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Author manuscript Br J Haematol. Author manuscript; available in PMC 2018 May 10.

Published in final edited form as:

Br J Haematol. 2015 June ; 169(6): 851–858. doi:10.1111/bjh.13383.

# Incidence and clinical features of extramedullary multiple myeloma in patients who underwent stem cell transplantation

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# Summary

Extramedullary disease (EMD), defined as an infiltrate of clonal plasma cells at an anatomic site distant from the bone marrow, is an uncommon manifestation of multiple myeloma. Six hundred and sixty-three consecutive patients with multiple myeloma who underwent stem cell transplantation between January 2005 and December 2011 were assessed for the presence of EMD. A cohort of 55 patients with biopsy-proven EMD was identified, comprising 8.3% of the total study population. EMD was present at the time of diagnosis in 14.5% of cases and at the time of relapse in 76% of patients. The most common EMD presentations at relapse were liver involvement and pleural effusions. EMD specimens had high expression of CD44 (92%) and moderate expression of CXCR4. The median overall survival from time of myeloma diagnosis was 4.1 years (95% confidence interval: 3.1, 5.1) and the median overall survival from time of EMD diagnosis was 1.3 years (95% confidence interval: 0.8, 2.3). This report demonstrates that the incidence of EMD has not increased with the introduction of novel agents and stem cell

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**Competing interests:** the authors have no competing interests

Authorship contribution

Mathew Weinstock: performed the research, designed the research, analysed the data and wrote the paper.

Yosra Aljawai: performed the research, designed the research, analysed the data and wrote the paper.

Elizabeth A. Morgan: performed the research, analysed the data and wrote the paper.

Jacob Laubach: designed the research and wrote the paper

Muriel Gannon: performed the research

Aldo M. Roccaro: designed the research and analysed the data

Cindy Varga: performed the research

Constantine S. Mitsiades: analysed the data and wrote the paper

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Irene M. Ghobrial: performed the research, designed the research, analysed the data and wrote the paper.

transplantation. The most common EMD presentations in the relapsed setting were liver and pleural fluid. The presence of CD44 and CXCR4 expression may represent new markers of EMD that warrant further investigation.

#### Keywords

Multiple myeloma; extramedullary; Novel agents; stem cell transplantation; CD44

# Introduction

Multiple myeloma (MM), a malignancy of terminally differentiated monoclonal plasma cells, continues to afflict a substantial proportion of the population, with an estimated incidence of 21,700 new cases diagnosed per year in the United States (Howlader *et al.* 2011). Although MM remains incurable, new therapeutic advances over the past 20 years have greatly improved clinical response rates, event-free survival and overall survival in myeloma (Rajkumar *et al.* 2006, Dimopoulos *et al.* 2007, Richardson *et al.* 2005, Attal *et al.* 1996).

Monoclonal plasma cells, which give rise to the clinical phenotype of MM, most often remain localized to the bone marrow environment. However, there exists a subset of patients with myeloma in whom pathogenic plasma cells can be found at distant anatomical sites, such as the liver, kidney, pleura, breast, testes, skin and meninges, among other tissues. These patients with extramedullary disease (EMD), strictly defined as a clonal plasmacytic infiltrate at an anatomic site distant from the bone marrow or adjacent soft tissue, appear to account for 6–7.5% of the total myeloma population in small series, and tend to have an unfavourable prognosis relative to MM with marrow-only disease (Bartel *et al.* 2009, Usmani *et al.* 2012).

At present, there are limited data regarding the basic characteristics of EMD, including incidence, prevalence, clinical characteristics, laboratory features and response to novel therapies. Several previous series investigating EMD have been hampered by definitional inconsistencies, as some authors have deemed EMD to reflect any extension of plasma cells from the marrow to adjacent soft tissues, while others have strictly defined it as the proliferation of plasma cells at anatomical sites physically separated from the bone marrow. For example, in the largest series of EMD to date (Varettoni *et al.* 2010) EMD was defined as extension from the marrow to the adjacent soft tissues in 85% of cases studied, while in recent analyses, EMD has been defined more narrowly, as evidence of plasma cells at distant locations (Short *et al.* 2011).

We have previously proposed that EMD should be defined strictly as the presence of an infiltrate of clonal plasma cells at an anatomic site distant from the bone marrow or adjacent soft tissue in a patient with underlying MM (Weinstock & Ghobrial 2012). Adherence to this definition will aid in the ongoing study of EMD and will permit accurate analysis of this entity's basic epidemiological, clinical, pathogenic, immunophenotypic, cytogenetic and prognostic features.

Here, we report an analysis of EMD incidence, laboratory features and response to therapy among a group of patients with MM who underwent autologous or allogeneic stem cell transplantation at a single, large, academic medical centre in the United States.

# Methods

Six-hundred and sixty-three consecutive patients with MM who were treated with either autologous or allogeneic haematopoietic stem cell transplantation at the Dana-Farber Cancer Institute (DFCI) between January 2005 and December 2011 were analysed for the presence or absence of EMD, as well as their laboratory characteristics, specific treatment regimens and response to therapy. Approval for this protocol was obtained from DFCI and was in accordance with the Declaration of Helsinki. All patients were treated with novel therapeutic agents, including thalidomide, lenalidomide or bortezomib. This cohort was chosen in order to most accurately reflect the current state of myeloma therapy, in which novel agents are routinely used in tandem with stem cell transplantation.

The diagnosis of plasma cell neoplasm was rendered on tissue or cytology sections as part of routine clinical care in accordance with the 2008 World Health Organization Classification system (McKenna & Kroft 2008). EMD relapse was defined as pathological or radiological evidence of EMD at any time following the initial diagnosis of MM. In accordance with the strict definitional criteria noted above, those patients with pathological or radiological evidence of neoplastic plasma cells in the soft tissues adjacent to the axial skeleton, including the epidural space, paraspinal soft tissue and calvarium, were deemed to have locally-advanced myeloma, but not EMD. Patients with plasma cell leukaemia were specifically excluded from this analysis.

Immunohistochemical studies, performed as part of this analysis, utilized the following antibodies: anti-CD138 (Mouse mAb, MI15, Dako, Carpinteria, CA), anti-CXCR4 (Rabbit mAb, UMB2, Abcam, Cambridge, MA), anti-CD56 (Mouse mAb, 123C3, Dako), anti-CD44 (Rat mAb, IM7, eBioscience, San Diego, CA), and anti-CCR6 (Mouse mAb, 53103, R&D Systems, Minneapolis, MN).

Statistical analysis: Patient characteristics were summarized using descriptive statistics. Kaplan Meier methods were used to estimate survival from the time of myeloma diagnosis and time from EMD diagnosis. We used R version 3.0.2 and the survival package for the analysis (http://www.R-project.org/).

# Results

#### Baseline demographics and characteristics of EMD presentation

Of the entire study cohort of 663 patients who underwent autologous or allogeneic stem cell transplantation for MM at a single, large academic medical centre in the United States between January 2005 and December 2011, 55 were found to have EMD at any time during their disease course (8.3%). The cohort of EMD patients showed a male predominance (63.6%). At the time of MM diagnosis, the cohort of EMD patients had a median haemoglobin of 117 g/l, a median creatinine of 88.4 µmol/l, median calcium of 2.4 mmol/l,

median albumin of 39 g/l, median LDH of 183.5 us/l, and median beta-2 microglobulin of 2.79 mg/l (Table I).

The median age at the time of diagnosis of EMD was 52 years (range 34–66). The median haemoglobin was 104 g/l and median LDH elevation was 283.5 u/l at the time of EMD diagnosis . Fifty-three % had immunoglobin G (IgG) heavy chain restriction and 21.8% had immunoglobin A (IgA) heavy chain restriction; 40.0% had lambda light chain restriction and 60.0% had kappa light chain restriction.

EMD was present at the time of MM diagnosis in 8/55 of these patients (14.5% of all EMD cases). Another 5/55 (9%) patients had EMD at the time of diagnosis and at relapse and 42/55 (76%) patients developed EMD at the time of disease relapse.

The most common locations for EMD at the time of diagnosis of MM were the head and neck soft tissue (31.6%), abdomen (26.3%), chest (21.1%) and central nervous system (12%) (Table II) and included cervical lymph nodes and oropharynx involvement. Involvement in the abdomen included myeloma involvement in the pancreas, peritoneum, kidney and ileum.

The most common locations for EMD at relapse were the abdomen (40%) and the chest (23.9%). The most common sites of involvement were lung and pleural effusions (16%) followed by liver involvement in 15% of the total EMD sites of disease involvement (Table II, Figures 1, 2).

#### Cytogenetic and Molecular characteristics

Cytogenetic analysis of clonal plasma cells in the bone marrow was available for 29/55 (52%) of the patients with EMD at the time of MM diagnosis. Of these, 15/29 had normal cytogenetics (51.7%) and 14/29 had abnormal cytogenetics (48.3%). Hyperdiploidy occurred in 4/14 (28.5%) cases. The most common cytogenetic abnormalities included deletion 13q or monosomy 13 that were present in 9/14 (64%) of cases. *IGH* rearrangements occurred in 6 cases (43%). Two cases (14%) presented with t(11;14), 1q amplification occurred in 1 case (7%) and 17p deletion occurred in 1 case (7%).

Immunohistochemical analysis was available for 11 patients with EMD, and 13 total samples were analysed. These specimens were obtained from various anatomical sites, including the liver, chest wall, abdomen, breast, oral mucosa, pleural fluid, brain and maxillary sinus. Results of immunohistochemical staining confirmed the presence of CD138-positive plasma cells in all examined EMD specimens. Twelve of the 13 EMD biopsy sites were strongly positive for CD44 (92.3%, 90% confidence interval [CI]: 86.4–99.6), 5/13 of the EMD biopsy sites were positive for CXCR4 (38.5%, 90% CI: 16.6–64.5) and 5 for CD56 staining (38.5%, 90% CI 16.6–64.5) (Table III, Figure 3).

#### Therapeutic interventions

The 55 EMD patients were treated with a median of 4 different treatment regimens prior to the development of EMD and a median of 5 total treatment regimens (Table IV). All 55 patients underwent autologous stem cell transplantation and 15 also underwent allogeneic

stem cell transplantation (27.3%). The median age at autologous stem cell transplantation was 54.2 years (range 35–69). The most commonly used combination therapy was dexamethasone/thalidomide (45.5%). Other common regimens included RVD (lenalidomide, bortezomib, dexamethasone; 41.8%), maintenance lenalidomide (36.3%), VD (bortezomib/ dexamethasone; 27.3%) and lenalidomide/dexamethasone (25.4%).

#### Survival analysis

At a median follow-up from time of MM diagnosis of 8.8 years, 41 patients have died. The median overall survival from time of myeloma diagnosis was 4.1 years, (95% CI: 3.1, 5.1), Figure 4A. The median follow-up time from EMD diagnosis was 4.4 years. The median overall survival from time of EMD diagnosis was 1.3 years (95% CI: 0.8,2.3), Figure 4B.

# Discussion

It has been known for many years that EMD portends a worse prognosis relative to marrowlocalized MM (Blade *et al.* 1996, Blade *et al.* 1994). Several characteristic laboratory, cytogenetic and immunophenotypic features of EMD have been identified in small series (Blade *et al* 2011). However, all studies of EMD that have been published to date have either been small, limited largely to the era prior to the introduction of novel immunomodulatory therapies for MM or have been hampered by inconsistencies in the definition of EMD itself. Our cohort represents a large series of patients with EMD that adheres to a strict definition of biopsy-proven EMD and spans the time of novel therapeutic agents and stem cell transplantation.

Previous series have suggested that EMD may be present in 15–20% of MM cases at the time of diagnosis, and another 15% during the course of their disease (Blade *et al.* 1996, Blade *et al.* 1994). However, these early studies have been limited to patients with rare and more aggressive MM phenotypes (such as less than 40 years of age at presentation or IgD myeloma), which may be inherently and artificially enriched for a higher proportion of cases of EMD. Other studies of EMD (Varettoni *et al.* 2010) may also overestimate the incidence of EMD because of an overly inclusive definition of this clinical entity.

Our series of EMD patients was selected by using a strict definition of EMD at anatomical sites that were non-contiguous with the bone marrow cavity, and included only those patients with biopsy-proven clonal plasma cell infiltrates. From this series, we determined an EMD incidence of 8.3% among all MM patients. This is consistent with several smaller, recent studies, which indicate an EMD incidence of 6–7.5% at the time of diagnosis (Short *et al.* 2011, Bartel *et al.* 2009).

Other smaller studies have suggested that patients with EMD may share characteristic laboratory features relative to patients with marrow-localized MM, including lower haemoglobin levels, higher LDH levels and increased rates of thrombocytopenia (Barlogie *et al.* 1989, Usmani *et al.* 2012). Our cohort of 55 patients presented with mild anaemia and mildly elevated LDH at the time of MM diagnosis and at the time of EMD development, suggesting that neither the haemoglobin level nor the LDH level is a reliable predictor of

MM patients who may already harbour EMD or will proceed to develop EMD during the course of their disease.

Interestingly, the presentation of EMD at the time of diagnosis was mostly in the head and neck area. Lung and pleural effusions as well as abdominal involvement, such as pancreatic or renal involvement, occurred in some cases. However, liver involvement did not occur in any cases at the time of diagnosis. This is in contrast with cases presenting after prior therapeutic interventions. In the relapsed EMD setting, the most common site of involvement was the liver followed by pleural fluid. These presentations may indicate specific tropism or homing of extramedullary myeloma clones that are more prone to trafficking to these sites. Further investigations into the mechanisms of specific tropism of these aggressive myeloma clones are required. Clinically, attention to liver involvement in patients with myeloma should be considered and routine abdominal imaging may be considered to detect those lesions before significant tumour progression and liver dysfunction occurs.

Several bone marrow cytogenetic abnormalities identified in this EMD cohort, particularly deletion of 13q, have been observed in another small, retrospective study of patients with EMD (Rasche *et al.* 2012). Our cytogenetic data were obtained from patients at the time of myeloma diagnosis and not at the time of development of EMD. Cytogenetic or fluorescence *in situ* hybridization studies from the EMD samples or the bone marrow were not available at the time of EMD diagnosis . Given the recent data of clonal evolution and heterogeneity, it would not be surprising to identify specific subclones that have a higher propensity for development of EMD. Therefore, further studies are necessary to refine and differentiate the genomic profile of EMD from that of marrow-localized MM.

The immunophenotypic characteristics of EMD have, as yet, remained poorly defined. A previous small series of seven patients with extramedullary MM reported that CD56, a membrane glycoprotein in the immunoglobulin family, is variably expressed in plasma cells resident in the bone marrow, but is absent in extramedullary plasma cells (Dahl *et al.* 2002). Several reports have therefore suggested that CD56 down-regulation may have a pathogenic role in the development of EMD (Blade *et al* 2011). In our series, 5 of the 13 analysed EMD specimens (38.5%) were found to be positive for CD56. Although the sample size is small, our findings suggest that CD56 down-regulation may not be as closely linked to EMD pathogenesis as previously described.

Other immunohistochemical studies have reported up-regulation of the cell adhesion molecule CD44 in EMD (Dahl *et al.* 2002). CD44 mediates binding of tumour cells to stroma and regulates interleukin-6 production (Stauder *et al.* 1996). Prior studies have shown variable expression of CD44 with about 73% of the cases showing 20% or more positive expression in one study (Zheng *et al*, 2013). Higher expression was present in patients with recurrent or more aggressive disease. In addition, expression of variant isoforms containing the 9v domain was shown to be associated with an advanced stage and progressive disease with shorter overall survival in MM (Stauder *et al.* 1996).

In our series we confirm that CD44 appears to be reliably over-expressed in EMD specimens (92.3%). However, we did not have matching bone marrow samples for all the EMD samples to compare the relative expression of CD44 in plasma cells present in the bone marrow compared to those present in the EMD sites. Therefore, future studies are required to determine the role of CD44 expression and the different isoforms in the localization of malignant plasma cells in EMD sites.

Additionally, in our series, we note that an increased number of EMD specimens (38.5%) were positive for the presence of CXCR4. This stands in contrast to pre-clinical murine data, in which CXCR4 and other chemokine receptors have been found to be down-regulated in the setting of EMD (Stessman *et al.* 2013). Further study of the immunohistochemical characteristics of EMD will be necessary for complete elucidation of the mechanisms that underpin the "metastatic" transition of marrow-localized MM into EMD.

Previous studies have suggested that the introduction of novel therapeutic agents may have contributed to an increased incidence of EMD over the past two decades (Raanani *et al.* 2007). However, it is unclear whether this observation is related to improved methods of radiological detection, increased overall survival of the entire MM population in general or greater awareness of EMD as a distinct clinical entity. Our series of cases gathered entirely within the era of routine stem cell transplantation, proteasome inhibition and immunomodulatory therapy, suggests that the incidence of EMD has remained fairly consistent in the time of modern MM therapy.

The survival of the patients in this series remains poor, with a median overall survival of 4.1 years and a median overall survival of 1.3 years from the time of diagnosis of EMD. This indicates an urgent need for the development of better therapeutic modalities that target this unique subset of patients.

In summary, EMD is an uncommon, but by no means rare, manifestation of MM. This cohort of patients 55 patients represents a large group of patients with EMD who were treated in the era of novel therapeutic strategies and autologous or allogeneic haematopoietic stem cell transplantation for MM at a single academic medical centre. The presence of surface markers of CXCR4 and CD44 may represent new biological markers for EMD but need further confirmation. Further study of these patients' molecular characteristics and responses to therapy will be necessary so that this distinct disease entity can continue to be characterized.

# Acknowledgments

This work was supported by Leukemia and Lymphoma Society and NIH/NCI R01CA181683

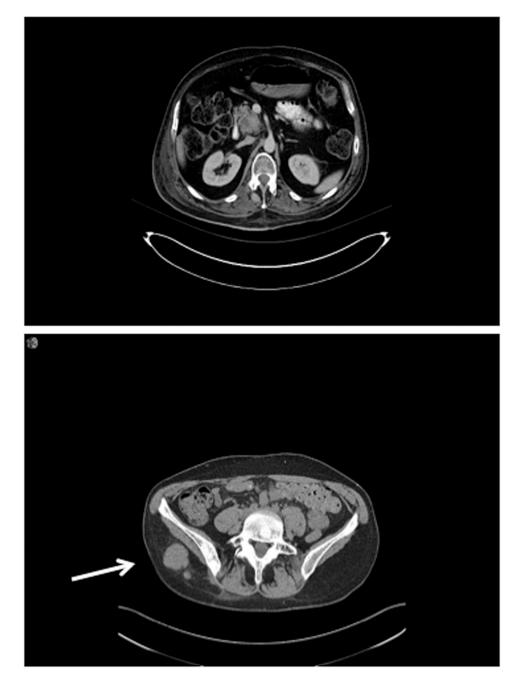
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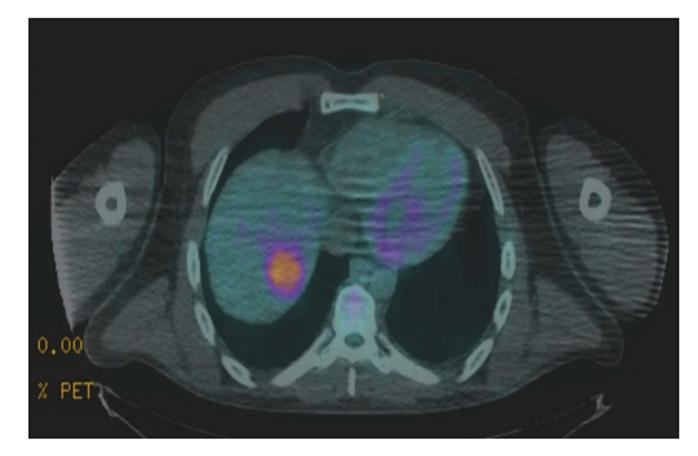
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poor survival in multiple myeloma patients treated with bortezomib. Leukemia. 2013 Oct; 27(10): 2075–7. [PubMed: 23728080]

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**Figure 1.** Right gluteal soft tissue mass, with biopsy-proven plasma cells.

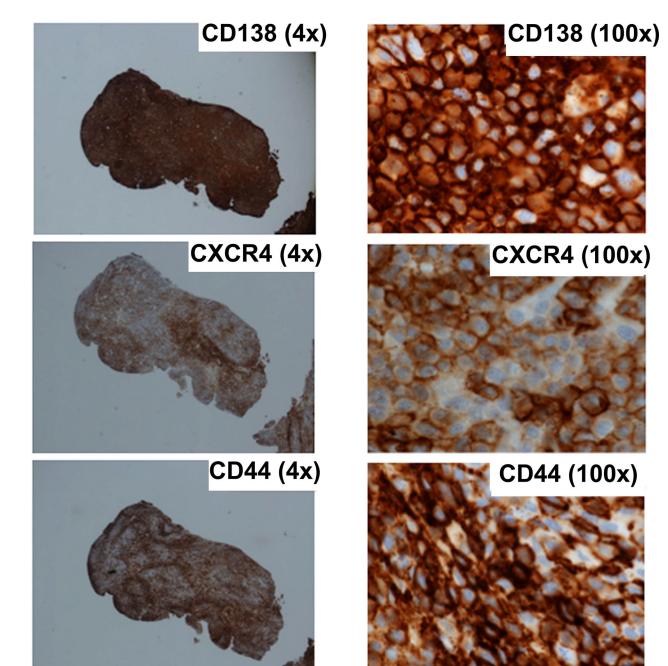


# Figure 2.

Positron emission tomography (PET) scan with liver mass demonstrating liver involvement with extramedullary myeloma.

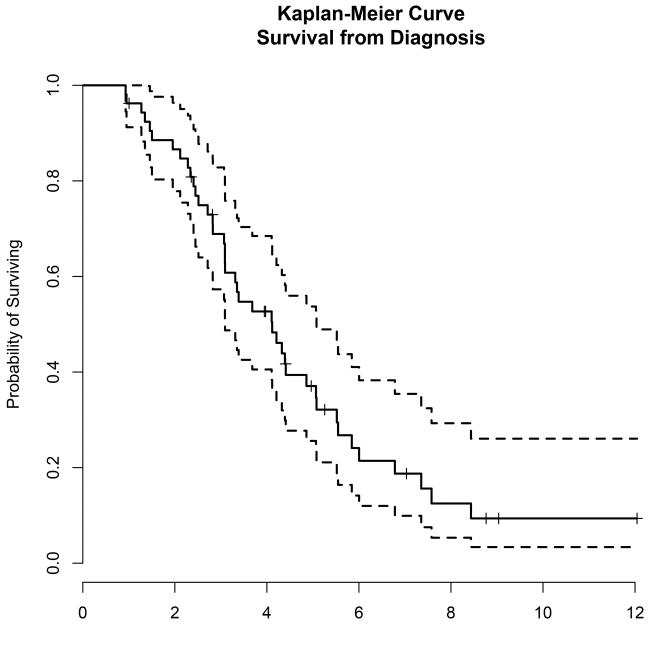
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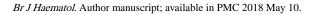


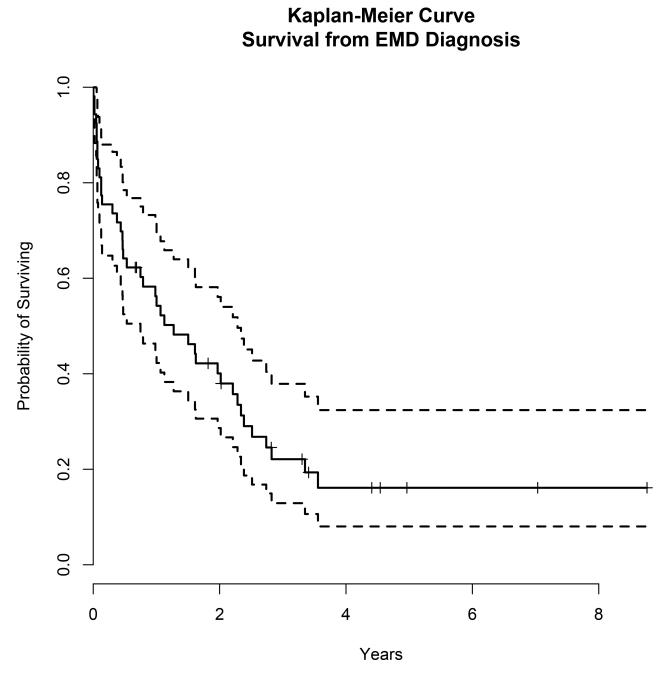
### Figure 3.

Immunohistochemistry straining of CD138, CD44 and CXCR4 of an extramedullary myeloma sample showing strong positive expression of CD138, CD44 and CXCR4. CD56 and CCR6 were negative in this sample (not shown).



Years





#### Figure 4.

Survival data. A) The median overall survival from time of myeloma diagnosis was 4.1 years, (95% confidence interval [CI]: 3.1, 5.1). B) The median overall survival from time of extramedullary disease (EMD) diagnosis was 1.3 years (95% CI: 0.8,2.3).

Table I

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Disease Characteristics of EMD patients at Time of MM Diagnosis

					1
	Variable	Z	Ū	% R	Reference
Gender					
	Male	35	9	63.6	
	Female	20	3	36.4	
Ig Heavy Chain		55			
	А	12	2	21.8	
	IJ	29	5	52.7	
	n/a	14	2	25.4	
Ig Light Chain		61			
	۲	22	4	40.0	
	¥	33	9	60.0	
Hb (g/l)		39			
	Median		117		
	Range		65-151	1	132 –167
Creatinine (µmol/l)		4			
	Median		88.4		
	Range		44.2–884	9	61.88-114.92
Calcium (mmol/l)		41			
	Median		2.4		
	Range		0.925-5	2	2.2-2.625
Albumin (g/l)		31			
	Median		39		
	Range		29–50	3	37–54
LDH (u/l)		20			
	Median		183.5		
	Range		105–622	1	107-231
β-2 microglobulin (mg/l)		30			
	Median		2.79		

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% Reference 0.2 - 71-20.8z Variable Range EMD, extramedullary disease; MM, multiple myeloma; Ig, immunoglobulin; Hb, haemoglobin; LDH, lactate dehydrogenase

Table II

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Anatomical Location of EMD

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nx15.3Sinus15.3Cranial Nerve15.3Cranial Nerve15.3Parotid15Larynx15.3Central Nervous System15.3Ista15.3Dura15.3Dura15.3Braincord15.315.3Braincord15.3Spinal Fluid115.3Spinal Fluid115.3Spinal Fluid15.3Spinal Fluid1Spinal Fluid1Spinal Fluid15.3Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid1Spinal Fluid	Lip	-	5.3	Gingiva	7	2.2
1   5.3   Cranial Nerve     Parotid   Parotid     Parotid   Larynx     Parotid   Cervical Lymph Node     I Nervous System   3   15.9   Central Nervous System     rood   1   5.3   Dura     cord   1   5.3   Brain     domal Return   1   Cerebrospinal Fluid     mod Pleural effusion   Netorical white matter     mod Pleural effusion   Mediastinum     mod Pleural effusion   Mediastinum     mod Pleural effusion   Mediastinum     mod Pleural effusion   Mediastinum     mod Pleural ef	Pharynx	-	5.3	Sinus	-	1.1
Panotid     Larynx     Larynx     Larynx     System     J Nervous System     J Nervous System     J S-3     I Nervous System     J S-3     Servical Lymph Node     J S-3     I Nervous System     rospinal Fluid     I S-3     Pura     Cord     I S-3     Pura     Pura     Spinal Fluid     Spinal extra-axial mass     I S-3     Pura     Spinal extra-axial mass     I S-3     Pura     Pura<	Tonsil	-	5.3	Cranial Nerve	-	1:1
Larynx     Cervical Lymph Node     al Nervous System   3   15.9   Central Nervous System     rospinal Fluid   1   5.3   Dura     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Spinal canal     Leptomeninges   1   Leptomeninges     Altra   21.1   Lung & pleural effusion     and pleural effusion   4   21.1   Lung & pleural effusion     Altra   21.1   Lung & pleural effusion     And pleural effusion   3   Arathar and     Altra   21.1   Lung & pleural effusion     And pleural effusion   Arathar and   Arathar and     Altra   21.1   Lung & pleural effusion     And pleural effusion   Arathar and   Arathar and     And pleural effusion   Arathar and   Arathar and     And pleural effusion   Arathar and <t< td=""><td></td><td></td><td></td><td>Parotid</td><td>-</td><td>1.1</td></t<>				Parotid	-	1.1
al Nervous System   Cervical Lymph Node     al Nervous System   3   15.9   Central Nervous System     rospinal Fluid   1   5.3   Dura     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   Subortical white matter     Mad pleural effusion   4   21.1   Lung & pleural effusion     Mad pleural effusion   4   21.1   Lung & pleural effusion     Mad pleural effusion   3   Mediastinum     Mad pleural effusion   3   Mediastinum     Mad pleural effusion   3   Mediastinum     Mad pleural effusion   3   3     Mad pleural effusion   3   3     Mad pleural effusion <t< td=""><td></td><td></td><td></td><td>Larynx</td><td>-</td><td>1.1</td></t<>				Larynx	-	1.1
al Nervous System   3   15.9   Central Nervous System     rospinal Fluid   1   5.3   Dura     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Brain     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5.3   Frontal extra-axial mass     Cord   1   5   Spinal canal     Cord   1   5   Shortcal     Cord   1   Spinal canal   End     Cord   1   Spinal canal   End     Cord   1   Leptomeninges   End     Cord   1   Subcortical white matter     Cord   4   21.1   Lung & pleural effusion     And pleural effusion   4   21.1   Lung & pleural effusion     Cord   2   2   Mediastinum     Cord   1   Lung & pleural effusion     Cord   1   Lung & pleural effusion     Cord   1   Subscritcal arch     Cord   1   Subscritcal arch     Cord </td <td></td> <td></td> <td></td> <td>Cervical Lymph Node</td> <td>-</td> <td>1.1</td>				Cervical Lymph Node	-	1.1
rospinal Fluid15.3DuraCord15.3Brain.15.3Frontal extra-axial mass15.3Frontal extra-axial mass22Cerebrospinal Fluid22Leptomeninges22Spinal canal21Leptomeninges22Subcortical white matter331.1Chest3421.1Lung & pleural effusion3421.1Lung & pleural effusion421.1Lung & pleural effusion55.3Abdomen	Central Nervous System	e	15.9	Central Nervous System	13	14.1
Cord15.3Brain15.3Frontal extra-axial mass15.3Frontal extra-axial mass15Cerebrospinal Fluid1Cerebrospinal Fluid1Spinal canal1Leptomeninges1Leptomeninges1Thecal1Thecal1Subcortical white matter121.1Lung & pleural effusion121.1Lung & pleural effusion121.1Lung & pleural effusion121.1Lung & pleural effusion11Subcortical arch1Addiastinum1Atalary Lymph Node1Abdomen226.3Abdomen226.3Abdomen	Cerebrospinal Fluid	-	5.3	Dura	4	4.3
1   5.3   Frontal extra-axial mass     Cerebrospinal Fluid   Cerebrospinal Fluid     Fondal extra-axial mass   Spinal canal     Fondal extra-axial extra   Spinal canal     Fondal extra   Subcortical white matter     and pleural effusion   4   21.1     And pleural effusion   Mediastinum     Fondal extra   Breast     Fondal extra   Axillary Lymph Node     Fondal extra   Stat	Spinal Cord	-	5.3	Brain	7	2.2
Cerebrospinal Fluid     Cerebrospinal Fluid     Spinal canal     Spinal canal     Leptomeninges     Leptomeninges     Thecal     Thecal     Thecal     Thecal     Thecal     Leptomeninges     Thecal     Leptomeninges     Thecal     Leptomeninges     Leptomenin	Brain	-	5.3	Frontal extra-axial mass	2	2.2
Spinal canal     Spinal canal     Leptomeninges     Leptomeninges     Thecal     Thecal     Subcortical white matter     Subcortical white matter     Label				Cerebrospinal Fluid	1	1.1
Leptomeninges     Thecal     Thecal     Thecal     Beboral     4     21.1     Chest     Pleural effusion     4     21.1     Lung & pleural effusion     Pleural effusion     Pleural effusion     A     21.1     Lung & pleural effusion     Pleural effusion     A     21.1     Lung & pleural effusion     Pleural effusion     A     21.1     Lung & pleural effusion     Pleural effusion     Pleural effusion     A     21.1     Lung & pleural effusion     Pleural effusion     Pleural effusion     Pleural effusion     Pleural effusion     A     A     A     A     A     A     A     A     A     A     A     A     A     A     A     B     A     A     A     A     A     A     A <t< td=""><td></td><td></td><td></td><td>Spinal canal</td><td>1</td><td>1.1</td></t<>				Spinal canal	1	1.1
Thecal     Thecal     Subcortical white matter     Subcortical white matter     1   21.1     Chest     Pleural of fusion     Nediastinum     Pleural of fusion     Pleural of fusion <td></td> <td></td> <td></td> <td>Leptomeninges</td> <td>1</td> <td>1.1</td>				Leptomeninges	1	1.1
Subcortical white matter     4   21.1     Chest     pleural effusion     4   21.1     Lung & pleural effusion     Mediastinum     Breast     Fracheal arch     A stillary Lymph Node     5   26.3				Thecal	1	1.1
4   21.1   Chest     pleural effusion   4   21.1   Lung & pleural effusion     notation   31.1   Lung & pleural effusion     notation   31.1   Lung & pleural effusion     notation   31.1   Lung & pleural effusion     notation   1   21.1     notation   1   Nediastinum				Subcortical white matter	1	1.1
pleural effusion   4   21.1   Lung & pleural effusion     Mediastinum     Breast     Tracheal arch     Axillary Lymph Node     5   26.3   Abdomen	Chest	4	21.1	Chest	22	23.9
Mediastinum Breast Tracheal arch Axillary Lymph Node 5 26.3 Abdomen	Lung and pleural effusion	4	21.1	Lung & pleural effusion	15	16.3
Breast Tracheal arch Axillary Lymph Node 5 26.3 Abdomen				Mediastinum	3	3.3
Tracheal arch Axillary Lymph Node 5 26.3 Abdomen				Breast	2	2.2
Axillary Lymph Node 5 26.3 Abdomen				Tracheal arch	1	1.1
5 26.3 Abdomen				Axillary Lymph Node	1	1.1
	Abdomen	S	26.3	Abdomen	40	43.5

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N $\%$ Pancreas1 $5.3$ LiverPancreas1 $5.3$ Peritoneal surfacePeritoneum1 $5.3$ Reinoreal surfaceKidney1 $5.3$ KidneySpleen1 $5.3$ MesenterySpleen1 $5.3$ Reinoreal surfaceInum1 $5.3$ MesenterySpleen1 $5.3$ Reinoreal surfaceInum1 $5.3$ Reinoreal surface <t< th=""><th>N       %       N         1       5.3       Liver       14         m       1       5.3       Peritoneal surface       7         m       1       5.3       Peritoneal surface       7         m       1       5.3       Ridney       6       7         m       1       5.3       Mesentery       7       7       7         m       5       Mesentery       1       7<!--</th--><th>EMD at Diagnosis of MM</th><th>of MM</th><th></th><th>EMD at Relapse</th><th>a</th><th></th></th></t<>	N       %       N         1       5.3       Liver       14         m       1       5.3       Peritoneal surface       7         m       1       5.3       Peritoneal surface       7         m       1       5.3       Ridney       6       7         m       1       5.3       Mesentery       7       7       7         m       5       Mesentery       1       7 </th <th>EMD at Diagnosis of MM</th> <th>of MM</th> <th></th> <th>EMD at Relapse</th> <th>a</th> <th></th>	EMD at Diagnosis of MM	of MM		EMD at Relapse	a	
1   5.3     m   1   5.3     1   5.3     1   5.3     1   5.3	I     5.3     Liver     14       mm     1     5.3     Peritoneal surface     7       m     1     5.3     Ridney     6       m     1     5.3     Kidney     6       m     1     5.3     Mesentery     7       m     1     5.3     Mesentery     3       m     5.3     Mesentery     3     3       m     5.3     Penetry     3     3       m     5.3     Penetry     3     3       m     Spleen     3     3     3     3       m     Spleen     3     3     3     3       m     1     1     1     3     3     3       m     1     1     1     1     3     3     3     3     3     3		Z	%		z	%
eum 1 5.3 1 5.3 1 5.3 1 5.3	eum       1       5.3       Peritoneal surface       7         1       5.3       Kidney       6         1       5.3       Kidney       6         1       5.3       Mesentery       5       6         1       5.3       Mesentery       5       7         1       5.3       Pancreas       2       2         1       5.3       Pancreas       2       2         1       Spleen       2       2       2         1       Para-aortic Lymph Node       1       1       1         1       Ureter       Ureter       1       1         1       Ureter       Buttock       1       1         1       Buttock       1       1       1         1       S.3       Other Soft Tissue       5       1	Pancreas	1	5.3	Liver	14	15.1
1 5.3 1 5.3 1 5.3	1     5.3     Kidney     6       1     5.3     Mesentery     3       1     5.3     Mesentery     3       1     5.3     Pancreas     3       1     5.3     Pancreas     2       2     Spleen     2       3     Spleen     1	Peritoneum	-	5.3	Peritoneal surface	٢	7.5
1 5.3 1 5.3	1       5.3       Mesentery       3         1       5.3       Pancreas       2         1       5.3       Pancreas       2         2       Spleen       2       2         2       Spleen       2       2         2       Para-aortic Lymph Node       1       1         2       Pelvic Lymph Node       1       1         3       Ureter       1       1         4       Lueter       1       1         4       Hatortebral       1       1         5       Other Soft Tissue       5       5	Kidney	-	5.3	Kidney	9	6.5
1 5.3	1       5.3       Pancreas       2         Spleen       2       Spleen       2         Para-aortic Lymph Node       1       1       1         Pelvic Lymph Node       1       1       1         Para-aortic Lymph Node       1       1       1	Spleen	-	5.3	Mesentery	ю	3.2
Spleen Para-aortic Lymph Node Pelvic Lymph Node Ureter Iliac Lymph Node Buttock Paravertebral	Spleen   2     Para-aortic Lymph Node   1     Pelvic Lymph Node   1     Ureter   1     Inac Lymph Node   1     Paravertebrah   1     Paravertebrah   1     Inac Lymph Node   1	Ileum	-	5.3	Pancreas	5	2.2
Para-aortic Lymph Node       Pelvic Lymph Node       Ureter       Iliac Lymph Node       Buttock       Paravertebral	Para-aortic Lymph Node   1     Pelvic Lymph Node   1     Ureter   1     Ureter   1     Buttock   1     Paravertebral   1     Softer Soft Tissue   5				Spleen	5	2.2
Pelvic Lymph Node Ureter Iliac Lymph Node Buttock Paravertebral	Pelvic Lymph Node   1     Ureter   1     Ureter   1     Buttock   1     Paravertebral   1     S.3   Other Soft Tissue   5				Para-aortic Lymph Node	-	1.1
Ureter Iliac Lymph Node Buttock Paravertebral	Ureter   1     Iliac Lymph Node   1     Buttock   1     Paravertebral   1     1   5.3   Other Soft Tissue   5				Pelvic Lymph Node	1	1.1
Iliac Lymph Node Buttock Paravertebral	Iliac Lymph Node   1     Buttock   1     Paravertebral   1     1   5.3   Other Soft Tissue   5				Ureter	1	1.1
Buttock Paravertebral	Buttock   1     Paravertebral   1     1   5.3   Other Soft Tissue   5				Iliac Lymph Node	1	1.1
Paravertebral	Paravertebral   1     1   5.3   Other Soft Tissue   5				Buttock	1	1.1
	1 5.3 Other Soft Tissue 5				Paravertebral	1	1.1
1 5.3		Other Soft Tissue	1	5.3	Other Soft Tissue	w	5.4

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EMD, extramedullary disease; MM, multiple myeloma

# Table III

Immunohistochemical Characteristics of a subset of 11 MM patients with EMD.

Patient	Location	CD138	CXCR4	CD56	CD44	CCR6
1	Liver	POS	NEG	NEG	POS	NEG
1	Chest Wall	POS	NEG	NEG	POS	NEG
7	Liver	POS	NEG	NEG	POS	NEG
3	Liver	POS	POS	NEG	POS	NEG
4	Chest Wall	POS	NEG	POS	POS	NEG
2 2	Chest Wall	POS	NEG	POS	POS	NEG
6	Abdominal mass	POS	NEG	NEG	POS	NEG
7	Abdominal mass	POS	POS	NEG	POS	NEG
8	Breast	POS	POS	NEG	NEG	NEG
6	Oral mucosa	POS	NEG	POS	POS	NEG
6	Pleural fluid	POS	NEG	POS	POS	NEG
10	Brain	POS	POS	NEG	POS	NEG
11	Maxillary sinus	POS	POS	POS	POS	NEG
Percenta	Percentage Positive (%)	100	38.46	38.46	92.31	0

Samples from 2 biopsy sites were analysed for Patients 1 and 9.

Br J Haematol. Author manuscript; available in PMC 2018 May 10.

EMD, extramedullary disease; MM, multiple myeloma

#### Table IV

# Treatment of patients with EMD

	Ν	%	Median
Autologous SCT	55	100	
Allogeneic SCT	15	27.3	
Dexamethasone/Thalidomide	25	45.5	
RVD	23	41.8	
Maintenance Lenalidomide	20	36.3	
Dexamethasone/Bortezomib	15	27.3	
Dexamethasone/Lenalidomide	14	25.4	
Lines of Therapy prior to EMD (n)			4
Total lines of Therapy (n)			5
Age at Autologous Transplant, years			54.2 (range 35-69)

EMD, extramedullary disease; SCT, stem cell transplantation; RVD, lenalidomide, bortezomib, dexamethasone