

Ambiguous roles of innate lymphoid cells in chronic development of liver diseases

Yue Shen, Jing Li, Si-Qi Wang, Wei Jiang

Yue Shen, Jing Li, Si-Qi Wang, Wei Jiang, Department of Gastroenterology, Zhongshan Hospital, Fudan University, Shanghai 200032, China

Jing Li, Department of Gastroenterology, Tongji Hospital, Tongji University, Shanghai 200000, China

ORCID number: Yue Shen (0000-0002-4771-9114); Jing Li (0000-0002-6815-495X); Si-Qi Wang (0000-0002-1905-1137); Wei Jiang (0000-0002-9354-6699).

Author contributions: Jiang W conceived this topic and organized the manuscript; Shen Y wrote the first draft of the manuscript and drew the figures; Li J and Wang SQ contributed to manuscript revision; all authors have read and approved the submitted version and are accountable for all aspects of the work.

Supported by National Nature Science Foundation of China, No. 81670541; and National Science and Technology Major Project, No. 2013ZX10002004 and No. 2017ZX10203202.

Conflict-of-interest statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Wei Jiang, MD, PhD, Academic Research, Chief Doctor, Professor, Department of Gastroenterology, Zhongshan Hospital, Fudan University, 180 Fenglin Road, Xuhui District, Shanghai 200032, China. jiang.wei@zs-hospital.sh.cn
Telephone: +86-21-64041990-2424
Fax: +86-21-64432583

Received: March 29, 2018

Peer-review started: March 30, 2018

First decision: April 19, 2018

Revised: April 25, 2018

Accepted: May 6, 2018

Article in press: May 6, 2018

Published online: May 14, 2018

Abstract

Innate lymphoid cells (ILCs) are defined as a distinct arm of innate immunity. According to their profile of secreted cytokines and lineage-specific transcriptional factors, ILCs can be categorized into the following three groups: group 1 ILCs (including natural killer (NK) cells and ILC1s) are dependent on T-bet and can produce interferon- γ ; group 2 ILCs (ILC2s) are dependent on GATA3 and can produce type 2 cytokines, including interleukin (IL)-5 and IL-13; and, group 3 ILCs (including lymphoid tissue-like cells and ILC3s) are dependent on ROR γ t and can produce IL-22 and IL-17. Collaborative with adaptive immunity, ILCs are highly reactive innate effectors that promptly orchestrate immunity, inflammation and tissue repair. Dysregulation of ILCs might result in inflammatory disorders. Evidence regarding the function of intrahepatic ILCs is emerging from longitudinal studies of inflammatory liver diseases wherein they exert both physiological and pathological functions, including immune homeostasis, defenses and surveillance. Their overall effect on the liver depends on the balance of their proinflammatory and antiinflammatory populations, specific microenvironment and stages of immune responses. Here, we review the current data about ILCs in chronic liver disease progression, to reveal their roles in different stages as well as to discuss their therapeutic potency as intervention targets.

Key words: Innate lymphoid cells; Chronic liver disease; Hepatitis; Liver fibrosis; Liver cancer

© The Author(s) 2018. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Innate lymphoid cells (ILCs), mirroring both the phenotypes and functions of T cells, have been defined as a distinct arm of innate immunity. There has been a marked increase in the studies investigating the dysregulation of ILCs in chronic liver pathologies. This manuscript presents a comprehensive overview of the state of ILCs, including the fundamental concepts as well as summarizing their ambiguous roles in the progression of the chronic liver hepatitis, fibrosis and carcinoma. It also provides an insight into the current research gaps and indicates the therapeutic potency and development direction of future research of ILCs.

Shen Y, Li J, Wang SQ, Jiang W. Ambiguous roles of innate lymphoid cells in chronic development of liver diseases. *World J Gastroenterol* 2018; 24(18): 1962-1977 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v24/i18/1962.htm> DOI: <http://dx.doi.org/10.3748/wjg.v24.i18.1962>

INTRODUCTION

Liver diseases usually evolve from inflammation to fibrosis, with cirrhosis manifested in the advanced stage, and serving as a well-determined major risk factor of liver cancer. Liver disease remains a major health problem, affecting millions of people worldwide. Ongoing chronic inflammation in the liver induced by infections, hepatotoxic drugs, autoimmunity, alcohol abuse or toxins will result in liver fibrosis, which is the consequence of an irreversible, progressive condition occurring in most types of chronic liver diseases and characterized by excessive deposition of extracellular matrix (ECM) proteins, mainly composed of collagen^[1]. The situation where ECM formation is prompted by activated hepatic stellate cells (HSCs) outweighs the collagen degradation by matrix metalloproteases (MMPs) and will lead to structural distortion of the normal liver tissue and functional impairment; furthermore, it is associated with an increased risk of cirrhosis, portal hypertension and subsequent liver failure and liver cancer^[2-4]. Tremendous efforts have been made to design strategies which could prevent liver disease progression.

Innate lymphoid cells (ILCs) are a recently identified group of mononuclear hematopoietic cells which encompass not only cytotoxic natural killer (NK) cells but also noncytotoxic ILCs, and are involved in immunity and tissue remodeling. Though characterized with lymphoid morphology, ILCs lack the rearranged antigen receptors and are defined as cell lineage marker-negative (Lin⁻) cells^[5]. ILCs mirror both the phenotypes and functions of T cells, for which noncytotoxic ILCs have been proposed as the innate counterparts of CD4⁺ T helper (Th) cells, whereas NK cells are considered

to be the innate equivalents of CD8⁺ cytotoxic T (Tc) cells^[6].

Group1 ILCs comprise both Eomes-dependent NK cells and T-bet-dependent ILC1s^[7]. Upon stimulation by interleukin (IL)-12, IL-15 and IL-18 derived from both myeloid cells and nonhematopoietic cells, the ILC1s can produce Th1 cell-associated cytokines, such as interferon (IFN)- γ and tumor necrosis factor (TNF)- α , which play critical roles in clearing intracellular pathogens^[8,9]. Distinguished from ILC1s, NK cells depend on Eomes to develop and exert their cytotoxic functions by secreting granzymes and perforin^[10,11]. Group 2 ILCs (ILC2s), being dependent on GATA3 and ROR- α and respond to epithelium-derived cytokines IL-25, IL-33 and thymic stroma lymphopoeitin (TSLP), can produce Th2 effector cytokines (IL-4, IL-5, IL-9, IL-13 and amphiregulin), thus playing a critical role in antihelminth immunity and allergic inflammation as well as tissue repair^[12-15]. Finally, group 3 ILCs are dependent on ROR γ t and mainly respond to myeloid cell-derived IL-1 β and IL-23. These ILCs include lymphoid tissue-like (LTi) cells and ILC3s, which can produce IL-22, IL-17, granulocyte macrophage colony-stimulating factor and TNF, thus showing great significance in antibacterial immunity^[16-18] (Figure 1).

Dysregulation of ILCs can cause severe inflammation and injury in gut^[19], lung^[20], skin^[21] and liver^[22]. During the past 5 years, a growing number of studies have investigated the roles of ILCs in inflammatory, fibrotic and cancerous liver diseases^[23-27]. Herein, we summarize the present knowledge of ILCs to reveal their complicated and versatile effects and the underlying mechanism in chronic liver diseases, in order to provide perspectives of new therapeutic strategies.

LIVER INFLAMMATION

Group 1 ILCs

There are two distinct NK populations in murine liver, CD49a⁻CD49b⁺ and CD49a⁺CD49b⁻ cells. The CD49a⁻CD49b⁺ subset represents conventional (c)NK cells, which circulate in the blood. The CD49a⁺CD49b⁻ subset has ever been considered as tissue-resident (tr)NK cells or ILC1s in previous studies, which are beside dendritic cells (DCs) localizing in the sinusoids of the portal area^[28,29]. Both cNK and trNK cells express natural cytotoxicity receptors and require IL-15 signaling for their development. Compared to cNK cells, trNK cells have relatively lower expression of CD11b, Ly49 receptors, CD43 and KLRG1, but higher expression of CXCR3 and CXCR6, which is a chemokine receptor for CXCL16 responsible for the enrichment of natural killer T (NKT) cells in the liver and can provide intravascular immune surveillance^[30,31]. In parallel, human livers contain CD56^{bright} and CD56^{dim} (accounting for 90%) NK cells, respectively representing the equivalents of murine cNK and trNK cells. With respect to function, CD56^{bright} NK cells are prominent cytokine producers,

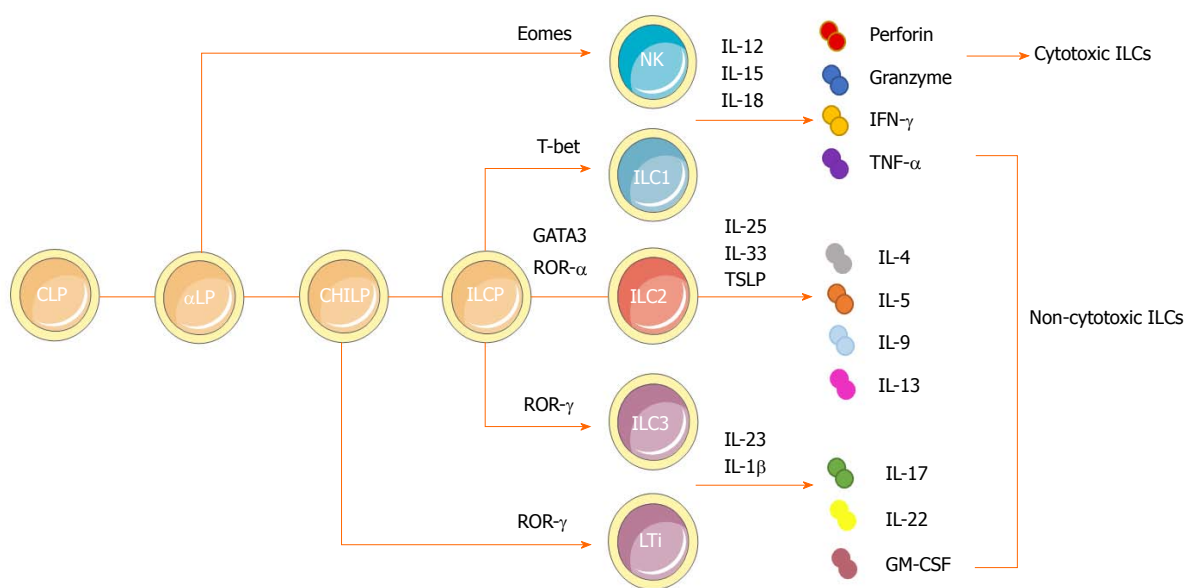


Figure 1 Developmental pathways and classification of innate lymphoid cells. ILCs are derived from a CLP. With the same phenotype as CLP as well as expressing $\alpha 4\beta 7$ integrin, α LP gives rise to cytotoxic NK cells and differentiates into a CHILP, which gives rise to all noncytotoxic ILCs. The transcription factor PLZF further divides the progeny of CHILPs into a PLZF⁺ ILCs that are restricted to ILCs except LTi cells and PLZF-independent LTi cells. Group 1 ILCs comprise both Eomes-dependent NK cells and T-bet-dependent ILC1s. They could produce IFN- γ and TNF- α in response to the stimulation by IL-12, IL-15 and IL-18. NK cells can also secrete granzymes and perforin to exert cytotoxic functions. Dependent on GATA3 and ROR- α as well as responsive to cytokines IL-25, IL-33 and TSLP, group 2 ILCs could produce Th2 effector cytokines (IL-4, IL-5, IL-9 and IL-13 and amphiregulin). Group 3 ILCs encompass both ROR γ -dependent LTi cells and ILC3s. They can produce IL-22, IL-17 and GM-CSF, mainly in response to IL-1 β and IL-23. α LP: α -lymphoid progenitor; CHILP: Common helper-like innate lymphoid progenitor; CLP: Common lymphoid progenitor; GM-CSF: Granulocyte macrophage colony-stimulating factor; IL: Interleukin; ILC: Innate lymphoid cell; ILCP: Innate lymphoid cell precursor; INF: Interferon; LTi: Lymphoid tissue-like; NK: Natural killer; PLZF: Promyeloid leukemia zinc finger; Th: T helper; TNF: Tumor necrosis factor; TSLP: Thymic stroma lymphopoietin.

whereas CD56^{dim} NK cells are efficient killers^[32].

Liver is an immune-tolerant organ predisposing for chronic infections of certain clinically significant pathogens. As an organ with predominant innate immunity, the liver is enriched with NK cells, which account for 25%-50% of total intrahepatic lymphocytes and are responsible for killing transformed cells and viruses. The cytotoxicity of NK cells is regulated by both cytokines and surface receptors^[33,34]. IFN- γ is one of the most prominent cytokines derived from NK cells to exert antiviral, antifibrotic and antitumorigenic effects. Furthermore, it has been demonstrated that cNK rather than trNK cell-derived IFN- γ promotes Th1 polarization and secondary CD8⁺ cytotoxic lymphocyte (CTL) responses, the major effectors for clearance of hepatic B virus (HBV) in transgenic HBV mouse models^[35]. Meanwhile, the interactions of NK cells with hepatocytes *via* the NKG2A inhibitory receptor could prime DCs to induce CD4⁺CD25⁺ regulatory T cells (Tregs), which will in turn up-regulate the expression of NKG2A on NK cells *via* IL-10 production, thus impairing the antiviral ability of NK cells^[36,37].

In the pathogenesis of chronic HBV infection (CHB), ILC1s have potential proinflammatory effects that mirror Th1 cells in adaptive immunity exactly. First, in patients with CHB, liver injury has been significantly associated with enhanced ILC1s' response, as reflected by markedly elevated levels of T-bet, IFN- γ and IL-12 signaling. Besides, decreased ILC1-produced IFN- γ has

been found to have a connection with the telbivudine-induced alleviation of liver injury in CHB patients^[23]. These results could be explained by the study of Krueger *et al.*^[38], in which it was demonstrated that CD49a⁺ ILC1s could inhibit expression of CXCL9, which was further required for robust accumulation of IFN- γ ⁺CD49b⁺ NK cells during the early phase of adenovirus infection. In this way, ILC1s played a role in maintaining the liver as a tolerogenic site as a result of increased expression of NKG2A receptors compared with NK cells, which would further suppress the activation of liver CD103⁺ DCs, thus interrupting the priming of antigen-specific, antiviral CD8⁺ T cells and the clearance of virus. The mechanism was found to be the same in hepatitis C virus infection for which NKG2A^{-/-} patients showed resistance^[39,40]. To conclude, ILC1s help to maintain the tolerance of liver in normal conditions, and blockage of NKG2A signaling to generate potent anti-viral CD8⁺ T cell responses required for the elimination of persistent liver pathogens may prove to be a novel therapeutic strategy (Figure 2A).

Group 2 ILCs

IL-33 belongs to the IL-1 superfamily, which is alarmin secreted by epithelial cells upon cellular stress and tissue damage. Upon binding to its specific heterodimeric receptor which comprises the ST2 and IL-1 receptor accessory protein, IL-33 is able to induce strong expression of Th2-like cytokines, thus balancing the Th1

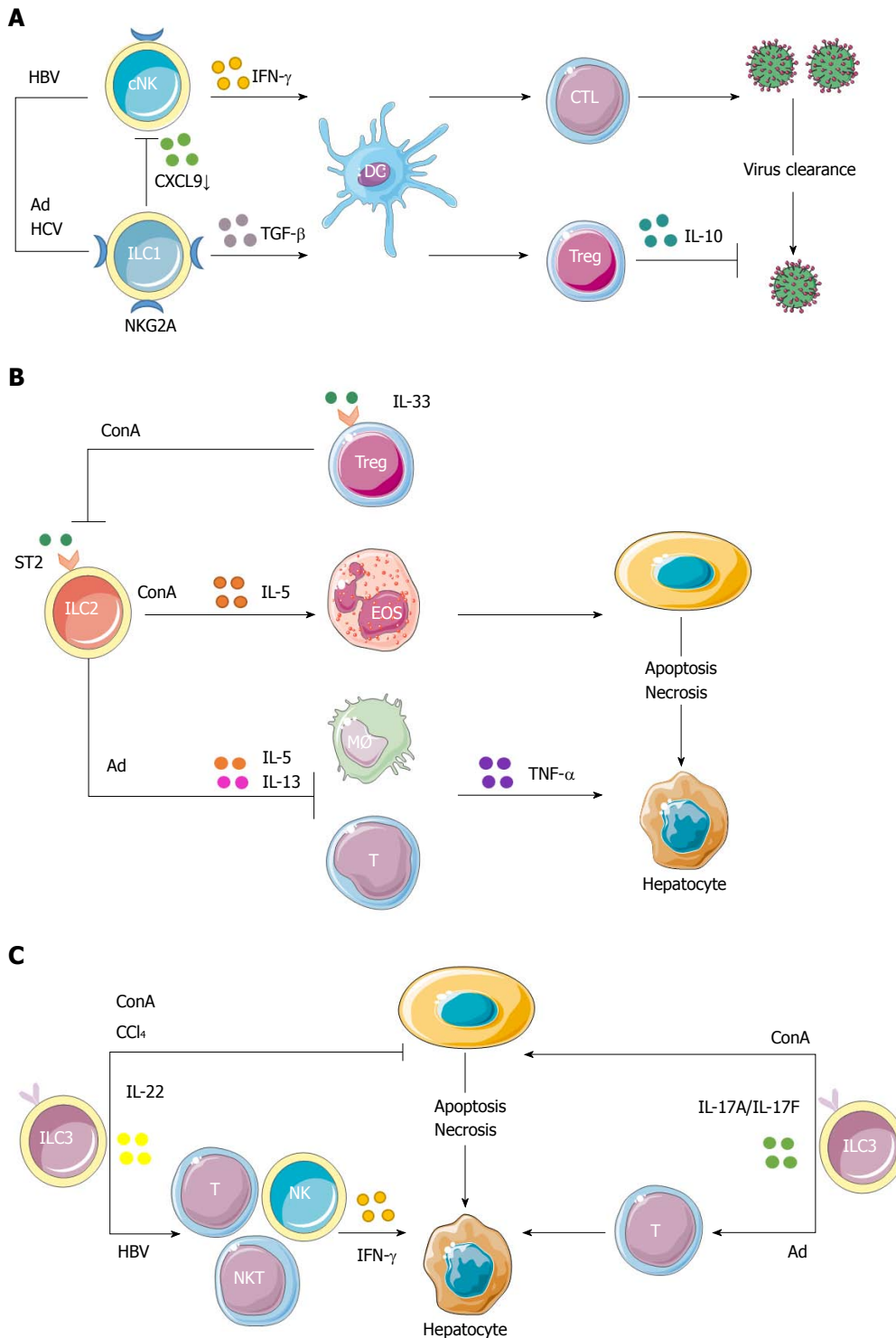


Figure 2 Protective and pathogenic roles of innate lymphoid cells in hepatic inflammation. A: cNK cells could produce IFN- γ to enhance the priming of CD8 $^+$ T cells to clear HBV. The interactions of NK cells with hepatocytes via NKG2A inhibitory receptor could prime DCs to induce CD4 $^+$ CD25 $^+$ Tregs, which would in turn up-regulate the expression of NKG2A on NK cells via IL-10 production, thus impairing the antiviral ability of NK cells. Because of increased expression of NKG2A on ILC1s in hepatic Ad as well as hepatitis C virus infection, ILC1s play a role in maintaining the liver as a tolerogenic site by inhibiting CXCL9 expression, which is required for the accumulation of cNK cells. This would further impair the activation of liver CD103 $^+$ DCs, thus interrupting the proliferation of virus-specific CD8 $^+$ T cells and the clearance of virus; B: In ConA-induced immune hepatitis, hepatic ILC2s could amplify inflammation through the expression of IL-5 to recruit eosinophils in response to IL-33 released upon liver tissue damage. The inflammatory activity of endogenous ILC2s in immune-mediated hepatitis might be regulated by IL-33-elicited ST2 $^+$ Tregs. Besides, in Ad-induced viral hepatitis, a strong expression of ILC2s was induced by IL-33 to exert a protective role through down-regulation of the hepatotoxic cytokine TNF- α in T cells and macrophages. Both the proinflammatory and protective roles of ILC2s in hepatitis are part of IL-33 action; C: In immune hepatitis, ILC3-derived IL-22 has a protective role in ConA- and carbon tetrachloride-induced hepatitis, while IL-17 plays a pathological role in ConA-induced hepatitis. Besides, Notch-mediated IL-22 is an important mediator of the inflammatory response in HBV infection, being responsible for the recruitment of antigen-nonspecific inflammatory cells into the liver and subsequent liver injury. In Ad-induced acute hepatitis, the IL-17A/F signaling is critical for adaptive T response and is responsible for affected lymphocyte infiltration and hepatic inflammation. Ad: Adenovirus; cNK: Conventional natural killer; ConA: Concanavalin A; DC: Dendritic cell; HBV: Hepatitis B virus; IL: Interleukin; ILC: Innate lymphoid cell; NK: Natural killer; Tregs: T regulatory cells.

immune response^[41,42]. Epithelial cells, hepatocytes and HSCs have been reported as the main sources of IL-33 in the liver^[43,44]. The expression of IL-33 shows significant connection with chronic liver diseases, such as persistent viral infection^[45-47], liver fibrosis^[43] and liver failure^[48].

It has been demonstrated that through interaction with ST2, IL-33 induces production of the inflammatory cytokine IFN- γ , as well as Fas-FasL interaction between hepatocytes and NKT cells in concanavalin A (ConA)-induced hepatitis, which is a well-established murine model of T-cell mediated hepatitis, resembling the pathology of immune-mediated hepatitis in humans^[49,50]. Neumann *et al*^[24] further demonstrated that IL-33-elicited ILC2s were also involved in the pathogenic process of murine immune-mediated hepatitis. In response to the release of IL-33 upon liver tissue damage induced by CD4⁺ T cells, hepatic ILC2s were found to be poised to produce type 2 cytokines, including IL-13 and IL-5. Recruitment of eosinophils induced by IL-5 could be one mechanism by which ILC2 amplifies inflammation during immune hepatitis, and IL-13 was indicated to have a prominent role in chronic diseases^[51]. Exogenous IL-33-elicited hepatic ILC2s appear to aggravate immune-mediated hepatitis, while the inflammatory activities of endogenous ILC2s might be regulated by IL-33-elicited ST2⁺ Tregs, which showed strong expansion in immune-mediated hepatitis as well. These findings are consistent with those of the previous study that revealed IL-33/ST2 axis to exert a protective role in ConA-induced hepatitis by preventing Th1 and Th17 cell-mediated hepatic immune responses, promoting IL-4 production of CD4⁺ liver-infiltrating T cells, elevating the total number of CD4⁺Foxp3⁺ Tregs together with affecting the expression of apoptotic or antiapoptotic proteins^[52]. These results suggest that the proinflammatory role of ILC2s in immune-mediated hepatitis is part of the action mechanism of IL-33. Multiple modules of the immune response should be taken into consideration when investigating its overall protective or pathogenic effect on the liver.

What's more, the IL33/ST2 axis has also been shown to play a crucial role in driving antiviral CD8⁺ and CD4⁺ T cell responses^[53,54]. On one hand, Liang *et al*^[55] demonstrated that, as a newly discovered damage-associated molecular pattern molecule, IL-33 can promote innate IFN- γ production by $\gamma\delta$ T cells and NK cells. It could also modulate DC responses to enhance the plurifunctionality of antiviral T cells in lymphocytic choriomeningitis virus-induced hepatitis in mice, while it was also further demonstrated that ILC2s were not involved in this process^[56]. On the other hand, IL-33 was able to directly engage multiple arms of immune mechanisms to mediate potent hepatoprotective effects in adenovirus-induced hepatitis, wherein strong CTL, CD4⁺ Th and B lymphocyte responses share common characteristics with a number of hepatotropic viruses, including hepatitis A virus, HBV, cytomegalovirus, herpes simplex and Epstein-Barr virus. It significantly inhibited

the expression of TNF- α in T cells and macrophages and induced a strong expression of IL-5- or IL-13-expressing-Lin⁻ nuocytes to further down-modulate the hepatotoxic cytokine TNF- α ^[57]. An increased number of Lin⁻13⁺ or Lin⁻5⁺ cells were found in the livers of Lin⁻ cells adoptively-transferred mice. Though the serum level of alanine aminotransferase and hepatic TNF- α presented a downward trend, there was no statistical significance compared with control groups^[58]. These results also suggest that the potential protective role of ILC2s in viral hepatitis might only be one facet of the complex mechanisms of IL-33, but this still remains to be further elucidated (Figure 2B).

Group 3 ILCs

Dependent on ROR γ t and IL-7, ILC3s induce the production of IL-17 and IL-22 upon stimulation by IL-23 and IL-1 β . IL-22 is a member of the IL-10 cytokine family and has a crucial role in inflammation, immune surveillance and tissue homeostasis. In the inflammatory context, IL-22 has both proinflammatory and protective properties^[59,60]. The proinflammatory nature of IL-22 has been shown in mouse models with diseases such as psoriasis and rheumatoid arthritis^[61,62]. In contrast, the protective role has been shown in inflammatory bowel disease^[63], hepatitis^[64] and pathogenic bacterial infection^[65,66]. Hepatocytes are important targets of IL-22, for it can induce the expression of acute-phase proteins, several antiapoptotic proteins and mitogenic proteins, to protect cells against liver tissue damage^[67-69]. IL-22 can also act on liver stem or progenitor cells, which are important in chronic and severe liver injury^[70].

In ConA-induced acute immune hepatitis, the expression of IL-23 combined with activated Notch signaling resulted in an aryl hydrocarbon receptor (AHR)-dependent production of IL-22, as well as in an ROR γ t-dependent production of IL-17. IL-22 was shown to play a protective role, while IL-17 was shown to be critical for the pathogenesis in liver tissue^[71]. The protective role of IL-22 in hepatitis was consistent with findings of a previous study that identified NKT and T cells, rather than ILC3s, as the main sources of IL-22^[64]. Later, it was confirmed by Abe *et al*^[72] that, combined with the suppression of IFN- γ from NKT cells induced by AHR, IL-22-producing ILC3s were also involved in the protective process in ConA-induced acute hepatic injury, as high IL-22 mRNA levels were found in CD3⁺Sca1⁺Thy1⁺ cells rather than in CD3⁺ T cells after stimulation by IL-23. Besides, the same results were obtained in ROR γ t^{-/-} mice; specifically, there was almost no IL-22 production in the hepatic mononuclear cells of *Ahr*^{-/-} or *Ahr* and *recombination activation gene (RAG)* double-negative mice, thus further suggesting that the major sources of IL-22 were both ROR γ t- and AHR-dependent ILC3s. In addition, the decreased frequency of IL-22-producing ILC3s (Lin⁻ SCA-1⁺ Thy1^{high} ILCs) was consistent with the severity of carbon tetrachloride-induced hepatitis of RAG-2^{-/-}*ROR γ t^{-/-} mice compared with that of RAG-2^{-/-} mice^[72].

Taken together, all the results of these studies considering ROR γ t⁺ hepatic ILC3s in immune-mediated hepatitis suggest their protective roles against liver injury *via* IL-22 production. Compared with IL-22-producing Th17 cells, which can play a protective role against liver injury as well, hepatic ILC3s may be able to act in the early innate immune response stage^[73]. On the contrary, IL-17, another ILC3s-derived cytokine, has shown a pathological role in ConA-induced hepatitis. The overexpression of IL-17A resulted in massive hepatocyte necrosis, and antiIL-17A blockage significantly ameliorated liver injury^[74]. In addition, Lafdil *et al.*^[75] showed that liver injury was alleviated in ConA-induced hepatitis among IL-17-deficient mice.

On the other hand, however, IL-22 was identified as a potent mediator of the inflammatory response in HBV infection, following the recognition of HBV by T cells in the liver^[67,76]. It was further confirmed that inhibition of Notch signaling *in vivo* would lead to decreased ILC22 and LTi4 cells, along with down-regulated expressions of IL-22 and related proinflammatory cytokines and chemokines in the liver. As a result, subsequent liver injury was alleviated due to blockage of the recruitment of antigen-nonspecific inflammatory cells into the liver, without affecting HBV antigen production in HBV infection^[77]. These results suggest the potential proinflammatory role of Notch-mediated IL-22 and provide a new potential therapeutic approach for the treatment of HBV. What's more, intrahepatic early IL-17 was found to be important for activating antigen presenting cells in viral infection, but the sources and regulation of IL-17 surges were not well defined at first^[78]. It was further shown that ILC3s, including a large proportion of NKP46⁻ ILC3s and a small part of the CD4⁺ LTi cells, secreted IL-17A and IL-17F shortly after adenovirus infection, in addition to γ δ T cells. In adenovirus-induced acute hepatitis, the IL-17A/F signaling was found to be critical for adaptive T response and was responsible for affected lymphocyte infiltration and hepatic inflammation, except in viral clearance. The study also revealed the existence of the compensatory IL-17F production for IL-17A deficiency underlying the previous contradictory result that IL-17A deficiency did not appear to thwart T cell activation and liver inflammation^[64,79]. Though there have been studies showing Th17-derived IL-17 causes liver damage by IL-23 activation, the role of ILC3-derived IL-17 remains to be further clarified in chronic infection models, such as for lymphocytic choriomeningitis virus and HBV infections^[80]. To conclude, the downstream effector cytokines of ILC3s may exert both proinflammatory and protective roles according to the specific microenvironment, and more studies are required to clarify their explicit role in liver inflammation (Figure 2C).

LIVER FIBROSIS

Group 1 ILCs

NK cells can directly decrease the proliferation and

activation as well as induce cell cycle arrest of HSCs through IFN- γ ^[81,82]. They can also induce apoptosis of activated HSCs through the TNF-related apoptosis-inducing ligand (TRAIL) and Fas ligand pathways^[83,84]. These interactions between NK cells and HSCs are regulated by NK cell receptors and cytokines. The activation of HSCs in response to hepatocyte damage leads to increased NK cell stimulation and decreased NK cell inhibition. Firstly, increased amounts of retinoic acid derived from activated HSCs was found to be associated with elevated expression of RAE-1, a ligand for the NKG2D-activating receptor. Together with MICA, RAE-1 could promote the killing of activated HSCs by NK cells^[85,86]. The NKP46 and NKP30 activating receptors have also been shown to be involved in the amelioration of liver fibrosis by inducing HSC killing by NK cells, in both humans and mice^[87,88]. Secondly, engagement of Ly49 inhibitory NK cell receptors was found to be reduced by the mechanism of siRNA-mediated silencing, as a result of down-regulated major histocompatibility complex (MHC) class I in activated HSCs^[89,90]. Besides, elevated surface expression of TRAIL in NK cells *via* IFN- α , simultaneous with increased expression of TRAIL receptors, in activated HSCs could also enhance HSC killing by NK cells^[91,92]. Instead, tumor growth factor (TGF)- β down-regulates the surface expression of NKG2D and 2B4 to suppress the antifibrotic role of NK cells^[93]. Whether trNK cells exert a protective role in liver fibrosis remains unclear. As a member of the IFN- γ -producing group 1 ILCs, these cells may contribute to the activation of NK cells by their production of IFN- γ , which is crucial for the antifibrotic roles of NK cells^[81].

Group 2 ILCs

In the study by Marvie *et al.*^[43], the over-expression of IL-33 was shown to be closely associated with hepatic fibrosis, in both human and mouse cases. Besides, type 2 cytokines including IL-4 and IL-13 have been considered as representatives of the most potent fibrogenic factors^[94]. As the major sources of Th2-type cytokines, ILC2s, which also require IL-33 for activation, were proposed as potent profibrogenic factors in hepatic fibrosis^[95]. It has been demonstrated that, in response to ST2-dependent signaling owing to chronic hepatocellular stress and tissue damage, IL-33 release leads to accumulation and activation of IL-13-producing liver resident ILC2s. The downstream effector cytokine IL-13 can further trigger the activation and transdifferentiation of HSCs in an IL-4Ra- and STAT6-dependent manner to induce potent fibrogenic responses, suggesting a pathogenic capacity of ILC2s in the context of the tissue damage response^[22]. In parallel, the study of human liver fibrosis by Forkel *et al.*^[96] has identified primary hepatocytes, HSCs and Kupffer cells as cellular sources of IL-33 and TSLP, which could further potentially cause the accumulation of ILC2s in fibrotic livers following TLR3-activation, as a model for hepatitis C infection. There was also a direct correlation found between the increase in

frequency of intrahepatic ILC2s and the severity of liver fibrosis. The induction of IL-13 by intrahepatic ILC2s in response to IL-33 and TSLP was also confirmed, suggesting the possibility of a similar mechanism in humans and mice^[96]. These results provide an avenue for investigation into the application of serum IL-33 as a possible noninvasive diagnostic biomarker for uncovering early inflammatory and fibrogenic events. Furthermore, targeting ILC2s may represent a novel therapeutic strategy for liver fibrosis treatment.

Group 3 ILCs

As one of the ILC3s' downstream effector cytokines, IL-22 has been shown to promote the survival and proliferation of epithelial cells (*e.g.*, hepatocytes), suggesting its possibility of involvement in liver fibrosis^[64,68]. Upon binding to IL-22R1 and IL-10R2 on HSCs, IL-22 can induce senescence of the HSCs following the activation of a STAT3/SOCS3/p53 signaling axis, which represents an important strategy to ameliorate liver fibrosis^[97]. It could also inhibit HSC apoptosis, without affecting HSC proliferation. By means of enhancement of *in vivo* clearance of senescent HSCs, most probably by NK cells, simultaneous to reduction of released tissue inhibitor of metalloprotease 2 to promote MMP activities and down-regulate the deposition of collagen, the senescence of activated HSCs played an important role in limiting liver fibrosis^[86,98]. The expression of α -smooth muscle actin was also down-regulated in response to IL-22, but this effect was not associated with senescent HSCs^[97]. Besides, elevated systemic IL-22 level - independent of age, liver-related complications, C-reactive protein, creatinine and model for end-stage liver disease score - could be predictive for reduced survival prognosis in patients with liver cirrhosis. Thus, it is possible that systemic IL-22 level could be applied as a negative indicator for evaluating the prognosis of advanced liver cirrhosis^[99]. Though no direct evidence has linked ILC3s with liver fibrosis, ILC3s may exert a protective role since they are the source of IL-22. Nonetheless, considering IL-22 can be produced by Th17 cells as well, it is important to use specific gene knock-out mice to determine which type of cell plays the pivotal role.

In experimental liver fibrosis, upon stimulation of IL-17A, both HSCs and Kupffer cells could produce TGF- β , TNF- α and IL-6 following the activation of STAT3 and nuclear factor- κ B. Accordingly, mouse models with IL-17A and IL-17RA deficiency have displayed reduced liver fibrosis, suggesting a profibrotic role^[100]. In addition, IL-17A can also exert an antifibrotic effect in normal fibroblast cultures directly, by down-regulating the expressions of collagen and connective tissue growth factor, which was shown to be impaired in the isolated primary fibroblasts from patients with scleroderma^[101]. Considering the complicated and versatile effects in fibrosis, it might be necessary to determine the exact roles of such cytokines in different stages of liver fibrosis (Figure 3).

LIVER CANCER

Group 1 ILCs

Considering the potent tumor surveillance properties of NK cells, a group of NK cell-associated genes in hepatocellular carcinoma (HCC) tissues was positively associated with prolonged survival^[102]. Evidence of dysfunction of NK cells in HCC has been observed, as well, suggesting a strong connection between NK cells and HCC progression^[103].

Although there currently is no direct evidence revealing connections between ILC1s and liver tumor immunity, the effects of their secreted cytokines have been extensively investigated, among which IFN- γ was shown to have prominent antiproliferative, antiangiogenic and proapoptotic effects against cancer cells^[104-106]. This cytokine can promote the up-regulation of MHC molecules to induce the priming as well as antigen processing and presentation of professional antigen presenting cells, and has been shown to increase the immunogenicity of tumor cells, thereby enhancing antitumor responses^[107,108]. In addition to promoting the polarization of CD4⁺ T cells into Th1 cells, it can also boost the responses of macrophages, NK cells and CTLs against tumor tissues^[109,110]. TNF- α , another cytokine secreted by ILC1s, can also play an antitumor role by interfering with angiogenesis, cellular growth and migration. Further, it can induce the recruitment of macrophages and DCs, as well as the generation of CTLs, leading to a strong antitumor immune response^[111,112]. Combined with the previously reported research findings, a recent study which showed the NK1.1⁺CD49a⁺CD103⁺ ILC1-like cells could lyse tumor cells, dependent on the activation of granzyme B and TRAIL, in an oncogene-induced cancer model also supports the protective function of ILC1s in antitumor immunity^[113]. However, its protective function can be hampered in cancer patients, as ILC1s of acute myeloid leukemia patients were found to be dramatically impaired in their production of IFN- γ and TNF- α compared to those of healthy control subjects^[114].

It was also demonstrated that both IFN- γ and TNF- α could play ambiguous roles in cancer immunity. The protumor function of IFN- γ involves increased proliferative and antiapoptotic signals, as well as escape of the tumor cells from recognition and cytolysis by CTLs and NK cells^[115]. TNF- α is also involved in tumor formation, growth and spread considering its versatile impacts on the expression of angiogenic and growth factors, cytokines, adhesion receptors and proteases^[111,116,117]. Recently, Gao *et al.*^[118] demonstrated that CD49a⁻CD49b⁺Eomes⁺ NK cells could convert into intermediate CD49a⁺CD49b⁺Eomes⁺ type 1 innate lymphoid cells (intILC1s) and CD49a⁺CD49b⁻Eomes⁻ ILC1s in tumor microenvironment in a TGF- β signaling-dependent manner. Strikingly, distinguishable from the potent tumor surveillance properties of NK cells, intILC1s and ILC1s were incapable of controlling local tumor growth

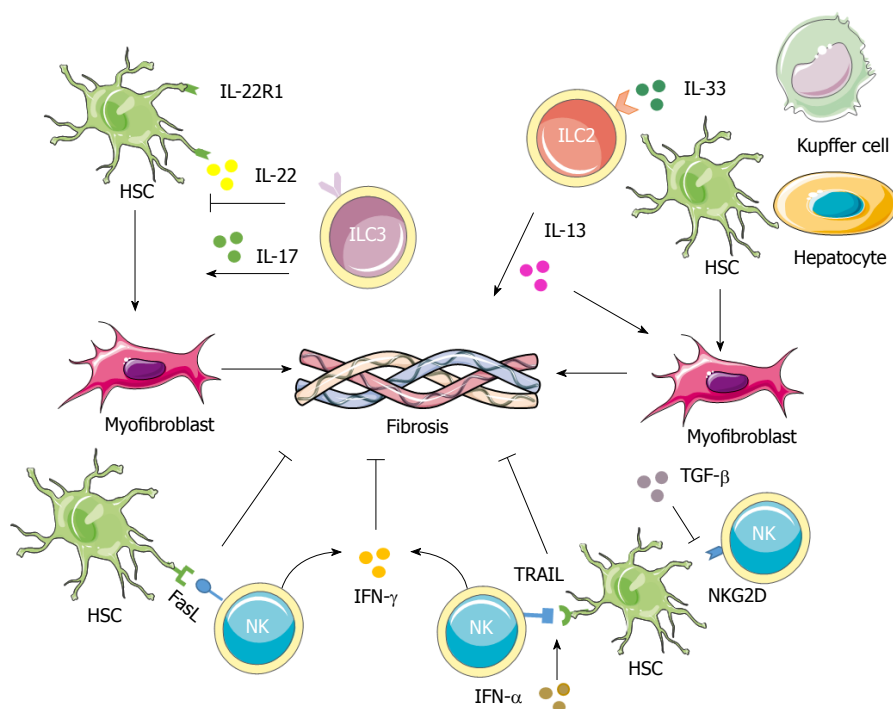


Figure 3 Contributions of innate lymphoid cells in liver fibrosis. NK cells can decrease the proliferation and activation as well as induce cell cycle arrest of HSCs through IFN- γ . They can also induce the apoptosis of activated HSCs through the TRAIL and Fas ligand pathways. The expression of RAE-1, which is the ligand for the NKG2D activating receptor, is increased on activated HSCs, thus promoting killing by NK cells. IFN- α could increase the surface expression of TRAIL on NK cells to enhance HSCs killing by NK cells, while TGF- β down-regulates the surface expression of NKG2D and 2B4 to suppress the antifibrotic role of NK cells. In both human and mouse cases, IL-33 released from hepatocytes, HSCs and Kupffer cells in response to chronic hepatocellular stress leads to the accumulation and activation of IL-13-producing liver resident ILC2s via ST2-dependent signaling. IL-13 further triggers the activation and transdifferentiation of HSCs into myofibroblasts, to induce potent fibrogenic responses. ILC3s play more complicated roles in fibrosis. On one hand, ILC3s exert an antifibrotic effect by inducing fibroblasts senescence through IL-22 signaling and by down-regulating the expressions of collagen and CTGF through IL-17 signaling. On the other hand, IL-17 can promote inflammation and induce activation of fibroblasts, indicating a profibrotic role for the ILC3s. CTGF: Connective tissue growth factor; HSC: Hepatic stellate cell; IL: Interleukin; IFN: Interferon; NK: Natural killer; TGF: Tumor growth factor; TRAIL: TNF-related apoptosis-inducing ligand.

and metastasis, uncovering an unknown mechanism by which tumors can escape surveillance by the innate immune system. This study also provided a new insight into the phenotypic and functional plasticity of tumor group 1 ILCs, while the precious roles and interactions of ILC1s in tumor microenvironment - especially in the liver - still needs to be further elucidated^[118,119].

Group 2 ILCs

Considering the potential profibrogenic properties of ILC2s elicited by IL-33 in hepatic fibrosis under the circumstances of tissue damage, it is possible that these cells are involved in the progression from liver fibrosis to cancer as well. The precise role of ILC2s in carcinogenesis remains unclear; however, it can be supported by the evidence that has emerged from studies addressing factors that trigger their activation and proliferation, as well as their downstream effector molecules.

When it comes to the liver, IL-33 was shown to be involved in the initiation of cancer, based on a previous study wherein increased expressions of Th2 cytokines and hepatic IL1RL1 mRNA encoding ST2 were detected in a subgroup of patients at the time of diagnosis of biliary atresia^[120]. ILC2s were further identified as important mediators of the IL-33-dependent proliferative

response for their production of high levels of IL-13, which in turn promoted cholangiocyte proliferation and epithelial hyperplasia in mice by involving the activation of IL-4R and the downstream target Stat6. Administration of IL-33 with constitutive activation of AKT and Yes-associated protein in biliary epithelium would lead to the development of cholangiocarcinoma, which resembles both the morphological and biochemical features of human disease in a mouse model of experimental carcinogenesis^[121,122]. Thus, the activation of the IL-33/ILC2s/IL-13 circuit may promote epithelial repair, and the disruption of IL-33 or other elements of the paracrine circuit may constitute potential new therapeutic targets against cholangiocarcinoma. Furthermore, the level of IL-33 was found to be increased in patients with HCC as well^[123].

IL-33 can also increase the intratumor accumulation of myeloid-derived suppressor cells (MDSCs), which require arginase and nitric oxide synthase II from IL-13 for their activation^[124,125]. Together with the pro-angiogenesis process, MDSCs can also produce TGF- β to support tumor progression^[126]. Besides, IL-13 can induce the polarization of macrophages to the M2 phenotype, and the production of growth and angiogenic factors to promote tumor initiation, progression and metastasis^[127,128]. It has also been demonstrated that

amphiregulin, another cytokine secreted by ILC2s, could enhance the activities of Tregs *in vivo*, which would further inhibit antitumor immune responses induced by DC vaccination^[129,130].

IL-33 also plays a role in antitumor immune responses *via* effector functions of both CD8⁺ T cells and ILC2s, dependent on its dose^[131,132]. The latter was confirmed in a study by Kim *et al.*^[133], which demonstrated that ILC2s were involved in IL-33-mediated antitumor responses. A massive amount of CXCR2 ligands released from ILC2s interacted with CXCR2 expressed by tumor cells by means of a dysfunctional angiogenesis/hypoxia/reactive oxygen species axis triggered by IL-33, subsequently leading to the apoptosis of active tumor cells^[133]. Taking these results into consideration, ILC2s could exert both immune suppression and antitumor functions according to different tumor microenvironments, while its precious role, especially in the context of human livers, remains to be further confirmed.

Group 3 ILCs

The protective role of ILC3s has been directly revealed for its contribution to the formation of protective tumor-associated tertiary lymphoid structures in non-small cell lung cancer (NSCLC)^[134]. The ILC3s are able to up-regulate adhesion molecules in the tumor microenvironment to enhance leukocyte invasion, and have been characterized as important mediators of the efficacy of a combination therapy of chemotherapy and tumor antigen-targeted monoclonal antibodies^[135]. Though there is no direct evidence linking ILC3s and liver cancer, the connection can be inferred according to their role in colorectal cancer, as well as the ambiguous roles of their downstream effector cytokines, including IL-22 and IL-17.

It has been demonstrated that ILC3-derived IL-22 is crucial for promoting bacterial inflammation-induced colorectal cancer in *Rag*^{-/-} mice through the activation of epithelial cells *via* STAT3 signaling^[136]. Furthermore, deficiency of soluble IL-22 binding protein (IL-22BP) secreted by immature DCs was found to be associated with increased colitis-associated colon cancer due to the aberrant proliferation induced by IL-22. However, IL-22 was demonstrated as important for colonic epithelial cell repair in the early stage of colitis, using the same model; in particular, *IL22*^{-/-} mice were shown to have enhanced cancer development^[137]. These results suggest that IL-22 is crucial for regulating intestinal tissue repair during the peak of damage, while prolonged IL-22 in the recovery phase would be expected to favor tumorigenesis.

Paralleling the dual effects of IL-22 on tumorigenesis of colitis-associated colon cancer, IL-22 has also been reported to induce tissue regeneration or tumorigenesis and metastasis in the liver. Firstly, characterized with the protective role of hepatocyte proliferation and tissue regeneration during hepatitis and after hepatectomy, the functions of IL-22 may be exploited

in liver cancer, as suggested by the significant up-regulation of IL-22 detected in human HCC tumor-infiltrated leukocytes^[70,138,139]. Besides, there is a positive correlation between IL-22 expression and the oncogenesis and staging of tumors, according to the finding that both IL-23 and IL-22BP are highly expressed in tumor tissue^[68]. Secondly, though the induction of MMP enables IL-22 to protect against tumor formation in chronic liver fibrotic diseases, by the same mechanism it can increase the metastatic capacity of established tumor cells by digesting ECM, invading surrounding tissue and escaping the primary site. This has been shown in the A549 lung carcinoma cell line and pancreatic cancer, while whether the same mechanism also exists in hepatic tumor tissue remains unknown^[97,140,141].

The same balance also exists in the antiviral activity and associated oncogenesis. IL-22 disturbs the establishment of chronic inflammation to prevent liver cancer. As was shown in acute infection of HBV, IL-22 acts as the mediator of an acute phase reaction to clear the virus *via* the recruitment of T cells^[142]. However, it plays a contrary role in the progressive diseases, as IL-22 level was elevated and high serum IL-22 level indicated a poor prognosis both in patients with HBV and hepatitis C virus-associated HCC, suggesting that expression of IL-22 during progressive disease may reflect increased aggressiveness of HCC instead of predisposal to cirrhosis^[76,143].

IL-22 can also influence the outcome of tumorigenesis by the mechanisms of pro- and antiinflammatory functions, angiogenesis, epithelial-mesenchymal transition, dysplasia and metabolic functions that remain less clear in the liver^[142]. All these results suggest whether the effect of IL-22 is tumorigenic or antitumorigenic seems to depend on the stage of their responses and the specific tumor microenvironment.

Thy1⁺IL-23R⁺ ILC3s are important for IL-23-induced initiation of gut tumorigenesis, as substantial inhibition of tumorigenesis in *RAG*^{-/-}*IL17*^{-/-} double knock-out mice provided evidence for an important contribution of IL-17 expression in ILC3s, which consistently occurred before the recruitment of overt inflammatory infiltrates^[144]. When it comes to liver, the connection between IL-17 and angiogenesis has emerged in the context of HCC^[145]. Besides, it has been shown to have protumor activity in proliferation, immune-resistance, tumorigenesis and metastasis as well^[146]. On the contrary, IL-17 also plays a vital role in antitumor activity *via* the stimulation of tumor-specific CTLs, which were associated with establishment of a tumor-protective immunity in hematopoietic cancer^[147]. These results also lead to the suggestion that there is a balance between protective CTLs' formation during the acute phase of hepatitis and angiogenic activity during the chronic phase, which would determine the outcome of tumors.

Overall, ample evidence has pointed towards ILC3s having an important role in tumor progression. The

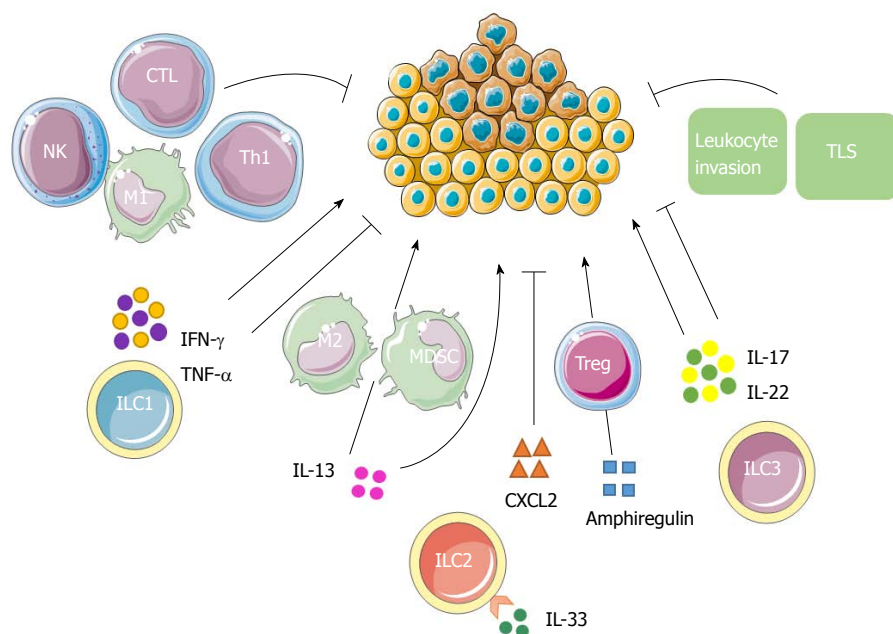


Figure 4 Possible innate lymphoid cell interactions in liver tumor immunity. ILC1s have both antitumor and protumor effects according to the properties of cytokines secreted. IFN- γ and TNF- α have antiproliferative, antiangiogenic and proapoptotic effects against cancer cells. In addition to promoting the polarization of CD4⁺ T cells into Th1 cells, they can also boost the responses of macrophages, NK cells and CTLs, leading to a strong antitumor response. On the contrary, their ambiguous roles enable them to enhance tumor formation, growth and spread. ILC2s may contribute to tumor progression either directly through the tumorigenic effects of IL-13, or indirectly by stimulating M2 macrophages and MDSCs through IL-13. The production of amphiregulin suggests that ILC2s may further inhibit antitumor immune responses either directly or via stimulation of Tregs. IL-33 could also induce the secretion of a massive amount of CXCR2 ligands from ILC2s as well as create a tumor microenvironment wherein tumor cells express CXCR2, leading to apoptosis of active tumor cells. ILC3s might promote antitumor responses by enhancing leukocyte invasion, promoting TLSs' induction and through the antitumor effects of IL-17 and IL-22. Conversely, ILC3s may promote tumor formation and progression by IL-17 and IL-22 as well according to the phase of responses and specific tumor microenvironment. CTL: Cytotoxic T lymphocyte; IL: Interleukin; ILC: Innate lymphoid cell; IFN: Interferon; MDSC: Myeloid-derived suppressor cell; NK: Natural killer; Th: T helper; TLS: Tertiary lymphoid structure; TNF: Tumor necrosis factor; Tregs: T regulatory cells.

elements consisting of specific tumor microenvironment as well as the timing of responses count when considering their ambiguous roles, while their explicit functions in humans, especially in human liver cancer, are incompletely understood and remain to be fully elucidated (Figure 4).

CONCLUSION AND FUTURE PERSPECTIVES

The recent research studies on the roles of ILCs in development of chronic liver diseases have made great progress, especially for hepatitis and liver fibrosis. Different or even the same ILC subsets have shown complex functions at a certain stage of chronic liver diseases. Also, the same ILC subsets have exhibited both pathological and protective functions during the dynamic development of chronic liver diseases. When considering their effects on liver, both the downstream effector cytokines and the molecules involved in the upstream signaling must be taken into consideration simultaneously, and more research is required to further elucidate the underlying mechanisms and signaling pathways. Apparently, there is a balance between the protective and pathological properties of ILCs, according to the specific liver tissue microenvironment at different

stages of liver diseases, whereby effector cytokines, surrounding interaction cells and functional state of cell receptors vary remarkably. What's more, the basis of the functions of ILCs and their downstream effector cytokines in hepatitis and liver fibrosis can represent the foundation of future research interests for investigating their roles in tumorigenesis.

Different methods have been applied to detect ILCs and measure their activities. Intrahepatic as well as peripheral blood mononuclear cells are isolated for further *in vitro* staining with fluorescence-labeled antibodies according to the cluster of differentiation on the surface of different ILC subsets and intracellular contents. Flow cytometry is further applied to detect the frequency and cellularity of ILCs and analyse the expression of their transcription factors and effector cytokines induced by PMA/ionomycin once they have been sorted *in vitro*. By observation of the differences of these factors between patients with chronic liver diseases and healthy control groups, their changes before and after the inducing factors and their consistency with liver injury, the researchers could validate the activities and functions of ILCs in the liver^[23,38]. Considering limited accessibility of primary intrahepatic ILCs, the expansion of cell lines of primary intrahepatic ILCs is also an alternative to assess the function of this

small cell population and to seek their secretion profile through the stimulation of PMA/ionomycin^[96]. Besides, there have been studies exploring the roles of ILCs in initiation of liver injury including both hepatitis and liver fibrosis by the mechanism of *in vivo* depletion of ILCs using specific antibodies^[22,73] or targeted transcription factor gene-deficient mice^[72]. The protective or pathological roles of ILCs are determined by comparison of the severity of liver injury before and after the depletion of ILCs as indicated by histological analysis of liver tissue and expression of liver injury serum biomarkers as well as inflammatory cytokines in *RAG1*^{-/-} mice which are reconstituted with CD4⁺ T cells. *In vivo* experiments to distribute the signaling pathway of ILCs through the blockade of upstream cytokines and surface receptors of ILCs *via* targeted gene-knock mice are also important methods, in which the expansion of ILCs and their expression of transcription factors and downstream effector cytokines are further detected by flow cytometry and quantitative real-time PCR analysis^[24,52]. Additionally, the activities of ILCs could also be monitored by transfer experiments, in which purified ILCs sorted by MACS/FACS are adoptively transferred into recipient mice before the challenge of stimulus including ConA and carbon tetrachloride to further investigate the function of ILCs in the liver^[22,24,58].

There still remains a lot to be fully elucidated. Firstly, the functions of some ILC subsets at a particular stage of chronic liver diseases have only been inferred by their downstream effector cytokines, while lacking direct and potent evidence. Secondly, given their distribution characteristics, evidence with regard to the functions of ILCs in tumorigenesis is emerging from studies that have mostly investigated chronic inflammation and the procarcinogenic role of secreted cytokines in skin, gut and lung, and less so for the liver. Thirdly, as the innate counterpart of CD4⁺ Th cells, the same effector cytokines can be produced by both adaptive lymphoid cells and ILCs. It is important to identify the sources of effector cytokines, while the results from the most recent *in vivo* studies were obtained from *RAG*^{-/-} mice or antibodies that are specific to ILCs' genes leading to broad immune deficiencies. Thus, it is necessary to apply ILCs' specific gene-knockout or transgenic models to reveal the precise and direct actions of each in the liver.

New strategies targeting ILCs have been designed for diagnosis and treatment, to prevent or stop the progression of chronic liver diseases. The inhibition of NKG2A receptors on ILC1s to further promote robust CD8⁺ T cell responses has been considered a potential therapeutic strategy against persistent liver pathogens in patients with hepatitis. Besides, for liver fibrosis treatment, serum IL-33 may be a possible noninvasive diagnostic biomarker for uncovering early inflammatory and fibrogenic events. Targeting ILC2s may represent a novel strategy as well. Further in-depth studies to elucidate the distinct and explicit effects of each of the ILC subsets at different stages of chronic liver diseases

are required in order to promote the exploration and realization of their therapeutic potency.

REFERENCES

- 1 **Bataller R**, Brenner DA. Liver fibrosis. *J Clin Invest* 2005; **115**: 209-218 [PMID: 15690074 DOI: 10.1172/JCI24282]
- 2 **Fattovich G**, Stroffolini T, Zagni I, Donato F. Hepatocellular carcinoma in cirrhosis: incidence and risk factors. *Gastroenterology* 2004; **127**: S35-S50 [PMID: 15508101]
- 3 **Wynn TA**. Common and unique mechanisms regulate fibrosis in various fibroproliferative diseases. *J Clin Invest* 2007; **117**: 524-529 [PMID: 17332879 DOI: 10.1172/JCI1487]
- 4 **Kew MC**. The role of cirrhosis in the etiology of hepatocellular carcinoma. *J Gastrointest Cancer* 2014; **45**: 12-21 [PMID: 24203525 DOI: 10.1007/s12029-013-9556-9]
- 5 **Spits H**, Cupedo T. Innate lymphoid cells: emerging insights in development, lineage relationships, and function. *Annu Rev Immunol* 2012; **30**: 647-675 [PMID: 22224763 DOI: 10.1146/annurev-immunol-020711-075053]
- 6 **Eberl G**, Colonna M, Di Santo JP, McKenzie AN. Innate lymphoid cells. Innate lymphoid cells: a new paradigm in immunology. *Science* 2015; **348**: aaa6566 [PMID: 25999512 DOI: 10.1126/science.aaa6566]
- 7 **Klose CSN**, Flach M, Möhle L, Rogell L, Hoyler T, Ebert K, Fabiunke C, Pfeifer D, Sexl V, Fonseca-Pereira D, Domingues RG, Veiga-Fernandes H, Arnold SJ, Busslinger M, Dunay IR, Tanriver Y, Diefenbach A. Differentiation of type 1 ILCs from a common progenitor to all helper-like innate lymphoid cell lineages. *Cell* 2014; **157**: 340-356 [PMID: 24725403 DOI: 10.1016/j.cell.2014.03.030]
- 8 **Fuchs A**, Vermi W, Lee JS, Lonardi S, Gilfillan S, Newberry RD, Cella M, Colonna M. Intraepithelial type 1 innate lymphoid cells are a unique subset of IL-12- and IL-15-responsive IFN- γ -producing cells. *Immunity* 2013; **38**: 769-781 [PMID: 23453631 DOI: 10.1016/j.immuni.2013.02.010]
- 9 **Bernink JH**, Peters CP, Munneke M, te Velde AA, Meijer SL, Weijer K, Hreggvidsdottir HS, Heinsbroek SE, Legrand N, Buskens CJ, Bemelman WA, Mjösberg JM, Spits H. Human type 1 innate lymphoid cells accumulate in inflamed mucosal tissues. *Nat Immunol* 2013; **14**: 221-229 [PMID: 23334791 DOI: 10.1038/ni.2534]
- 10 **Constantinides MG**, McDonald BD, Verhoef PA, Bendelac A. A committed precursor to innate lymphoid cells. *Nature* 2014; **508**: 397-401 [PMID: 24509713 DOI: 10.1038/nature13047]
- 11 **Daussy C**, Faure F, Mayol K, Viel S, Gasteiger G, Charrier E, Biennu J, Henry T, Debien E, Hasan UA, Marvel J, Yoh K, Takahashi S, Prinz I, de Bernard S, Buffat L, Walzer T. T-bet and Eomes instruct the development of two distinct natural killer cell lineages in the liver and in the bone marrow. *J Exp Med* 2014; **211**: 563-577 [PMID: 24516120 DOI: 10.1084/jem.20131560]
- 12 **Neill DR**, Wong SH, Bellosi A, Flynn RJ, Daly M, Langford TK, Bucks C, Kane CM, Fallon PG, Pannell R, Jolin HE, McKenzie AN. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature* 2010; **464**: 1367-1370 [PMID: 20200518 DOI: 10.1038/nature08900]
- 13 **Price AE**, Liang HE, Sullivan BM, Reinhardt RL, Eislely CJ, Erle DJ, Locksley RM. Systemically dispersed innate IL-13-expressing cells in type 2 immunity. *Proc Natl Acad Sci USA* 2010; **107**: 11489-11494 [PMID: 20534524 DOI: 10.1073/pnas.1003988107]
- 14 **Wong SH**, Walker JA, Jolin HE, Drynan LF, Hams E, Camelo A, Barlow JL, Neill DR, Panova V, Koch U, Radtke F, Hardman CS, Hwang YY, Fallon PG, McKenzie AN. Transcription factor ROR α is critical for nuocyte development. *Nat Immunol* 2012; **13**: 229-236 [PMID: 22267218 DOI: 10.1038/ni.2208]
- 15 **Licona-Limón P**, Kim LK, Palm NW, Flavell RA. TH2, allergy and group 2 innate lymphoid cells. *Nat Immunol* 2013; **14**: 536-542 [PMID: 23685824 DOI: 10.1038/ni.2617]
- 16 **Sawa S**, Lochner M, Satoh-Takayama N, Dulauroy S, Bérard

- M, Kleinschek M, Cua D, Di Santo JP, Eberl G. ROR γ t⁺ innate lymphoid cells regulate intestinal homeostasis by integrating negative signals from the symbiotic microbiota. *Nat Immunol* 2011; **12**: 320-326 [PMID: 21336274 DOI: 10.1038/ni.2002]
- 17 **Mortha A**, Chudnovskiy A, Hashimoto D, Bogunovic M, Spencer SP, Belkaid Y, Merad M. Microbiota-dependent crosstalk between macrophages and ILC3 promotes intestinal homeostasis. *Science* 2014; **343**: 1249288 [PMID: 24625929 DOI: 10.1126/science.1249288]
- 18 **Eberl G**, Marmon S, Sunshine MJ, Rennert PD, Choi Y, Littman DR. An essential function for the nuclear receptor ROR γ (t) in the generation of fetal lymphoid tissue inducer cells. *Nat Immunol* 2004; **5**: 64-73 [PMID: 14691482 DOI: 10.1038/ni1022]
- 19 **Buonocore S**, Ahern PP, Uhlig HH, Ivanov II, Littman DR, Maloy KJ, Powrie F. Innate lymphoid cells drive interleukin-23-dependent innate intestinal pathology. *Nature* 2010; **464**: 1371-1375 [PMID: 20393462 DOI: 10.1038/nature08949]
- 20 **Halim TY**, Krauss RH, Sun AC, Takei F. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. *Immunity* 2012; **36**: 451-463 [PMID: 22425247 DOI: 10.1016/j.immuni.2011.12.020]
- 21 **Kim BS**, Siracusa MC, Saenz SA, Noti M, Monticelli LA, Sonnenberg GF, Hepworth MR, Van Voorhees AS, Comeau MR, Artis D. TSLP elicits IL-33-independent innate lymphoid cell responses to promote skin inflammation. *Sci Transl Med* 2013; **5**: 170ra16 [PMID: 23363980 DOI: 10.1126/scitranslmed.3005374]
- 22 **McHedlidze T**, Waldner M, Zopf S, Walker J, Rankin AL, Schuchmann M, Voehringer D, McKenzie AN, Neurath MF, Pflanz S, Wirtz S. Interleukin-33-dependent innate lymphoid cells mediate hepatic fibrosis. *Immunity* 2013; **39**: 357-371 [PMID: 23954132 DOI: 10.1016/j.immuni.2013.07.018]
- 23 **Yang Z**, Tang T, Wei X, Yang S, Tian Z. Type 1 innate lymphoid cells contribute to the pathogenesis of chronic hepatitis B. *Innate Immun* 2015; **21**: 665-673 [PMID: 25977358 DOI: 10.1177/1753425915586074]
- 24 **Neumann K**, Karimi K, Meiners J, Voeltlaue R, Steinmann S, Dammernann W, Lüth S, Asghari F, Wegscheid C, Horst AK, Tieg G. A Proinflammatory Role of Type 2 Innate Lymphoid Cells in Murine Immune-Mediated Hepatitis. *J Immunol* 2017; **198**: 128-137 [PMID: 27872212 DOI: 10.4049/jimmunol.1600418]
- 25 **Zhang Y**, Tang J, Tian Z, van Velkinburgh JC, Song J, Wu Y, Ni B. Innate Lymphoid Cells: A Promising New Regulator in Fibrotic Diseases. *Int Rev Immunol* 2016; **35**: 399-414 [PMID: 26222510 DOI: 10.3109/08830185.2015.1068304]
- 26 **van Beek JJP**, Martens AWJ, Bakdash G, de Vries IJM. Innate Lymphoid Cells in Tumor Immunity. *Biomedicines* 2016; **4**: pii: E7 [PMID: 28536374 DOI: 10.3390/biomedicines4010007]
- 27 **Vallentin B**, Barlogis V, Piperoglou C, Cypowyj S, Zucchini N, Chéné M, Navarro F, Farnarier C, Vivier E, Vély F. Innate Lymphoid Cells in Cancer. *Cancer Immunol Res* 2015; **3**: 1109-1114 [PMID: 26438443 DOI: 10.1158/2326-6066.CIR-15-0222]
- 28 **Peng H**, Jiang X, Chen Y, Sojka DK, Wei H, Gao X, Sun R, Yokoyama WM, Tian Z. Liver-resident NK cells confer adaptive immunity in skin-contact inflammation. *J Clin Invest* 2013; **123**: 1444-1456 [PMID: 23524967 DOI: 10.1172/JCI66381]
- 29 **Sojka DK**, Plougastel-Douglas B, Yang L, Pak-Wittel MA, Artyomov MN, Ivanova Y, Zhong C, Chase JM, Rothman PB, Yu J, Riley JK, Zhu J, Tian Z, Yokoyama WM. Tissue-resident natural killer (NK) cells are cell lineages distinct from thymic and conventional splenic NK cells. *Elife* 2014; **3**: e01659 [PMID: 24714492]
- 30 **Knolle PA**, Wöhlleber D. Immunological functions of liver sinusoidal endothelial cells. *Cell Mol Immunol* 2016; **13**: 347-353 [PMID: 27041636 DOI: 10.1038/cmi.2016.5]
- 31 **Geissmann F**, Cameron TO, Sidobre S, Manlongat N, Kronenberg M, Briskin MJ, Dustin ML, Littman DR. Intravascular immune surveillance by CXCR6⁺ NKT cells patrolling liver sinusoids. *PLoS Biol* 2005; **3**: e113 [PMID: 15799695 DOI: 10.1371/journal.pbio.0030113]
- 32 **Björkström NK**, Kekäläinen E, Mjösberg J. Tissue-specific effector functions of innate lymphoid cells. *Immunology* 2013; **139**: 416-427 [PMID: 23489335 DOI: 10.1111/imm.12098]
- 33 **Watzl C**. How to trigger a killer: modulation of natural killer cell reactivity on many levels. *Adv Immunol* 2014; **124**: 137-170 [PMID: 25175775 DOI: 10.1016/B978-0-12-800147-9.00005-4]
- 34 **Sun C**, Sun H, Zhang C, Tian Z. NK cell receptor imbalance and NK cell dysfunction in HBV infection and hepatocellular carcinoma. *Cell Mol Immunol* 2015; **12**: 292-302 [PMID: 25308752 DOI: 10.1038/cmi.2014.91]
- 35 **Zheng M**, Sun R, Wei H, Tian Z. NK Cells Help Induce Anti-Hepatitis B Virus CD8⁺ T Cell Immunity in Mice. *J Immunol* 2016; **196**: 4122-4131 [PMID: 27183639 DOI: 10.4049/jimmunol.1500846]
- 36 **Jinushi M**, Takehara T, Tatsumi T, Yamaguchi S, Sakamori R, Hiramatsu N, Kanto T, Ohkawa K, Hayashi N. Natural killer cell and hepatic cell interaction via NKG2A leads to dendritic cell-mediated induction of CD4 CD25 T cells with PD-1-dependent regulatory activities. *Immunology* 2007; **120**: 73-82 [PMID: 17052247 DOI: 10.1111/j.1365-2567.2006.02479.x]
- 37 **Li F**, Wei H, Wei H, Gao Y, Xu L, Yin W, Sun R, Tian Z. Blocking the natural killer cell inhibitory receptor NKG2A increases activity of human natural killer cells and clears hepatitis B virus infection in mice. *Gastroenterology* 2013; **144**: 392-401 [PMID: 23103614 DOI: 10.1053/j.gastro.2012.10.039]
- 38 **Krueger PD**, Narayanan S, Surette FA, Brown MG, Sung SJ, Hahn YS. Murine liver-resident group 1 innate lymphoid cells regulate optimal priming of anti-viral CD8⁺ T cells. *J Leukoc Biol* 2017; **101**: 329-338 [PMID: 27493244 DOI: 10.1189/jlb.3A0516-225R]
- 39 **Nattermann J**, Feldmann G, Ahlenstiel G, Langhans B, Sauerbruch T, Spengler U. Surface expression and cytolytic function of natural killer cell receptors is altered in chronic hepatitis C. *Gut* 2006; **55**: 869-877 [PMID: 16322112 DOI: 10.1136/gut.2005.076463]
- 40 **Thoens R**, Berger C, Trippler M, Siemann H, Lutterbeck M, Broering R, Schlaak J, Heinemann FM, Heinold A, Nattermann J, Scherbaum N, Alter G, Timm J. KIR2DL3⁺NKG2A⁺ natural killer cells are associated with protection from productive hepatitis C virus infection in people who inject drugs. *J Hepatol* 2014; **61**: 475-481 [PMID: 24780303 DOI: 10.1016/j.jhep.2014.04.020]
- 41 **Schmitz J**, Owyang A, Oldham E, Song Y, Murphy E, McClanahan TK, Zurawski G, Moshrefi M, Qin J, Li X, Gorman DM, Bazan JF, Kastelein RA. IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. *Immunity* 2005; **23**: 479-490 [PMID: 16286016 DOI: 10.1016/j.immuni.2005.09.015]
- 42 **Cayrol C**, Girard JP. IL-33: an alarmin cytokine with crucial roles in innate immunity, inflammation and allergy. *Curr Opin Immunol* 2014; **31**: 31-37 [PMID: 25278425 DOI: 10.1016/j.coi.2014.09.004]
- 43 **Marvie P**, Lisbonne M, L'helgoualc'h A, Rauch M, Turlin B, Preisser L, Bourd-Boittin K, Théret N, Gascan H, Piquet-Pellorce C, Samson M. Interleukin-33 overexpression is associated with liver fibrosis in mice and humans. *J Cell Mol Med* 2010; **14**: 1726-1739 [PMID: 19508382 DOI: 10.1111/j.1582-4934.2009.00801.x]
- 44 **Arshad MI**, Piquet-Pellorce C, L'helgoualc'h A, Rauch M, Patrat-Delon S, Ezan F, Lucas-Clerc C, Nabti S, Lehuen A, Cubero FJ, Girard JP, Trautwein C, Samson M. TRAIL but not FasL and TNF α , regulates IL-33 expression in murine hepatocytes during acute hepatitis. *Hepatology* 2012; **56**: 2353-2362 [PMID: 22961755 DOI: 10.1002/hep.25893]
- 45 **Wang J**, Cai Y, Ji H, Feng J, Ayana DA, Niu J, Jiang Y. Serum IL-33 levels are associated with liver damage in patients with chronic hepatitis B. *J Interferon Cytokine Res* 2012; **32**: 248-253 [PMID: 22304300 DOI: 10.1089/jir.2011.0109]
- 46 **Wang J**, Zhao P, Guo H, Sun X, Jiang Z, Xu L, Feng J, Niu J, Jiang Y. Serum IL-33 levels are associated with liver damage in patients with chronic hepatitis C. *Mediators Inflamm* 2012; **2012**: 819636 [PMID: 22315510 DOI: 10.1155/2012/819636]

- 47 **Kurowska-Stolarska M**, Hueber A, Stolarski B, McInnes IB. Interleukin-33: a novel mediator with a role in distinct disease pathologies. *J Intern Med* 2011; **269**: 29-35 [PMID: 21158975 DOI: 10.1111/j.1365-2796.2010.02316.x]
- 48 **Roth GA**, Zimmermann M, Lubczyk BA, Pilz J, Faybik P, Hetz H, Hacker S, Mangold A, Bacher A, Krenn CG, Ankersmit HJ. Up-regulation of interleukin 33 and soluble ST2 serum levels in liver failure. *J Surg Res* 2010; **163**: e79-e83 [PMID: 20638676 DOI: 10.1016/j.jss.2010.04.004]
- 49 **Chen J**, Duan L, Xiong A, Zhang H, Zheng F, Tan Z, Gong F, Fang M. Blockade of IL-33 ameliorates Con A-induced hepatic injury by reducing NKT cell activation and IFN- γ production in mice. *J Mol Med (Berl)* 2012; **90**: 1505-1515 [PMID: 22983503 DOI: 10.1007/s00109-012-0938-4]
- 50 **Tiegs G**, Hentschel J, Wendel A. A T cell-dependent experimental liver injury in mice inducible by concanavalin A. *J Clin Invest* 1992; **90**: 196-203 [PMID: 1634608 DOI: 10.1172/JCI115836]
- 51 **Gieseck RL 3rd**, Ramalingam TR, Hart KM, Vannella KM, Cantu DA, Lu WY, Ferreira-González S, Forbes SJ, Vallier L, Wynn TA. Interleukin-13 Activates Distinct Cellular Pathways Leading to Ductular Reaction, Steatosis, and Fibrosis. *Immunity* 2016; **45**: 145-158 [PMID: 27421703 DOI: 10.1016/j.immuni.2016.06.009]
- 52 **Volarevic V**, Mitrovic M, Milovanovic M, Zelen I, Nikolic I, Mitrovic S, Pejnovic N, Arsenijevic N, Lukic ML. Protective role of IL-33/ST2 axis in Con A-induced hepatitis. *J Hepatol* 2012; **56**: 26-33 [PMID: 21703183 DOI: 10.1016/j.jhep.2011.03.022]
- 53 **Bonilla WV**, Fröhlich A, Senn K, Kallert S, Fernandez M, Johnson S, Kreuzfeldt M, Hegazy AN, Schrick C, Fallon PG, Klemenz R, Nakae S, Adler H, Merkler D, Löhning M, Pinschewer DD. The alarmin interleukin-33 drives protective antiviral CD8⁺ T cell responses. *Science* 2012; **335**: 984-989 [PMID: 22323740 DOI: 10.1126/science.1215418]
- 54 **Baumann C**, Bonilla WV, Fröhlich A, Helmstetter C, Peine M, Hegazy AN, Pinschewer DD, Löhning M. T-bet- and STAT4-dependent IL-33 receptor expression directly promotes antiviral Th1 cell responses. *Proc Natl Acad Sci USA* 2015; **112**: 4056-4061 [PMID: 25829541 DOI: 10.1073/pnas.1418549112]
- 55 **Carriere V**, Roussel L, Ortega N, Lacorre DA, Americh L, Aguilar L, Bouche G, Girard JP. IL-33, the IL-1-like cytokine ligand for ST2 receptor, is a chromatin-associated nuclear factor in vivo. *Proc Natl Acad Sci U S A* 2007; **104**: 282-287 [PMID: 17185418 DOI: 10.1073/pnas.0606854104]
- 56 **Liang Y**, Jie Z, Hou L, Yi P, Wang W, Kwota Z, Salvato M, de Waal Malefyt R, Soong L, Sun J. IL-33 promotes innate IFN- γ production and modulates dendritic cell response in LCMV-induced hepatitis in mice. *Eur J Immunol* 2015; **45**: 3052-3063 [PMID: 26249267 DOI: 10.1002/eji.201545696]
- 57 **Liu ZX**, Govindarajan S, Okamoto S, Dennert G. Fas- and tumor necrosis factor receptor 1-dependent but not perforin-dependent pathways cause injury in livers infected with an adenovirus construct in mice. *Hepatology* 2000; **31**: 665-673 [PMID: 10706557 DOI: 10.1002/hep.510310317]
- 58 **Liang Y**, Jie Z, Hou L, Aguilar-Valenzuela R, Vu D, Soong L, Sun J. IL-33 induces neutrophils and modulates liver injury in viral hepatitis. *J Immunol* 2013; **190**: 5666-5675 [PMID: 23630360 DOI: 10.4049/jimmunol.1300117]
- 59 **O'Connor W Jr**, Zenewicz LA, Flavell RA. The dual nature of T(H)17 cells: shifting the focus to function. *Nat Immunol* 2010; **11**: 471-476 [PMID: 20485275 DOI: 10.1038/ni.1882]
- 60 **Geboes L**, Dumoutier L, Kelchtermans H, Schurgers E, Mitera T, Renaud JC, Mathys P. Proinflammatory role of the Th17 cytokine interleukin-22 in collagen-induced arthritis in C57BL/6 mice. *Arthritis Rheum* 2009; **60**: 390-395 [PMID: 19180498 DOI: 10.1002/art.24220]
- 61 **Boniface K**, Bernard FX, Garcia M, Gurney AL, Lecron JC, Morel F. IL-22 inhibits epidermal differentiation and induces proinflammatory gene expression and migration of human keratinocytes. *J Immunol* 2005; **174**: 3695-3702 [PMID: 15749908]
- 62 **Wolk K**, Witte E, Wallace E, Döcke WD, Kunz S, Asadullah K, Volk HD, Sterry W, Sabat R. IL-22 regulates the expression of genes responsible for antimicrobial defense, cellular differentiation, and mobility in keratinocytes: a potential role in psoriasis. *Eur J Immunol* 2006; **36**: 1309-1323 [PMID: 16619290 DOI: 10.1002/eji.200535503]
- 63 **Zenewicz LA**, Yancopoulos GD, Valenzuela DM, Murphy AJ, Stevens S, Flavell RA. Innate and adaptive interleukin-22 protects mice from inflammatory bowel disease. *Immunity* 2008; **29**: 947-957 [PMID: 19100701 DOI: 10.1016/j.immuni.2008.11.003]
- 64 **Zenewicz LA**, Yancopoulos GD, Valenzuela DM, Murphy AJ, Karow M, Flavell RA. Interleukin-22 but not interleukin-17 provides protection to hepatocytes during acute liver inflammation. *Immunity* 2007; **27**: 647-659 [PMID: 17919941 DOI: 10.1016/j.immuni.2007.07.023]
- 65 **Schulz SM**, Köhler G, Schütze N, Knauer J, Straubinger RK, Chackerian AA, Witte E, Wolk K, Sabat R, Iwakura Y, Holscher C, Müller U, Kastelein RA, Alber G. Protective immunity to systemic infection with attenuated *Salmonella enterica* serovar enteritidis in the absence of IL-12 is associated with IL-23-dependent IL-22, but not IL-17. *J Immunol* 2008; **181**: 7891-7901 [PMID: 19017979]
- 66 **Zheng Y**, Valdez PA, Danilenko DM, Hu Y, Sa SM, Gong Q, Abbas AR, Modrusan Z, Ghilardi N, de Sauvage FJ, Ouyang W. Interleukin-22 mediates early host defense against attaching and effacing bacterial pathogens. *Nat Med* 2008; **14**: 282-289 [PMID: 18264109 DOI: 10.1038/nm1720]
- 67 **Zhang Y**, Cobleigh MA, Lian JQ, Huang CX, Booth CJ, Bai XF, Robek MD. A proinflammatory role for interleukin-22 in the immune response to hepatitis B virus. *Gastroenterology* 2011; **141**: 1897-1906 [PMID: 21708106 DOI: 10.1053/j.gastro.2011.06.051]
- 68 **Radaeva S**, Sun R, Pan HN, Hong F, Gao B. Interleukin 22 (IL-22) plays a protective role in T cell-mediated murine hepatitis: IL-22 is a survival factor for hepatocytes via STAT3 activation. *Hepatology* 2004; **39**: 1332-1342 [PMID: 15122762 DOI: 10.1002/hep.20184]
- 69 **Park O**, Wang H, Weng H, Feigenbaum L, Li H, Yin S, Ki SH, Yoo SH, Dooley S, Wang FS, Young HA, Gao B. In vivo consequences of liver-specific interleukin-22 expression in mice: Implications for human liver disease progression. *Hepatology* 2011; **54**: 252-261 [PMID: 21465510 DOI: 10.1002/hep.24339]
- 70 **Feng D**, Kong X, Weng H, Park O, Wang H, Dooley S, Gershwin ME, Gao B. Interleukin-22 promotes proliferation of liver stem/progenitor cells in mice and patients with chronic hepatitis B virus infection. *Gastroenterology* 2012; **143**: 188-198.e7 [PMID: 22484119 DOI: 10.1053/j.gastro.2012.03.044]
- 71 **Xu M**, Morishima N, Mizoguchi I, Chiba Y, Fujita K, Kuroda M, Iwakura Y, Cua DJ, Yasutomo K, Mizoguchi J, Yoshimoto T. Regulation of the development of acute hepatitis by IL-23 through IL-22 and IL-17 production. *Eur J Immunol* 2011; **41**: 2828-2839 [PMID: 21953641 DOI: 10.1002/eji.201141291]
- 72 **Abe H**, Kimura A, Tsuruta S, Fukaya T, Sakaguchi R, Morita R, Sekiya T, Shichita T, Chayama K, Fujii-Kuriyama Y, Yoshimura A. Aryl hydrocarbon receptor plays protective roles in ConA-induced hepatic injury by both suppressing IFN- γ expression and inducing IL-22. *Int Immunol* 2014; **26**: 129-137 [PMID: 24150244 DOI: 10.1093/intimm/dxt049]
- 73 **Matsumoto A**, Kanai T, Mikami Y, Chu PS, Nakamoto N, Ebinuma H, Saito H, Sato T, Yagita H, Hibi T. IL-22-producing ROR γ t-dependent innate lymphoid cells play a novel protective role in murine acute hepatitis. *PLoS One* 2013; **8**: e62853 [PMID: 23626860 DOI: 10.1371/journal.pone.0062853]
- 74 **Yan S**, Wang L, Liu N, Wang Y, Chu Y. Critical role of interleukin-17/interleukin-17 receptor axis in mediating Con A-induced hepatitis. *Immunol Cell Biol* 2012; **90**: 421-428 [PMID: 21691280 DOI: 10.1038/icb.2011.59]
- 75 **Lafdil F**, Wang H, Park O, Zhang W, Moritoki Y, Yin S, Fu XY, Gershwin ME, Lian ZX, Gao B. Myeloid STAT3 inhibits T cell-mediated hepatitis by regulating T helper 1 cytokine and interleukin-17 production. *Gastroenterology* 2009; **137**: 2125-2135.e1-2 [PMID: 19686746 DOI: 10.1053/j.gastro.2009.08.004]
- 76 **Zhao J**, Zhang Z, Luan Y, Zou Z, Sun Y, Li Y, Jin L, Zhou C, Fu J, Gao B, Fu Y, Wang FS. Pathological functions of interleukin-22

- in chronic liver inflammation and fibrosis with hepatitis B virus infection by promoting T helper 17 cell recruitment. *Hepatology* 2014; **59**: 1331-1342 [PMID: 24677193 DOI: 10.1002/hep.26916]
- 77 **Wei X**, Wang JP, Hao CQ, Yang XF, Wang LX, Huang CX, Bai XF, Lian JQ, Zhang Y. Notch Signaling Contributes to Liver Inflammation by Regulation of Interleukin-22-Producing Cells in Hepatitis B Virus Infection. *Front Cell Infect Microbiol* 2016; **6**: 132 [PMID: 27800305 DOI: 10.3389/fcimb.2016.00132]
- 78 **Hou L**, Jie Z, Desai M, Liang Y, Soong L, Wang T, Sun J. Early IL-17 production by intrahepatic T cells is important for adaptive immune responses in viral hepatitis. *J Immunol* 2013; **190**: 621-629 [PMID: 23233727 DOI: 10.4049/jimmunol.1201970]
- 79 **Jie Z**, Liang Y, Hou L, Dong C, Iwakura Y, Soong L, Cong Y, Sun J. Intrahepatic innate lymphoid cells secrete IL-17A and IL-17F that are crucial for T cell priming in viral infection. *J Immunol* 2014; **192**: 3289-3300 [PMID: 24600029 DOI: 10.4049/jimmunol.1303281]
- 80 **Wang Q**, Zhou J, Zhang B, Tian Z, Tang J, Zheng Y, Huang Z, Tian Y, Jia Z, Tang Y, van Velkinburgh JC, Mao Q, Bian X, Ping Y, Ni B, Wu Y. Hepatitis B virus induces IL-23 production in antigen presenting cells and causes liver damage via the IL-23/IL-17 axis. *PLoS Pathog* 2013; **9**: e1003410 [PMID: 23825942 DOI: 10.1371/journal.ppat.1003410]
- 81 **Jeong WI**, Park O, Radaeva S, Gao B. STAT1 inhibits liver fibrosis in mice by inhibiting stellate cell proliferation and stimulating NK cell cytotoxicity. *Hepatology* 2006; **44**: 1441-1451 [PMID: 17133483 DOI: 10.1002/hep.21419]
- 82 **Gao B**, Radaeva S, Jeong WI. Activation of natural killer cells inhibits liver fibrosis: a novel strategy to treat liver fibrosis. *Expert Rev Gastroenterol Hepatol* 2007; **1**: 173-180 [PMID: 19072444 DOI: 10.1586/17474124.1.1.173]
- 83 **Radaeva S**, Sun R, Jaruga B, Nguyen VT, Tian Z, Gao B. Natural killer cells ameliorate liver fibrosis by killing activated stellate cells in NKG2D-dependent and tumor necrosis factor-related apoptosis-inducing ligand-dependent manners. *Gastroenterology* 2006; **130**: 435-452 [PMID: 16472598 DOI: 10.1053/j.gastro.2005.10.055]
- 84 **Glässner A**, Eisenhardt M, Krämer B, Körner C, Coenen M, Sauerbruch T, Spengler U, Nattermann J. NK cells from HCV-infected patients effectively induce apoptosis of activated primary human hepatic stellate cells in a TRAIL-, FasL- and NKG2D-dependent manner. *Lab Invest* 2012; **92**: 967-977 [PMID: 22449797 DOI: 10.1038/labinvest.2012.54]
- 85 **Radaeva S**, Wang L, Radaeva S, Jeong WI, Park O, Gao B. Retinoic acid signaling sensitizes hepatic stellate cells to NK cell killing via upregulation of NK cell activating ligand RAE1. *Am J Physiol Gastrointest Liver Physiol* 2007; **293**: G809-G816 [PMID: 17673545 DOI: 10.1152/ajpgi.00212.2007]
- 86 **Krizhanovsky V**, Yon M, Dickins RA, Hearn S, Simon J, Miething C, Yee H, Zender L, Lowe SW. Senescence of activated stellate cells limits liver fibrosis. *Cell* 2008; **134**: 657-667 [PMID: 18724938 DOI: 10.1016/j.cell.2008.06.049]
- 87 **Gur C**, Doron S, Kfir-Erenfeld S, Horwitz E, Abu-Tair L, Safadi R, Mandelboim O. NKp46-mediated killing of human and mouse hepatic stellate cells attenuates liver fibrosis. *Gut* 2012; **61**: 885-893 [PMID: 22198715 DOI: 10.1136/gutjnl-2011-301400]
- 88 **Mantovani S**, Mele D, Oliviero B, Barbarini G, Varchetta S, Mondelli MU. NKp30 isoforms in patients with chronic hepatitis C virus infection. *Immunology* 2015; **146**: 234-242 [PMID: 26094914 DOI: 10.1111/imm.12495]
- 89 **Melhem A**, Muhanna N, Bishara A, Alvarez CE, Ilan Y, Bishara T, Horani A, Nassar M, Friedman SL, Safadi R. Anti-fibrotic activity of NK cells in experimental liver injury through killing of activated HSC. *J Hepatol* 2006; **45**: 60-71 [PMID: 16515819 DOI: 10.1016/j.jhep.2005.12.025]
- 90 **Muhanna N**, Abu Tair L, Doron S, Amer J, Azzeh M, Mahamid M, Friedman S, Safadi R. Amelioration of hepatic fibrosis by NK cell activation. *Gut* 2011; **60**: 90-98 [PMID: 20660699 DOI: 10.1136/gut.2010.211136]
- 91 **Stegmann KA**, Björkström NK, Veber H, Ciesek S, Riese P, Wiegand J, Hadem J, Suneetha PV, Jaroszewicz J, Wang C, Schlaphoff V, Fytli P, Cornberg M, Manns MP, Geffers R, Pietschmann T, Guzmán CA, Ljunggren HG, Wedemeyer H. Interferon-alpha-induced TRAIL on natural killer cells is associated with control of hepatitis C virus infection. *Gastroenterology* 2010; **138**: 1885-1897 [PMID: 20334827 DOI: 10.1053/j.gastro.2010.01.051]
- 92 **Ahlenstiel G**, Edlich B, Hogdal LJ, Rotman Y, Nouredin M, Feld JJ, Holz LE, Titerence RH, Liang TJ, Rehermann B. Early changes in natural killer cell function indicate virologic response to interferon therapy for hepatitis C. *Gastroenterology* 2011; **141**: 1231-1239, 1239.e1-1239.e2 [PMID: 21741920 DOI: 10.1053/j.gastro.2011.06.069]
- 93 **Sun C**, Fu B, Gao Y, Liao X, Sun R, Tian Z, Wei H. TGF- β 1 down-regulation of NKG2D/DAP10 and 2B4/SAP expression on human NK cells contributes to HBV persistence. *PLoS Pathog* 2012; **8**: e1002594 [PMID: 22438812 DOI: 10.1371/journal.ppat.1002594]
- 94 **Hams E**, Armstrong ME, Barlow JL, Saunders SP, Schwartz C, Cooke G, Fahy RJ, Crotty TB, Hirani N, Flynn RJ, Voehringer D, McKenzie AN, Donnelly SC, Fallon PG. IL-25 and type 2 innate lymphoid cells induce pulmonary fibrosis. *Proc Natl Acad Sci USA* 2014; **111**: 367-372 [PMID: 24344271 DOI: 10.1073/pnas.1315854111]
- 95 **Hammerich L**, Tacke F. Interleukins in chronic liver disease: lessons learned from experimental mouse models. *Clin Exp Gastroenterol* 2014; **7**: 297-306 [PMID: 25214799 DOI: 10.2147/CEG.S43737]
- 96 **Forkel M**, Berglin L, Kekäläinen E, Carlsson A, Svedin E, Michaëlsson J, Nagasawa M, Erjefält JS, Mori M, Flodström-Tullberg M, Bergquist A, Ljunggren HG, Westgren M, Lindfors U, Friberg D, Jorns C, Ellis E, Björkström NK, Mjösberg J. Composition and functionality of the intrahepatic innate lymphoid cell-compartment in human nonfibrotic and fibrotic livers. *Eur J Immunol* 2017; **47**: 1280-1294 [PMID: 28613415 DOI: 10.1002/eji.201646890]
- 97 **Kong X**, Feng D, Wang H, Hong F, Bertola A, Wang FS, Gao B. Interleukin-22 induces hepatic stellate cell senescence and restricts liver fibrosis in mice. *Hepatology* 2012; **56**: 1150-1159 [PMID: 22473749 DOI: 10.1002/hep.25744]
- 98 **Schnabl B**, Purbeck CA, Choi YH, Hagedorn CH, Brenner D. Replicative senescence of activated human hepatic stellate cells is accompanied by a pronounced inflammatory but less fibrogenic phenotype. *Hepatology* 2003; **37**: 653-664 [PMID: 12601363 DOI: 10.1053/jhep.2003.50097]
- 99 **Kronenberger B**, Rudloff I, Bachmann M, Brunner F, Kapper L, Filmann N, Waidmann O, Herrmann E, Pfeilschifter J, Zeuzem S, Piiper A, Mühl H. Interleukin-22 predicts severity and death in advanced liver cirrhosis: a prospective cohort study. *BMC Med* 2012; **10**: 102 [PMID: 22967278 DOI: 10.1186/1741-7015-10-102]
- 100 **Tan Z**, Qian X, Jiang R, Liu Q, Wang Y, Chen C, Wang X, Ryffel B, Sun B. IL-17A plays a critical role in the pathogenesis of liver fibrosis through hepatic stellate cell activation. *J Immunol* 2013; **191**: 1835-1844 [PMID: 23842754 DOI: 10.4049/jimmunol.1203013]
- 101 **Nakashima T**, Jinnin M, Yamane K, Honda N, Kajihara I, Makino T, Masuguchi S, Fukushima S, Okamoto Y, Hasegawa M, Fujimoto M, Ihn H. Impaired IL-17 signaling pathway contributes to the increased collagen expression in scleroderma fibroblasts. *J Immunol* 2012; **188**: 3573-3583 [PMID: 22403442 DOI: 10.4049/jimmunol.1100591]
- 102 **Chew V**, Chen J, Lee D, Loh E, Lee J, Lim KH, Weber A, Slinkamenac K, Poon RT, Yang H, Ooi LL, Toh HC, Heikenwalder M, Ng IO, Nardin A, Abastado JP. Chemokine-driven lymphocyte infiltration: an early intratumoural event determining long-term survival in resectable hepatocellular carcinoma. *Gut* 2012; **61**: 427-438 [PMID: 21930732 DOI: 10.1136/gutjnl-2011-300509]
- 103 **Sun C**, Sun HY, Xiao WH, Zhang C, Tian ZG. Natural killer cell dysfunction in hepatocellular carcinoma and NK cell-based immunotherapy. *Acta Pharmacol Sin* 2015; **36**: 1191-1199 [PMID: 26073325 DOI: 10.1038/aps.2015.41]
- 104 **Levy EM**, Roberti MP, Mordoh J. Natural killer cells in human cancer: from biological functions to clinical applications. *J*

- Biomed Biotechnol* 2011; **2011**: 676198 [PMID: 21541191 DOI: 10.1155/2011/676198]
- 105 **Hobeika AC**, Etienne W, Torres BA, Johnson HM, Subramaniam PS. IFN-gamma induction of p21(WAF1) is required for cell cycle inhibition and suppression of apoptosis. *J Interferon Cytokine Res* 1999; **19**: 1351-1361 [PMID: 10638704 DOI: 10.1089/107999099312812]
- 106 **Beatty G**, Paterson Y. IFN-gamma-dependent inhibition of tumor angiogenesis by tumor-infiltrating CD4+ T cells requires tumor responsiveness to IFN-gamma. *J Immunol* 2001; **166**: 2276-2282 [PMID: 11160282]
- 107 **Seliger B**, Ruiz-Cabello F, Garrido F. IFN inducibility of major histocompatibility antigens in tumors. *Adv Cancer Res* 2008; **101**: 249-276 [PMID: 19055946 DOI: 10.1016/S0065-230X(08)00407-7]
- 108 **Martini M**, Testi MG, Pasetto M, Picchio MC, Innamorati G, Mazzocco M, Ugel S, Cingarlani S, Bronte V, Zanovello P, Krampera M, Mosna F, Cestari T, Riviera AP, Brutti N, Barbieri O, Matera L, Tridente G, Colombatti M, Sartoris S. IFN-gamma-mediated upmodulation of MHC class I expression activates tumor-specific immune response in a mouse model of prostate cancer. *Vaccine* 2010; **28**: 3548-3557 [PMID: 20304037 DOI: 10.1016/j.vaccine.2010.03.007]
- 109 **Whitmire JK**, Tan JT, Whitton JL. Interferon-gamma acts directly on CD8+ T cells to increase their abundance during virus infection. *J Exp Med* 2005; **201**: 1053-1059 [PMID: 15809350 DOI: 10.1084/jem.20041463]
- 110 **Reiter Z**. Interferon- γ a major regulator of natural killer cell-mediated cytotoxicity. *J Interferon Res* 1993; **13**: 247-257 [PMID: 7693829 DOI: 10.1089/jir.1993.13.247]
- 111 **Balkwill F**. Tumour necrosis factor and cancer. *Nat Rev Cancer* 2009; **9**: 361-371 [PMID: 19343034 DOI: 10.1038/nrc2628]
- 112 **Calzascia T**, Pellegrini M, Hall H, Sabbagh L, Ono N, Elford AR, Mak TW, Ohashi PS. TNF-alpha is critical for antitumor but not antiviral T cell immunity in mice. *J Clin Invest* 2007; **117**: 3833-3845 [PMID: 17992258 DOI: 10.1172/JCI32567]
- 113 **Dadi S**, Chhangawala S, Whitlock BM, Franklin RA, Luo CT, Oh SA, Toure A, Pritykin Y, Huse M, Leslie CS, Li MO. Cancer Immunoreveillance by Tissue-Resident Innate Lymphoid Cells and Innate-like T Cells. *Cell* 2016; **164**: 365-377 [PMID: 26806130 DOI: 10.1016/j.cell.2016.01.002]
- 114 **Trabanelli S**, Curti A, Lecciso M, Salomé B, Riether C, Ochsenbein A, Romero P, Jandus C. CD127+ innate lymphoid cells are dysregulated in treatment naïve acute myeloid leukemia patients at diagnosis. *Haematologica* 2015; **100**: e257-e260 [PMID: 25710455 DOI: 10.3324/haematol.2014.119602]
- 115 **Zaidi MR**, Merlino G. The two faces of interferon- γ in cancer. *Clin Cancer Res* 2011; **17**: 6118-6124 [PMID: 21705455 DOI: 10.1158/1078-0432.CCR-11-0482]
- 116 **Leibovich SJ**, Polverini PJ, Shepard HM, Wiseman DM, Shively V, Nuseir N. Macrophage-induced angiogenesis is mediated by tumour necrosis factor-alpha. *Nature* 1987; **329**: 630-632 [PMID: 2443857 DOI: 10.1038/329630a0]
- 117 **Wang H**, Wang HS, Zhou BH, Li CL, Zhang F, Wang XF, Zhang G, Bu XZ, Cai SH, Du J. Epithelial-mesenchymal transition (EMT) induced by TNF- α requires AKT/GSK-3 β -mediated stabilization of snail in colorectal cancer. *PLoS One* 2013; **8**: e56664 [PMID: 23431386 DOI: 10.1371/journal.pone.0056664]
- 118 **Gao Y**, Souza-Fonseca-Guimaraes F, Bald T, Ng SS, Young A, Ngui SF, Rautela J, Straube J, Waddell N, Blake SJ, Yan J, Bartholin L, Lee JS, Vivier E, Takeda K, Messaoudene M, Zitvogel L, Teng MWL, Belz GT, Engwerda CR, Huntington ND, Nakamura K, Hölzel M, Smyth MJ. Tumor immunoevasion by the conversion of effector NK cells into type 1 innate lymphoid cells. *Nat Immunol* 2017; **18**: 1004-1015 [PMID: 28759001 DOI: 10.1038/ni.3800]
- 119 **Cortez VS**, Ulland TK, Cervantes-Barragan L, Bando JK, Robinette ML, Wang Q, White AJ, Gilfillan S, Cella M, Colonna M. SMAD4 impedes the conversion of NK cells into ILC1-like cells by curtailing non-canonical TGF- β signaling. *Nat Immunol* 2017; **18**: 995-1003 [PMID: 28759002 DOI: 10.1038/ni.3809]
- 120 **Li J**, Bessho K, Shivakumar P, Mourya R, Mohanty SK, Dos Santos JL, Miura IK, Porta G, Bezerra JA. Th2 signals induce epithelial injury in mice and are compatible with the biliary atresia phenotype. *J Clin Invest* 2011; **121**: 4244-4256 [PMID: 22005305 DOI: 10.1172/JCI57728]
- 121 **Li J**, Razumilava N, Gores GJ, Walters S, Mizuochi T, Mourya R, Bessho K, Wang YH, Glaser SS, Shivakumar P, Bezerra JA. Biliary repair and carcinogenesis are mediated by IL-33-dependent cholangiocyte proliferation. *J Clin Invest* 2014; **124**: 3241-3251 [PMID: 24892809 DOI: 10.1172/JCI73742]
- 122 **Yamada D**, Rizvi S, Razumilava N, Bronk SF, Davila JI, Champion MD, Borad MJ, Bezerra JA, Chen X, Gores GJ. IL-33 facilitates oncogene-induced cholangiocarcinoma in mice by an interleukin-6-sensitive mechanism. *Hepatology* 2015; **61**: 1627-1642 [PMID: 25580681 DOI: 10.1002/hep.27687]
- 123 **Zhang P**, Liu XK, Chu Z, Ye JC, Li KL, Zhuang WL, Yang DJ, Jiang YF. Detection of interleukin-33 in serum and carcinoma tissue from patients with hepatocellular carcinoma and its clinical implications. *J Int Med Res* 2012; **40**: 1654-1661 [PMID: 23206447 DOI: 10.1177/030006051204000504]
- 124 **Brunner SM**, Schiechl G, Falk W, Schlitt HJ, Geissler EK, Fichtner-Feigl S. Interleukin-33 prolongs allograft survival during chronic cardiac rejection. *Transpl Int* 2011; **24**: 1027-1039 [PMID: 21797940 DOI: 10.1111/j.1432-2277.2011.01306.x]
- 125 **Gabitass RF**, Annels NE, Stocken DD, Pandha HA, Middleton GW. Elevated myeloid-derived suppressor cells in pancreatic, esophageal and gastric cancer are an independent prognostic factor and are associated with significant elevation of the Th2 cytokine interleukin-13. *Cancer Immunol Immunother* 2011; **60**: 1419-1430 [PMID: 21644036 DOI: 10.1007/s00262-011-1028-0]
- 126 **Terabe M**, Matsui S, Park JM, Mamura M, Noben-Trauth N, Donaldson DD, Chen W, Wahl SM, Ledbetter S, Pratt B, Letterio JJ, Paul WE, Berzofsky JA. Transforming growth factor-beta production and myeloid cells are an effector mechanism through which CD1d-restricted T cells block cytotoxic T lymphocyte-mediated tumor immunosurveillance: abrogation prevents tumor recurrence. *J Exp Med* 2003; **198**: 1741-1752 [PMID: 14657224 DOI: 10.1084/jem.20022227]
- 127 **Dhakal M**, Hardaway JC, Guloglu FB, Miller MM, Hoeman CM, Zaghoulani AA, Wan X, Rowland LM, Cascio JA, Sherman MP, Zaghoulani H. IL-13R α 1 is a surface marker for M2 macrophages influencing their differentiation and function. *Eur J Immunol* 2014; **44**: 842-855 [PMID: 24281978 DOI: 10.1002/eji.201343755]
- 128 **Qian BZ**, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* 2010; **141**: 39-51 [PMID: 20371344 DOI: 10.1016/j.cell.2010.03.014]
- 129 **Monticelli LA**, Osborne LC, Noti M, Tran SV, Zaiss DM, Artis D. IL-33 promotes an innate immune pathway of intestinal tissue protection dependent on amphiregulin-EGFR interactions. *Proc Natl Acad Sci U S A* 2015; **112**: 10762-10767 [PMID: 26243875 DOI: 10.1073/pnas.1509070112]
- 130 **Zaiss DM**, van Loosdregt J, Gorlani A, Bekker CP, Gröne A, Sibilina M, van Bergen en Henegouwen PM, Roovers RC, Coffey PJ, Sijts AJ. Amphiregulin enhances regulatory T cell-suppressive function via the epidermal growth factor receptor. *Immunity* 2013; **38**: 275-284 [PMID: 23333074 DOI: 10.1016/j.immuni.2012.09.023]
- 131 **Gao X**, Wang X, Yang Q, Zhao X, Wen W, Li G, Lu J, Qin W, Qi Y, Xie F, Jiang J, Wu C, Zhang X, Chen X, Turnquist H, Zhu Y, Lu B. Tumoral expression of IL-33 inhibits tumor growth and modifies the tumor microenvironment through CD8+ T and NK cells. *J Immunol* 2015; **194**: 438-445 [PMID: 25429071 DOI: 10.4049/jimmunol.1401344]
- 132 **Villarreal DO**, Wise MC, Walters JN, Reuschel EL, Choi MJ, Obeng-Adjei N, Yan J, Morrow MP, Weiner DB. Alarmin IL-33 acts as an immunoadjuvant to enhance antigen-specific tumor immunity. *Cancer Res* 2014; **74**: 1789-1800 [PMID: 24448242 DOI: 10.1158/0008-5472.CAN-13-2729]
- 133 **Kim J**, Kim W, Moon UJ, Kim HJ, Choi HJ, Sin JI, Park NH, Cho

- HR, Kwon B. Intratumorally Establishing Type 2 Innate Lymphoid Cells Blocks Tumor Growth. *J Immunol* 2016; **196**: 2410-2423 [PMID: 26829987 DOI: 10.4049/jimmunol.1501730]
- 134 **Carrega P**, Loiacono F, Di Carlo E, Scaramuccia A, Mora M, Conte R, Benelli R, Spaggiari GM, Cantoni C, Campana S, Bonaccorsi I, Morandi B, Truini M, Mingari MC, Moretta L, Ferlazzo G. NCR(+)ILC3 concentrate in human lung cancer and associate with intratumoral lymphoid structures. *Nat Commun* 2015; **6**: 8280 [PMID: 26395069 DOI: 10.1038/ncomms9280]
- 135 **Moskalenko M**, Pan M, Fu Y, de Moll EH, Hashimoto D, Mortha A, Leboeuf M, Jayaraman P, Bernardo S, Sikora AG, Wolchok J, Bhardwaj N, Merad M, Saenger Y. Requirement for innate immunity and CD90⁺ NK1.1⁺ lymphocytes to treat established melanoma with chemo-immunotherapy. *Cancer Immunol Res* 2015; **3**: 296-304 [PMID: 25600438 DOI: 10.1158/2326-6066.CIR-14-0120]
- 136 **Kirchberger S**, Royston DJ, Boulard O, Thornton E, Franchini F, Szabady RL, Harrison O, Powrie F. Innate lymphoid cells sustain colon cancer through production of interleukin-22 in a mouse model. *J Exp Med* 2013; **210**: 917-931 [PMID: 23589566 DOI: 10.1084/jem.20122308]
- 137 **Huber S**, Gagliani N, Zenewicz LA, Huber FJ, Bosurgi L, Hu B, Hedl M, Zhang W, O'Connor W Jr, Murphy AJ, Valenzuela DM, Yancopoulos GD, Booth CJ, Cho JH, Ouyang W, Abraham C, Flavell RA. IL-22BP is regulated by the inflammasome and modulates tumorigenesis in the intestine. *Nature* 2012; **491**: 259-263 [PMID: 23075849 DOI: 10.1038/nature11535]
- 138 **Jiang R**, Tan Z, Deng L, Chen Y, Xia Y, Gao Y, Wang X, Sun B. Interleukin-22 promotes human hepatocellular carcinoma by activation of STAT3. *Hepatology* 2011; **54**: 900-909 [PMID: 21674558 DOI: 10.1002/hep.24486]
- 139 **Ren X**, Hu B, Colletti LM. IL-22 is involved in liver regeneration after hepatectomy. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G74-G80 [PMID: 19875704 DOI: 10.1152/ajpgi.00075.2009]
- 140 **Ye ZJ**, Zhou Q, Yin W, Yuan ML, Yang WB, Xiang F, Zhang JC, Xin JB, Xiong XZ, Shi HZ. Interleukin 22-producing CD4⁺ T cells in malignant pleural effusion. *Cancer Lett* 2012; **326**: 23-32 [PMID: 22809567 DOI: 10.1016/j.canlet.2012.07.013]
- 141 **Nestle FO**, Kaplan DH, Barker J. Psoriasis. *N Engl J Med* 2009; **361**: 496-509 [PMID: 19641206 DOI: 10.1056/NEJMra0804595]
- 142 **Lim C**, Savan R. The role of the IL-22/IL-22R1 axis in cancer. *Cytokine Growth Factor Rev* 2014; **25**: 257-271 [PMID: 24856143 DOI: 10.1016/j.cytogfr.2014.04.005]
- 143 **Waidmann O**, Kronenberger B, Scheiermann P, Köberle V, Mühl H, Piiper A. Interleukin-22 serum levels are a negative prognostic indicator in patients with hepatocellular carcinoma. *Hepatology* 2014; **59**: 1207 [PMID: 23729376 DOI: 10.1002/hep.26528]
- 144 **Chan IH**, Jain R, Tessmer MS, Gorman D, Mangadu R, Sathe M, Vives F, Moon C, Penaflor E, Turner S, Ayanoglu G, Chang C, Basham B, Mumm JB, Pierce RH, Yearley JH, McClanahan TK, Phillips JH, Cua DJ, Bowman EP, Kastelein RA, LaFace D. Interleukin-23 is sufficient to induce rapid de novo gut tumorigenesis, independent of carcinogens, through activation of innate lymphoid cells. *Mucosal Immunol* 2014; **7**: 842-856 [PMID: 24280935 DOI: 10.1038/mi.2013.101]
- 145 **Zhang JP**, Yan J, Xu J, Pang XH, Chen MS, Li L, Wu C, Li SP, Zheng L. Increased intratumoral IL-17-producing cells correlate with poor survival in hepatocellular carcinoma patients. *J Hepatol* 2009; **50**: 980-989 [PMID: 19329213 DOI: 10.1016/j.jhep.2008.12.033]
- 146 **Song Y**, Yang JM. Role of interleukin (IL)-17 and T-helper (Th)17 cells in cancer. *Biochem Biophys Res Commun* 2017; **493**: 1-8 [PMID: 28859982 DOI: 10.1016/j.bbrc.2017.08.109]
- 147 **Satoh T**, Tajima M, Wakita D, Kitamura H, Nishimura T. The development of IL-17/IFN- γ -double producing CTLs from Tc17 cells is driven by epigenetic suppression of Soes3 gene promoter. *Eur J Immunol* 2012; **42**: 2329-2342 [PMID: 22674086 DOI: 10.1002/eji.201142240]

P- Reviewer: Carvalho-Filho RJ, Osna NA **S- Editor:** Gong ZM
L- Editor: A **E- Editor:** Huang Y





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>



ISSN 1007-9327

