B, exenatide-infusion did not affect plasma-UA, but increased fractional-UE_{UA} (+0.76±0.38%, P=0.049) and absolute-UE_{UA} (+0.75±0.27mg/min/1.73m², P=0.007), compared to placebo. In regression analyses, both parameters were explained by changes in urine-pH, and in part, by changes in UE_{Na}. In study-C, liraglutide-treatment did not affect plasma-UA, UE_{UA}, UE_{Na}, or urine-pH compared to placebo. In study-D, lixisenatide-treatment increased UE_{Na} and urine-pH from baseline, but did not affect plasma-UA or UE_{UA}.

Conclusion—Immediate exenatide-infusion increases UE_{UA} in overweight healthy males and T2DM-patients, likely via inhibiting Na⁺/H⁺-exchanger type-3 in the renal proximal tubule. Prolonged treatment with a long-acting or short-acting GLP-1RA does not affect plasma-UA or UE_{UA} in T2DM patients with normal plasma UA-levels and relatively low cardiovascular risk. Our results suggest that the cardio-renal benefits of GLP-1RA are not mediated through changes in UA.

Introduction

In humans, uric acid (UA) is the final breakdown product of purine nucleotides, which originate from exogenous dietary sources, and endogenous de novo synthesis and tissue catabolism.^{1, 2} UA-formation from purines occurs mainly in the liver, with a prominent role for xanthine oxidase, the rate-limiting enzyme in UA-biosynthesis.^{3, 4} Normally, >70% of UA is excreted into the urine (with a fractional excretion of 5 to 10%), while the remainder is excreted through the intestines and perspiration.^{3, 5, 6} Hyperuricemia is usually defined as a plasma UA-level $>6.8 \text{ mg/dL} (405 \mu \text{mol/L}; \text{ corresponding with its solubility-point})^6$ and can be caused by reduced excretion by the kidney (90% of hyperuricemic cases) or UAoverproduction in the liver.³ Hyperuricemia can result in deposition of UA-crystals, leading to gout and nephrolithiasis. Moreover, evidence suggests that increased plasma-UA-levels even in the high-normal range-4, ⁷ are an independent risk-factor for impaired fasting glucose, diabetes mellitus, hypertension, heart failure, coronary heart disease and chronic kidney disease.⁸ Accordingly, the indication for plasma UA lowering therapy –either with xanthine-oxidase inhibitors or uricosuric agents- may be broader than gout or UA nephrolithiasis. Interestingly, in studies in diverse populations, including type 2 diabetes mellitus (T2DM) patients,^{4, 9–11} plasma-UA lowering (therapy) improved (surrogate) cardiorenal end points, although dedicated large-sized trials are needed to firmly establish the causation between UA lowering and long-term clinical outcomes.44

Over the last three decades, multiple antihyperglycaemic drug-classes with different modes of action have been successfully licensed for the treatment of hyperglycaemia in T2DM. The results of recent long-term cardiovascular safety trials in thousands of at-risk T2DM patients indicate benefit of two of these drug-classes on cardiovascular and renal events and mortality: glucagon-like peptide-1 (GLP-1) receptor agonists (RA) (i.e. liraglutide^{12, 13} and semaglutide,¹⁴ while exenatide once-weekly tended [p=0.06] to improve cardiovascular risk¹⁵) and sodium-glucose co-transporter-2 (SGLT2) inhibitors (i.e. empagliflozin^{16, 17} and canagliflozin¹⁸). Delineation of the underlying cardio-renal protective mechanisms may

have immediate relevance for the prevention and treatment of diabetes-related complications, but many uncertainties in this regard remain. For SGLT2 inhibitors, some have suggested that benefits are in-part explained by their ability to reduce serum UA-levels, likely through increased urinary UA-excretion.¹⁹ The effect of GLP-1RAs on plasma UA-levels and urinary-excretion of UA (UE_{UA}) in humans are hitherto unknown. In rats with streptozotocin induced (type 1)-diabetes, daily GLP-1-peptide injections for 10 weeks reduced serum UA-concentrations by 58% compared to untreated diabetic rats.²⁰

As GLP-1RA administration inhibits Na⁺/H⁺-exchanger type-3 (NHE3)-activity in the proximal tubule, causing natriuresis and alkalization of the urine, $^{21-24}$ we hypothesise that this antihyperglycaemic drug-class may reduce plasma-UA levels by increasing urinary UA-excretion in humans with and without T2DM.^{25–27} To explore the effects of currently available GLP-1RAs –with clinically relevant differences in their pharmacokinetic profile (see below)– on plasma-UA and UE_{UA}, we performed a *post-hoc* analysis of four clinical trials, which assessed the effects of these drugs on kidney physiology.^{21, 22, 24, 28} First, we assessed the *immediate* effects by means of administering intravenous exenatide-administration in healthy overweight males and T2DM patients. Second, as immediate effects on tubular UA-handling could be offset during chronic intervention, we also examined the effects of *prolonged* GLP-1RA therapy in overweight T2DM patients. The pharmacokinetic profile of a GLP-1RA is known to have different effects on gastric emptying rate, urinary sodium excretion and urinary alkalization during prolonged administration.^{22, 28–30}

Material and methods

Trial designs

Post-hoc analyses of four phase-4 clinical intervention trials, all conducted at the Diabetes Center of the VU University Medical Center (VUMC) in Amsterdam (The Netherlands) between May 2013 and April 2017. All trials were primarily designed to assess the effects of immediate or prolonged GLP-1RA-administration on kidney physiology in overweight healthy males and T2DM patients.

A detailed overview of trial-designs and testing-procedures is shown in Figure 1. Study-A was an acute, non-blinded, placebo-controlled trial, examining the effect of placebo (isotonic saline) and subsequent exenatide-infusion at therapeutic concentrations in healthy overweight males.²¹ Study-B was an acute, randomised, double-blind, placebo-controlled, parallel-group trial, examining the effect of exenatide-infusion in T2DM patients.²⁴ Patients from Study-B were subsequently enrolled in Study-C, which was a 12-week, randomised, double-blind, placebo-controlled trial, examining the effect of subcutaneous injections with liraglutide in T2DM patients.²⁸ Study-D was an 8-week, randomised, open-label, comparator-controlled trial, examining the effect of subcutaneous injections with lixisenatide versus once-daily titrated insulin glulisine in T2DM patients.²³

All trials were registered at ClinicalTrials.gov (ID's: NCT01744236 [Studies A-C] and NCT02276196 [Study-D]. Trials were approved by the local ethic review board, and conducted in accordance with the Declaration of Helsinki and International Conference on

Harmonization on Good Clinical Practice. All subjects provided written informed consent before any trial-related activities.

Detailed descriptions of participants, interventions and randomisation procedures has been reported.^{21, 23, 24, 28, 31} A brief description is provided the Supplemental methods.

Study protocols

Two days prior to testing procedures, participants were instructed to adhere to a controlled intake of protein (1.5–2.0 g/kg/day) and sodium (~150-200 mmol/day). In addition, prior to the experiments, participants were instructed to refrain from vigorous physical activity and alcohol consumption for 24 hours, and avoid caffeine-containing products or nicotine for 12 hours. After an overnight fast, and before arriving at the CRU of the Diabetes Center VUMC at 07:30 AM, patients consumed 500 mL of tap water to stimulate diuresis. T2DM patients delayed their morning medications, with the exception of metformin and thyroid hormone replacement therapy. Participants assumed a semi-recumbent position in a temperature controlled-room $(23.0 \pm 1.0 \,^{\circ}\text{C})$ throughout testing procedures. Intravenous catheters were inserted in an antecubital vein of both forearms to allow intermittent blood sampling (non-dominant side) and infusion of the kidney tracer substances (dominant side). Blood and urine was obtained for outcome variables as described in detail.^{21, 23, 24, 28} Subsequently, the kidney tests commenced as previously described in detail.^{21, 23, 24, 28} In all studies, a bolus of inulin was administered over 10 min, which was followed by a continuous infusion of 90 min, which served as an equilibration period.

In Study-A, subjects subsequently underwent two consecutive urine collection periods of 45 min during placebo-infusion, which were repeated after 60 min infusion with exenatide (Figure 1).²¹ In Study-B, participants subsequently underwent two urine collection periods of 45 min at baseline, which were repeated after 60 min infusion with exenatide or placebo (Figure 1).²⁴ In Study-C, the baseline measurements of Study-B were repeated after 12 weeks treatment with liraglutide or placebo (Figure 1).²⁸ In Study-D, participants consumed a standardized mixed breakfast consisting of bread with cheese and jam, and milk (416 kcal; 22 g protein, 48 g carbohydrates and 14 g fat) 35 minutes after the start of the inulin infusion (Figure 1).²³ After the equilibration period, patients underwent two urine collection periods of 45 minutes at baseline, which were repeated at week-8. After 8 weeks lixisenatide or insulin glulisine was administered 30 and 10 min before the standardized breakfast, respectively.

In all studies, diuresis was prompted by oral intake of 10 mL/kg (maximum 1,000 mL) tap water during the inulin equilibration period, followed by an intake of 200 mL/h for the remainder of the testing day. Before and after each urine collection period, blood samples were taken to obtain the outcome variables as described in detail.^{21, 23, 24, 28} UA-excretion was measured over the second of two consecutive urine collection periods in Studies-A to C, and over both of the two urine collection periods in Study-D.

Biochemical analyses

UA was measured as urate using an enzymatic colorimetric test (Cobas-C501, Roche Diagnostics, Indianapolis, IN) and urine-pH was buffered to >8.0 with NaOH. Inulin was

analysed as described previously.²⁴ Routine blood determinations (HbA_{1c}, glucose, sodium, osmolality, and creatinine) were performed at baseline and after prolonged therapy, using conventional assay methods by the Department of Clinical Chemistry in the VUMC. Venous blood glucose was measured throughout the kidney tests using an YSI-2300 STAT glucose analyzer (YSI Life Sciences, Yellow Springs, OH). Insulin was determined using an immunometric assay (ADVIA Centaur-XP Immunoassay System; Siemens Healthcare, Erlangen, Germany). Cross-reactivity of the assay with insulin glulisine (low) and insulin glargine (high) has been described.²² Urine-pH was measured by hand-held VARIO® 2V00 pH-meter and SenTix-V electrode (Wissenschaftlich-Technische Werkstätten GmbH, Weilheim, Germany).

Outcomes

In all studies, plasma-UA, fractional and absolute UE of UA (UE_{UA}) and sodium (UE_{Na}), GFR, urine-pH, urine-osmolality, insulin (serum or plasma) and glucose (blood or plasma) were measured. Urinary-glucose was determined in Study-B to D. In Study-C, fractional-UE_{UA} was additionally assessed at weeks 0, 2 and 12.

Calculations

Fractional UE_{UA} and UE_{Na} were calculated by using inulin as reference substance, unless stated otherwise. Absolute electrolyte excretions were calculated by multiplying urinary-electrolyte concentrations by urine flow. GFR was calculated from inulin clearances. GFR and absolute electrolyte excretion were corrected for body surface area using the Mosteller formula.^{21, 23, 24, 28}

Statistical methods

No *a priori* sample-size power calculation was performed for these *post-hoc* exploratory analyses. Statistical analyses were performed in the per protocol population of the separate trials as described,^{21, 23, 24, 28} using SPSS Statistics for Windows, V22.0 (IBM Corp, Armonk, NY). Log10-transformation of non-Gaussian distributed data was carried out to achieve normality prior to analyses. Paired *t*-test or Wilcoxon signed-rank test (as appropriate) were used to identify single effects of exenatide versus placebo in Study-A and within-group comparisons in Study-D. Multivariable linear regression models adjusting for corresponding baseline values only were used to identify effects of exenatide versus placebo (Study-B), liraglutide versus placebo Study-C) or lixisenatide versus insulin glulisine (Study-D). The Mann-Whitney test was used to assess between-group differences in urinaryglucose in Study-D. Pearson's correlation or Spearman signed rank test (as appropriate) was used to assess associations. A two-sided P < 0.05 was considered statically significant for all comparisons. Data are presented as least square mean ± SEM or, in case of a non-Gaussian distribution, as median [interquartile range], unless stated otherwise.

Results

Study-A

The study population consisted of 9 subjects (Table 1).²¹ Plasma-UA was 6.33 [6.05-7.05] mg/dL (2 subjects hyperuricemic) during placebo, and increased upon exenatide-infusion to

6.48 [6.09-7.08] mg/dL (+0.07±0.02 mg/dL, P=0.04). Exenatide did not significantly affect fractional-UE_{UA} (+0.82±0.47%, P=0.2), but increased absolute-UE_{UA} compared to placebo-infusion (+1.58±0.65 mg/min/1.73m², P=0.02; Figure 2). As previously reported, exenatide increased GFR, fractional-UE_{Na}, absolute-UE_{Na}, urine-pH (from 6.53 [6.03-6.90] to 7.04 [6.89-7.34], P=0.005) and osmolality (Supplemental Table 1).²¹ Exenatide reduced plasma glucose, without affecting plasma insulin concentrations (Supplemental Table 1).²¹

Changes in fractional-UE_{UA} and absolute-UE_{UA} correlated with changes in urine-pH (r: 0.90, P=0.001 and r: 0.82, P=0.007, respectively) (Figure 3A–B), but not with changes in plasma-UA (r: -0.20, P=0.6 and r: -0.35, P=0.4), glucose (r: 0.58, P=0.1 and r: 0.27, P=0.5) or insulin (r: 0.17, P=0.7 and r: 0.26, P=0.5). Changes in fractional-UE_{UA} correlated with changes in fractional-UE_{Na} (r: 0.75, P=0.019) (Figure 3C). Changes in fractional-UE_{UA} absolute-UE_{UA} and plasma-UA did not correlate with baseline plasma-UA (all P>0.3).

Study-B

The study population consisted of 52 patients (Table 1).²⁴ Mean± SD baseline UA was $5.50\pm1.04 \text{ mg/dL}$ (9 patients hyperuricemic) and did not change during immediate exenatide-infusion compared to placebo (-0.01±0.02 mg/dL, P=0.8). While UA levels were not affected, exenatide-infusion increased fractional-UE_{UA} (+0.76±0.38%, P=0.049) and absolute-UE_{UA} (+0.75±0.27 mg/min/1.73m², P=0.007), compared to placebo (Figure 2). As previously reported, exenatide increased fractional-UE_{Na}, absolute-UE_{Na}, urine-pH (+0.72±0.11, P<0.001) and urinary-osmolality (Supplemental Table 2).²⁴ Urinary-glucose at baseline was <0.11 mmol/L in all but one patient (0.97 mmol/L) and was not affected by exenatide.²⁴ Exenatide-infusion reduced blood glucose during the kidney tests, and increased serum insulin concentrations, compared to placebo (Supplemental Table 2). There were no changes in plasma renin concentrations (PRC).²⁴

In regression analyses, exenatide-induced alterations in fractional-UE_{UA} and absolute-UE_{UA} were largely explained by the change in urine-pH (regression-coefficient reduced to $0.01\pm0.45\%$ [P=0.9] and to -0.03 ± 0.28 mg/min/ $1.73m^2$ [P=0.9], respectively), and in-part by respective changes in fractional-UE_{Na} (to $0.58\pm0.37\%$, P=0.1) and absolute-UE_{Na} (to 0.50 ± 0.25 mg/min/ $1.73m^2$, P=0.049). Correcting for the change in blood glucose partially lowered the effect of exenatide on UE_{UA} (to $0.67\pm0.64\%$ [P=0.3] and to 0.56 ± 0.48 mg/min/ $1.73m^2$ [P=0.3]). When correcting for exenatide-induced change in insulin, the regression-coefficient of fractional-UE_{UA} and absolute-UE_{UA} increased to $1.11\pm0.45\%$ (P=0.02) and 0.95 ± 0.31 mg/min/ $1.73m^2$ (P=0.004), respectively. In the exenatide-arm, changes in fractional-UE_{UA} and plasma-UA did not correlate with baseline UA levels (all P>0.6).

Study-C

The study population consisted of 36 patients (Table 1).²⁸ Mean±SD baseline UA was $5.39\pm1.07 \text{ mg/dL}$ (6 patients hyperuricemic). After 12-week treatment with liraglutide, no alterations in UA (P=0.8; Figure 4A) or UE_{UA} (Figure 2 and Figure 4B) were observed compared to placebo. As previously reported, GFR, UE_{Na}, PRC, urine-pH, urine-osmolality and urinary-glucose excretion were not affected after 12 weeks of treatment. (Supplemental

Table 3).²⁸ Yet, compared to placebo, liraglutide reduced HbA_{1c} by $1.3\pm0.2\%$ (p=0.001), plasma glucose by 1.3 ± 0.3 mmol/L (p<0.001), and bodyweight by 1.9 ± 0.7 kg (P=0.009).²⁸ There were no changes in PRC.²⁸ At Week-2, no changes in UA (P=0.8), creatinine-based fractional-UE_{UA} (-1.55±0.88%, P=0.09), or fractional-UE_{Na} were observed in the liraglutide compared to the placebo-arm (Figure 4A–B).²⁸ In the liraglutide-, but not in the placebo-arm, changes from baseline to Week-12 in plasma-UA correlated with baseline plasma-UA (r: -0.52, P=0.022) (Figure 5), while fractional-UE_{UA} or absolute-UE_{UA} did not correlate with baseline plasma-UA (both P>0.5) or with changes in plasma-UA (both P>0.06). No correlations were observed in the liraglutide- or placebo-arm between changes from baseline to Week-2 in creatinine-based fractional-UE_{UA} and baseline plasma-UA (P>0.6).

Study-D

The study population consisted of 35 patients (Table 1). At baseline, mean±SD fasting UA was 5.68±1.19 mg/dL (7 patients hyperuricemic), which decreased after the meal by 0.09 ± 0.04 mg/dL (P=0.03). Prior to randomisation, creatinine-based fractional-UE_{IIA} tended to increase slightly after the meal by 0.02±0.01% (P=0.051). After 8 weeks of treatment, fasting and postprandial UA did not differ between or within treatment arms (P>0.1 for all). Within-group, lixisenatide treatment did not affect postprandial fractional-UE_{UA} (P=0.6) or absolute- UE_{UA} (P=0.9), while insulin glulisine reduced fractional-UE_{UA} (-1.52±0.64; P=0.03) and tended to reduce absolute-UE_{UA} (-0.65 mg/min/1.73m²; P=0.052). This resulted in significant between-group differences (Figure 2). As previously reported, urinepH and fractional-UE_{Na} increased with lixisenatide compared to insulin glulisine in the postprandial state, while GFR was not affected (Supplemental Table 4).²³ Postprandial urinary-glucose did not differ between- (P=0.2) or within-groups (both P>0.6). Compared to insulin glulisine, lixisenatide did not change HbA1c (P=0.9), but decreased blood glucose during the kidney tests by 1.8 ± 0.5 mmol/L (P=0.002). Within-group, insulin concentrations decreased with lixisenatide (Supplemental Table 4).²³ Bodyweight decreased by 1.4±0.6 kg with lixisenatide compared to insulin glulisine (P=0.03).²³ No within- or between-group differences in postprandial PRC or angiotensin-II were observed.²³ In the lixisenatide- and insulin glulisine-arm, changes from baseline to Week-8 in plasma-UA correlated with fasting baseline plasma-UA (r: -0.52, P=0.031 and r: -0.52, P=0.041, respectively) (Figure 5B). Fasting or postprandial fractional- UE_{UA} did not correlate with baseline plasma-UA (all P>0.08) or with changes in plasma-UA (all P>0.3). Changes in absolute-UE_{UA} correlated with baseline plasma-UA in the lixisenatide-arm (r: -0.51, P=0.038) (Figure 5C), but not in the insulin glulisine-arm (r: 0.30, P=0.3), while changes in absolute-UE_{UA} did not correlate with changes in plasma-UA in neither treatment group (both P>0.3).

Discussion

We are the first to determine the immediate and chronic effects of different GLP-1RAs on UA-levels and kidney UA-clearance in humans, using *post-hoc* analyses of four controlled clinical intervention trials. We demonstrate that GLP-1RA exenatide-infusion immediately increases UE_{UA} in healthy overweight males and T2DM patients. Plasma UA-levels increased slightly in healthy subjects, but were unaffected in the T2DM population. In contrast, prolonged treatment with two pharmacologically different GLP-1RA (i.e. 12-weeks

of liraglutide [long-acting agent], or 8-weeks of lixisenatide [short-acting agent]) did not affect UE_{UA} or plasma UA-levels in T2DM patients (of whom ~20% fulfilled the criteria of hyperuricemia).

Recent landmark cardiovascular safety trials indicate that GLP-1RA provide cardiovascular and renal benefit compared to placebo (i.e. standard of care) in at-risk T2DM patients after prolonged therapy.^{13–15, 32} However, underlying cardio-renal protective mechanisms remain incompletely understood. Hyperuricemia is frequently observed in patients with pre- or recent-onset T2DM,33 and increased levels of UA are thought to partly mediate cardiovascular and renal risk in T2DM.^{4, 8} As GLP-1-peptide administration for 10-weeks in diabetic rats reduces UA-levels,²⁰ and GLP-1RA exhibit clinically relevant tubular effects in humans,²¹⁻²⁴ we hypothesised that GLP-1RA administration lowers plasma-UA by increasing kidney UA-clearance.²⁵⁻²⁷ Although intravenous exenatide administration rapidly increased UE_{UA} in overweight healthy subjects (Study-A) and T2DM patients (Study-B), no reduction were observed in UA-levels. This could be explained by the short duration of the studies, although we cannot exclude that exenatide concurrently increased UA-production. Furthermore, upon prolonged treatment with either a short-acting or long-acting GLP-1RA, we did not observe effects on UA-clearance or UA-levels. As such, our results indicate that it is unlikely that UA is a relevant clinical mediator of the recently described cardio-renal benefits of GLP-1RAs in T2DM patients.

Although the tubular handling of UA in humans is incompletely understood, we propose several mechanisms by which exenatide-infusion could have immediately increased UE_{UA} in our studied populations, based on correlation and regression analyses. The effect seems to large extent driven by a concurrent increase in urine-pH, and in part, by an increase in UE_{Na}. Inhibition of NHE3 by GLP-1RA²¹⁻²⁴ may have decreased pH-dependent organic acid transporter (OAT)-4-activity,²⁵ thereby reducing kidney UA-reabsorption. In addition, urinary alkalisation^{26, 27} or increased $UE_{Na}^{2, 34}$ may have contributed directly to the uricosuric effect of acute exenatide administration. While high concentrations of urinaryglucose may stimulate UE_{UA} via the tubular GLUT9 transporter, $^{2, 35, 36}$ we did not observe any relevant glycosuria in our T2DM patient population at baseline or after treatment, which makes such a pathway unlikely. Furthermore, statistical adjustment for the observed decrease in blood glucose, which could result in modest reductions in glycosuria and consequent UA-excretion, did not increase UE_{UA}. We also suggest that the exenatideinduced increase in insulin blunted the increase in UE_{UA}, because insulin is known to augment tubular UA-reabsorption.^{2, 35} Finally, GLP-1RA can decrease renin-angiotensinaldosterone-system (RAAS)-activity,^{37–39} which could increase UE_{IIA} by lowering of angiotensin-II.² However, as we and others^{40, 41} did not observe changes in PRC or angiotensin-II, there seems no important role for RAAS-involvement, although some contribution of intrarenal RAAS cannot be excluded.

In order to explore the sustainability of the immediate GLP-1RA-induced effect on UE_{UE} , and to determine whether this translates into plasma UA-lowering upon long-term treatment, we analysed the data of two clinical intervention trials involving structurally different GLP-1RAs (Study-C and Study-D). GLP-1RAs can be categorized as either short-acting/ prandial compounds, which provide short-lived receptor activation (e.g. exenatide and

lixisenatide) or as long-acting compounds (e.g. exenatide once-weekly and liraglutide), which activate GLP-1 receptors continuously. The pharmacokinetic differences between these drugs are known to lead to clinically relevant differences in their pharmacodynamic profiles. For example, due to their continuous receptor stimulation and resultant tachyphylaxis, the ability of long-acting GLP-1RAs to decrease gastric emptying rate and to inhibit areas in the central nervous system involved in hedonic feeding are not sustained during prolonged treatment.^{29, 30} In line with these observations, we recently suggested that tachyphylaxis and/or induction of compensatory mechanisms at the tubular level explains that only short-acting GLP-1RA harbor sustained natriuretic and urine alkalizing actions during prolonged treatment.^{23, 28} We hypothesised that such divergent actions could also emerge with respect to tubular UA-handling. Indeed, 12-week treatment with the long-acting GLP-1RA liraglutide compared to placebo did not affect UE_{UA}. Interestingly, although 8week treatment with the short-acting GLP-1RA lixisenatide increased UE_{UA} compared to insulin glulisine, this was entirely mediated by an anti-uricosuric effect in the insulin glulisine-arm. Thus, these results do not indicate that lixisenatide affects UE_{UA}. Finally, UAlevels were unaffected by either GLP-1RAs in our prolonged intervention studies despite decreases in bodyweight (potentially improving insulin resistance).⁴²

In comparison, the initial stimulating effects of an SGLT2 inhibitor on UE_{UA} also seems to wane upon prolonged intervention, as suggested by a mechanistic study in healthy subjects and *in vitro* transport experiments.⁴³ However, in contrast to current GLP-1RA data, chronic SGLT2 inhibitor therapy does have a sustained suppressive effect on plasma-UA,³⁶ suggesting that these drugs establish a new steady state in UA-metabolism and clearance.

Notably, our correlation analyses suggest that prolonged GLP-1RA treatment may lower plasma-UA in T2DM patients with higher baseline plasma-UA levels. However, as a similar relation is observed in patients treated with insulin glulisine, a glucose-dependent mechanism is suggested.³⁵ In line, individual responses in renal UA-handling did not associate with changes in plasma-UA, although the isolated correlation of changes in absolute-UE_{UA} with plasma-UA in patients treated with lixisenatide merits further research.

The described studies are not without limitations. First, intestinal UA-excretion or UAformation was not assessed. Such measurements are technically challenging, and intestinal excretion was not expected to be affected substantially.³ UA-formation could theoretically be influenced by treatment through alterations in circulating fructose levels.³⁵ We did not assess fructose levels in these trials, but emphasise that this would be confounded by glucose-lowering. Second, we did not measure UA-crystallization as the increment in urinepH dramatically improves UA solubility (especially when pH >6.0-6.5),⁴⁴ rendering the likelihood of precipitation negligible. It is tempting to speculate that this would potentially also prevent UA-mediated tubulopathy.³⁵ However, we did not observe any effects on glomerular and tubular injury markers (albumin-creatinine ratio, KIM-1 or NGAL) as previously reported in T2DM patients without overt nephropathy.^{23, 24, 28} Third, since the intervention conditions in Study-A were performed sequentially, we cannot exclude timedependent effects. This may explain the somewhat unexpected slight increase in plasma-UA after immediate exenatide-infusion in healthy subjects. Fourth, our T2DM population had a prolonged history of diabetes and we did not specifically include patients with (symptomatic

or asymptomatic) hyperuricemia, which does not allow generalisation of the current data to patients with a shorter diabetes-history and/or higher UA-levels at baseline.^{33, 45} Fifth, T2DM patients in Study-B and Study-C had relatively low cardiovascular risk, while the duration of follow-up in Study-C and Study-D was relatively short, which may hamper direct generalisation to the recently published GLP-1RA cardiovascular outcome trials. Interestingly, the association between UA levels and progression of renal disease in diabetes appears, however, to be present only in healthier subjects, i.e. those with earlier stages of chronic kidney disease (CKD),⁴⁶ and is lost in more advanced CKD.^{47, 48} Sixth, effects of lixisenatide were investigated in the postprandial state and an active comparator was used instead of placebo, which may have led to a more heterogeneous response and hampers direct comparison with the other reported studies that were performed in the fasting state. Seventh, lixisenatide markedly slows gastric emptying rate,²³ which could affect purine uptake in the postprandial state. However, the effects of the mixed breakfast on plasma-UA and UE_{UA} were minor at baseline. Finally, although we did not test prolonged treatment effects of exenatide twice-daily, the drug is structurally similar to lixisenatide, and harbors comparable pharmacokinetic and pharmacodynamic properties.⁴⁹ As such, we believe it is unlikely that prolonged treatment this formulation of exenatide would result in differential effects on kidney UA-clearance and/or UA-levels.

In conclusion, immediate exenatide-infusion increases UE_{UA} in healthy overweight males and T2DM patients by increasing UE_{UA} , possibly mediated by GLP-1RA–induced NHE3inhibition in the proximal tubule and/or consequent urine alkalization. Prolonged treatment with the long-acting GLP-1RA liraglutide or the short-acting GLP-1RA lixisenatide does not affect UE_{UA} or UA-levels in T2DM patients with normal plasma UA-levels and relatively low cardiovascular risk. Our results indicate that it is unlikely that UA is a relevant mediator of the cardiovascular and renal benefits of GLP-1RA.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Figure 1. Trial designs





Data are mean \pm SEM, median [IQR], or baseline-corrected mean difference \pm SEM. Wilcoxon signed-rank test was used to examine effects of exenatide versus placebo in Study-A. Multiple linear regression corrected for correponding baseline values were used to determine exenatide-induced effects compared to placebo in Study-B, liraglutide-induced effects compared to placebo in Study-C, and lixisenatide-induced effects compared to insulin glulisine in Study-D. Paired t-test was used for within-group comparisons in Study-D.



Figure 3. Correlations of within-group changes in healthy overweight males (Study-A) Spearman signed rank test (A and B) or Pearson's correlation (C) was used to assess correlations.



Figure 4. Effect of 12-week treatment with liraglutide in T2DM patients on creatinine-based fractional uric acid-excretion (A) and plasma uric acid (B)

Data are median [IQR]. Multiple linear regression corrected for correponding baseline values was used to examine liraglutide-induced effects compared to placebo. No significant differences were found.





Spearman signed rank test was used to assess correlations.

Table 1

Baseline clinical and biochemical characteristics

	Study-A (N=9)	Study-B (N=52)	Study-C (N=36)	Study-D (N=35)
Clinical characteristics				
Male sex, $n(\%)$	9 (100)	39 (77)	27 (75)	23 (66)
Age, years	22.0 [22.0–23.0]	62.9 ± 6.9	63.0 ± 7.0	61.7 ± 6.6
Weight, kg	96.0 (7.0)	99.0 ± 14.3	101.2 ± 14.9	97.5 ± 16.4
BMI, kg/m ²	$29.3 \pm (1.7)$	31.1 [28.3–33.6]	31.2 [29.2–33.3]	31.5 ± 4.0
Systolic BP, mm Hg	113 [107–123]	135 ± 15	137 ± 16	134 ± 16
Diastolic BP, mm Hg	67 [61–71]	76 ± 6	77 ± 6	76 ± 9
Biochemistry				
HbA _{1c} , %	5.1 [5.0-5.3]	7.3 ± 0.6	7.4 ± 0.7	8.0 ± 0.9
HbA _{1c} , mmol/mol	32 [31–34]	57 ± 7	58 ± 8	64 ± 9
Fasting plasma glucose, mmol/L	4.9 [4.4–5.2]	8.4 ± 1.5	8.6 ± 1.7	7.0 ± 2.0
HOMA2-IR	0.67 [0.59–1.47]	1.7 [1.2–2.5]	1.6 [1.1–2.4]	N/A*
Plasma-UA, mg/dL	6.33 [6.05-7.05]	5.50±1.04	5.39±1.07	5.68±1.19
Hyperuricemia [#] , $n(\%)$	2 (22)	9 (17)	6 (17)	7 (20)
Diabetes history characteristics				
Type 2 diabetes duration, years	N/A	7 [4–12]	8 [4–12]	13 ± 7
Metformin use, $n(\%)$	N/A	49 (94)	34 (94)	32 (91)
Sulphonylurea use, $n(\%)$	N/A	23 (44)	15 (42)	0 (0)
Insulin use, <i>n</i> (%)	N/A	0 (0)	0 (0)	35 (100)
Xanthine-oxidase inhibitor use, $n(\%)$	0 (0)	1 (2)	1 (3)	2 (6)
Antihypertensive medication use, $n(\%)$	N/A	34 (65)	26 (72)	22 (63)
RAS inhibitor use $\ddagger, n(\%)$	N/A	32 (62)	20 (56)	22 (63)

Data are shown as mean \pm SD, median [IQR] or absolute (%).

<u>Abbreviations</u>: BMI=body mass index. BP=blood pressure. HbA_{1c}=glycated hemoglobin. NA=not applicable. HOMA2-IR=updated homeostatic model assessment-insulin resistance. RAS=renin-angiotensin-system. UA=uric acid.

* Not calculated due to interference with insulin glargine.

[#]Defined as plasma-UA >6.8 mg/dL⁶. Distribution of hyperuricemic cases is as follows: Study-B, 5 randomised to placebo and 4 to exenatide; Study-C, 2 randomised to placebo and 4 to liraglutide; Study-D, 6 randomised to insulin glulisine and 1 to lixisenatide.

[‡]The use of losartan was as follows: Study-B, 1 randomised to placebo and 1 to exenatide; Study-C, 1 randomised to placebo and 1 to liraglutide; Study-D, 1 randomised to insulin glulisine and 2 to lixisenatide.