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## The PYHIN protein p205 regulates the inflammasome by controlling *Asc* expression

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### Abstract

Members of the interferon-inducible PYHIN protein family such as Absent in Melanoma-2 and Interferon Gamma Inducible Protein (IFI) 16 bind double-stranded DNA (dsDNA) and form caspase-1 activating inflammasomes, important in immunity to cytosolic bacteria, DNA viruses or Human Immunodeficiency Virus. IFI16 has also been shown to regulate transcription of type I Interferons during Herpes Simplex Virus infection. The role of other members of the PYHIN protein family in the regulation of immune responses is much less clear. Here, we identified an immune regulatory function for a member of the murine PYHIN protein family, p205 (also called Ifi205). Examination of immune responses induced by dsDNA and other microbial ligands in bone marrow derived macrophages lacking p205 revealed that inflammasome activation by dsDNA as well as ligands that engage the NLRP3 inflammasome was severely compromised in these cells. Further analysis revealed that p205 knockdown cells showed reduced expression of *Asc* at the protein and RNA level. p205 knockdown resulted in reduced binding of actively transcribing RNA Polymerase II to the endogenous *Asc* gene resulting in decreased transcription and processing of *Asc* pre-mRNA. Deletion of p205 in B16 melanoma cells using CRISPR/Cas9 showed similar loss of *Asc* expression. Ectopic expression of p205 induced expression of an *Asc* promoter-luciferase reporter gene. Together these findings suggest that p205 controls expression of *Asc* mRNA to regulate inflammasome responses. These findings expand on our understanding of immune regulatory roles for the PYHIN protein family.

### Introduction

Foreign nucleic acids play a critical role in initiating innate and adaptive immune responses. Nucleic acid (NA) sensors expressed in distinct cellular compartments survey the extracellular and intracellular environment for signs of infection and initiate immune

defenses against bacterial, viral and eukaryotic pathogens (1). NA sensors include RNA Sensors such as TLR3, 7/8 and RIG-I/MDA5 (2–4) as well as DNA sensors such as TLR9, cGAS, AIM2 and IFI16 (5–8). In addition, recognition of foreign nucleic acids especially dsDNA leads to assembly of an inflammasome, a large caspase-1 activating multiprotein complex that controls the proteolytic processing and release of IL-1 $\beta$  (9). Inflammasome activation also results in a proinflammatory form of cell death called pyroptosis (10). While most inflammasomes are composed of members of the NLR family, the dsDNA-activated inflammasome is formed following dsDNA binding to a PYHIN protein, Absent in Melanoma-2 (AIM2). Work from several labs including our own has defined AIM2 as a cytosolic DNA binding innate immune receptor (7, 11–13). AIM2 binds pathogen-derived dsDNA that accumulates in the cytosol during infection with DNA viruses or cytosolic bacterial pathogens (14, 15). In some instances, AIM2 can also recognize host dsDNA that gains access to the cytosol leading to autoinflammation (16). The related PYHIN protein IFI16 also forms an inflammasome during infection with Kaposi's Sarcoma Herpes Virus (KSHV) and Human Immunodeficiency Virus 1 (HIV1) (17, 18).

The PYHIN proteins were first characterized as a family of interferon (IFN) inducible proteins that are predominantly nuclear localized (19). PYHINs are constitutively expressed in different hematopoietic cell types, although most members of this family are also inducible by type I IFN in non-hematopoietic cells (19). Phylogenetic analysis of the mammalian PYHIN proteins also called the AIM2-like receptors or ALRs show strong evolutionary and functional diversity (20, 21). The murine PYHIN locus has undergone extensive gene duplication with more than 13 members encoded in the murine genome in contrast to the human gene locus with 5 genes including MNDA, PYHIN1, POP3, IFI16 and AIM2 (19, 22). PYHIN proteins have been implicated in a wide-range of cellular processes including transcription, tumor suppression, cell cycle, cell growth, differentiation and cell death (19). The majority of the PYHIN proteins share the same structural domains. They contain an N-terminal  $\alpha$ -helical domain known as the Pysin domain capable of homotypic protein-protein interactions and one or more HIN200 domains in their C-terminus, except murine p208 and human POP3 that contain only a Pysin domain and murine p202 that contains only two HIN200 domains (19, 20). Most PYHIN proteins contain a nuclear localization signal in their N-terminus that can be either monopartite, bipartite or both. Some family members also contain a nuclear export sequence that enables them to shuttle between the nucleus and the cytosol. Aim2 is the most conserved family member. Unlike the other PYHINs, AIM2 is localized in the cytosol. Phylogenetic analysis indicates that, with the exception of AIM2, there is a complete lack of orthology among mammalian ALRs (20, 21).

While the role of AIM2 and IFI16 in dsDNA recognition and immune activation has been well established, the role of other members of the PYHIN protein family, especially those in the mouse remains unclear. Recently, genetic studies in mice lacking the entire PYHIN locus and analysis of type I IFN induction following dsDNA treatment in cells from these animals demonstrated no clear link to dsDNA recognition and induction of type I IFN in murine myeloid cells (23). It remains to be defined therefore whether the PYHINs have other immune regulatory functions. In this study we examined the contribution of the murine PYHIN protein p205 (also known as Ifi205) in innate immunity. p205 is primarily expressed in macrophages and granulocytes (24). p205 shares the highest homology with p204/Ifi204

and the human PYHIN protein, MNDA (25). p205 has been implicated as a positive regulator of cell growth, and differentiation during hematopoiesis, adipogenesis and osteogenesis (26–28). To evaluate a possible immune function for p205, we investigated the ability of p205 to control immune gene expression. Using a series of loss of function and gain of function approaches combined with functional studies we demonstrate that p205 controls expression of the inflammasome adapter molecule Asc and as such regulates inflammasome dependent processing of IL-1 $\beta$ . This effect of p205 was not related to the prior work linking AIM2 and IFI16 to sensing of foreign DNA. Rather we find that p205 works in the nucleus to control Asc gene expression. These findings add to our understanding of PYHIN proteins in innate immunity, expanding their functions beyond dsDNA sensing to innate immune gene regulation.

## Material and Methods

### Reagents and Plasmids

Lipopolysaccharide (LPS) and poly-dAdT (pdAdT) were obtained from Sigma-Aldrich and Immunostimulatory DNA oligonucleotides were synthesized as described (8). Nigericin and ATP were from Invivogen and Sigma respectively. Polyinosinic-polycytidylic acid (poly I:C) was obtained from Invivogen. Sendai virus (Cantrell strain) was purchased from Charles River Laboratories (Wilmington, MA). Lipofectamine 2000® Transfection Reagent was from Invitrogen. GeneJuice was from Novagen (Madison, WI). Universal type I IFN and IFN- $\gamma$  were from PBL Interferon Source (Piscataway, NJ) and PeproTech (315-05), respectively. *S. typhimurium* (SL1344 lab strain) was from M. O’Riordan. The plasmids used were p65-pCMV4, c/EBP $\beta$ -pcDNA (Addgene), pGL3-enhancer luciferase reporter (Promega). Other plasmids such as Asc in pMSCVneo (Clontech), p205-HA in pRZ-retro, Aim2-FLAG, p204-HA, p205-HA in pEF-BOS or HA-tagged HIN-p205, PYD-p205 and H/ P-p205 in pMSCV-PIG (Addgene) were made in the lab.

### Cell culture, Stimulations, ELISA and Cell death assays

Primary bone-marrow derived macrophages (BMDM) from C57BL/6J mice, cultured with L929 supernatant as a source for MCSF, were transformed using CreJ2 virus to make immortalized BMDM (iBMDM). The transformation process does not change their ability to induce type I IFNs or other immune genes. The cells were cultured in DMEM with 10% FCS and PenStrep. Antibiotics for selection were used as required. Cells were primed with repurified LPS at 100 ng/ml for 2–3h and stimulated with *Salmonella* sp., Nigericin or ATP for 1h, pdAdT and Interferon Stimulatory DNA (ISD) for 6h, poly I:C and Sendai virus for overnight. Cells infected with *Salmonella typhimurium* and media containing gentamicin (100  $\mu$ g/ml) was added to kill the extracellular bacteria. Supernatants from the stimulated cells were analyzed for the cytokines, IL-1 $\beta$  (eBiosciences) and IFN- $\beta$  by ELISA. Cell death was measured by quantitating the amount of LDH, which is released into the supernatant upon cell lysis, using CytoTox96 Non Radio Cytotoxicity Assay (Promega) kit. 10% Triton-X was added to the cells as a representation of 100% cell death.

### shRNA mediated silencing

The shRNA sequences targeting p205 were cloned into a lentiviral pLKO.1 TRC cloning vector. Two separate shRNA sequences for p205 used either targeted the coding region (KD CDS) (TRCN0000095887) or the 3' untranslated region (KD 3'UTR) (TRCN0000095884). HEK 293T cells were transfected with 4 µg shRNA along with 3 µg pSPAX (gag/pol) and 1 µg pMD2 (VSV-G) plasmids for production of lentiviral particles. Viral particles were collected at 48 h, filtered and added to immortalized BMDM. As controls, BMDM were transduced with either an empty pLKO.1 vector (EV) or pLKO.1 containing an shRNA sequence targeting GFP (GFP shRNA). The cells were selected for effective transduction by selection with puromycin (5 µg/ml).

### qRT-PCR and Nanostring

RNA was extracted using Qiagen RNeasy Kit. cDNA was synthesized from 1 µg total RNA using either iScript cDNA synthesis kit (Bio-Rad) or QuantiTect Reverse Transcription Kit (Qiagen). Quantitative RT-PCR was performed using iQ SYBR Green supermix (Bio-Rad) or QuantiNova SYBR Green PCR Kit (Qiagen). Primers were constructed to respective genes (Supplemental Table 1). Target genes expressions are relative to housekeeping genes expression and were normalized to respective controls. The levels of p204, p205, Mnda, Mndal and Aim2 were measured using nCounter (Nanostring). Briefly, endogenous RNA transcript counting was performed on total RNA hybridized to a custom gene expression CodeSet and analyzed on an nCounter Digital Analyzer (29). The counts were normalized to internal spike-in and endogenous controls per Nanostring Technologies' specifications. The heatmap was generated using the Morpheus software.

### Western Blot and Co-immunoprecipitation

For detecting caspase-1 and IL-1 $\beta$  in the supernatants and lysates, 20% vol RIPA buffer and 30% vol SDS-loading dye were added directly to the wells containing cells and media. The samples were boiled at 100°C for 15'–30' and were run on 13% polyacrylamide gels. Cytosolic and nuclear fractions for protein detection were prepared by using Active Motif Nuclear Extract Kit. Co-immunoprecipitation assays were carried out with  $5 \times 10^6$  cells (treated or untreated) in ice-cold RIPA buffer (without SDS and DOC) using Protein A Dynabeads (Novex/Life Technologies, Cat# 10001D) conjugated with specific antibodies, and immunoblotted for proteins of interest. Antibodies used were against Asc (Santa Cruz, Cat# sc-22514-R or Cell Signaling, Cat# 67824), IL-1 $\beta$  (R&D Systems, Cat# AF-401-NA), caspase-1 (Adipogen, Cat# AG-20B-0042), Aim2 (eBioscience, 14-6008), Nlrp3 (Enzo Life Sciences), Histone 3 (Abcam, Cat# ab1791), c/EBP $\beta$  (Santa Cruz, Cat# sc-150), Usf2 (Santa Cruz, Cat# sc-862), Gapdh (Sigma, Cat# G9295),  $\beta$ -actin (Sigma, Cat# A3854) HA-tag (Anti-HA-Peroxidase; Roche, Cat# 12 013 819 001), and FLAG-tag (Sigma, Cat# A8592). An affinity purified polyclonal antibody against p205 was generated using the following peptide: AGLDRLINFCERVPTL-amide) was generated (21<sup>st</sup> Century Biochemicals).

### Confocal Microscopy

HEK 293T cells were transfected with p205 tagged with CFP and 24h post-transfection, the cells were washed with PBS and stained with Acridine Orange and/or Cholera toxin-B (CtxB) (ThermoFisher) and were visualized by confocal microscopy (Leica 8000) for localization.

### Luciferase assays

The *Asc* gene promoter from -2000 +10 bp was cloned into pGL3-Enhancer reporter vector upstream of the firefly luciferase gene. The promoter-reporter construct was either transfected alone or co-transfected with plasmids expressing p205, p204, Aim2, p205 deletion mutants, p65/RelA or c/EBP $\beta$  or a combination thereof in HEK 293T cells. A plasmid expressing Renilla luciferase gene under thymidine kinase (pGL4-TK Renilla) promoter was included as transfection efficiency control. Data is represented as fold change over *Asc* reporter construct alone, and relative to transfection efficiency.

### Chromatin Immunoprecipitation

Cells were fixed with 1% formaldehyde, lysed and sheared. The DNA was quantified, and 5  $\mu$ g of total chromatin was immunoprecipitated with specific antibodies and Dynabeads Protein G (Novex/Life Technologies, Cat#10009D). The DNA was then reverse crosslinked, purified and quantitated by qPCR amplification with primers at *Asc*, *Gapdh* genes (Supplemental Table 1). Antibodies used were against total RNA Polymerase II (RNAPII; Active Motif Cat# 39097), phospho Serine-2 RNA Polymerase II (Ser2 RNAPII; Abcam Cat# ab5095), phospho Serine-5 RNA Polymerase II (Ser5 RNAPII; Abcam Cat# ab5131) or IgG isotype (Abcam, Cat# AB37415 and Cell Signaling, 5415).

### CRISPR/Cas9-mediated gene knockout

B16 mouse melanoma cells were cultured in DMEM containing 10% FBS, 0.5% Ciprofloxacin and 0.0075 %  $\beta$ -mercaptoethanol and transfected with 200 ng of plasmid containing mCherry-Cas9 and a U6 promoter-driven gRNA against p205 (Target Sequence: ATGAAGCCGAAGATGAGACCTGG) using Lipofectamine 2000<sup>®</sup> according to manufacturer's protocol (30). Two days after transfection cells were sorted for mCherry expression. Positive cells were plated at limiting dilution to obtain single cell clones. Genotyping of the B16 clones was conducted by deep sequencing (Illumina, MiSeq) as previously described (31) using the following primer sequences: 5' - ACACTCTTTCCCTACACGACGctctccgatctCGTGAAGAAGATCAAGGCATCTG-3' and 5' - TGACTGGAGTTCAGACGTGTGctctccgatctAAATCTCAGGGAGAAGTGGGGGA-3' (uppercase letters: 1<sup>st</sup> PCR adapter sequences, lowercase letters: linker sequences, uppercase letters: target site specific primer sequences). Cell clones harboring all-allelic frame shift mutations were then selected as p205 KO cell clones.

### Statistics

ELISA and luciferase assays are presented as mean  $\pm$  SD from three independent biological replicates and are representative of at least three separate experiments. Data was analyzed

using two-way ANOVA by Prism 6 Software (GraphPad, San Diego, CA). The  $p$  values  $< 0.05$  were considered significant (\* $p < 0.05$ , \*\* $p < 0.001$ , \*\*\* $p < 0.005$ , \*\*\*\* $p < 0.0001$ ) and n.s. = non-significant, unless otherwise indicated.

## Results

### Lipopolysaccharide (LPS) and Interferon (IFN)-inducible p205 localizes to the nucleus in resting macrophages

The *p205* gene is encoded on chromosome 1q, the last of the 13 consecutive genes of the mouse PYHIN locus. *p205* is encoded on the reverse strand and flanked by *p202b* at the 5'-end and several olfactory receptor genes as well as the *Spta1* gene at its 3'-end. *p205* is expressed in murine primary bone marrow derived macrophages and treatment of these cells with LPS, type-I IFN and IFN- $\gamma$  treatments further unregulated its expression, similar to other PYHIN genes such as *Mnda* and *p204* (Figure 1A, B and C). The inducible expression of *p205* was as robust as the well-characterized immune genes such as *Il6* or *Cxcl10* that are used as positive controls.

To evaluate the localization of *p205* in macrophages, we overexpressed hemagglutinin (HA) tagged *p205* into immortalized BMDM and performed subcellular fractionation to purify nuclear and cytosolic extracts. Analysis of these fractions by immunoblotting for HA tagged *p205* in two independently transduced BMDM showed that *p205* is primarily localized in the nucleus (Figure 1D). We also confirmed the nuclear localization of *p205* using confocal microscopy by imaging a CFP-tagged *p205* construct expressed in HEK 293T cells (Figure 1E). Treatment of primary BMDM with LPS, type-I IFN and IFN- $\gamma$  increased *p205* protein expression in the nucleus (Figure 1F). Further, we stimulated primary BMDM with type-I IFN at different time points and separated the cell lysates into nuclear and cytosolic fractions, which were immunoblotted for endogenous *p205*. Upon IFN-treatment, *p205* expression was robustly increased over time in the nucleus (Figure 1G).

### Knockdown of p205 in bone marrow derived macrophages results in impaired inflammasome activation

To determine the potential immune function of *p205*, we generated immortalized murine BMDM expressing shRNA-targeting *p205*. BMDM were transduced with pLKO.1 lentiviral particles either containing short hairpin RNA (shRNA) targeting *p205* or GFP as a control. Two shRNA sequences for *p205* were used which targeted the 3' untranslated region (KD 3'UTR) (Figure 2A) and the coding region (KD CDS) (Figure 2B) of the mRNA, respectively. The level of *p205* was evaluated by quantitative PCR (qPCR). The shRNA targeting the coding region specifically knocked down *p205* mRNA with no apparent effect on the two most closely related murine PYHINs, murine *Mnda* or *p204* mRNA. This effect was also evaluated using Nanostring where *p205* mRNA was reduced with little impact on 4 other related PYHIN genes (*p204*, *Mnda*, *Mnda1* and *Aim2*) (Figure 2C).

Both of these macrophage cell lines with *p205* knockdown were then tested for their ability to respond to different immune stimuli and induce cytokine gene expression. We first evaluated the inducible expression of type-I IFN since prior work had identified other

PYHIN proteins such as IFI16 and p204 as regulators of type I IFN gene expression. iBMDM expressing shRNA targeting p205 or GFP were stimulated with LPS, pdAdT, Interferon Stimulatory DNA (ISD), Sendai virus (SV) or poly I:C, all of which induced the type I IFN, IFN $\beta$ , the levels of which were measured by ELISA. For most of these ligands the inducible expression of IFN $\beta$  was largely unaffected by the absence of p205 (Figure 2D). However, stimulation of the p205 knockdown macrophages with pdAdT, a synthetic dsDNA mimetic showed an enhanced IFN- $\beta$  response compared to the control cells. p205 knockdown cells were also tested for IL-1 $\beta$  production since PYHINs such as AIM2 and IFI16 form inflammasome complexes that regulate IL1 $\beta$  production. The cell lines were treated for two hours with 200 ng/mL of LPS which acts as Signal 1 to prime cells allowing transcriptional induction of pro-IL1 $\beta$ . These cells were then challenged with transfected pdAdT or ISD or Nigericin, which activate the Aim2 and Nlrp3 inflammasomes, respectively. Macrophages lacking p205 had reduced levels of IL-1 $\beta$  in response to all of these stimuli (Figure 2E).

To understand whether the diminished levels of p205 impacted on either the priming (signal 1) or processing of pro-IL-1 $\beta$  (signal 2), the supernatants and lysates from the stimulated macrophages were tested for levels of both the inactive and proteolytically processed forms of both caspase-1 and IL-1 $\beta$ . Cells lacking p205 expressed similar levels of pro-IL-1 $\beta$  following stimulation with LPS, ruling out an effect of p205 on TLR4 signaling and transcription of pro-IL1 $\beta$  (Figure 2F top panel). In contrast the levels of mature processed IL-1 $\beta$  were significantly reduced in the knockdown cells (Figure 2F middle upper panel). Moreover, the activated form of caspase-1 was also undetectable in cells with shRNA knockdown for p205 (Figure 2F, bottom panel), despite comparable amounts of pro-caspase-1 between the control and the knockdown cell lines (Figure 2F middle lower panel). Collectively these observations indicate that knockdown of p205 impacted inflammasome dependent activation of caspase-1 leading to reduced processing of pro-IL1 $\beta$ .

### Reduced expression of p205 leads to reduced expression of Asc

To understand the mechanistic basis for the defect in inflammasome activation in cells expressing shRNA-p205, we first measured the expression of Nlrp3 and Aim2 as well as the adapter protein Asc (Apoptosis-associated speck-like molecule containing CARD domain) in these LPS-stimulated macrophages. While there was no difference in the protein levels of either Nlrp3 or Aim2, expression of the Asc protein was completely absent in the p205 knockdown cell lines (Figure 3A). We further confirmed the loss of p205 protein in these cell lines using a newly generated antibody generated against p205, and the LPS-stimulated macrophages lacking p205 showed no Asc protein expression (Figure 3B). The expression of Asc was further investigated by measuring its transcript levels. These studies indicated that cells with shRNA knockdown of p205 had decreased levels of Asc mRNA (Figure 3C). Collectively, these findings indicate that p205 regulates Asc mRNA levels. To discern whether the defective inflammasome responses we observed in the p205-shRNA iBMDM was specifically due to the absence of Asc, we restored Asc expression in these cells using retroviral transduction. The Asc cDNA was cloned into a retroviral vector, pMSCV and viral particles were transduced in p205 knockdown macrophages to renew Asc expression. p205-shRNA cells transduced with the retrovirus had restored Asc levels comparable to that seen

in control cell lines (Figure 3D). The reconstituted stable cell lines were then tested for IL-1 $\beta$  release by ELISA. IL-1 $\beta$  release by the macrophages following LPS priming and pdAdT or Nigercin treatment, was fully restored upon re-expression of Asc in these cells (Figure 3E). *Salmonella typhimurium* also activates the inflammasome. When *Salmonella* is in its log phase of growth i.e. the bacteria are actively dividing they are recognized by Nlr4, while when in their non-dividing or stationary growth phase, *Salmonella* is primarily recognized by Nlrp3. Unlike Nlrp3 or Aim2, Nlr4 itself contains a CARD domain, and can directly recruit pro-caspase 1 as well as engage Asc to do so. Thus, Nlr4 can activate the inflammasome in a manner that is only partially dependent on Asc (32). The p205 knockdown cells were challenged with *Salmonella* in their stationary phase and log phase after LPS stimulation, for one hour. *Salmonella* in the stationary phase (overnight culture) that primarily engages Nlrp3 to activate the inflammasome showed minimal IL-1 $\beta$  production by ELISA. But when the same cells were infected with the actively dividing *Salmonella* (4–6hr culture), the cells retained part of their inflammasome activating capability that was again rescued to its full potential upon reconstitution with Asc (Figure 3F). Further, stimulation with LPS and pdAdT in the Asc-reconstituted macrophages showed processing of caspase 1 and IL-1 $\beta$  into their active forms signifying a functional inflammasome (Figure 3G).

Inflammasome activation also leads to pyroptotic cell death (10), which can be monitored by examining the release of lactate dehydrogenase enzyme (LDH). Poly dAdT treatment of macrophages leads to Aim2 inflammasome dependent release of LDH. Consistent with the other effects on inflammasome function, the release of LDH was also significantly affected in cells with p205 knockdown. Restoration of Asc in these knockdown cell lines rendered these cells susceptible to cell death similar to the levels seen in the control cell line (Figure 3H). Altogether, these findings attribute the loss of inflammasome responses in p205-knockdown cells directly to p205-dependent effects on Asc expression.

### **Reconstitution of p205 expression in macrophages restores Asc expression and inflammasome activation**

We next restored expression of p205 in the knockdown cells using a HA-tagged retroviral vector. The levels of p205 gene expression were measured by qPCR (Figure 4A). The reconstitution of p205 restored Asc mRNA expression (Figure 4B). Moreover, reconstitution with p205 restored Asc protein expression in these knockdown cell lines (Figure 4C). p205 protein expression was detected using an anti-HA antibody. Cell lysates from GFP shRNA, p205 KD 3'UTR cell lines along with the p205 KD 3'UTR cell line reconstituted with p205-HA were separated into nuclear and cytoplasmic fractions. The reconstituted p205 was found to be primarily nuclear. Levels of Asc in both the nucleus and cytoplasm increased in cells expressing the p205-HA. Levels of Aim2 were largely unchanged upon p205 reconstitution. We also ectopically expressed the HA-tagged p205 in wild-type immortalized BMDM, which resulted in increased Asc mRNA levels compared to that found in control cell lines (Figure 4D). Analysis of IL-1 $\beta$  release and cell death in these cells further demonstrated that reconstitution of p205 in the knockdown cells restored inflammasome dependent responses (Figure 4E and F). Altogether, these findings provide further evidence for a role for p205 as a regulator of Asc expression and inflammasome responses.



## RNA polymerase II occupancy at the *Asc* locus is reduced in macrophages expressing p205-shRNA

The reduced mRNA levels of *Asc* in cells lacking p205 could be due to either effects on *Asc* gene transcription or *Asc* mRNA stability. To further explore how p205 impacted *Asc* mRNA levels we performed chromatin immunoprecipitation assays to explore the binding of RNA Polymerase II (RNA Pol II) to the endogenous *Asc* promoter in both the GFP shRNA and p205 KD BMDM. Figure 5A shows a schematic of the *Asc* gene locus and the location of the primers used to evaluate RNA Pol II binding. Using antibodies to RNA Pol II and an isotype control IgG, we evaluated RNA Pol II binding to the endogenous *Asc* promoter. RNA Pol II binding was considerably lower in the p205 KD cell lines using a series of primers as indicated (Figure 5B). We also expanded this analysis using antibodies to RNA Pol II phosphorylated either at position Serine 5 (Serine-5P RNA Pol II) or at Serine 2 (Serine-2P RNA Pol II) in its C-terminal domain. The RNA Pol II carboxy-terminal domain (CTD) contains multiple repeats of a heptapeptide sequence, YSPTSPS that is differentially phosphorylated to control the function of the polymerase (33). When preferentially phosphorylated at Serine 5 alone, there is only abortive transcription where the RNA Pol II can initiate, but not elongate or process functional mRNA (33). The change from an initiating to an actively elongating RNA Pol II occurs with the increase in phosphorylation of the Serine 2 residue on the CTD repeats (33). Consistent with the findings with antibodies to total RNA Pol II, both Serine-5P RNA Pol II (Figure 5C) and Serine-2P RNA Pol II (Figure 5D) distributions on the *Asc* gene were considerably lower in the p205-shRNA cells compared to the GFP shRNA BMDM. The recruitment of total RNA Pol II, Serine-5P RNA Pol II and Serine-2P RNA Pol II to the housekeeping gene, *Gapdh*, was equivalent between the GFP shRNA and p205 KD macrophages (Figure 5E).

Although p205 KD BMDM showed reduced levels of RNA Pol II recruited at the endogenous loci, the levels of *Asc* mRNA appeared to be affected more severely than the ChIP data indicated. Recent studies have shown that RNA Pol II is also a major player in successful mRNA processing and splicing. Transcription and splicing are dependent on each other- a phenomenon termed co-transcriptional splicing (34). Hence, to understand if p205 was regulating both transcription as well as mRNA processing of the *Asc* transcript, we measured the levels of *Asc* pre-mRNA as well as that of the processed and mature mRNA in cells with and without p205. By designing primers that specifically measure pre-mRNA and mature mRNA (Supplemental Table 1), we found that p205 KD BMDM had very low levels of the *Asc* pre mRNA and little mature *Asc* mRNA being made (Figure 5F). We conclude from this data that p205 controls *Asc* expression both at the level of gene transcription as well as processing of mRNA.

## p205 drives expression from an *Asc* promoter-reporter construct in a dose-dependent manner

To gain further insight into the mechanisms involved in the transcription of *Asc*, we cloned the *Asc* promoter (-2000 to +10 *Asc* gene) as a reporter gene upstream of a firefly luciferase gene. This promoter-reporter construct was transfected into HEK 293T cells together with increasing concentrations of p205. Ectopic expression of p205 led to a dose dependent increase in the *Asc*-luciferase reporter gene. This effect was specific to p205, as transfection

of two related PYHIN proteins p204 and Aim2 had no effect (Figure 6A). We also determined the role of specific domains of p205 on reporter gene activity by generating and testing deletion mutants of p205 that either lacked the pyrin domain (PYD), the HIN domain (HIN) or both (H/P). p205 mutants lacking either the HIN or PYD domain failed to induce expression from the *Asc* promoter (Figure 6B) indicating that both HIN and PYD domains were important.

Analysis of the *Asc* promoter for binding sites for transcription factors revealed putative binding sites for c/EBP $\beta$  and NF- $\kappa$ B amongst others. Previously, Parsons et al. had shown that TNF $\alpha$  treatment could enhance *Asc* expression through p65/RelA in MCF7 cell lines (35). Furthermore, p205 has been shown to co-localize and bind directly with the CCAAT/enhancer binding protein- $\beta$  (c/EBP $\beta$ ) in mouse adipose-derived stem cells (27). Furthermore, c/EBP $\beta$  has also been implicated in the induction of several proinflammatory genes in macrophages (36). Therefore we tested the effect of ectopic expression of c/EBP $\beta$  or p65/RelA on *Asc* reporter gene expression. In both cases these transcription factors increased *Asc* luciferase reporter gene expression in a dose dependent manner (Figure 6C, left panel). In addition, co-expressing p205 together with c/EBP $\beta$  or p65/RelA, further enhanced *Asc* reporter gene expression (Figure 6C, middle panel). Transfection of c/EBP $\beta$  and p65/RelA together with increasing concentrations of p205 further synergized to drive more robust expression from the *Asc* gene reporter (Figure 6C, right panel). Furthermore, transiently transfected HA-tagged p205 was found to interact with overexpressed c/EBP $\beta$  in HEK 293T cells (Figure 6D) as well as endogenously in both resting and LPS-stimulated (200ng/ml for 3h or 6h) wild-type macrophages (Figure 6E) by co-immunoprecipitation assays. Together these observations indicate that p205 can increase *Asc* reporter gene activation and can collaborate with c/EBP $\beta$  or p65/RelA to do so.

### Knockout of p205 using CRISPR/Cas9 impairs *Asc* expression

Having defined p205 as a regulator of *Asc* gene expression using shRNA-based knockdown approaches, we also wanted to evaluate the impact of p205 on *Asc* gene expression using an independent strategy and in a different cell-type that expresses p205. We used B16 melanoma cell lines to generate CRISPR/Cas9 mediated knockout of p205. Plasmids expressing Cas9 and the sgRNA targeting specific regions of p205 were transfected into B16 melanoma cells to generate p205 knockout (p205 KO) cells. A clone of these cells was produced and evaluated for p205 deficiency using sequencing, qPCR and Western blot. Levels of p205 mRNA following overnight IFN- $\gamma$  stimulation were undetectable in these cells (Figure 7A). In addition, there was no discernible p205 protein expression in p205 knockout cells stimulated with Sendai virus (Figure 7B). Similar to what we had observed in our macrophage system, we found that cells lacking p205 had a loss of mature *Asc* mRNA and protein (Figure 7C and D). These observations further support our shRNA-based studies and underscore the importance of p205 as a regulator of *Asc* expression.

### Discussion

Studies over the last decade have defined the importance of AIM2, a cytosolic DNA binding protein as a regulator of caspase-1 activity and proteolytic processing of IL-1 $\beta$  and IL18

(37). The role of the PYHINs in sensing dsDNA was further highlighted by the identification of IFI16 as a regulator of type I IFN gene transcription following HSV-1 infection (8).

Early studies had also linked members of the murine PYHIN family to type I IFN gene regulation. Knockdown of mouse Ifi203 and mouse Ifi204 have been shown to dampen the IFN response to infection with multiple pathogens, including HSV-1, human immunodeficiency virus (HIV), murine leukemia virus (MLV), *Francisella tularensis*, and Mycobacterium tuberculosis (8, 38, 39). Despite these studies, a recent genetic study from Gray et al. using mice lacking the entire PYHIN locus, lacking all 13 PYHIN genes, found that there was no change in the IFN signature when challenged with various immunostimulatory DNA ligands, DNA virus infection and lentivirus infection (23). This body of work indicates that the murine PYHIN proteins play limited roles in DNA ligand recognition at least in the myeloid cells and fibroblasts tested in these studies, raising the possibility that the PYHINs have alternative functions in these cells.

Prior to the discovery of AIM2 and IFI16 as sensors of microbial DNA, PYHIN proteins were shown to regulate cell growth, differentiation, tumor suppression and DNA damage responses (25). IFI16, IFIX, p202, and p204 regulate cell cycle transcription factors such as p53, p21, pRb, and E2F resulting in cell cycle arrest (40). p202 acts as a transcriptional repressor targeting NF- $\kappa$ B (41), AP-1 (42, 43), MYOD1 (44), and myogenin (19, 43). p204 also regulates gene expression during monocyte/macrophage differentiation and osteoblast differentiation (45–47).

Here we expand upon these studies and define p205, a murine PYHIN protein as an additional regulator of gene expression in innate immunity. Using a series of loss of function approaches (both shRNA and CRISPR/Cas9 mediated gene editing), we report that p205 regulates expression of the inflammasome adapter protein Asc and in doing so controls inflammasome activation pathways broadly. Cells lacking p205 failed to activate caspase-1 and control inflammasome dependent processing of IL1 $\beta$  in response to multiple ligands that engage the Aim2 inflammasome as well as the Nlrp3 inflammasome. The abrogated inflammasome activation in the p205 KD macrophages upon pdAdT stimulation also likely explained the enhanced IFN $\beta$  levels detected in these cells, since prior work from our lab and others have shown that activation of the Aim2 inflammasome by intracellular DNA antagonizes the type I IFN pathway (14, 48). In cells with defects in inflammasome responses, there is a more robust dsDNA driven induction of type I IFNs.

By carefully measuring expression levels of key components of the Aim2 and Nlrp3 pathways, we found that cells lacking p205 had reduced expression of Asc. This effect was observed at both the protein and mRNA levels. The compromised inflammasome dependent responses observed in these cells could be fully rescued by ectopically expressing either p205 or Asc itself. Cells lacking p205 had reduced RNA Polymerase II binding to the *Asc* gene indicating that p205 functioned in part to control *Asc* gene transcription. In addition, by comparing the levels of the *Asc* pre-mRNA to those of the mature transcript, we could also observe additional effects on *Asc* mRNA processing. It is broadly accepted that the CTD of RNA Pol II is involved in efficient transcription as well as mRNA processing (34), and the Serine-2 phosphorylated polymerase determines the rate of mRNA elongation,

spliceosome assembly and splicing efficiency (49). Thus we deduced from our ChIP experiments and qPCR analysis of *Asc* pre-mRNA and mature transcripts, that the absence of p205 not only affects *Asc* gene transcription but also affected the processing of the immature *Asc* mRNA.

Further, we show that in regulating gene expression, p205 cooperates with both c/EBP $\beta$  and p65/RelA to drive *Asc* expression. The transcription factor c/EBP $\beta$  is important in controlling macrophage differentiation. c/EBP $\beta$  remains fully active in resting macrophages and stays positioned on target genes, ready to stimulate transcription with other inducible transcription factors (36). p205 has previously been shown to interact with c/EBP $\beta$  in adipocytes. p205 interacts with c/EBP $\beta$  in unstimulated macrophages as well, and maintains basal expression of *Asc*, while under stimulated conditions, LPS-inducible p205 may interact with both c/EBP $\beta$  and activated NF- $\kappa$ B transcription factors to further enhance expression.

Previous studies have shown that ASC, originally identified as TMS1 (Target of methylation induced silencing-1) is influenced by DNA methylation. The *ASC* gene contains a 600-bp long CpG island located near the transcription start site and the methylation status of this CpG island correlates with the expression level of ASC/TMS1. The methylation status of the CpG island, as well as promoter proximal pausing of RNA Polymerase II in multiple cancer cell lines and tissues result in reduced expression of ASC (50–58). Our studies indicate that *Asc* levels are influenced by p205 in murine macrophages adding additional understanding to the regulation of this important immune gene. Our findings expand on our understanding of transcriptional regulatory roles for the PYHIN proteins. p205 has previously been shown to control p21<sup>CIP/WAF</sup> gene expression via p53 in Saos2 cell lines and to impact gene expression in adipogenesis and osteogenesis via transcriptional mechanisms. p205 acts as a transactivator synergizing with p53 to induce expression of the cell cycle inhibitor p21 to inhibit cell growth. In adipocytes, p205 interacts with c/EBP $\beta$  and c/EBP $\alpha$  to further activate the transcriptional activity of c/EBP $\alpha$  and PPAR $\gamma$ . All of these studies help establish evidence for PYHIN proteins as regulators of gene expression. The discovery of p205 as a regulator of inflammasomes via transcriptional regulation of *Asc* further supports a role for the PYHIN proteins as regulators of gene expression and in addition emphasizes the importance of PYHIN family members in innate immunity.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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## References

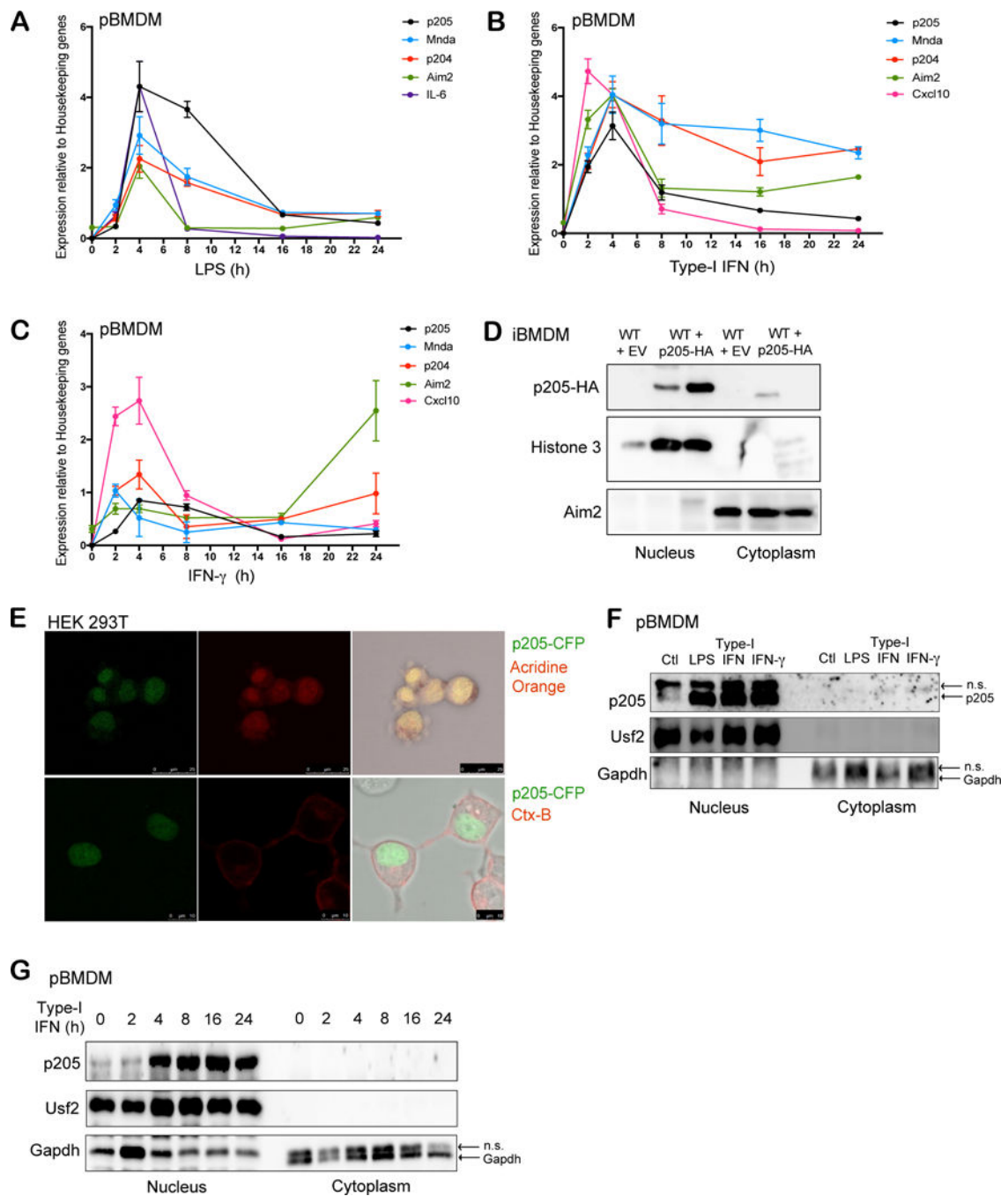
1. Sharma S, Fitzgerald KA. Innate immune sensing of DNA. *PLoS pathogens*. 2011; 7:e1001310. [PubMed: 21533068]
2. Yoneyama M, Kikuchi M, Natsukawa T, Shinobu N, Imaizumi T, Miyagishi M, Taira K, Akira S, Fujita T. The RNA helicase RIG-I has an essential function in double-stranded RNA-induced innate antiviral responses. *Nature immunology*. 2004; 5:730–737. [PubMed: 15208624]
3. Heil F, Hemmi H, Hochrein H, Ampenberger F, Kirschning C, Akira S, Lipford G, Wagner H, Bauer S. Species-specific recognition of single-stranded RNA via toll-like receptor 7 and 8. *Science*. 2004; 303:1526–1529. [PubMed: 14976262]
4. Alexopoulou L, Holt AC, Medzhitov R, Flavell RA. Recognition of double-stranded RNA and activation of NF-kappaB by Toll-like receptor 3. *Nature*. 2001; 413:732–738. [PubMed: 11607032]
5. Hemmi H, Takeuchi O, Kawai T, Kaisho T, Sato S, Sanjo H, Matsumoto M, Hoshino K, Wagner H, Takeda K, Akira S. A Toll-like receptor recognizes bacterial DNA. *Nature*. 2000; 408:740–745. [PubMed: 11130078]
6. Sun L, Wu J, Du F, Chen X, Chen ZJ. Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway. *Science*. 2013; 339:786–791. [PubMed: 23258413]
7. Hornung V, Ablasser A, Charrel-Dennis M, Bauernfeind F, Horvath G, Caffrey DR, Latz E, Fitzgerald KA. AIM2 recognizes cytosolic dsDNA and forms a caspase-1-activating inflammasome with ASC. *Nature*. 2009; 458:514–518. [PubMed: 19158675]
8. Unterholzner L, Keating SE, Baran M, Horan KA, Jensen SB, Sharma S, Sirois CM, Jin T, Latz E, Xiao TS, Fitzgerald KA, Paludan SR, Bowie AG. IFI16 is an innate immune sensor for intracellular DNA. *Nature immunology*. 2010; 11:997–1004. [PubMed: 20890285]
9. Schroder K, Tschopp J. The inflammasomes. *Cell*. 2010; 140:821–832. [PubMed: 20303873]
10. Bergsbaken T, Fink SL, Cookson BT. Pyroptosis: host cell death and inflammation. *Nature reviews Microbiology*. 2009; 7:99–109. [PubMed: 19148178]
11. Burckstummer T, Baumann C, Bluml S, Dixit E, Durnberger G, Jahn H, Planyavsky M, Bilban M, Colinge J, Bennett KL, Superti-Furga G. An orthogonal proteomic-genomic screen identifies AIM2 as a cytoplasmic DNA sensor for the inflammasome. *Nature immunology*. 2009; 10:266–272. [PubMed: 19158679]
12. Fernandes-Alnemri T, Yu JW, Datta P, Wu J, Alnemri ES. AIM2 activates the inflammasome and cell death in response to cytoplasmic DNA. *Nature*. 2009; 458:509–513. [PubMed: 19158676]
13. Roberts TL, Idris A, Dunn JA, Kelly GM, Burnton CM, Hodgson S, Hardy LL, Garceau V, Sweet MJ, Ross IL, Hume DA, Stacey KJ. HIN-200 proteins regulate caspase activation in response to foreign cytoplasmic DNA. *Science*. 2009; 323:1057–1060. [PubMed: 19131592]
14. Rathinam VA, Jiang Z, Waggoner SN, Sharma S, Cole LE, Waggoner L, Vanaja SK, Monks BG, Ganesan S, Latz E, Hornung V, Vogel SN, Szomolanyi-Tsuda E, Fitzgerald KA. The AIM2 inflammasome is essential for host defense against cytosolic bacteria and DNA viruses. *Nature immunology*. 2010; 11:395–402. [PubMed: 20351692]
15. Fernandes-Alnemri T, Yu JW, Juliana C, Solorzano L, Kang S, Wu J, Datta P, McCormick M, Huang L, McDermott E, Eisenlohr L, Landel CP, Alnemri ES. The AIM2 inflammasome is critical for innate immunity to *Francisella tularensis*. *Nature immunology*. 2010; 11:385–393. [PubMed: 20351693]
16. Baum R, Sharma S, Carpenter S, Li QZ, Busto P, Fitzgerald KA, Marshak-Rothstein A, Gravallese EM. Cutting edge: AIM2 and endosomal TLRs differentially regulate arthritis and autoantibody production in DNase II-deficient mice. *Journal of immunology (Baltimore, Md : 1950)*. 2015; 194:873–877.
17. Kerur N, Veettil MV, Sharma-Walia N, Bottero V, Sadagopan S, Otageri P, Chandran B. IFI16 acts as a nuclear pathogen sensor to induce the inflammasome in response to Kaposi Sarcoma-associated herpesvirus infection. *Cell Host Microbe*. 2011; 9:363–375. [PubMed: 21575908]
18. Monroe KM, Yang Z, Johnson JR, Geng X, Doitsh G, Krogan NJ, Greene WC. IFI16 DNA sensor is required for death of lymphoid CD4 T cells abortively infected with HIV. *Science*. 2014; 343:428–432. [PubMed: 24356113]

19. Schattgen SA, Fitzgerald KA. The PYHIN protein family as mediators of host defenses. *Immunological reviews*. 2011; 243:109–118. [PubMed: 21884171]
20. Cridland JA, Curley EZ, Wykes MN, Schroder K, Sweet MJ, Roberts TL, Ragan MA, Kassahn KS, Stacey KJ. The mammalian PYHIN gene family: phylogeny, evolution and expression. *BMC evolutionary biology*. 2012; 12:140. [PubMed: 22871040]
21. Brunette RL, Young JM, Whitley DG, Brodsky IE, Malik HS, Stetson DB. Extensive evolutionary and functional diversity among mammalian AIM2-like receptors. *J Exp Med*. 2012; 209:1969–1983. [PubMed: 23045604]
22. Connolly DJ, Bowie AG. The emerging role of human PYHIN proteins in innate immunity: implications for health and disease. *Biochemical pharmacology*. 2014; 92:405–414. [PubMed: 25199457]
23. Gray EE, Winship D, Snyder JM, Child SJ, Geballe AP, Stetson DB. The AIM2-like Receptors Are Dispensable for the Interferon Response to Intracellular DNA. *Immunity*. 2016; 45:255–266. [PubMed: 27496731]
24. Weiler SR, Gooya JM, Ortiz M, Tsai S, Collins SJ, Keller JR. D3: a gene induced during myeloid cell differentiation of Linlo c-Kit+ Sca-1(+) progenitor cells. *Blood*. 1999; 93:527–536. [PubMed: 9885214]
25. Asefa B, Klarmann KD, Copeland NG, Gilbert DJ, Jenkins NA, Keller JR. The interferon-inducible p200 family of proteins: a perspective on their roles in cell cycle regulation and differentiation. *Blood cells, molecules & diseases*. 2004; 32:155–167.
26. Li H, Jiao Y, Zhang L, Wang C, Zhang X, Guo H, Xu H. The interferon-inducible protein p205 acts as an activator in osteoblast differentiation of mouse BMSCs. *Differentiation*. 2016; 92:318–325. [PubMed: 26965419]
27. Liu F, Jiao Y, Zhu Z, Sun C, Li H. Interferon-inducible protein 205 (p205) plays a role in adipogenic differentiation of mouse adipose-derived stem cells. *Mol Cell Endocrinol*. 2014; 392:80–89. [PubMed: 24859602]
28. Dermott JM, Gooya JM, Asefa B, Weiler SR, Smith M, Keller JR. Inhibition of growth by p205: a nuclear protein and putative tumor suppressor expressed during myeloid cell differentiation. *Stem cells (Dayton, Ohio)*. 2004; 22:832–848.
29. Geiss GK, Bumgarner RE, Birditt B, Dahl T, Dowidar N, Dunaway DL, Fell HP, Ferree S, George RD, Grogan T, James JJ, Maysuria M, Mitton JD, Oliveri P, Osborn JL, Peng T, Ratcliffe AL, Webster PJ, Davidson EH, Hood L, Dimitrov K. Direct multiplexed measurement of gene expression with color-coded probe pairs. *Nat Biotechnol*. 2008; 26:317–325. [PubMed: 18278033]
30. Schmidt T, Schmid-Burgk JL, Hornung V. Synthesis of an arrayed sgRNA library targeting the human genome. *Scientific reports*. 2015; 5:14987. [PubMed: 26446710]
31. Schmid-Burgk JL, Schmidt T, Gaidt MM, Pelka K, Latz E, Ebert TS, Hornung V. OutKnocker: a web tool for rapid and simple genotyping of designer nuclease edited cell lines. *Genome research*. 2014; 24:1719–1723. [PubMed: 25186908]
32. Broz P, Newton K, Lamkanfi M, Mariathasan S, Dixit VM, Monack DM. Redundant roles for inflammasome receptors NLRP3 and NLRC4 in host defense against Salmonella. *J Exp Med*. 2010; 207:1745–1755. [PubMed: 20603313]
33. Phatnani HP, Greenleaf AL. Phosphorylation and functions of the RNA polymerase II CTD. *Genes Dev*. 2006; 20:2922–2936. [PubMed: 17079683]
34. Hsin JP, Manley JL. The RNA polymerase II CTD coordinates transcription and RNA processing. *Genes Dev*. 2012; 26:2119–2137. [PubMed: 23028141]
35. Parsons MJ, Vertino PM. Dual role of TMS1/ASC in death receptor signaling. *Oncogene*. 2006; 25:6948–6958. [PubMed: 16715133]
36. Bradley MN, Zhou L, Smale ST. C/EBPbeta regulation in lipopolysaccharide-stimulated macrophages. *Molecular and cellular biology*. 2003; 23:4841–4858. [PubMed: 12832471]
37. Rathinam VA, Vanaja SK, Fitzgerald KA. Regulation of inflammasome signaling. *Nature immunology*. 2012; 13:333–342. [PubMed: 22430786]
38. Conrady CD, Zheng M, Fitzgerald KA, Liu C, Carr DJ. Resistance to HSV-1 infection in the epithelium resides with the novel innate sensor, IFI-16. *Mucosal Immunol*. 2012; 5:173–183. [PubMed: 22236996]

39. Storek KM, Gertsvolf NA, Ohlson MB, Monack DM. cGAS and Ifi204 cooperate to produce type I IFNs in response to Francisella infection. *Journal of immunology* (Baltimore, Md : 1950). 2015; 194:3236–3245.
40. Ludlow LE, Johnstone RW, Clarke CJ. The HIN-200 family: more than interferon-inducible genes? *Experimental cell research*. 2005; 308:1–17. [PubMed: 15896773]
41. Ma XY, Wang H, Ding B, Zhong H, Ghosh S, Lengyel P. The interferon-inducible p202a protein modulates NF-kappaB activity by inhibiting the binding to DNA of p50/p65 heterodimers and p65 homodimers while enhancing the binding of p50 homodimers. *The Journal of biological chemistry*. 2003; 278:23008–23019. [PubMed: 12676938]
42. Datta B, Li B, Choubey D, Nallur G, Lengyel P. p202, an interferon-inducible modulator of transcription, inhibits transcriptional activation by the p53 tumor suppressor protein, and a segment from the p53-binding protein 1 that binds to p202 overcomes this inhibition. *The Journal of biological chemistry*. 1996; 271:27544–27555. [PubMed: 8910340]
43. Min W, Ghosh S, Lengyel P. The interferon-inducible p202 protein as a modulator of transcription: inhibition of NF-kappa B, c-Fos, and c-Jun activities. *Molecular and cellular biology*. 1996; 16:359–368. [PubMed: 8524315]
44. Datta B, Min W, Burma S, Lengyel P. Increase in p202 expression during skeletal muscle differentiation: inhibition of MyoD protein expression and activity by p202. *Molecular and cellular biology*. 1998; 18:1074–1083. [PubMed: 9448005]
45. Lembo M, Sacchi C, Zappador C, Bellomo G, Gaboli M, Pandolfi PP, Gariglio M, Landolfo S. Inhibition of cell proliferation by the interferon-inducible 204 gene, a member of the Ifi 200 cluster. *Oncogene*. 1998; 16:1543–1551. [PubMed: 9569021]
46. Liu CJ, Chang E, Yu J, Carlson CS, Prazak L, Yu XP, Ding B, Lengyel P, Di Cesare PE. The interferon-inducible p204 protein acts as a transcriptional coactivator of Cbfa1 and enhances osteoblast differentiation. *The Journal of biological chemistry*. 2005; 280:2788–2796. [PubMed: 15557274]
47. Liu C, Wang H, Zhao Z, Yu S, Lu YB, Meyer J, Chatterjee G, Deschamps S, Roe BA, Lengyel P. MyoD-dependent induction during myoblast differentiation of p204, a protein also inducible by interferon. *Molecular and cellular biology*. 2000; 20:7024–7036. [PubMed: 10958697]
48. Corrales L, Woo SR, Williams JB, McWhirter SM, Dubensky TW Jr, Gajewski TF. Antagonism of the STING Pathway via Activation of the AIM2 Inflammasome by Intracellular DNA. *Journal of immunology* (Baltimore, Md : 1950). 2016; 196:3191–3198.
49. Hirose Y, Tacke R, Manley JL. Phosphorylated RNA polymerase II stimulates pre-mRNA splicing. *Genes Dev*. 1999; 13:1234–1239. [PubMed: 10346811]
50. Collard RL, Harya NS, Monzon FA, Maier CE, O’Keefe DS. Methylation of the ASC gene promoter is associated with aggressive prostate cancer. *Prostate*. 2006; 66:687–695. [PubMed: 16425203]
51. Das PM, Ramachandran K, Vanwert J, Ferdinand L, Gopisetty G, Reis IM, Singal R. Methylation mediated silencing of TMS1/ASC gene in prostate cancer. *Molecular cancer*. 2006; 5:28. [PubMed: 16848908]
52. Machida EO, Brock MV, Hooker CM, Nakayama J, Ishida A, Amano J, Picchi MA, Belinsky SA, Herman JG, Taniguchi S, Baylin SB. Hypermethylation of ASC/TMS1 is a sputum marker for late-stage lung cancer. *Cancer research*. 2006; 66:6210–6218. [PubMed: 16778195]
53. Ohtsuka T, Liu XF, Koga Y, Kitajima Y, Nakafusa Y, Ha CW, Lee SW, Miyazaki K. Methylation-induced silencing of ASC and the effect of expressed ASC on p53-mediated chemosensitivity in colorectal cancer. *Oncogene*. 2006; 25:1807–1811. [PubMed: 16331272]
54. Riojas MA, Guo M, Glockner SC, Machida EO, Baylin SB, Ahuja N. Methylation-induced silencing of ASC/TMS1, a pro-apoptotic gene, is a late-stage event in colorectal cancer. *Cancer biology & therapy*. 2007; 6:1710–1716. [PubMed: 17986858]
55. Wu L, Zhang C, Wang X, Ding X, Deng J, Liang H. Methylation of ASC/TMS1 promoter is associated with poor prognosis of patients with gastric cancer. *Clinical & translational oncology : official publication of the Federation of Spanish Oncology Societies and of the National Cancer Institute of Mexico*. 2016; 18:296–303.

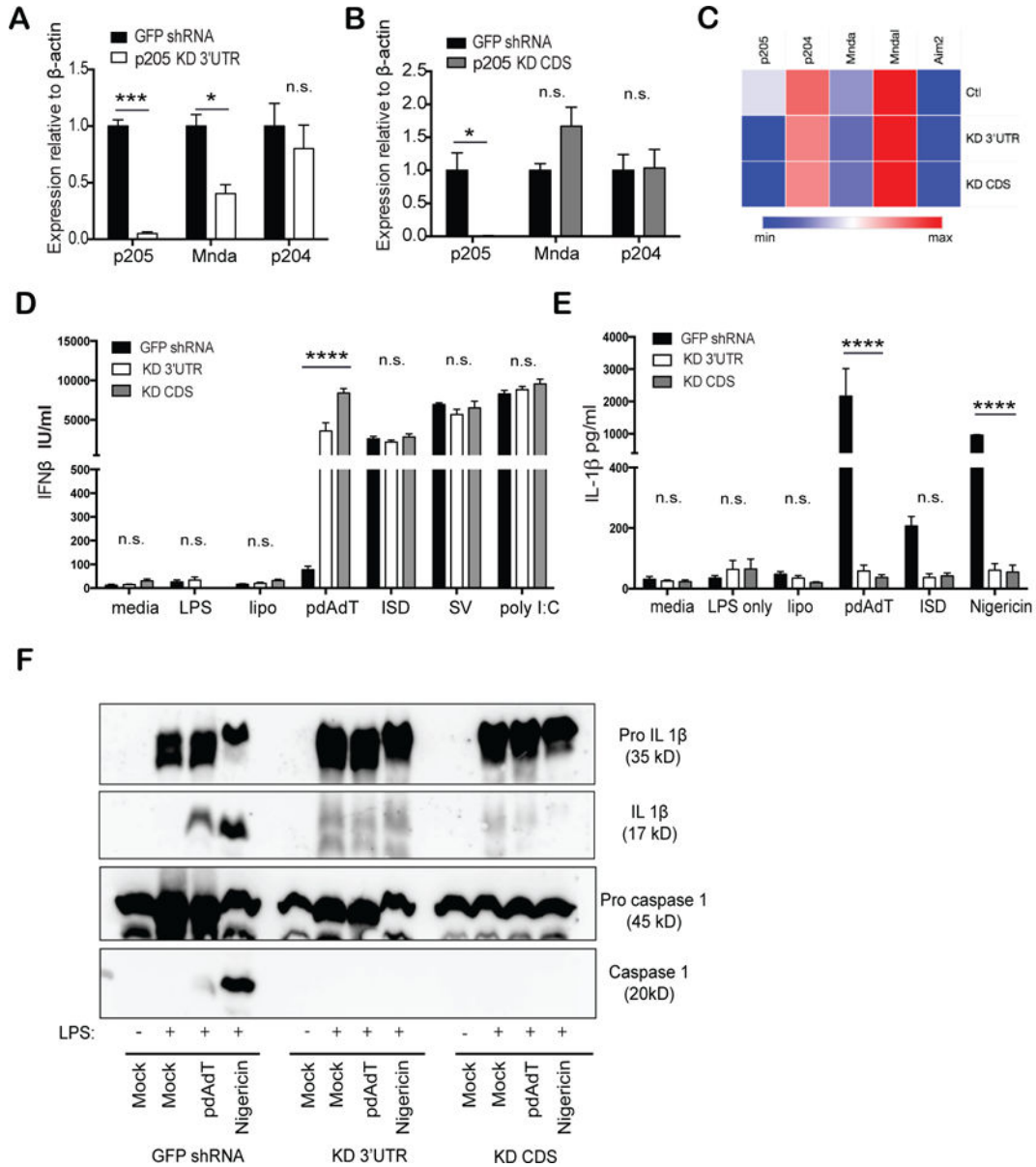
56. Levine JJ, Stimson-Crider KM, Vertino PM. Effects of methylation on expression of TMS1/ASC in human breast cancer cells. *Oncogene*. 2003; 22:3475–3488. [PubMed: 12776200]
57. Moriai R, Tsuji N, Kobayashi D, Yagihashi A, Namiki Y, Takahashi H, Watanabe N. A proapoptotic caspase recruitment domain protein gene, TMS1, is hypermethylated in human breast and gastric cancers. *Anticancer research*. 2002; 22:4163–4168. [PubMed: 12553049]
58. Virmani A, Rathi A, Sugio K, Sathyanarayana UG, Toyooka S, Kischel FC, Tonk V, Padar A, Takahashi T, Roth JA, Euhus DM, Minna JD, Gazdar AF. Aberrant methylation of TMS1 in small cell, non small cell lung cancer and breast cancer. *International journal of cancer*. 2003; 106:198–204. [PubMed: 12800194]





**Figure 1. p205 is induced by LPS, Type I and II IFN and localizes to the nucleus**  
 Primary BMDMs stimulated with (A) LPS (200 ng/ml), (B) Type-I IFN (100 U/ml) or (C) IFN- $\gamma$  (20 ng/ml) at different time points (0, 2, 4, 8, 16, 24 h) were tested for p205 mRNA expression as well as other PYHIN genes, *Mnda*, *p204* and *Aim2*. Levels of IL-6 or Cxcl10 mRNA were included as positive controls. Gene expression is reported relative to a combination of three housekeeping genes *Gapdh*, *Hprt*,  $\beta$ -*actin*. (D) Immunoblot analysis of the nuclear and cytoplasmic fractions of wild-type BMDM transduced with empty vector (EV) and wild-type BMDM overexpressing HA-tagged p205 using anti-HA antibody.

Histone 3 and Aim2 were used as controls for nuclear and cytosolic extracts respectively. **(E)** Confocal microscopy of CFP-tagged p205 (green) in HEK 293T cells stained for nucleus using Acridine orange in first panel and Cholera Toxin B (CtxB) staining plasma membrane in second panel. Data is representative of two independent experiments. **(F)** Primary BMDM untreated (Ctl) or treated with LPS for 6h, type-I IFN for 16h or IFN- $\gamma$  for 16h were separated into nuclear and cytosolic fractions and immunoblotted for endogenous p205 (n.s.- non-specific band). **(G)** Western blot analysis of endogenous p205 expression in the nuclear and cytosolic extracts of primary macrophages treated with type-I IFN as indicated. Usf2 and Gapdh were used as controls for nuclear and cytosolic fractions respectively (n.s.- non-specific band).



**Figure 2. p205 knockdown using shRNA results in impaired inflammasomes activation**  
 BMDM transduced with shRNA targeting either (A) 3'UTR or (B) CDS of p205 gene were inspected for expression of p205, Mnda and p204 mRNA relative to β-actin mRNA and normalized to expression in GFP shRNA BMDM. (C) Heatmap of PYHIN gene expression in p205 KD 3'UTR and KD CDS BMDM compared to control (Ctl) BMDM. The p205 knockdown BMDMs were primed with LPS (200ng/ml) for 3h and then stimulated with transfected pdAdT (1μg/ml for 6h), transfected ISD (3μM for 6h), Nigericin (10μM for 1h), ATP (5μM) or, stimulated alone with Sendai virus (SV; overnight) or poly I:C (overnight). Secreted (D) IFNβ and (E) IL-1β levels were assessed by ELISA. (F) GFP shRNA CTL, p205 KD 3'UTR and KD CDS were primed with LPS (200ng/ml) for 3h and then stimulated with pdAdT (1 μg/ml for 6h) or Nigericin (10 μM for 1h) and the supernatants

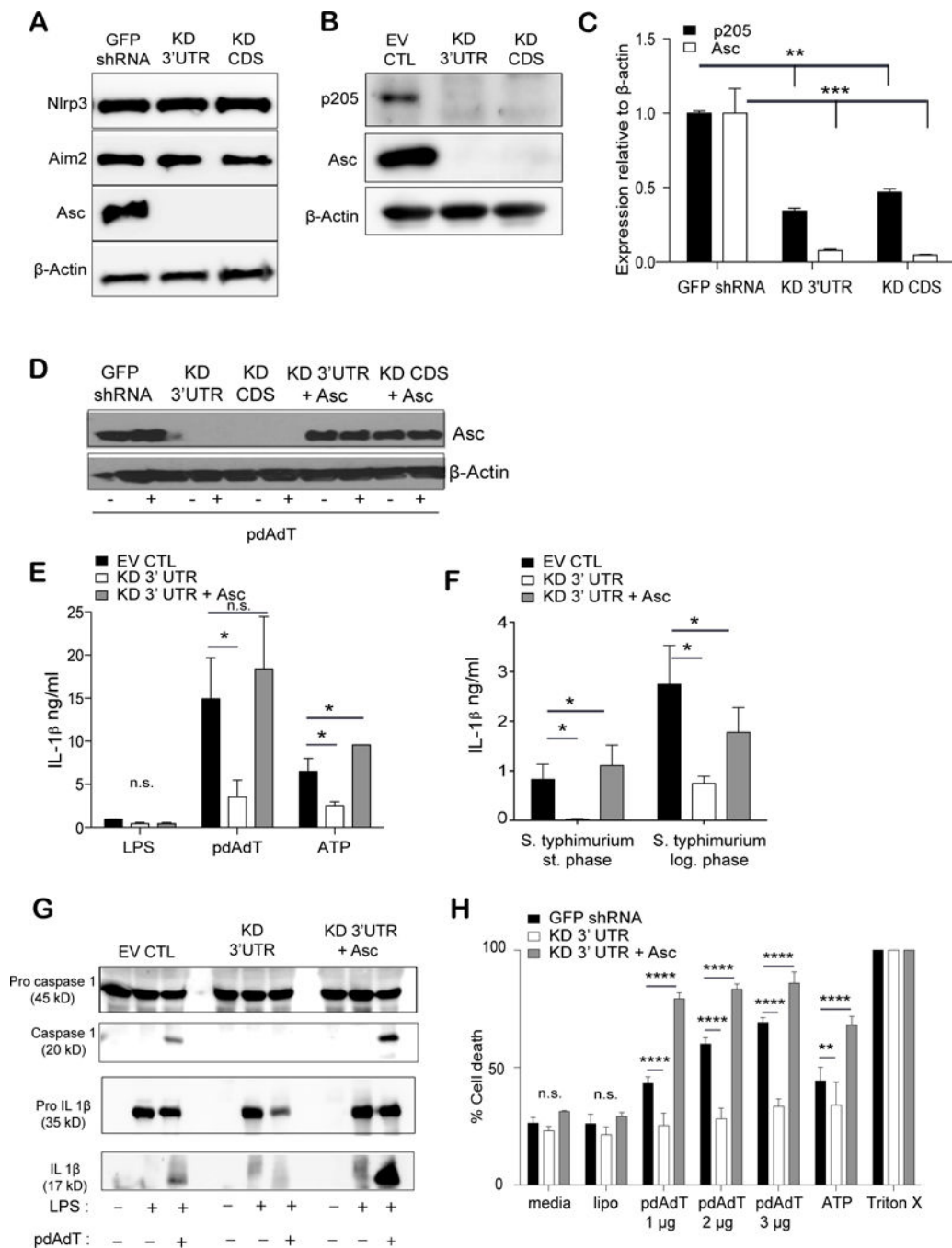
and the lysates from the macrophages were immunoblotted for pro-IL1 $\beta$  (35 kD), cleaved form of IL-1 $\beta$  (p17), pro-caspase 1 (45kD) and the active subunit of caspase 1(p20).

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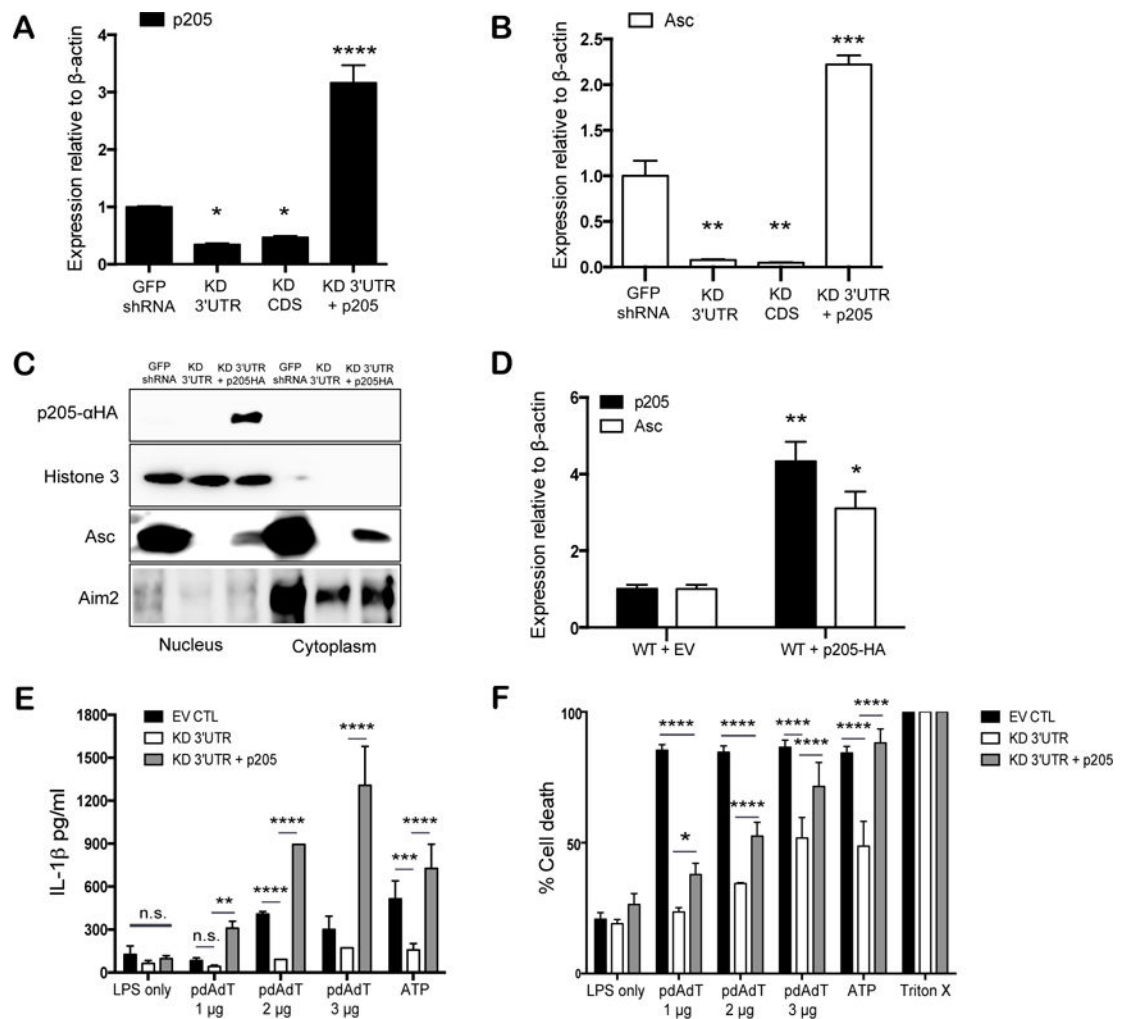
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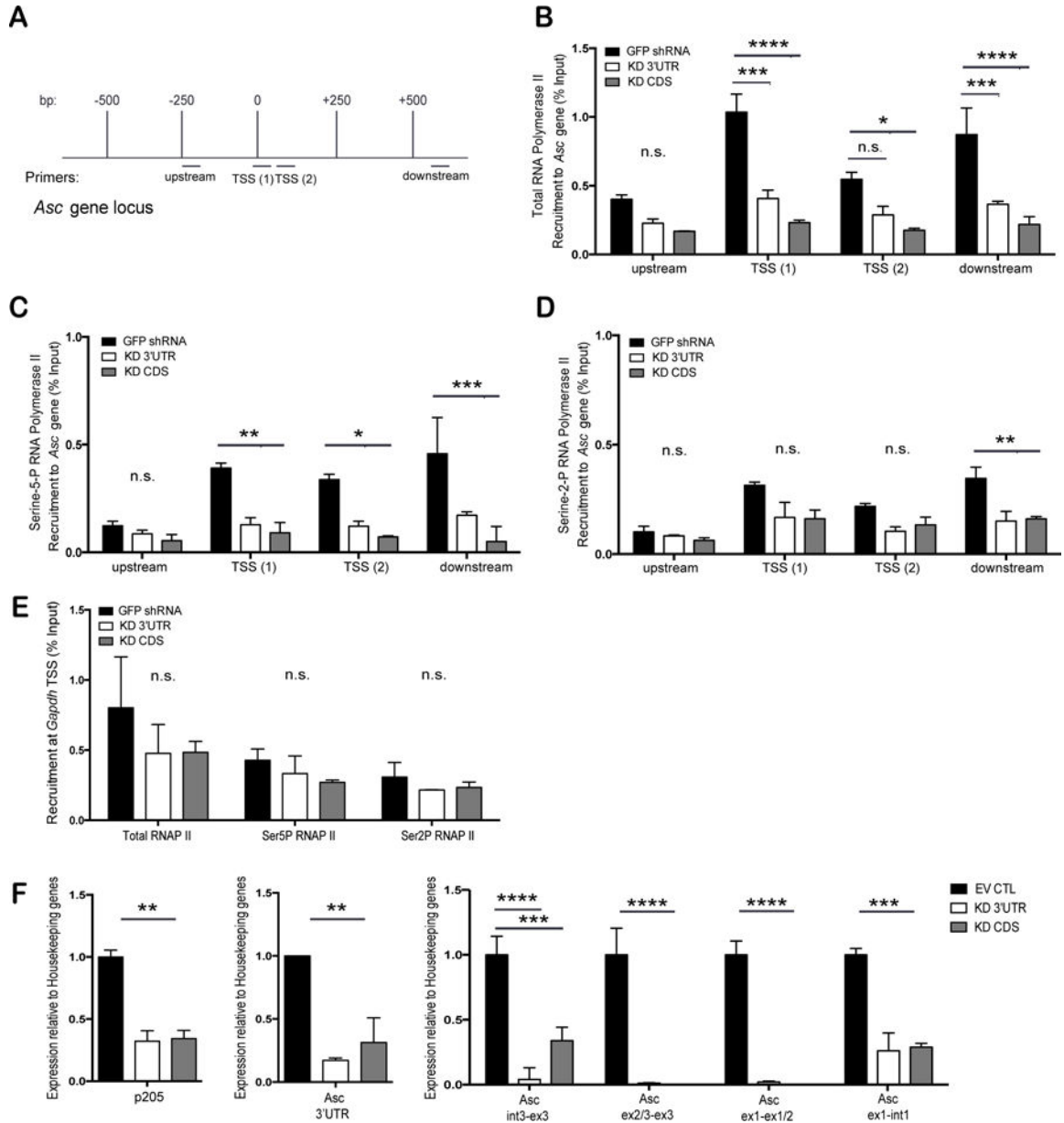
**Figure 3. Loss of p205 leads to a defect in Asc expression**

(A) Levels of Nlrp3, Aim2, Asc and  $\beta$ -actin proteins were elucidated by Western blot in LPS stimulated (200 ng/ml) GFP shRNA CTL, p205 KD 3'UTR and KD CDS macrophages. (B) Immunoblot of p205, Asc and  $\beta$ -actin proteins in LPS-stimulated (200 ng/ml for 6h) p205 knockdown macrophages. (C) Levels of p205 and Asc mRNA (relative to  $\beta$ -actin and normalized to GFP shRNA BMDM) were detected by qPCR in the same cell lines. (D) Western blot analysis of Asc overexpression in the p205 knockdown BMDM either left untreated or treated with LPS treated (200 ng/ml for 3h). Asc reconstituted cell lines tested

for IL-1 $\beta$  production (**E**) on stimulation with LPS (200 ng/ml for 3h) and pdAdT (1  $\mu$ g/ml for 6h) or ATP (5  $\mu$ M for 1h) (left) and (**F**) with overnight culture (stationary phase) or log phase culture of *Salmonella* sp. by ELISA. (**G**) Levels of caspase 1 and IL-1 $\beta$  processing in the Asc reconstituted cells were detected by Western Blot and (**H**) cell death was measured by amounts of LDH released with LPS (200 ng/ml for 3h) and pdAdT (1  $\mu$ g, 2  $\mu$ g, or 3  $\mu$ g per ml for 6h) and/or ATP (5  $\mu$ M for 1h) stimulation.

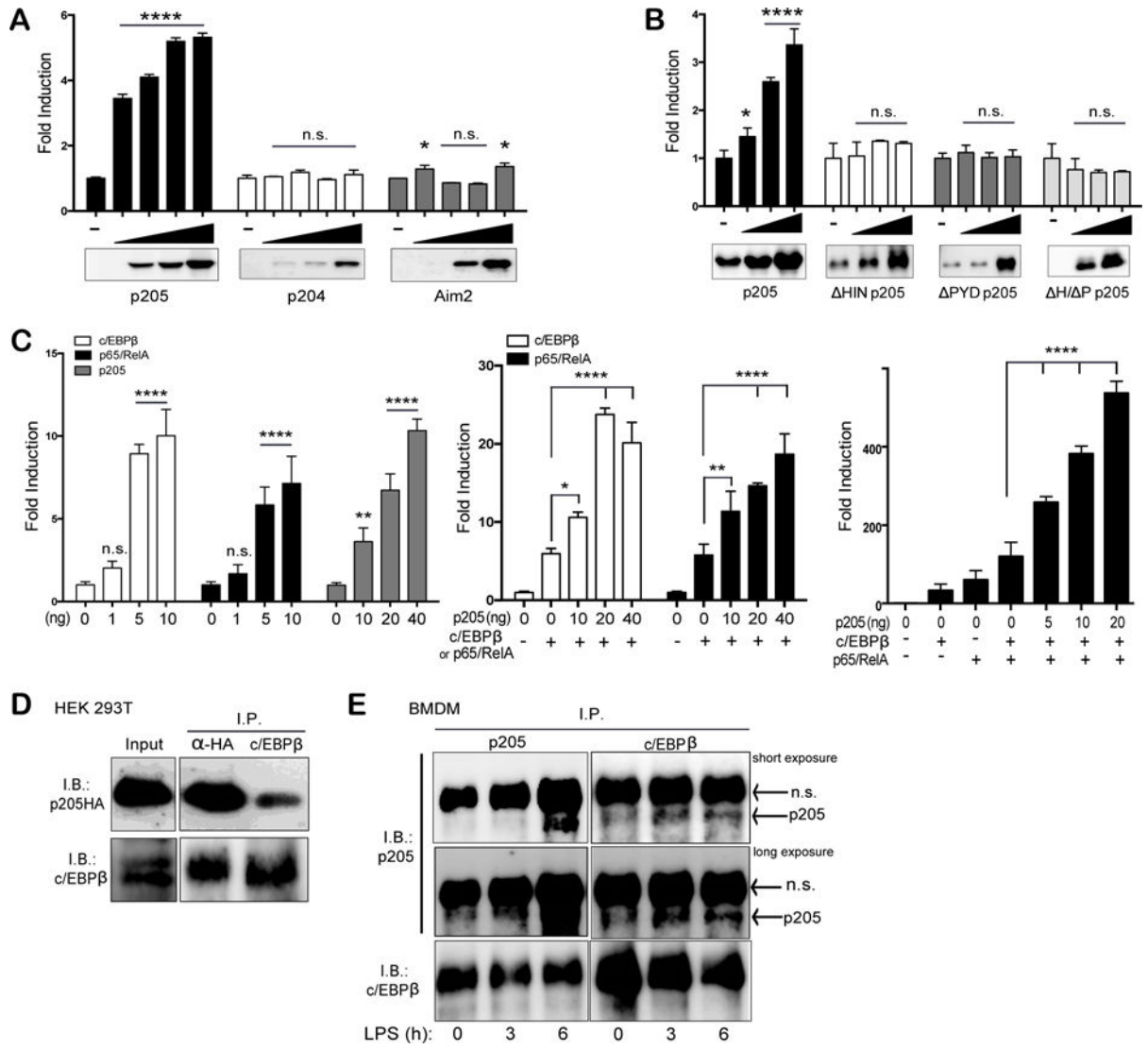


**Figure 4. Reconstitution of p205 rescues Asc expression and inflammasome activation**  
 (A) p205 and (B) Asc mRNA expression relative to  $\beta$ -actin and normalized to GFP shRNA BMDM were measured in BMDMs reconstituted with p205. (C) Western blot analysis of nuclear and cytoplasmic fractions of p205 reconstituted cell lines to detect p205-HA, Histone H3, Asc and Aim2. (D) p205 and Asc mRNA levels by qPCR relative to  $\beta$ -actin in p205- overexpressing BMDM. The rescued cells were tested for (E) IL1 $\beta$  production and (F) cell death with LPS (200ng/ml for 3h) and pdAdT (1  $\mu$ g, 2  $\mu$ g, or 3 $\mu$ g per ml for 6h) or ATP (5  $\mu$ M for 1h) by ELISA and by measuring amount of LDH released respectively.

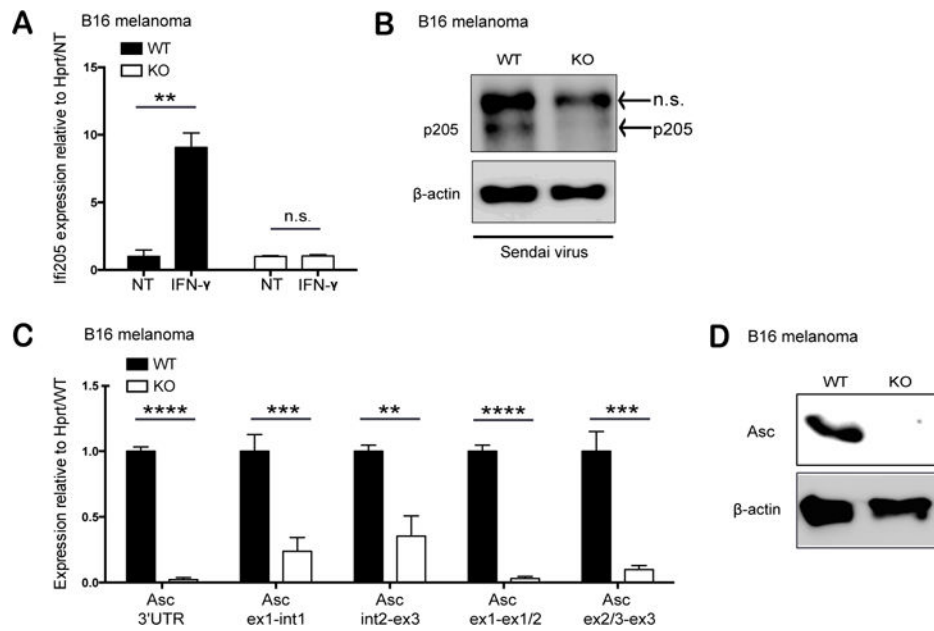


**Figure 5. p205 modulates transcription and splicing from the endogenous *Asc* gene**  
**(A)** Schematic of the *Asc* gene locus and Chromatin-IP primers locations on the gene. **(B)** Recruitment of total RNA Pol II to endogenous *Asc* gene in GFP shRNA CTL, p205 KD 3'UTR or KD CDS macrophages. Recruitment of **(C)** Serine-5-P RNA Pol II and **(D)** Serine-2-P RNA Pol II on endogenous *Asc* gene. **(E)** Recruitment of total, Serine-5-P or Serine-2-P RNA Pol II to the *Gapdh* transcription start site. All values are represented as percent fraction of total input DNA. Data was calculated against the IgG isotype control and is representative of three independent experiments. **(D)** qRT-PCR analysis of p205, nascent or mature *Asc* mRNA expression (relative to *Hprt* and *Gapdh*, and normalized to EV CTL BMDM) in p205 knockdown macrophages.





**Figure 6. p205 drives expression from an *Asc* promoter-luciferase reporter construct**  
 HEK 293T cells were transfected with a mixture containing 10 ng of TK-Renilla luciferase with 1 ng of *Asc* promoter firefly luciferase reporter and increasing amounts of (A) p205-HA, p204-HA or Aim2-FLAG, or (B) with full length p205 or p205 deletion mutants as indicated. (C) Transfection of increasing concentrations of c/EBPβ, p65/RelA and p205 alone or, co-transfection of increasing concentrations of p205-HA with either c/EBPβ or p65/RelA with *Asc* promoter-reporter or, transfection of the *Asc* promoter-reporter with increasing concentrations of p205 with both c/EBPβ and p65/RelA, as indicated. All luciferase values were measured and normalized to Renilla values. Values are displayed as fold change over the *Asc* reporter construct alone. Data is representative of three independent experiments. (D) Co-immunoprecipitation and immunoblot of overexpressed HA-tagged p205 and c/EBPβ in HEK 293T cells using anti-HA and anti-c/EBPβ antibodies. (E) Co-immunoprecipitation and Western blot of endogenous p205 and c/EBPβ in LPS-stimulated BMDM using antibodies against p205 and c/EBPβ. Data is representative of two independent experiments (n.s.- non-specific band).



**Figure 7. CRISPR KO of p205 leads to reduced Asc expression in B16 melanoma**

(A) Expression of p205 mRNA in IFN-stimulated wild type (WT) and p205 knockout (KO) B16 melanoma cell lines relative to Hprt and normalized to non-treated (NT). (B) p205 and  $\beta$ -actin protein expression in WT and p205 KO cell lines stimulated with Sendai virus (n.s.-non-specific band) (C) Mature mRNA and pre-mRNA profile of Asc expression (relative to Hprt and normalized to WT) and (D) Western blot analysis of Asc and  $\beta$ -actin in WT and p205 KO B16 melanoma cell lines.