


Closed-loop regulation of arterial pressure after acute brain death

Kristian Soltesz¹  · Trygve Sjöberg² · Tomas Jansson³ · Rolf Johansson¹ · Anders Robertsson¹ · Audrius Paskevicius² · Quiming Liao² · Guangqi Qin² · Stig Steen²

Received: 17 March 2017 / Accepted: 30 May 2017 / Published online: 10 June 2017
© The Author(s) 2017. This article is an open access publication

Abstract The purpose of this concept study was to investigate the possibility of automatic mean arterial pressure (MAP) regulation in a porcine heart-beating brain death (BD) model. Hemodynamic stability of BD donors is necessary for maintaining acceptable quality of donated organs for transplantation. Manual stabilization is challenging, due to the lack of vasomotor function in BD donors. Closed-loop stabilization therefore has the potential of increasing availability of acceptable donor organs, and serves to indicate feasibility within less demanding patient groups. A dynamic model of nitroglycerine pharmacology, suitable for controller synthesis, was identified from an experiment involving an anesthetized pig, using a gradient-based output error method. The model was used to synthesize a robust PID controller for hypertension prevention, evaluated in a second experiment, on a second, brain dead, pig. Hypotension was simultaneously prevented using closed-loop controlled infusion of noradrenaline, by means of a previously published controller. A linear model of low order, with variable (uncertain) gain, was sufficient to describe the dynamics to be controlled. The robustly tuned PID controller utilized in the second experiment kept the MAP within a user-defined range. The system was able to prevent hypertension, exceeding a reference of 100 mmHg by more than

10%, during 98% of a 12 h experiment. This early work demonstrates feasibility of the investigated modelling and control synthesis approach, for the purpose of maintaining normotension in a porcine BD model. There remains a need to characterize individual variability, in order to ensure robust performance over the expected population.

Keywords Hemodynamics · Blood pressure · Closed-loop control · Brain death

1 Introduction

1.1 Pathophysiology and treatment of heart-beating brain dead donors

Heart-beating brain dead (BD) donors are the largest source of donor organs [1]. Upon acute BD, homeostasis is lost, leading to the rapid degradation of body organs. The incidence of BD is associated with a dramatic increase in plasma catecholamine levels. This effect, resulting in a brief (<30 min) episode of hypertension is referred to as the catecholamine storm [2]. As catecholamines are eliminated from the blood stream, blood vessels lose their tone, resulting in severe hypotension. In the absence of treatment, the body experiences systemic circulatory collapse, leading to a loss of adequate organ perfusion. Relatedly, plasma concentration of cortisol, antidiuretic hormone (vasopressin), and thyroid hormones decrease upon BD [2].

In order to compensate for the mentioned changes, BD donors are subject to hormone therapy. Internationally recognized guidelines for treatment of BD donors are provided in [3, 4]. This study is based on local (Swedish) recommendations [5], derived from the aforementioned guidelines. Prevention of hypotension is the most

✉ Kristian Soltesz
kristian@control.lth.se

¹ Department Automatic Control, Lund University, P.O. Box 118, SE-221 00 Lund, Sweden

² Department Cardiothoracic Surgery, Lund University and Skåne University Hospital, Lund, Sweden

³ Department Clinical Sciences Lund, Biomedical Engineering, Lund University, Lund, Sweden and Medical Services, Skåne University Hospital, Lund, Sweden

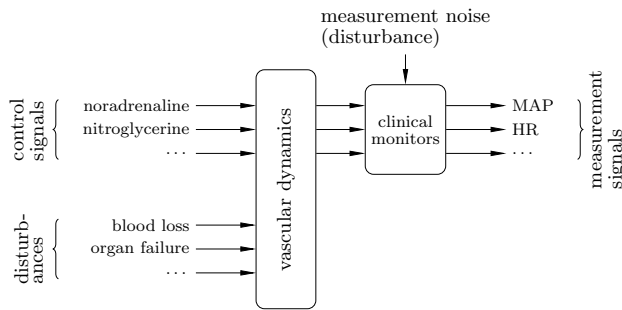


Fig. 1 The BD organ donor, illustrated from a control system perspective

challenging of the treatment goals. The guidelines dictate maintenance of a mean arterial pressure (MAP) exceeding 60–70 mmHg, while remaining below 100 mmHg. This can be partially achieved by intravenous infusion of noradrenaline. However, infusion rates exceeding 0.05 $\mu\text{g}/\text{min}/(\text{kg body weight})$, are not feasible due to elevated risk of cardiac graft dysfunction, and higher early and late recipient mortality. Consequently, hormonal therapy is complemented by forced fluid therapy, with the adverse effect of drastically increasing the risk of lung edema [6]. It has been demonstrated in a porcine BD model, that a sufficient MAP can be maintained throughout a 24 h treatment window,¹ in the absence of forced fluid therapy, by blocking neural catecholamine reuptake [7].

While previous research [7, 8] has focused on maintaining a sufficient MAP, clinical care of BD donors would additionally benefit from the capability to avoid periods of excessive hypertension. Most notably, the catecholamine storm associated with BD, results in decreased tissue perfusion due to drastically increased systemic (SVR) and pulmonary (PVR) vascular resistance [2]. It has been observed in a primate BD model, that the subsequent fall of PVR is more abrupt than that of SVR, resulting in blood pooling in the lungs, and consequently an increased risk of pulmonary edema [9].

A further motivation for this work is that sustained episodes of elevated arterial pressure, exceeding the recommended upper limit of MAP >100 mmHg, occurred spontaneously after 5–10 h in the utilized porcine BD model, during all experiments underlying [10]. These episodes could not be explained by the combined administration of catecholamines and reuptake blocker, as they did not cease upon halted administration of the mentioned drugs.

¹ The authors are based in Sweden, where treatment of BD donors is limited to 24 h by national regulations.

1.2 Closed-loop hemodynamic management

While the human body is rich in closed-loop control systems (vasomotor and blood glucose regulation, to mention two), synthetic counterparts are still rare in clinical medicine. Studies aimed at closed-loop control of anesthetic depth and analgesia by means of EEG-guided infusion of anesthetic and analgesic drugs have proven successful [11, 12]. The purpose of such systems is to alleviate clinicians of repetitive low-level tasks, such as intermittent infusion rate adjustments.

Following successful studies on closed-loop anesthesia systems, the prospect of closed-loop controlled hemodynamic stabilization, including the control of intravascular volume, has been proposed as a challenging interdisciplinary research problem [12, 13]. Relatedly, potential clinical benefit of closed-loop controlled blood pressure management was highlighted in a recent *in silico* simulation study [14]. A comprehensive review of the current state of the art regarding closed-loop controlled hemodynamic management in humans is provided in [15]. The review reveals that apart from efforts in hypertension managements using sodium nitroprusside in the 1980s and early 1990s, there exist very few reports directly related to the work presented herein.

The thesis underlying the work presented herein, is that relatively simple system identification and robust control methods are sufficient to control MAP in a porcine BD model. The use of a porcine model is motivated by the fact that large animal models are the closest available approximation of human cardiovascular physiology, see e.g. [16].

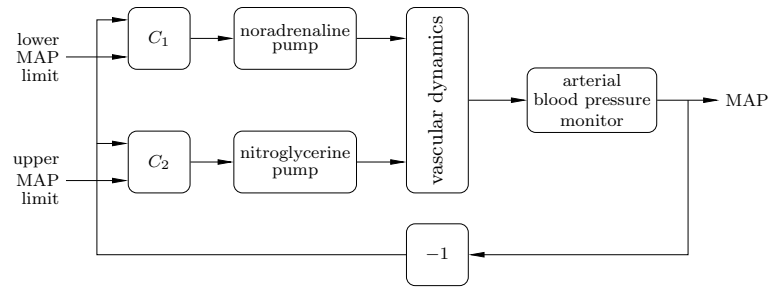
To investigate the thesis, a control system with two loops, schematically depicted in Fig. 2 was constructed. In this context, the process to be controlled are the vascular dynamics, conceptually illustrated by Fig. 1. One of the loops prevents hypotension by means of noradrenaline infusion, as described in [10]. A second controller, being the focus of this paper, is simultaneously acting to prevent hypertension, by means of nitroglycerine infusion. Nitroglycerine is a potent vasodilator, suited for the task, as it lacks side effect, which would limit its administration to BD donors [17].

2 Methods

2.1 Experimental setup

A modular control system, enabling simultaneous computer control of eight infusion pumps (Carefusion Alaris TIVA; BD, Franklin Lakes, NJ) was constructed. It is shown in Fig. 3, with two infusion pumps attached. In addition to the pumps, the system has connection ports for invasive blood

Fig. 2 Control system configuration, used during the close-loop controlled experiment. The controller C_1 administers noradrenaline to prevent hypotension, while C_2 administers nitroglycerine to prevent hypertension. Remaining drugs of Table 1 were not under closed-loop control



pressure transducers (DTXPlus; Argon Medical, Plano, TX). Signal processing, control and GUI are implemented on a PC, communicating with the pumps and sensors via RS232 over a USB cable.

For the experiments described herein, the system was fitted with five infusion pumps, according to Table 1. Arterial pressure measurements were collected at 100 Hz, and used to compute mean arterial pressure (MAP), as well as diastolic and systolic pressure. All measurement and control signals were displayed on the graphical user interface, and logged at 1 Hz.

2.2 Animal preparation

Two experiments, each involving one 40 kg Swedish native breed pig, were conducted, as subsequently described. The



Fig. 3 Prototype closed-loop hemodynamic stabilization system, with two (out of eight possible) computer controlled infusion pumps attached. Connectors for blood pressure transducers and user interface PC are situated on the backside of the column

Table 1 Drugs per syringe pump

Pump	Drug	Conc. (mg/ml)
1	Noradrenaline	0.02
2	Cocaine	0.02
3	Nitroglycerine	5.0
4	Adrenaline	0.02
5	Desmopressin	0.72

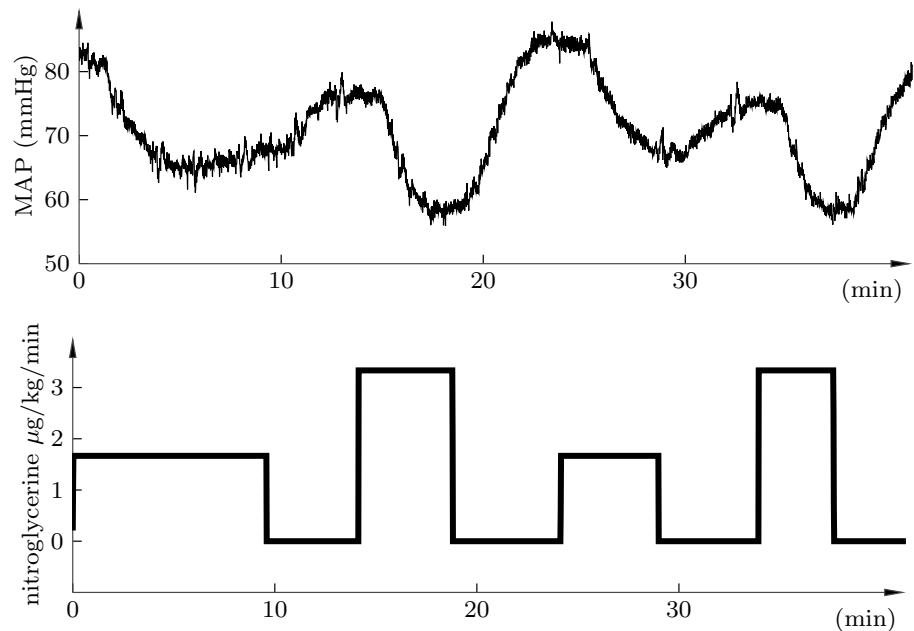
Prevention of hypotension by means of closed-loop co-administration of noradrenaline and cocaine was studied in [10]. This work focused on simultaneously preventing hypertension through closed-loop controlled infusion of nitroglycerine

animals were subject to anesthesia, induced through intramuscular injection of atropine (0.5 mg) (Atropine; Mylan AB, Stockholm, Sweden), xylazine (100 mg) (Rompun Vet; Bayer A.B., Solna, Sweden), and ketamine (800 mg) (Ketaminol Vet.; Intervet, Boxmeer, Netherlands). Fentanyl (160 µg) (Fentanyl B.; Braun, Melsungen, Germany) and midazolam (16 mg) (Midazolam Panpharma; Panpharma S.A., Trittau, Germany) were given intravenously before tracheostomy. Anesthesia and muscle relaxation were maintained with continuous intravenous infusion of a mixture of ketamine (10 mg/kg/h) and rocuronium (1.5 mg/kg/h) (Fresenius Kabi, Uppsala, Sweden).

The animals were placed in supine position, and mechanically ventilated (Servo Ventilator 300; Siemens AB, Solna, Sweden), using volume-controlled and pressure regulated ventilation, with a minute volume of 100–150 ml/(kg body weight) at 20 breaths per minute. Positive end-expiratory pressure (PEEP) was adjusted to 5 cm H₂O, inspired oxygen fraction was 0.5, and end tidal CO₂ was kept between 4.5 and 5.5 kPa through the minute volume regulation.

A suprapubic cystostomy was performed to catheterize the urine bladder. A urimeter was subsequently connected via a Foley catheter, in order to monitor urine production. A saline solution (9 mg/ml) (Sodium chloride; Baxter Medical AB, Kista, Sweden) was continuously infused (3 ml/kg/h) to compensate for fluid losses due to urination, respiration and perspiration, as well as bleeding. Heating of the table was manually adjusted to maintain a body core temperature of 36–38 °C.

Fig. 4 Upper plot shows time evolution of MAP, resulting from the infusion profile of the lower plot. Step sizes correspond to infusion rate changes of 5 ml/h, and 10 ml/h, respectively. MAP was stable at the start time of the experiment



Subsequently, the animals were connected to the control system; lines from the syringe pumps were connected to the line carrying Ringer-acetate (Baxter Viaflo; Baxter Medical AB, Kista, Sweden), and catheters for continuous blood pressure measurement were inserted in the ascending aorta and superior vena cava, respectively, and connected to the system shown in Fig. 3. The pressure transducers were continuously rinsed with a Ringer-heparin solution (5 IU/ml) (Heparin LEO; Vianex Factory A, Athens, Greece) to prevent coagulation on the substrate. Arterial blood gas samples were drawn and analyzed intermittently (ABL 700; Radiometer, Copenhagen, Denmark), to ensure normal values.

Following the preparation phase, the two experiments differed, as explained below, under the “System identification” and “Closed-loop demonstration” sections, respectively. In both experiments, total BD was induced by surgical decapitation between C_2 and C_3 , as explained in [7].

2.3 System identification

The purpose of the first experiment was to obtain a dynamical model relating nitroglycerine infusion rate to MAP. In both porcines and humans, nitroglycerine has very short time-to-peak effect, and half-life (both on a sub-minute or minute time scale). While this makes manual titration challenging, it becomes an advantage in the closed-loop control framework—it allows the controller to react faster to both disturbances and reference changes, as it does not have to ‘wait’ for slow drug dynamics.

The identification experiment conducted, comprised a sequence of alternating positive and negative step changes of nitroglycerine infusion, as shown in Fig. 4.

Pharmacokinetics and dynamics are traditionally described by compartment models. They constitute a subclass of linear time-invariant systems, which can be defined by *transfer functions* in the frequency domain. For our purposes (closed-loop control), the physiological interpretation of individual compartments is not of explicit interest. Therefore, it may suffice to assert a low-order model structure.

$$P(s) = \frac{K}{(sT_1 + 1)(sT_2 + 1)} e^{-sL}, \quad (1)$$

where K ($\Delta\text{mmHg}/\Delta\text{ml/h}$) denotes the static gain, T_1 (s), and T_2 (s) are time constants explaining the temporal evolution of the response, and L (s) is a pure time delay between change of infusion rate and noticeable effect in the MAP.

A superimposition of the positive and negative changes in MAP are shown in Fig. 5, in grey, together with corresponding responses of identified second-order time-delayed transfer functions of the form (1), in black. While the identified gain parameter K varied between the responses; $K \in \{0.77, 0.98, 1.06, 1.10, 1.21, 1.42, 1.68, 1.78\}$, the same dynamic parameters; $T_1 = 30$, $T_2 = 70$, and $L = 40$, were able to accurately describe the dynamics, as shown in Fig. 5.

2.4 Closed-loop demonstration

With a set of dynamic models (one model per K -value) explaining the relation between changes in nitroglycerine

Fig. 5 Superimposition of the positive (*upper*) and negative (*lower*) step responses of Fig. 4 (*grey*), together with corresponding responses of the identified models (*black*). The static gain of each model is shown explicitly in the figure. Values marked *Asterisk* correspond to the larger of the two step sizes of Fig. 4

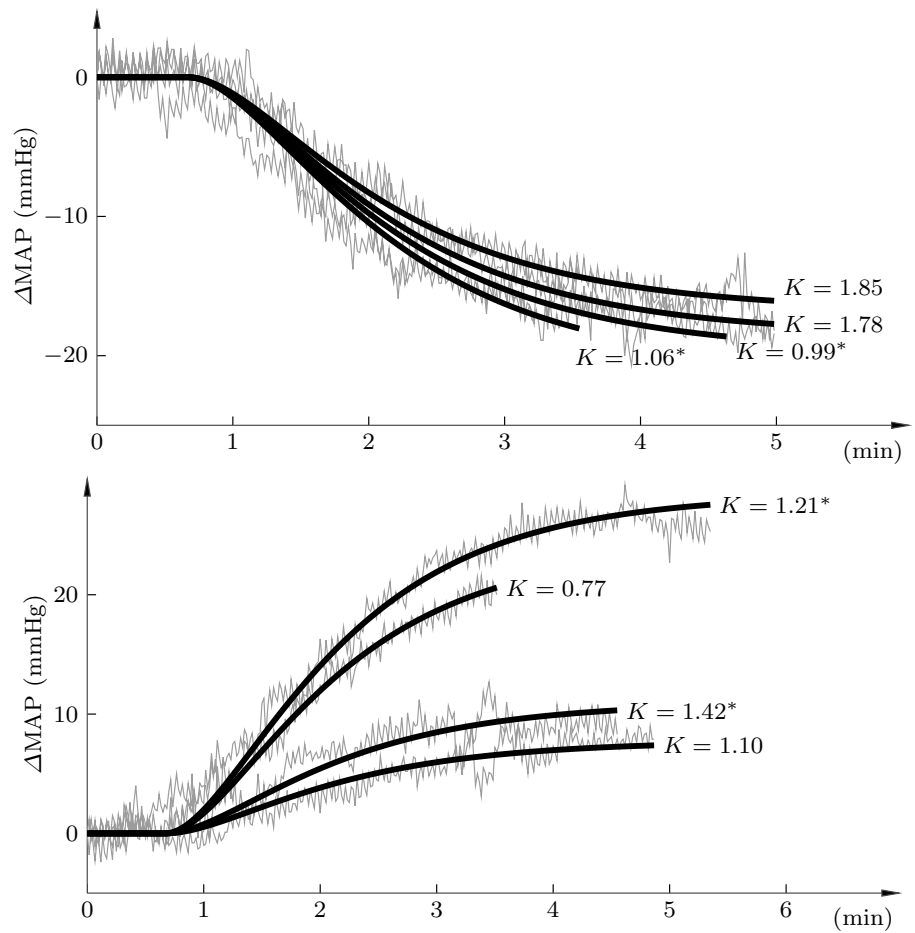


Fig. 6 Nitroglycerine infusion (*bottom*) and resulting MAP (*top, black*). Vertical grey line shows division between closed-loop control (*left*), and halted nitroglycerine infusion (*right*). Dashed lines indicate limits of the desired MAP range [5]. Marks show: **a** missing data due replacement of arterial pressure transducer; **b** infusion pause due to syringe replacement

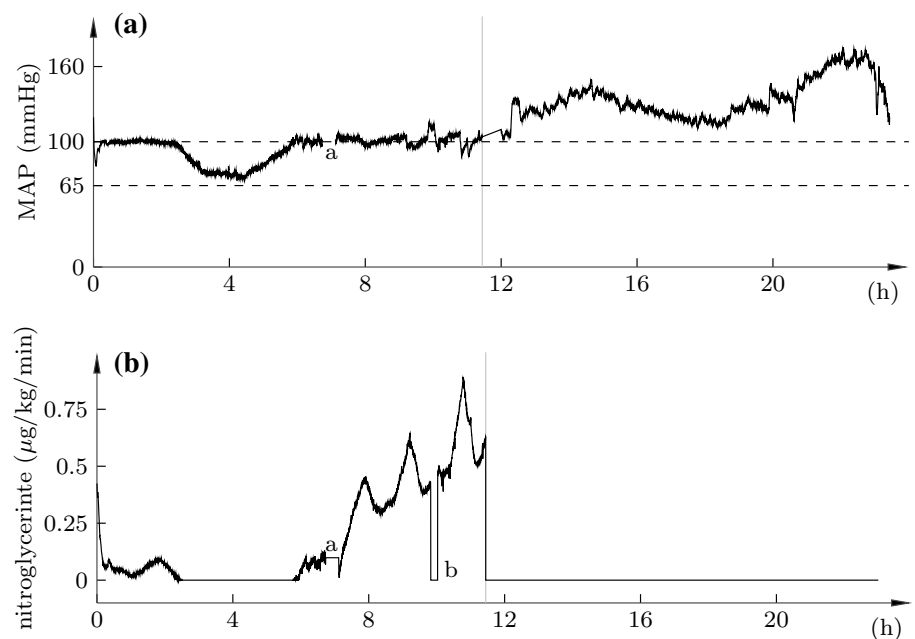


Fig. 7 Central venous pressure throughout the 24 h experiment

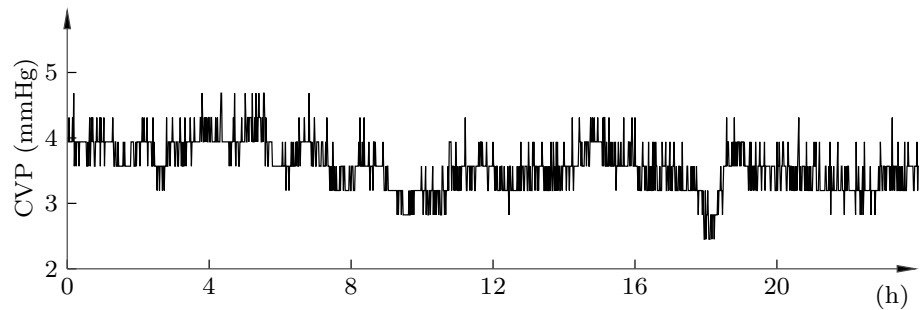
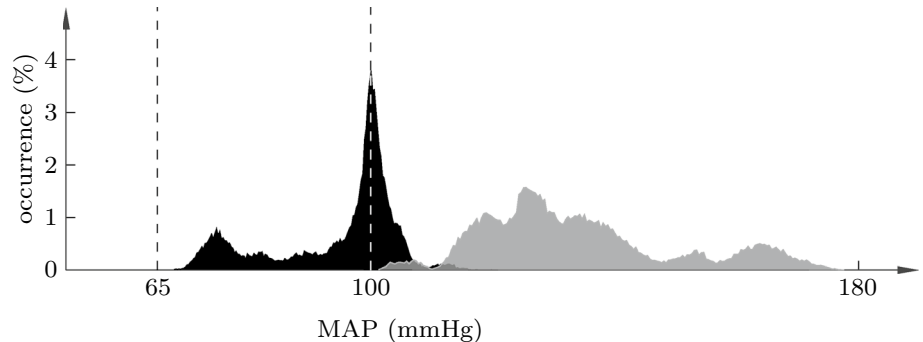


Fig. 8 MAP distribution during the 12 h episodes with (black) and without (grey) closed-loop control. Dashed lines indicate the recommended [5] 65–100 mmHg range



infusion rate and MAP, it was possible to synthesize a closed-loop controller. Owing to its simplicity and reliability, a PID controller,

$$C(s) = k_p + k_i s + \frac{k_d}{s}, \quad (2)$$

was chosen, where, k_p (ml/s/mmHg), k_i (ml/mmHg) and k_d (ml/s/mmHg/s) are constant parameters.

In order to attenuate high-frequency measurement noise, the controller was connected in series with a second-order low-pass filter with transfer function $(sT_f + 1)^{-2}$, where $T_f = 8$ s was chosen to facilitate a trade-off between noise attenuation and controller performance.

Tuning of the PID controller for the identified model set was performed using an optimization-based algorithm, as explained in detail in [10]. Both a PID controller ($k_p = -0.41$, $k_i = -6.64 \times 10^{-3}$, $k_d = -20.3$) and a PI controller ($k_p = -0.21$, $k_i = -2.7 \times 10^{-3}$, $k_d = 0$) were obtained. The PID controller resulted in superior disturbance rejection properties (load step integrated absolute error) for the model set, and was hence chosen in favor of its PI counterpart.

The controller and filter were discretized and implemented in the control system of Fig. 3. Clamping integrator anti-windup was added, as described in [18]. The system was connected to the second pig prior to decapitation. Nitroglycerine was administered under closed-loop control, with the controller described above, and an (upper) MAP reference of 100 mmHg, to prevent hypertension.

Noradrenaline was simultaneously administered under closed-loop control, with a (lower) MAP reference of 65 mmHg, as described in [10]. Cocaine was infused at a fixed ratio (1 mg/mg) to noradrenaline. Desmopressin administration was manually adjusted to maintain normal urine production [5].

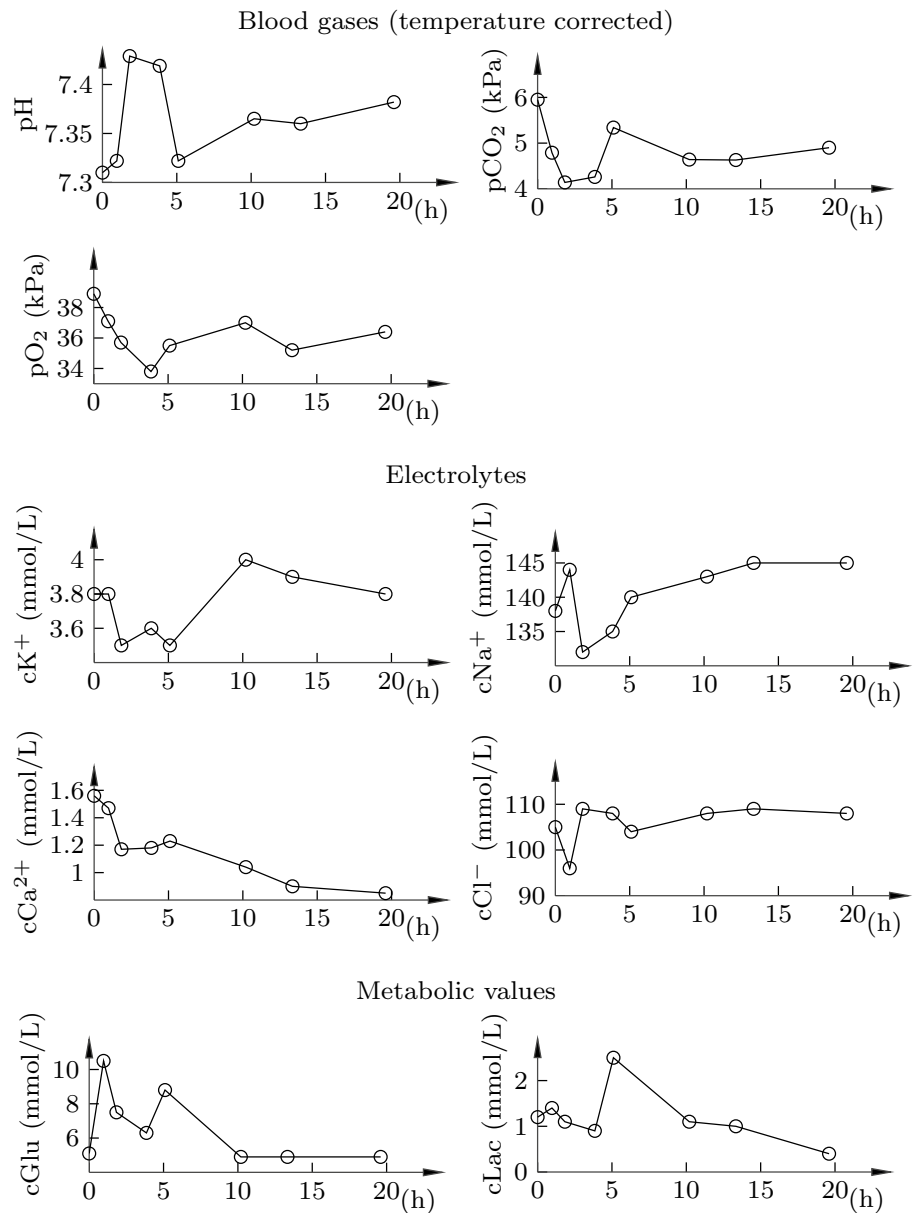
The 24 h treatment window was divided into two episodes of 12 h each. The first of these is more challenging from a control perspective due to the catecholamine storm, and associated hypertension, with subsequent hypotension and circulatory collapse in absence of therapy. Consequently, closed-loop controlled drug administration was enabled during the first 12 h episode. Nitroglycerine infusion was ceased during the second 12 h episode, in order to evaluate the effectiveness of the closed-loop controlled drug administration.

3 Results

The outcome of the full 24 h experiment is shown in Fig. 6. The vertical grey line indicated the division between the first 12 h of closed-loop control, and the subsequent 12 h of halted nitroglycerine infusion. The top plot shows the MAP (solid), and the recommended [5] MAP limits (dashed). The bottom plot shows the corresponding nitroglycerine infusion profile.

The corresponding central venous pressure, not subject to closed-loop control, is shown in Fig. 7. It remained

Fig. 9 Arterial blood gas analysis results. Circles indicate sampling instances



≤10 mmHg, as per the recommendation [5], throughout the experiment.

Decapitation was performed at t = 0. Owing to the responsiveness of the closed-loop nitroglycerine controller, the elevation of arterial pressure associated with the catecholamine storm was almost completely eliminated. This is to be compared with the response in absence of the controller, where previous publications reported MAP elevations to 170 mmHg [7], and 200 mmHg [10], respectively.

The MAP fell below 100 mmHg at t = 2 h 23 min, resulting in the closed-loop controller ceasing the administration of nitroglycerine. Unlike in previously conducted experiments [7, 10], the MAP did subsequently not fall below 65 mmHg. Consequently, neither noradrenaline nor

cocaine were administered by the system throughout the experiment.

The increased nitroglycerine demand, starting at t = 5 h 47 min of the first episode, indicates that MAP would have exceeded 100 mmHg, in absence of closed-loop controlled nitroglycerine. Further support for this are the witnessed MAP increases during the time of syringe change, marked a in Fig. 6, and throughout the second 12 h episode (grey), following termination of closed-loop controlled drug administration.

The MAP stayed within the recommended range of 65–100 mmHg 61% of the time, and in the range 65–110 mmHg 98% of the time, during the 12 h of closed-loop control, and 1% of the time during the subsequent 12 h

in absence of closed-loop control. A histogram showing the distribution of time-in-range is shown in Fig. 8. The average nitroglycerine infusion rate during the 12 h episode under closed-loop control was 0.17 g/kg/min.

As seen in Fig. 9, normal blood gas values were maintained throughout the experiment.

4 Discussion

The results support the thesis that simple modelling and control techniques are adequate maintaining normotension in the studied porcine BD model. The sufficiency of low-order linear models is supported by the fits shown in Fig. 5, and indirectly by the control performance of Fig. 6, which additionally indicates sufficiency of using a PID controller.

The static gain variations seen for both nitroglycerine (Fig. 4) and noradrenaline [10] response models has been met by tuning the controller in a robust, or conservative, fashion, at the cost of degraded performance (i.e., error responsiveness). An alternative to this paradigm is *adaptation*, where the static gain parameter is estimated online throughout the experiment, and the controller parameters updated accordingly.

Another finding is the spontaneous arterial pressure increase, witnessed in the experiments underlying this and previous [10] work. Whether this phenomenon is specific to our porcine BD model, should be investigated. Closed-loop control of a vasodilator, such as nitroglycerine, is motivated regardless, as it can eliminate the potentially harmful episode of severe hypertension, associated with the catecholamine storm.

As with concept studies, this work is limited by sample size. While the utilized controller tuning technique is capable of handling model sets as well as models with unstructured uncertainty [19], it remains to characterize the underlying individual variability. A related question is exactly how well (individual and population) porcine models translate to human physiology.

Funding The research was funded by VINNOVA (Grant 2016-01909), and Hans-Gabriel och Alice Trolle-Wachtmeisters Stiftelse för Medicinsk Forskning (Grant Number N/A). Authors affiliated with Lund University Department Automatic Control would like to acknowledge the LCCC and ELLIIT research centra.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval The experiments were run under ethic approval M174-15, issued by the *Malmö/Lunds regionala djurförsöksetiska nämnd* (REB).

Research involving with human and animal rights The animals received humane care in compliance with both EU [20] and US [21] guidelines.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

1. European directorate for the quality of medicines and healthcare (EDQM), Newsletter transplant—international figures on donation and transplantation 2015, Council of Europe, Technical Report 2016. Volume 21, ISSN: 2171-4118.
2. Ranasinghe AM, Bonser RS. Endocrine changes in brain death and transplantation. *Best Pract Res Clin Endocrinol Metab.* 2011;25(5):799–912.
3. McKeown DW, Bonser RS, Kellum JA. Management of the heartbeating brain-dead organ donor. *Br J Anaesth.* 2012;108(1):96–107.
4. Council of Europe, Guide to the quality and safety of organs for transplantation. 6th ed. Strasbourg, France: European directorate for the quality of medicines & healthcare (EDQM). 2016. ISBN: 978-92-871-8287-6.
5. The [Swedish] national board of health and welfare (Socialstyrelsen). Socialstyrelsens föreskrifter om donation och tillvaratagande av vävnader och av celler, Technical Report. 2009, document SOSFS 2009:30.
6. Steen S, Ingemansson R, Eriksson L, Pierre L, Algotsson L, Wierup P, Liao Q, Eyjolfsson A, Gustafsson R, Sjöberg T. First human transplantation of a nonacceptable donor lung after reconditioning ex vivo. *Ann Thorac Surg.* 2007;83:2191–5.
7. Steen S, Sjöberg T, Liao Q, Bozovic G, Wohlfart B. Pharmacological normalization of circulation after acute brain death. *Acta Anaesthesiol Scand.* 2012;56(8):1006–12.
8. Steen S, Paskevicius A, Liao Q, Sjöberg T. Safe orthopic transplantation of hearts harvested 24 hours after brain death and preserved for 24 hours. *Scand Cardiovasc J.* 2016;50(3):193–200.
9. Novitzky D, Wicomb WN, Rose AG, Cooper DKC, Reichart B. Pathophysiology of pulmonary edema following experimental brain death in the chacma baboon. *Ann Thorac Surg.* 1987;43(3):288–94.
10. Soltesz K, Sturk C, Paskevicius A, Liao Q, Qin G, Sjöberg T, Steen S. Closed-loop prevention of hypotension in the heartbeating brain-dead porcine model. *Trans Biomed Eng.* 2016. doi:10.1109/TBME.2016.2602228.
11. Dumont GA, Ansermino JM. Closed-loop control of anesthesia: a primer for anesthesiologists. *Anesth Analg.* 2013;117(5):1130–8.
12. Guen M, Liu N, Chazot T, Fischler M. Closed-loop anesthesia. *Minerva Anesthesiol.* 2016;582(5):573–81.
13. Rinehart J, Liu N, Alexander B, Cannesson M. Closed-loop systems in anesthesia: is there a potential for closed-loop fluid management and hemodynamics optimization? *Anesth Analg.* 2012;114(1):130–43.
14. Rinehart J, Ma M, Calderon MD, Cannesson M. Feasibility of automated titration of vasopressor infusions using a novel closed-loop controller. *J Clin Monit Comput.* 2017. doi:10.1007/s10877-017-9981-6.

15. Bahn E, Wu WD, Rinehart J. Principles of pharmacologic hemodynamic management and closed-loop systems. *Best Pract Res Clin Endocrinol Metab.* 2014;28(4):453–62.
16. Chang KT, Min KT. Progress in molecular biology and translational science. Animal models of human disease. *Prog Mol Biol Transl Sci.* 2011;100:xv.
17. Hill NS, Antman EM, Green LH, Alpert JS. Intravenous nitroglycerine. a review of pharmacology, indications, therapeutic effects and complications. *Chest.* 1981;79(1):69–76.
18. Åström KJ, Hägglund T. Advanced PID control. Research Triangle Park, NC 22709: ISA The instrumentation, Systems, and Automation Society; 2006. ISBN: 978-1-556-17942-6.
19. Soltesz K, van Heusden K, Hast M, Ansermino JM, Dumont GA. A synthesis method for automatic handling of inter-patient variability in closed-loop anesthesia. In *IEEE American Control Conference (ACC): Boston, USA; 2016*, pp. 4877–82.
20. The European parliament On the protection of animals used for scientific purpose, Council of Europe, Technical Reports; 2010, directive 2010/63/EU.
21. National research council of the national academies. Guide for the care and use of laboratory animals. Institute for laboratory animal research: Technical Reports; 2011. ISBN 978-0-309-15401-7.