

High-Resolution Adhesion Kinetics of EGCG-Exposed Tumor Cells on Biomimetic Interfaces: Comparative Monitoring of Cell Viability Using Label-Free Biosensor and Classic End-Point Assays

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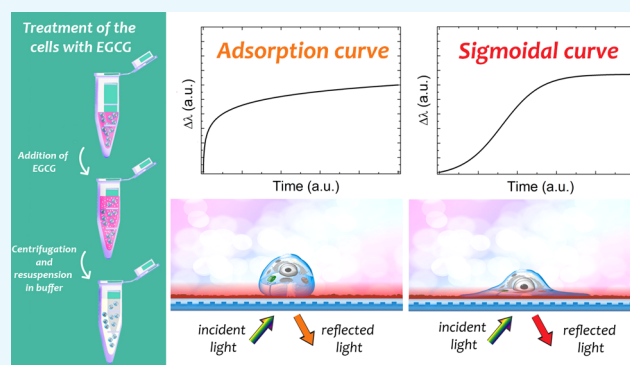
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ABSTRACT: A high-throughput label-free resonant waveguide grating biosensor, the Epic BenchTop, was utilized to in situ monitor the adhesion process of cancer cells on Arg-Gly-Asp tripeptide displaying biomimetic polymer surfaces. Using highly adherent human cervical adenocarcinoma (HeLa) cells as a model system, cell adhesion kinetic data with outstanding temporal resolution were obtained. We found that pre-exposing the cells to various concentrations of the main extract of green tea, the (–)-epigallocatechin gallate (EGCG), largely affected the temporal evolution of the adhesion process. For unexposed and low dosed cells, sigmoid shaped spreading kinetics was recorded. Higher dose of EGCG resulted in a complete absence of the sigmoidal character, and displayed adsorption-like kinetics. By using the first derivatives of the kinetic curves, a simple model was developed to quantify the sigmoidal character and the transition from sigmoidal to adsorption-like kinetics. The calculations showed that the transition happened at EGCG concentration of around 60 $\mu\text{g}/\text{mL}$. Using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide end-point assay, we concluded that EGCG is cytostatic but not cytotoxic. The effect of EGCG was also characterized by flow cytometry. We concluded that, using the introduced label-free methodology, the shape of the cell adhesion kinetic curves can be used to quantify in vitro cell viability in a fast, cost-effective, and highly sensitive manner.



INTRODUCTION

Natural compounds are becoming more and more popular in biomedicine, especially in cancer treatment and to develop novel antimicrobial agents.^{1–4} Tea catechins, especially (–)-epigallocatechin gallate (EGCG), have been shown to have various health benefits, for example, anti-metastasis, anticancer, anti-inflammatory, and antioxidant properties, and can prevent cardiovascular disease as well.^{5–8} EGCG is one of the most studied active substances, and many studies observed its effects on several cancer and normal cell types, and in animal models.⁴ This compound has significant impact on cell adhesion and movement, apoptosis, and proliferation, generally by altering gene expression.^{4,5,9–11} Tea polyphenols are well known for their antioxidant activities, too.^{5,12} Among them, EGCG is the most effective compound interacting with reactive oxygen species.¹³ EGCG and other catechins are unstable at

high temperature and under alkaline and neutral conditions; EGCG oxidizes and dimerizes easily^{5,12,14} at pH above 7.^{5,12,14} In an aqueous solution, it changes from noncolored at around natural pH to yellow at higher pH; the absorption in the UV range becomes more pronounced.^{4,5,15}

Determination of cell viability is a critical step in screening the efficacy of compounds, when evaluating the response to cytotoxic moiety. Flow cytometry is a sensitive and mainstream method to determine compound-induced cytotoxic effects and cell death. The main advantage of the method allows the analysis on a per-cell basis using fluorescent dyes to enter viable or dead cells. Propidium-iodide (PI) is a polar, fluorescent

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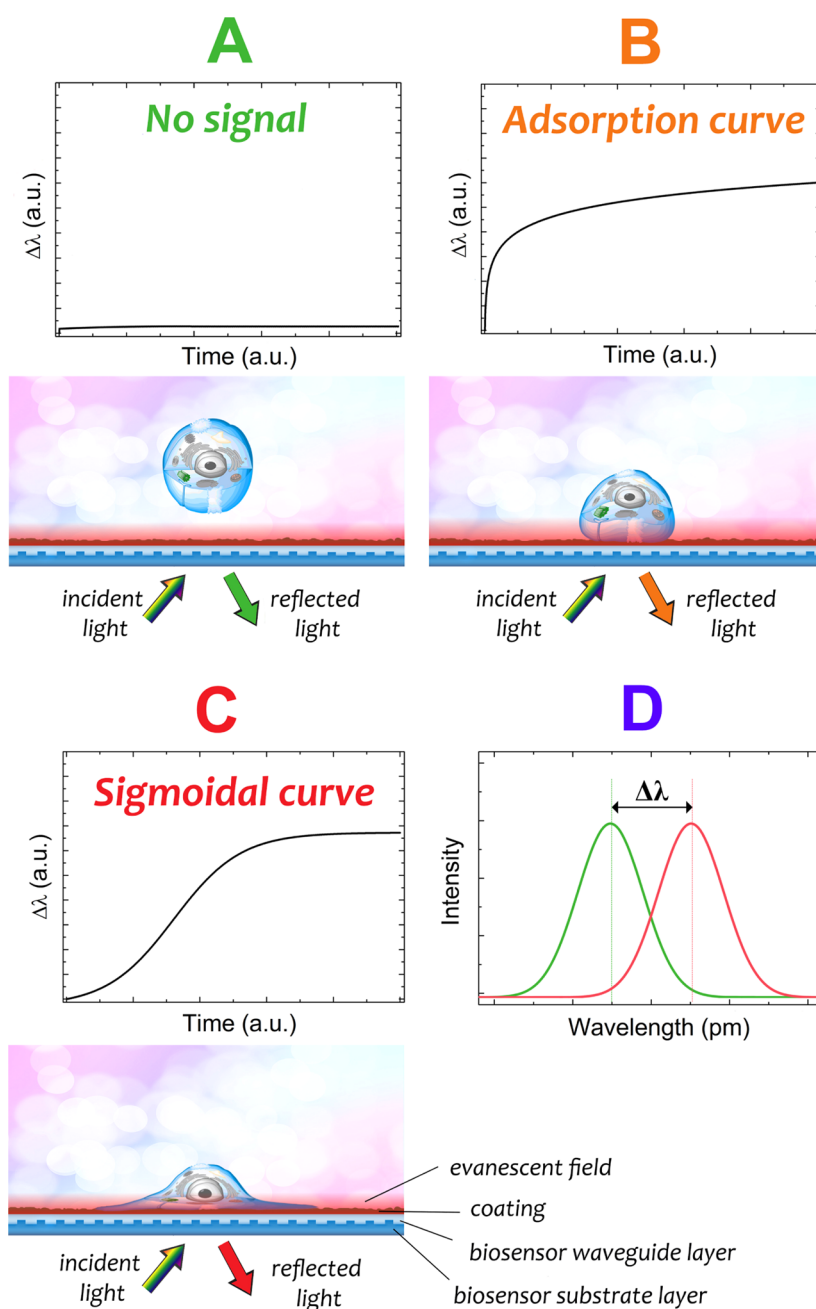


Figure 1. Schematic illustration of the working principle of the Epic BT biosensor. An optical biosensor found in each well is illuminated by a broadband light source. The biosensor contains a grating coupled planar optical waveguide and its surface can be coated with a protein or polymer of interest. Light coupled into the waveguide creates a resonant mode and a corresponding evanescent field penetrating to a depth of 150 nm above the sensor surface. Any refractive index change inside the evanescent field shifts the resonant wavelength of light reflected back from the biosensor. The biosensor signal is the wavelength shift of the reflected light. (A) If there is no change in the evanescent field, there is no signal. (B) The cell attaches but does not adhere onto the biosensor surface. This phenomenon represents an adsorption kinetic curve, which indicates a nonliving, “dead” process. Note, it also occurs when proteins adsorb onto the surface. (C) The cells adhere onto the biosensor surface. This phenomenon represents a sigmoidal kinetic curve, which indicates a “living” process. (D) Schematic graph of the difference ($\Delta\lambda$) between the resonant peak wavelengths of the reflected lights. The equipment plots the $\Delta\lambda$ data in time during the experiment (see C and B).

compound and can only enter cells that lack membrane integrity. After PI staining, nonviable cells show a bright red fluorescence, whereas viable cells remain nonfluorescent.¹⁶ Using the plate-based 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) method, the membrane permeability and mitochondrial activity of the cells were determined in metabolically active cells.¹⁷

However, most of the label-based assays have serious disadvantages, for example, labeling techniques use fluorescent

markers that may affect normal cell behavior and the imaging time is often limited by the low signal and the bleaching of the marker.⁵

Detection of cellular adhesion is of significant diagnostic and basic research utility. Changes in cell adhesion can be a sign for various illnesses; for example, the variety of integrins, a major group of cell adhesion receptors that bind to the extracellular matrix, changes during tumor transformation.⁵ Measurement of the effect of bioactive substances on the adhesion of tumor cells

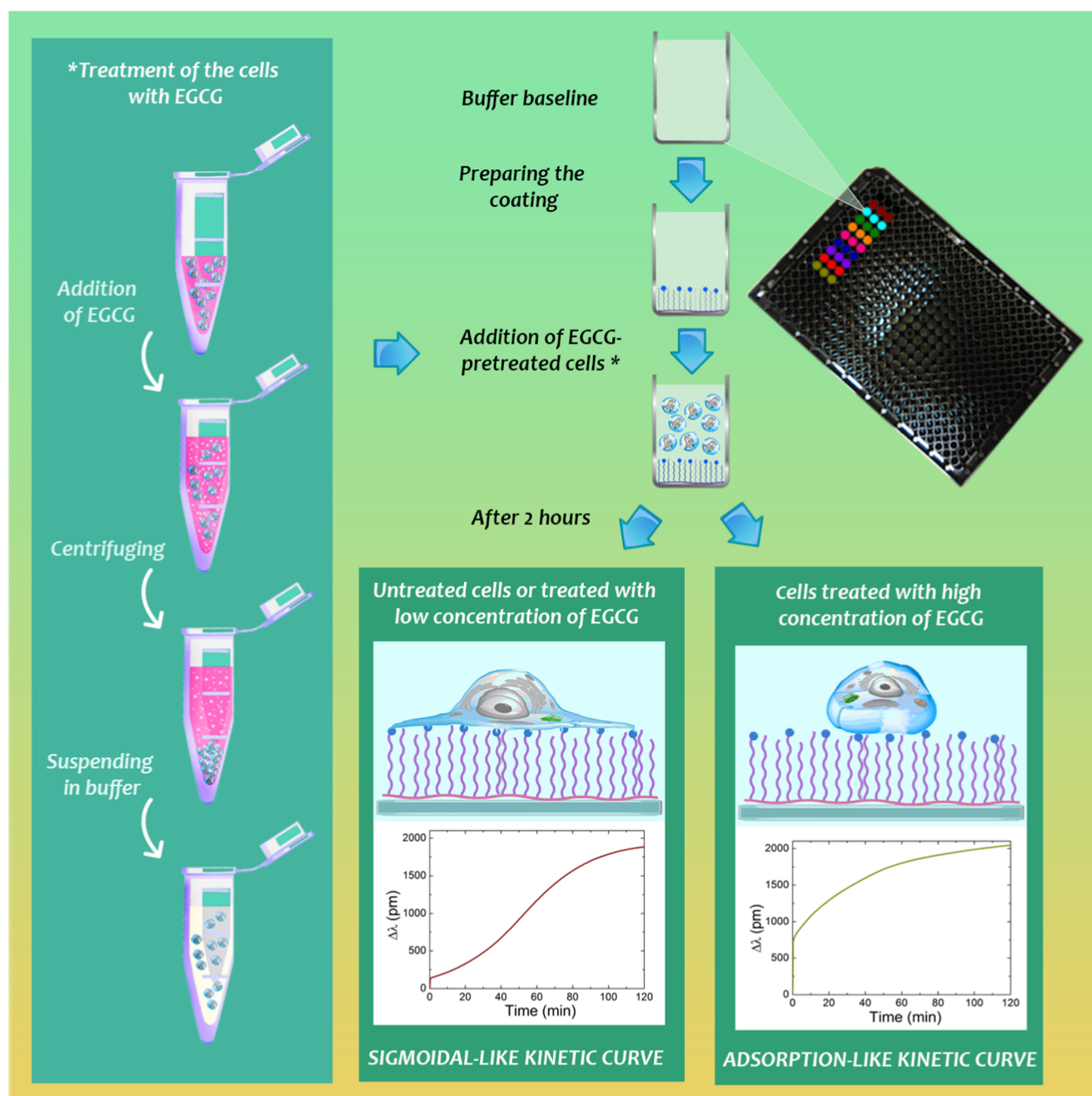


Figure 2. Summarized schematic illustration of the label-free method employing the Epic BT biosensor. Cells were pre-exposed to various concentrations of EGCG and then transferred to the biosensor wells to record their adhesion and spreading on the RGD displaying polymer surfaces. As the most striking observation, cell spreading with low concentration of EGCG or without EGCG shows sigmoidal-like kinetic curves (active, living process), whereas treatments with high concentration of EGCG result in adsorption-like kinetic curves (dead process).

can be an effective tool in the design of antineoplastic pharmaceuticals.⁵ A wide range of previously existing and well-documented, conventional label-based experimental methods are available to assess cellular processes such as in vitro cell viability and adhesion.^{5,18–25}

Label-free biosensors, not requiring the usage of dyes, have the ability to become a routine tool for measuring cell adhesion, spreading, proliferation, signalization, and cytotoxicity as well.⁵ These techniques are especially promising when the real-time kinetics of interactions have to be investigated.

In the measurements of label-free techniques, biomimetic surfaces are usually applied as coatings to create circumstances that resemble the real biological conditions. The biomimetic

surfaces mimic the materials that occur in vivo, but these artificial substrates are simpler to hand, they need less preparations, and the created coatings are more reproducible.

Poly(*l*-lysine)-*graft*-poly(ethylene glycol) (PLL-*g*-PEG) is a biomimetic polycationic copolymer with poly(ethylene glycol) (PEG) chains covalently grafted onto a positively charged PLL backbone. It spontaneously adsorbs from the aqueous solution via electrostatic interactions onto negatively charged surfaces such as tissue culture polystyrene, TiO₂, Nb₂O₅, and SiO₂.^{5,26,27} However, several studies have shown that it can also adsorb onto nonpolar, hydrophobic poly(dimethylsiloxane) (PDMS) surfaces from the aqueous solutions. The PEG side chains stretch into the bulk aqueous solution to generate a brushlike

conformation due to its high affinity for water. Recent studies have shown that a PLL-g-PEG layer on metal oxide surfaces displays resistance to a nonspecific adsorption of proteins and cell adhesion.^{5,27,28}

The copolymer and its cell adhesive, functionalized counterpart with Arg-Gly-Asp (RGD) motif, PLL-g-PEG-RGD, can be mixed to vary the cell adhesion ligand density and to promote cell adhesion and spreading on the coated surfaces.⁵ Here, we use the mixture of PLL-g-PEG and PLL-g-PEG-RGD to create highly cell adhesive surfaces in a reproducible manner.^{5,29,30}

In the present work, we investigate the adhesion kinetics of EGCG pre-exposed cells on PLL-g-PEG-RGD as biomimetic surfaces using the Epic BenchTop (BT) resonant waveguide grating (RWG) biosensor. The Epic BT employs resonant waveguide gratings arranged in a 96- or 384-well array format. In each well, the resonant wavelength of the waveguide is monitored, which is sensitive to refractive index variations above the sensing surface. This surface-sensitive high-throughput label-free technique was previously demonstrated to monitor cell signaling and adhesion.^{5,30–35} Thanks to its excellent temporal resolution, the cellular adhesion can be recorded with unprecedented data quality.³⁰ This feature is exploited in the present work by investigating the shapes of the cell adhesion kinetic curves in detail. A simple model is created to quantify the shape of the kinetic curves. The results of the label-free investigations are compared with classical MTT assay and flow cytometry employing PI staining. Our work demonstrates a novel way to measure cell viability in a fast, label-free, and highly sensitive manner.

RESULTS AND DISCUSSION

In vitro effect of EGCG on cellular viability was accessed by three different methods using HeLa cells: (i) to analyze real-time cell adhesion kinetics, (ii) to determine metabolic activity using colorimetric MTT assay, and (iii) to quantify dead or dying cells by cytofluorometry with PI staining.

The plate-based EPIC system is suitable for observing adhesive parameters in real time as physiologically relevant and quantitative data related to the cellular fitness and viability. The optical core of the sensing procedure is schematically illustrated in Figure 1. Using classic end-point tetrazolium reduction, the cytostatic or cytotoxic activity can be determined through enzymatic activity as a marker of viable cells generating a signal that is proportional to the number of viable cells.

Adhesion Kinetics of EGCG Pre-Exposed Cells Recorded by the Optical Biosensor. The schematic illustration of the employed methodology is summarized in Figure 2. One can observe an interesting phenomenon on the kinetic curves measured by Epic BT.

If the cells can well adhere and spread, sigmoidal-like curves can be monitored as a characteristic pattern of living cells (living process). In contrast, at higher concentration, adsorption-like curves imply that the observed change is due to dying or dead cells (dead process). Note, the adsorption-like curves are typically observed when discrete objects are adsorbing on a solid surface. In this case, the main driving force is the unoccupied surface area; therefore, the adsorption is fastest at the very beginning, when the free surface area is maximal. In contrast, the sigmoidal kinetics require an active, self-catalytic, and self-controlled mechanism, a unique characteristics of the living cells.

In these measurements, we both monitored the sigmoidal- and adsorption-like kinetic curves, depending on the

concentration of the EGCG. At values lower than the added 40 $\mu\text{g}/\text{mL}$ of EGCG, we received spreading curves (active, living process), and above this value, adsorption curves (dead process) (Figure 3). (Supposedly, due to the EGCG, the

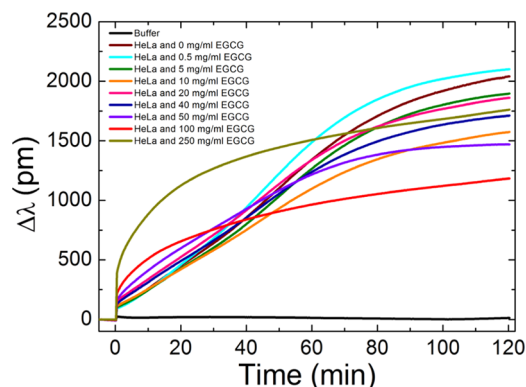


Figure 3. Kinetic cell adhesion curves recorded by Epic BT. The plotted lines represent the averaged values of the three parallel experiments. For obtaining these data, the cells were preincubated with solutions having different EGCG concentrations. After, these treated cells were pipetted into the biosensor wells and cell adhesion was monitored for 2 h. It is clearly seen that increasing the EGCG concentration decreases the sigmoidal character of the kinetic curves. For the highest EGCG concentrations, the sigmoidal character is completely missing and adsorption-like curves are recorded.

cells—instead of active adhesion—may even secrete some cell-specific compounds at higher extract concentrations.) This border concentration is approximately 40–50 $\mu\text{g}/\text{mL}$, above which the EGCG has harmful effects on the cancer cells.

Following the EGCG preincubation, our observation was that the size of the cells shrunk and their shape, spreading, and adhesive properties also changed. The microscopy images (Figure 4) clearly show that at low EGCG concentrations (0–40 $\mu\text{g}/\text{mL}$), the cells could well adhere and spread; after the 2 h period, the cells almost formed a homogeneous layer on the RGD displaying a biosensor surface. However, at higher EGCG concentrations of 40–50 $\mu\text{g}/\text{mL}$, the shape of the cells changed; they are not as spread as the other cells with lower concentrations or without green tea extract. At higher quantities, the cells remained rounded. The observation of the same phenomenon can be followed on the kinetic curves as mentioned earlier.

In Vitro Cytotoxic and Cytostatic Activity of the Compound EGCG on HeLa Cell Culture (MTT assay). Tetrazolium salts are widely used for in vitro assessment of metabolic activity of cells. 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) reduction is one of the most frequently used methods for measuring cytotoxicity and cell proliferation.³⁶ HeLa cells were treated with EGCG at 0.064–500 $\mu\text{g}/\text{mL}$ concentration range and the viability of the cells was determined by using the MTT assay.

The 50% inhibitory concentration (IC_{50}) values were determined from the dose–response curves. We found that EGCG exhibited a modest, but not negligible cytotoxic effect on HeLa cells ($\text{IC}_{50} > 500 \mu\text{g}/\text{mL}$). From the dose–response curves, it can be observed that the results obtained with the Epic BT measurements are reconcilable: the transition concentration is probably 20–30 $\mu\text{g}/\text{mL}$ in the MTT cell cytotoxicity assay as well. The cytostasis assay (at 0.064–1000

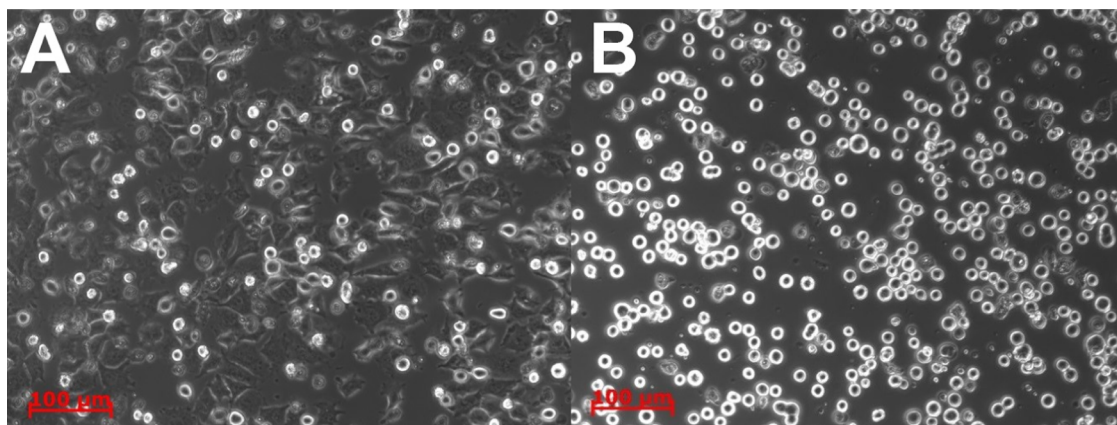


Figure 4. Microscopic images of the cells after the measurement in the Epic BT microplate wells. (A) Cells without EGCG treatment could spread onto the surface and form adhesive contacts. (B) Cells treated with 250 $\mu\text{g}/\text{mL}$ EGCG (highest concentration) remained rounded and spreading was inhibited. Scale bar: 100 μm . The differences between the cells treated by the lowest EGCG concentrations cannot be detected by phase contrast microscopy (images not shown). Note, in contrast, the label-free kinetic curves well demonstrate the differences even at low EGCG concentrations (see Figure 3 for more details).

$\mu\text{g}/\text{mL}$ concentration range) shows higher inhibitory concentrations.

Evaluation of Viability Profiles of EGCG-Treated HeLa Cells by Flow Cytometry. The cells were exposed to EGCG for 1.5 h. Following the exposure and removal of EGCG by washing and trypsinization of the cells, they were transferred to the FACS tubes and stained with PI. As HeLa is an adherent culture, when preparing the samples for assay, the cells are carefully harvested to avoid the loss or damage of a certain subpopulations, and the cells are thoroughly processed to single-cell suspension before PI staining. The data shown are typical of those obtained from repeated experiments. The spontaneous cell death due to sample preparation was determined for untreated control cells (5–10% of death). For maximum lysis (100% of death), the cells were incubated at 37 $^{\circ}\text{C}$ with 20 μL of saponin at 0.3 mg/mL (data not shown). A threshold was set on the forward scatter to exclude cell debris or any artifacts. The gates that were obtained with FACSDiva software and used for data acquisition. The gated dead cells were displayed on a histogram of log-scale red fluorescence of PI to enable the differentiation between dead and live HeLa cells. The cell death was expressed as the percentage of PI positive cells.

Quantification of the Shape of the Label-Free Kinetic Curves: Comparison with Classical Cell Viability Assays. We analyzed the kinetic curves of the Epic BT in more detail by using the following ideas. First, the kinetic data were differentiated (first derivative) and the following parameter was calculated.

$$\Delta = \left. \frac{d\Delta\lambda}{dt} \right|_{\text{max}} - \left. \frac{d\Delta\lambda}{dt} \right|_{t=0} \quad (1)$$

$\left. \frac{d\Delta\lambda}{dt} \right|_{\text{max}}$ is the first derivative maximal value, $\left. \frac{d\Delta\lambda}{dt} \right|_{t=0}$ is the first derivative at the $t = 0$ timepoint (see Figures 6 and 7 for more details). Note, in the actual calculations, the first derivative recorded at 2.5 min was used instead of the first derivative at $t = 0$ due to practical reasons.

The parameter Δ characterizes the shape of the kinetic curves in a straightforward manner. Simply, if $\Delta > 0$ pm/min, the recorded curve has a sigmoidal character. For all adsorption-like curves (having the maximum of the first

derivative at $t = 0$), the parameter Δ is zero. Therefore, these two types of behaviors can be well distinguished.

The schematic illustration of the method is seen in Figure 5 with all of the relevant parameters highlighted. The time dependency of the first derivatives for the actual biosensor measurements are plotted in Figure 6.

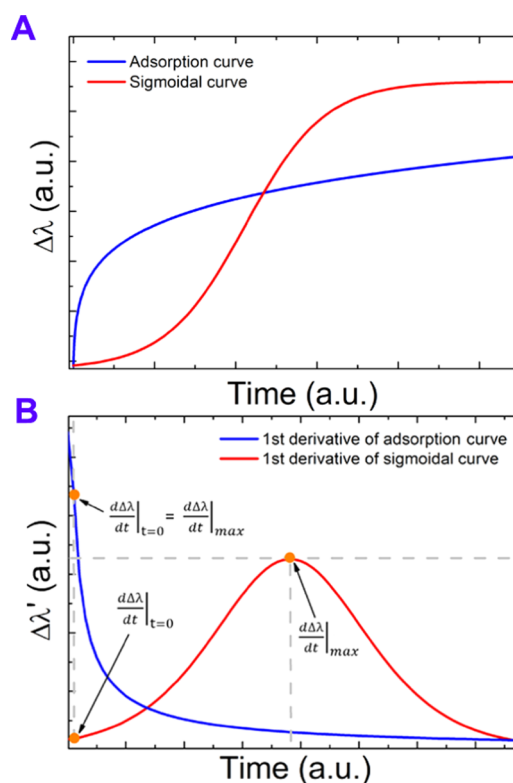


Figure 5. Schematic illustration of the calculation method to reveal the transition EGCG concentration from the Epic BT data from sigmoidal- to adsorption-like curves. (A) Typical sigmoidal- and adsorption-like kinetic curves. (B) The first derivative of the curves plotted in (A) with the parameters appearing in the equation of Δ highlighted. In case of adsorption-like curves, the parameter Δ is 0 because the first derivative at $t = 0$ timepoint is equal to the first derivative's maximum value.

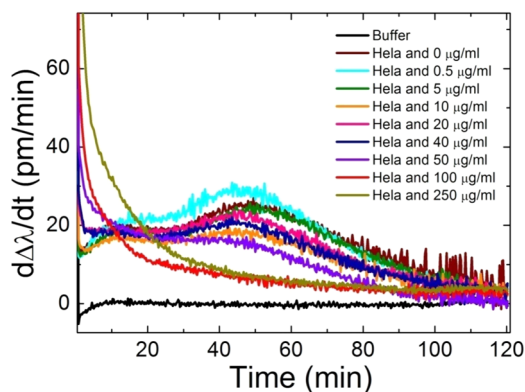


Figure 6. First derivatives of the kinetic curves recorded by Epic BT (and plotted in Figure 4). Data corresponding to HeLa cells treated by EGCG from 0 to 250 $\mu\text{g}/\text{mL}$ concentrations and a buffer control is shown. It is clearly seen that the character of the kinetic curves drastically changes as the EGCG concentration is increased.

Next, the Δ values for various EGCG concentrations were plotted with the concentration data at the x -axis (logarithmic scale), and the data points were fitted with a linear curve (Figure 7A). The Δ values are 0 pm/min at concentrations higher than 40 $\mu\text{g}/\text{mL}$ and >0 at lower concentrations. The exact transition concentration is $60 \pm 40 \mu\text{g}/\text{mL}$, calculated by linear extrapolation (Figure 7A). The error of the calculation is estimated from the standard errors of the linear fit. To compare the label-free data with the findings of the labeling methods, we plotted the MTT cytostasis (Figure 7B), cytotoxicity (Figure 7C), and flow cytometry results (Figure 7D).

The calculated transition concentration is compatible with the MTT assay because this is the point where the fitted dose–response curve (cytostatic activity) starts to increase as well. From the results shown in Figure 7, the novel method based on Epic BT seems to be much more sensitive because it shows evident dose dependence even under effective, transition concentration as well, whereas the other two MTT curves do not show a difference. Therefore, there is a dose-dependent noncytotoxic effect as well, which can be clearly monitored and highlighted by Epic BT. Therefore, the above protocol might serve as an alternative way to quantitatively characterize cell viability in a completely label-free and highly sensitive manner.

In general, the Epic BT assay was more sensitive than the flow cytometric or MTT assay. The MTT assay gave results approximately comparable to those from the flow cytometric assay. In comparing the flow cytometric with the MTT assay, we find higher standard deviations with the flow cytometric assay (standard deviation less than 10–15% of mean versus 6–8% for the MTT assay).

Moreover, Epic BT is a high-throughput and fast method. Only 1 h would have been enough to receive the maximum values of the derivative curves (see Figure 6). These features make this measurement technique highly cost-effective too, when compared with standard biosensing equipment having a single measurement channel. Significant amount of time can be saved by performing hundreds of experiments in a single run. Moreover, the status of the cells and the condition of the reagents are always the same in these parallel performed experiments. It should be also emphasized that the results obtained by the biosensor are not affected by any molecules typically used for labeling and possibly even interacting with the investigated compounds.

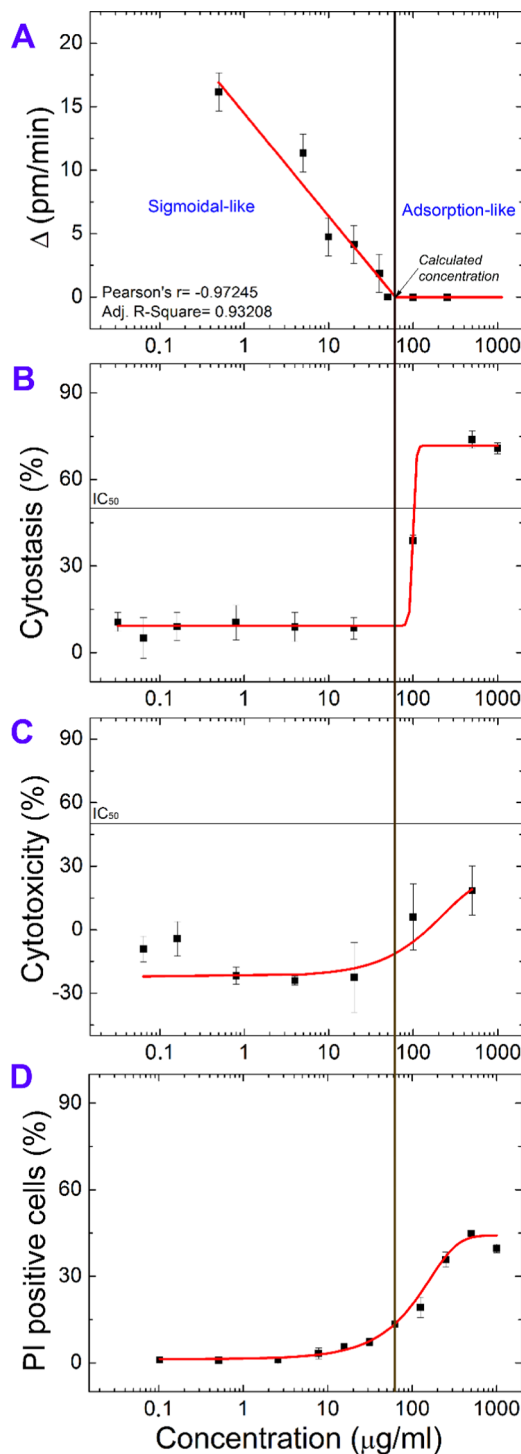


Figure 7. Calculated dose–response curves and the transition concentration compared with the results of the MTT and cytofluorimetric assays. (A) Plotted and fitted Δ values. On the left, $\Delta > 0$ pm/min, indicating an active cell spreading process. On the right, at higher EGCG concentrations, Δ values are 0 pm/min, showing an adsorption-like process. The transition concentration between these two types of behaviors is $60 \pm 40 \mu\text{g}/\text{mL}$, calculated by linear extrapolation. (B) Result of the in vitro cytostatic activity assay. It shows higher inhibitory concentrations than cytotoxicity test. (C) Result of the in vitro cytotoxicity assay (MTT assay). EGCG exhibited a modest, but not negligible cytotoxic effect on HeLa cells ($\text{IC}_{50} > 500 \mu\text{g}/\text{mL}$). (D) Result of the in vitro flow cytometry assay. Cell death rate was expressed as the percentage of PI positive cells.

CONCLUSIONS

Cell-based assays are widely used to determine if a compound has a cytostatic activity or a direct cytotoxic effect leading to cell death. To access cellular viability plays an important role in the functional characterization of a molecule. There are wide variety of cell viability methods that provide accurate picture of cytostatic and cytotoxic activity, such as colometric assays and cytofluorimetric analysis of individual cells. Previous studies have shown that EGCG can inhibit cell adhesion,^{5,23,24,37} and this extract has an obstructive effect not just on cancer cell adhesion but also on cell movement and can induce apoptosis too,^{9,23,24,37–39} whereas EGCG has an influence on cell viability as well. The classic colorimetric end-point method to evince this effect is the MTT assay; however, both MTT- and 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfo-phenyl)-2H-tetrazolium (MTS)-based assays underestimate the antiproliferative effect of EGCG. This phenomenon is probably due to the increased activity of mitochondrial dehydrogenase in response to the EGCG treatment, as well as the intrinsic potential of EGCG to reduce MTT and MTS and increase the formation of formazan.¹⁷ Thus, a careful evaluation of the applied method is essential for the in vitro assessment of cell viability and proliferation¹⁷ after EGCG treatment.

To monitor compound-induced, real-time dose response, label-free technologies were demonstrated to be robust, but sensitive tools with high statistical significance and reduced artifacts.^{32,40,41}

In this study, the Epic BT—a novel high-throughput label-free RWG biosensor—was applied to study the effect of EGCG on the kinetics of cancer cell adhesion. The recorded real-time adhesion curves were compared with the data of MTT and flow cytometric cell viability assays.

On the basis of the kinetic information, we identified normal spreading curves, i.e., an active process of living HeLa cells, at EGCG concentrations lower than 60 $\mu\text{g}/\text{mL}$. Above this concentration, we observed simple adsorption curves lacking normal cell spreading, i.e., a cytostatic effect on HeLa cells is seen. We calculated that $60 \pm 40 \mu\text{g}/\text{mL}$ is a transition concentration between the two types of kinetic curves by developing a straightforward mathematical methodology. This concentration value is the starting point of the increasing part of the fitted cell viability cytostatic curve obtained from the MTT assays. Our results show that the high-throughput Epic BT biosensor can be successfully applied in cytotoxicity assays with a superior sensitivity.

Our technique offers a noninvasive, label-free, cost-effective platform to characterize cell viability with a high sensitivity and an accurate correlation with the conventional (classical) tetrazolium method. Furthermore, this appliance distinguishes the differences between the efficacy caused by very low EGCG concentrations as well, whereas the effects of low concentrations were too small to be reliably defined by the colorimetric or flow cytometry methods. These data points were represented as scattered flat sections (plateaus) on the respective graphs. The intensity of the measured signal (effect of EGCG based on absorbance or intracellular fluorescence intensity) is not detectable or difficult to quantify; the small changes produced by low concentrations of EGCG become lost within the base noise.

Using the label-free method, the observation range was considerably extended due to the enhanced signal-to-noise ratios. The data evaluation and the quantification of the

differences are simpler in the case of Epic BT as well. Thus, it may have an important role in biomedical research, for example, in early or second stage drug candidate testing due to its predictive ability. In our opinion, another possible scope of the introduced methodology is the testing of cytostatic or cytotoxic compounds on different tumor cells even from patients to find drugs that are harmful to the drug-resistant tumor cells as well. One would be able to measure a lot of active substances simultaneously in a very short time (in approximately 1 h); 16 drugs with their 8 concentrations or 8 drugs with their 16 concentrations in the case of a 384-well plate in triplicates. In this measurement/diagnostic, the spread cells could be separated from the dead ones in the first run, and then the survived cells could be treated with other active compounds in the second run.

MATERIALS AND METHODS

Synthetic Polymer Solutions. The synthetic copolymers, poly(L-lysine)-*graft*-poly(ethylene glycol) (PLL-*g*-PEG, [PLL(20)-*g*(3.5)-PEG(2)]) (hereafter PP) and its RGD functionalized counterpart, PLL-*g*-PEG/PEG-GGGYGRGDSP (PLL-*g*-PEG-RGD [PLL(20)-*g*(3.5)-PEG(2.3)/PEG(3.4)-RGD]) (hereafter PPR), were obtained as powders from SuSoS AG, Dübendorf, Switzerland. The materials were stored at $-20 \text{ }^\circ\text{C}$ until use. Each powder was then dissolved in 10 mM 4-(2-hydroxyethyl)-1-piperazine ethanesulfonic acid (HEPES, Sigma-Aldrich Chemie GmbH, Schelldorf, Germany) at pH 7.4 to make stock the solutions with a concentration of 1.0 mg/mL. The coating solution with RGD motifs and PLL-*g*-PEG were prepared by mixing the two 1 mg/mL stock solutions (hereafter PP/PPR).^{5,30}

Preparation of EGCG Solutions. The EGCG powder (Sigma-Aldrich Chemie GmbH, Schelldorf, Germany) was solved in 10 mM HEPES at pH 7.4, or in an assay buffer (Hank's Balanced Salt Solution (HBSS, Sigma-Aldrich Chemie GmbH, Schelldorf, Germany) containing 20 mM HEPES, pH 7.0, hereafter HBSS-HEPES). The concentration of the dissolved EGCG solution was 1, 10, 20, 40, 80, 100, 200, and 500 $\mu\text{g}/\text{mL}$, resulted in a final concentration of 0.5, 5, 10, 20, 40, 50, 100, and 250 $\mu\text{g}/\text{mL}$, respectively, in the biosensor wells.

Cell Culture and Cell Adhesion Assay Buffer. HeLa cells (ECACC 93021013 human, cervix, epitheloid, carcinoma) were routinely cultured in tissue culture polystyrene Petri dishes (Greiner Bio-One International GmbH, Kremsmünster, Austria) placed in a humidified incubator ($37 \text{ }^\circ\text{C}$, 5% CO_2). The cells were maintained in Dulbecco's modified Eagle's medium (DMEM), supplemented with 10% fetal bovine serum (FBS, Biowest SAS, France), 4 mM L-glutamine, 0.25 $\mu\text{g}/\text{mL}$ amphotericin B, 100 U/mL penicillin, and 100 $\mu\text{g}/\text{mL}$ streptomycin solution. On reaching 80% confluence, the cells were detached every 3–4 days using 0.05% (w/v) trypsin and 0.02% (w/v) ethylenediaminetetraacetic acid (EDTA) solution and not used beyond passage 20.^{6,38} The cell adhesion assay buffer was prepared by adding 20 mM HEPES to HBSS and adjusted to pH 7.0 with a 1 mM NaOH.^{5,30}

Epic Benchtop Resonant Waveguide Grating Biosensor. We used the Epic BenchTop (BT) system (Corning Incorporated, Corning, NY), a next-generation resonant waveguide grating (RWG) imager biosensor. It accepts 96- or 384-well Society for Biomolecular Screening standard format biosensor microplates. The bottom of the microplate is a planar optical waveguide—a thin, high refractive index, transparent

dielectric layer (waveguide layer, made of the biocompatible material niobium pentoxide) deposited on a thicker glass substratum.^{5,30} At the central position of each well, a 2×2 mm² optical grating is embedded into the optical structure to enable the interrogation of the TM₀ waveguide mode using a near-infrared electromagnetic radiation. Thus, individually addressable biosensors are created in each well of the microplate. All wells of an Epic microplate are simultaneously interrogated every 3 s by sweeping the illuminating wavelength through a range of 15 000 pm with 0.25 pm precision (in the range of 825–840 nm).^{5,30} Waveguide mode excitation only happens at a certain wavelength, called the resonant wavelength (λ). The refractive index variation in an approximately 150 nm thick layer (in the probing depth of the so-called evanescent wave) over the biosensor surface shifts the resonant wavelength to λ' . The refractive index variation can be caused by, for example, bulk refractive index change, molecular adsorption, cell spreading, or dynamic mass redistribution inside the cells. The primary signal output of the Epic BT system is the shift in the resonant wavelength ($\Delta\lambda = \lambda' - \lambda$) in each well.^{5,30}

EGCG Pretreatment of Cancer Cells and the Biosensor Measurement. Wells were given 30 μ L assay buffer to prewet the sensors and establish a baseline with the RWG imager. Following the stabilization of the biosensor signal, the measurement was stopped and the buffer was replaced with 30 μ L of the coating solution and incubated for 0.5 h while gently shaking at room temperature.³⁰ The coating solutions were then removed and the wells were rinsed three times with 30 μ L of the assay buffer, the polymers adsorbed to the surface irreversibly. Wells were then dosed with 30 μ L assay buffer for the fourth time to establish a new baseline for the subsequent room temperature cell spreading assay.³⁰ HeLa cells were trypsinized with $1\times$ prewarmed trypsin–EDTA. Trypsin was removed before complete detachment of HeLa cells and its activity arrested by adding culture medium containing 10% FBS.³⁰ Harvested cells were centrifuged at 380g for 6 min and the cell pellet was resuspended in assay buffer with intensive pipetting. Cell suspensions and EGCG solutions (in different concentrations) were pipetted in 1:1 ratio into 9 eppendorf tubes (with a total volume of 340 μ L in each tube), and they were incubated together for 3 min. Later, we centrifuged them again for 6 min at 380g. We removed the supernatant from the cells, and the assay buffer was added to the cells. The 20 000 treated cells/well were pipetted into the biosensor wells, without replacing the plate from the Epic BT. Spreading was monitored until saturation of the biosensor signals. Averaging every 5 subsequent data points, the effective sampling rate was $1/15$ s⁻¹.^{30,30} At the end of the biosensor measurement, the microplate was placed under a Zeiss Observer microscope to visually observe and image the cells with a $20\times$ objective (all wells were completely filled up with assay buffer and covered with a microscope slide to improve imaging quality by eliminating the disturbing meniscus).³⁰ All experiments were done in triplicates in three different wells at room temperature, and only the averaged data were used for further processing.^{5,30}

MTT Cell Viability Assay. HeLa cells were plated into a 96-well plate with an initial cell number of 12 500 per well. After 24 h incubation at 37 °C, the cells were treated with the compound EGCG in 200 μ L serum-free medium or in 200 μ L HBSS–HEPES buffer. The cells were incubated with the compounds at 0.064 to 500 or 1000 μ g/mL concentration range for 2 h. The control cells were treated with serum-free medium only or with HBSS–HEPES buffer at 37 °C for 2 h.

After washing the cells three times with serum-free medium or HBSS–HEPES buffer, the viability was determined by MTT.²⁵ A 45 μ L MTT solution (2 mg/mL) was added to each well.⁴² After 3 h of incubation, the cells were centrifuged for 5 min (380g) and supernatant was removed. The obtained formazan crystals were dissolved in dimethyl sulfoxide and optical density (OD) of the samples was measured at $\lambda = 540$ and 620 nm using enzyme-linked immunosorbent reader (iEMS Reader, Labsystems, Finland). The OD₆₂₀ values were subtracted from the OD₅₄₀ values. In the case of cytostatic effect determination, the cells were cultivated for further 72 h in DMEM medium at 37 °C after the washing steps. After 72 h, the viability was determined by the MTT.

The percent of cytotoxicity and cytostatic effect was calculated using the following equation

$$\begin{aligned} & \text{cytotoxicity/cytostasis \%} \\ & = [1 - (\text{OD}_{\text{treated}}/\text{OD}_{\text{control}})] \times 100 \end{aligned} \quad (2)$$

where OD_{treated} and OD_{control} correspond to the optical densities of the treated and the control cells, respectively. In each case, two independent experiments were carried out with four parallel measurements. The 50% inhibitory concentration (IC₅₀) values were determined from the dose–response curves, which were defined using a MicrocalTM Origin1 (version 8.6) software.

Flow Cytometry. Before cell plating, the plates were coated with PP/PPR 1:1 v/v (total volume of 10 μ L, shaking for 3 min, after washing with 200 μ L of HBSS buffer three times).

HeLa cells (1×10^5 cells per well) were plated in 24-well plates. The cells were stimulated with the medium alone, or with EGCG (0.0039–1 mg/mL) solutions for 1.5 h. After treatment, the supernatants were removed and the cells were washed with serum-free DMEM media and then the adherent cells were treated with 100 μ L trypsin solution (Sigma, 2.5 g/L porcine trypsin solution in 0.9% sodium chloride), trypsin digestion was blocked using HPMI buffer (20 mM HEPES, 132 mM NaCl, 3.5 mM KCl, 0.5 mM MgCl₂, 5 mM glucose, 1 mM CaCl₂ [pH 7.4])⁴³ supplemented with 10% (wt/vol) fetal bovine serum (FBS, Sigma) and the cells were collected by centrifugation at 300g for 5 min and then washed twice with the serum-free HPMI buffer. The pellets were resuspended in the HPMI buffer.

The cell viability was assessed using 10 μ g/mL PI solution (for the gating strategy, fluorescence parameters were gated using unstained and the PI-stained untreated control cells, and 10 000 cells were counted for each treatment). PI was excited at 488 nm, using a BD LSR II flow cytometer (BD Biosciences, San Jose, CA) Coherent Sapphire, 22 mW laser. After staining, the intracellular fluorescence intensity of the cells was measured on channel PE LP550 and the data were analyzed with a FACSDiva 5.0 software. All measurements were performed in triplicates.

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Author Contributions

R.H. established the research line and supervised the current work. B.P. conducted all biosensor experiments. R.U.-S. and

A.G.N. helped in the experiments. I.S. and S.B. maintained the cell culture. B.S. took part in biology-related discussions. S.B. conducted the MTT and flow cytometry experiments. The methodology to quantify the kinetic curves was developed by R.H. B.P. and R.H. analyzed the data. B.P. prepared the manuscript figures and tables. The manuscript text was written by B.P., S.B., and R.H. All authors reviewed and commented on the manuscript.

Notes

The authors declare no competing financial interest.

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