



# The Application and Limitation of Universal Chloroplast Markers in Discriminating East Asian Evergreen Oaks

Mengxiao Yan<sup>1</sup>, Yanshi Xiong<sup>1</sup>, Ruibin Liu<sup>1,2</sup>, Min Deng<sup>1\*</sup> and Jiaojiao Song<sup>1,3</sup>

<sup>1</sup> Shanghai Chenshan Plant Science Research Center, Chinese Academy of Sciences, Shanghai Chenshan Botanical Garden, Shanghai, China, <sup>2</sup> College of Life and Environmental Sciences, Shanghai Normal University, Shanghai, China, <sup>3</sup> College of Life Sciences, Shangrao Normal University, Shangrao, China

## OPEN ACCESS

### Edited by:

Rosane Garcia Collevatti,  
Universidade Federal de Goiás, Brazil

### Reviewed by:

Andrea Cecilia Premoli,  
Universidad Nacional del Comahue,  
Argentina

Glaucia Salles Cortopassi Buso,  
Embrapa Genetic Resources  
and Biotechnology, Brazil

### \*Correspondence:

Min Deng  
dengmin@sibs.ac.cn

### Specialty section:

This article was submitted to  
Evolutionary and Population Genetics,  
a section of the journal  
Frontiers in Plant Science

**Received:** 26 January 2018

**Accepted:** 11 April 2018

**Published:** 08 May 2018

### Citation:

Yan M, Xiong Y, Liu R, Deng M and  
Song J (2018) The Application  
and Limitation of Universal  
Chloroplast Markers in Discriminating  
East Asian Evergreen Oaks.  
Front. Plant Sci. 9:569.  
doi: 10.3389/fpls.2018.00569

The East Asian subtropics mostly occupied by evergreen broad-leaved forests (EBLFs), is one of the global diversity centers for evergreen oaks. Evergreen oaks are keystone canopy trees in EBLFs with important ecosystem function and crucial significance for regional biodiversity conservation. However, the species composition and diversity of Asian evergreen oaks are poorly understood. Here, we test whether the four chloroplast markers *atpI-atpH*, *matK*, *psbA-trnH*, and *ycf1*, can discriminate the two evergreen oak sections in Asia – *Cyclobalanopsis* and *Ilex*. Two hundred and seventy-two individuals representing 57 species were scanned and 17 species from other oaks sections were included for phylogenetic reconstruction. The genetic diversity of the *Quercus* sections was also compared. Overall, we found that universal chloroplast DNA (*cpDNA*) barcoding markers could resolve two clades in *Quercus*, i.e., subgenus *Cerris* (Old World Clade) and subgenus *Quercus* (New World Clade). The chloroplast markers distinguished the main sections, with few exceptions. Each *cpDNA* region showed no barcoding gap and none of them provided good resolution at the species level. The best species resolution (27.78%) was obtained when three or four markers were combined and analyzed using BLAST. The high conservation of the *cpDNA* and complicated evolutionary patterns, due to incomplete lineage sorting, interspecific hybridization and introgressions may hinder the ability of *cpDNA* markers to discriminate different species. When comparing diversification pattern across *Quercus* sections (*Cyclobalanopsis*, *Ilex*, *Cerris*, *Quercus*, and *Protobalanus*), we found that section *Ilex* was the most genetically diverse, and section *Cyclobalanopsis* was lower genetically diverse. This diversification pattern may have resulted from the interplay of the Eurasia Cenozoic tectonic movements, climate changes and different niches of their ancestral lineages.

**Keywords:** *cpDNA*, DNA barcoding, genetic diversity, *Quercus*, section *Cyclobalanopsis*, section *Ilex*

## INTRODUCTION

Understanding the biodiversity of ecosystems is critical for revealing the biome assembly and its function in different ecosystems (Hebert et al., 2003; Lahaye et al., 2008; Pitman et al., 2008). Taxonomy based on morphological features has enriched our understanding of biodiversity for over 300 years, and provide an inventory of global biodiversity. However, the morphology based

**Abbreviations:** *cpDNA*, chloroplast DNA; EBLF, evergreen broad-leaved forest.

taxonomy system has shortcomings (Hebert et al., 2003; Vilgalys, 2003; Gonzalez et al., 2009; Goldstein and DeSalle, 2011). For example, the key diagnostic traits may be under selection or subjected to parallel evolution providing false information on the identity of the taxa. Complementing techniques such as DNA sequences under neutral evolution have become an efficient way to identify species (Hollingsworth, 2007; CBOL Plant Working Group, 2009; Valentini et al., 2009; Hollingsworth et al., 2016). DNA barcoding has become instrumental in plant research for identification of cryptic species (Bickford et al., 2007; Miwa et al., 2009; Liu et al., 2011), biodiversity assessments (Lahaye et al., 2008; Gonzalez et al., 2009; von Crautlein et al., 2011; Yan et al., 2015), community phylogeny (Kress et al., 2009, 2010), conservation biology (Laiou et al., 2013; Veldman et al., 2014; Liu et al., 2015; Shapcott et al., 2015), invasive biology (Armstrong and Ball, 2005; Bleeker et al., 2008; Newmaster and Ragupathy, 2009; Van De Wiel et al., 2009), and disease and pest management (Ball and Armstrong, 2006; Lee et al., 2012; Montecchio and Faccoli, 2014; Shapcott et al., 2015). At present, the resolution of several candidate DNA barcodes has been tested in many plant groups (Clerc-Blain et al., 2010; Gao et al., 2010; Liu et al., 2011; Yu et al., 2011; Wang D.Y. et al., 2017). However, there is a challenge for DNA barcoding to discriminate closely related species, especially for those with shallow phylogeny and complex evolutionary history (Newmaster et al., 2008; Chen et al., 2015; Yan et al., 2015), such as species-rich genera, *Quercus*.

The genus *Quercus* (Fagaceae) has ca. 400–500 species, which are widely distributed in the warm temperate forests of the Northern Hemisphere (Camus, 1936–1954; Nixon, 1993). Many species of the genus are the canopy or dominant trees in regional EBLFs and play an important role in the function and service of ecosystem (Fang and Yoda, 1991; Nixon, 2006; Petit et al., 2013). The species identification and taxonomy in oaks are notoriously difficult (Manos et al., 1999; Simeone et al., 2013; Hubert et al., 2014), especially for Asian evergreen oaks that are composed of two species-rich sections (*Cyclobalanopsis* and *Ilex*) (Denk and Grimm, 2010; Denk et al., 2017). Species of section *Cyclobalanopsis* are the dominant trees in Asian (sub)tropical EBLFs with ca. 90–120 species (Denk and Grimm, 2010; Denk et al., 2017). Species of section *Ilex* are widely distributed in Eurasian low-middle latitude habitats comprising ca. 35–36 species (Denk and Grimm, 2010; Simeone et al., 2016). Unfortunately, the wide range of EBLFs in East Asia is now greatly diminished as a result of intensified human activities (Cao and Zhang, 1997; Tang, 2010). At least 1/3 oak species in China are now endangered or threatened due to the severe habitats loss and human activities (Deng et al., 2013a). Therefore, understanding the species composition of East Asian evergreen oak forests is important for future conservation efforts. Although the Flora of China and other regional floristic works had been finished for decades (e.g., Huang et al., 1999; Phengklai et al., 2008), the oak taxonomy and their systematic placement are still not well resolved, in part due to the high level of intra- and inter-species genetic variation. Even the fine anatomy features of the East Asian evergreen oaks, such as leaf epidermal features (Deng et al., 2014, 2017b), pollen morphology (Denk and Grimm, 2009; Deng et al., 2013b; Denk and Tekleva, 2014), and wood

anatomy (Zhao et al., 2007) had been comprehensively studied in recent years. But these works do not provide useful diagnostic features to clarify species identity, and suggest paraphyletic evolution of taxonomical traits. Moreover, the interspecies gene flow has blunted genetic integrity and reduced the ability to distinguish some East Asian evergreen oaks (Tamaki and Okada, 2014; An et al., 2017).

Chloroplast DNA sequences have been applied for phylogenetic analysis and DNA barcoding of species of section *Ilex* in the Mediterranean (Piredda et al., 2011; Simeone et al., 2013) and China (Yang et al., 2017), providing insights into the evolutionary history of oak taxa. However, these studies included limited taxa and geographic regions (Europe or small region of China), therefore did not provide enough information on DNA barcoding for evergreen oaks. For the species-rich section *Cyclobalanopsis* in East Asia, little is known about evolution of chloroplast genome and phylogeny. So far, only the intergenic region of the chloroplast *trnT-trnL* was applied to distinguish six species of section *Cyclobalanopsis* distributed in Japan, and proved to have moderate efficiency (Ohyama et al., 2001). Recent studies using *cpDNA* show that the species of section *Cyclobalanopsis*, e.g., *Q. glauca* (Xu et al., 2015), *Q. schottkyana* (Jiang et al., 2016), *Q. arbutifolia* (Xu et al., 2016), and *Q. kerrii* (Jiang et al., 2018), contain high variable regions to infer population history, but the resolution of *cpDNA* markers in discriminating section *Cyclobalanopsis* species is still unknown.

Evergreen broad-leaved forests potentially cover a wide zone of monsoon-dominated regions in East Asia (Fang and Yoda, 1991). This region, as a global biodiversity hotspot, has the world's richest flora harboring remarkable array of endemic, relic and endangered species (Myers et al., 2000; Yang et al., 2004; Lopez-Pujol et al., 2006), especially with many endemic and endangered oak species (Menitsky, 1984; Luo and Zhou, 2000; Deng et al., 2013a). Effective DNA barcodes for species identification, conservation and resource utilization are extremely necessary in East Asian evergreen oaks. A further DNA barcoding study with comprehensive sampling on species population from the main distribution ranges is needed. In addition, comparing of the genetic diversity patterns across the main lineages in *Quercus* can provide more information to infer the driving forces which might contribute to their divergence. Furthermore, the knowledge on biodiversity inventory in East Asian subtropics is far from enough, and new species are continuously being identified in recent years (e.g., Xiao et al., 2015; Liang et al., 2016; Jiang et al., 2017; Wang C.W. et al., 2017). DNA barcoding of keystone species in this region can provide crucial insight into the mechanism of plant community assembly and evolutionary trajectory of the East Asian regional biota.

In this study, we comprehensively collected East Asian evergreen oaks aiming to address the following issues: (1) to reveal the discrimination ability of *cpDNA* barcode markers in East Asian evergreen oaks; (2) to compare the genetic diversity of the main sections in *Quercus*; (3) to explore the possible applications of *cpDNA* markers into the taxonomic and phylogenetic approaches. Our study provides insights into the species identity of Asian oaks and important information for biogeographic and population genetics of these unique oaks.

## MATERIALS AND METHODS

### Ethics Statement

Sampling of oak species and other Fagaceae plants were granted and supported by National Forestry Bureau of China and Local National Nature Reserves.

### Plant Materials

One hundred and forty-seven individuals belonging to 29 species of *Quercus* section *Cyclobalanopsis* from East Asia and 125 individuals of the 28 species of section *Ilex* from East Asia were included in our studies. The sampling range covered the key distribution range of East Asian evergreen oaks in China, Japan, Vietnam, and Nepal. Additionally, individuals of five species of section *Ilex* from the Mediterranean were analyzed. The final dataset also included four species of section *Cerris* (18 individuals, from Eurasia), seven species of section *Quercus* (11 individuals, from North America and Eurasia), five species of the American section *Lobatae* (7 individuals), and one species (1 individual) of section *Protobalanus* from western North America. One individual of *Lithocarpus henryi* was used as outgroup to root the tree of genus *Quercus*. Sequences of eight species (16 individuals) were downloaded from GenBank. Detailed information of samples used in this study is listed in Supplementary Table S1.

Healthy leaves of each individual were collected and dried instantly in silica gel for DNA extraction. Voucher specimens of each individual were deposited in the Herbarium of the Shanghai Chenshan Botanical Garden (CSH).

### DNA Extraction, PCR, and Sequencing Protocols

Genomic DNA was extracted using the modified CTAB method (Doyle and Doyle, 1987). Four *cpDNA* regions, the *psbA-trnH* intergenic spacer, a part of the *matK* gene, the *atpI-atpH* intergenic spacer and a portion of the *ycf1* region were amplified using PCR and bidirectionally sequenced for analyses. PCR reactions were performed according to previously described method (Xu et al., 2015). The primer sequences used to amplify these regions are summarized in Supplementary Table S2. The sequences obtained in this study have been uploaded to GenBank (accession numbers: MH058100-MH059477).

Sequencher 4.01 (Gene Codes Corp., Ann Arbor, MI, United States) was used to assemble and edit sequences. The DNA sequences were aligned using the package Muscle (Edgar, 2004) implemented in MEGA 7.0.21<sup>1</sup> (Kumar et al., 2016) with subsequent manual adjustment.

### DNA Barcoding Analyses

DNA polymorphisms were examined using DnaSP 5.10 (Librado and Rozas, 2009). To evaluate species discrimination success, four widely used methods, genetic distance-based, similarity-based, tree-based and diagnostic method, were applied to the four *cpDNA* markers and all their possible combinations.

Genetic distance-based method: *p*-distances of the four plastid regions were calculated in MEGA 7.0.21 (Kumar et al., 2016) (see footnote 1). We measured three parameters to determine intraspecific variation (Meyer and Paulay, 2005; Lahaye et al., 2008). First, we calculated average intraspecific difference between all samples collected. Second, we measured theta ( $\theta$ ), which means the average *p*-distance within each species. Theta eliminates biases resulted by unequal sampling between species. Lastly, we calculated average coalescent depth, which is the maximum intraspecific distance within each species. Average interspecific distance and smallest interspecific distance were used to characterize interspecific divergence (Meier et al., 2008). The distribution of intraspecific versus interspecific variability was compared using DNA barcoding gaps. Differentiations between the intraspecific and interspecific *p*-distance for each candidate barcode were also compared using the Wilcoxon signed rank tests in R 3.2.3 (R Core Team, 2015). To explore potential species groups, the obtained *p*-distance matrices for each barcode candidate and all combinations were analyzed using the Automatic Barcode Gap Discovery (ABGD) under default setting (Puillandre et al., 2012). If conspecific individuals were partitioned into the same group without sequences from other species, the species was considered as successfully delimited.

Similarity-based method: Sequences of the four candidate barcodes and all possible combinations were built as 15 local reference databases using NCBI-blast-2.6.0+ (Camacho et al., 2009; Tao, 2010). Each barcode sequence was then queried using the blastn command against its local reference database. When all individuals of a species had a top hit to conspecific individuals, species discrimination was considered successful.

Tree-based method: Bayesian trees were constructed using MrBayes 3.2.6 (Ronquist et al., 2012). Model of substitution was selected by Modeltest 3.7 (Posada and Crandall, 1998) based on the akaike information criterion (AIC). Two parallel Markov Chain Monte Carlo (MCMC) runs were performed for 20 million generations. The trees were sampled every 1,000 generations and inspected in Tracer v1.6<sup>2</sup>. The first 15% trees were discarded as burn-in. Species discrimination was considered as successful only when all the conspecific individuals formed a monophyletic clade.

Diagnostic method: BLOG 2.0 (Barcoding with LOGic) was used for species identification under a Logic Mining method, which identifies the species in terms of location of key diagnostic nucleotides from the training sequences to classify species (Weitschek et al., 2013). Each candidate barcode and combination were tested with a single file input, with 80% of the sequences as the training data, and 20% as the test data. A species was considered as successfully delimited if conspecific individuals were correctly identified in both training and test datasets.

### Phylogeographical and Genetic Diversity Analysis

Haplotypes were extracted by DnaSP 5.10 (Librado and Rozas, 2009). To measure the level of genetic variation, variable sites,

<sup>1</sup><http://www.megasoftware.net/>

<sup>2</sup><http://tree.bio.ed.ac.uk/software/tracer/>

average pairwise differences per base pair between sequence (nucleotide diversity) (Nei and Li, 1979) and haplotype diversity (Hd) were calculated using DnaSP 5.10 (Librado and Rozas, 2009). The genetic distances (*p*-distances) of each section were calculated in MEGA 7.0.21 (Kumar et al., 2016) (see footnote 1) for further comparison of genetic diversity. A principal coordinate analysis (PCoA) was performed using GenALEx 6.5 (Peakall and Smouse, 2012) to illustrate both the similarity between different sections and the genetic diversity of each section.

## RESULTS

### PCR Success and Sequence Characteristics

The four chloroplast candidate barcodes showed high amplification success rates (100%) at the species level. At individual level, *matK*, *psbA-trnH*, and *ycf1* had high success rates (92.2–100%) (Table 1), while the *atpI-atpH* had the lowest success rate, potentially due to the long PCR product that was being amplified.

A total of 1,439 sequences were available for further analysis, of which 1,375 were newly generated in this study. Among the four *cpDNA* markers, the aligned lengths ranged from 554 bp for the *matK* to 1,187 bp for the *atpI-atpH*. The *atpI-atpH* region showed the highest number of variable sites, while the *matK* and *psbA-trnH* had the lowest variable sites (Table 1). The *atpI-atpH* region also had the highest number of indels (99), followed by *ycf1* (43), *psbA-trnH* (26) and *matK* (3). Among the four chloroplast loci, *psbA-trnH* was the highest haplotype diversity, followed by *atpI-atpH*, *ycf1* and *matK* (in descending order of Hd).

### Genetic Divergence Within and Between Species

*PsbA-trnH* exhibited the highest level of intra-species variation, followed by *ycf1*, *atpI-atpH* and *matK*. When comparing the interspecific genetic divergence among the four candidate barcodes, *psbA-trnH* region exhibited the highest interspecific divergence, followed by *ycf1*, *atpI-atpH*, and *matK* (Table 1). This analysis demonstrated that *psbA-trnH* sequences provide the most suitable DNA barcodes.

### DNA Barcoding Gap Assessment

The distribution of intraspecific and interspecific variation of four single markers lacked a distinct gap. Among the single locus, *psbA-trnH* had the highest variation between the distribution range of interspecific and intraspecific distances (Figure 1). Wilcoxon rank sum tests show that the intraspecific distances were always significantly lower than the interspecific distances (Table 2). However, the maximum intraspecific distances of four loci were higher than the minimum interspecific distance (Table 2).

### Species Identification Efficiency of Chloroplast Loci

Rates of species resolution depended on the analytic methods used (Figure 2 and Supplementary Table S3). Overall, BLAST provided the highest species discrimination rates (6.76–27.78%) among single marker or combined markers, followed by distance-based method (6.76–21.62%) and tree-based (0–20.27%) method. BLOG provided the lowest species resolution (1.35–12.16%) overall (Figure 2 and Supplementary Table S3). The diagnostic nucleotide of the highest species identification identified by BLOG is listed in Supplementary Table S4.

Among the four single markers, *psbA-trnH* region showed the highest species discrimination rate for all methods except the distance-based method. When distance-based method was used, *ycf1* exhibited the highest species discrimination rate (Figure 2 and Supplementary Table S3). In general, combinations with more loci could increase the probability that a species is identified correctly. Using three different combinations (*atpI-atpH* + *matK* + *psbA-trnH*, *atpI-atpH* + *matK* + *ycf1* and *atpI-atpH* + *matK* + *psbA-trnH* + *ycf1*) with BLAST, they all generated the highest species identification rate (27.78%) (Figure 2 and Supplementary Table S3).

### Phylogenetic Relationship

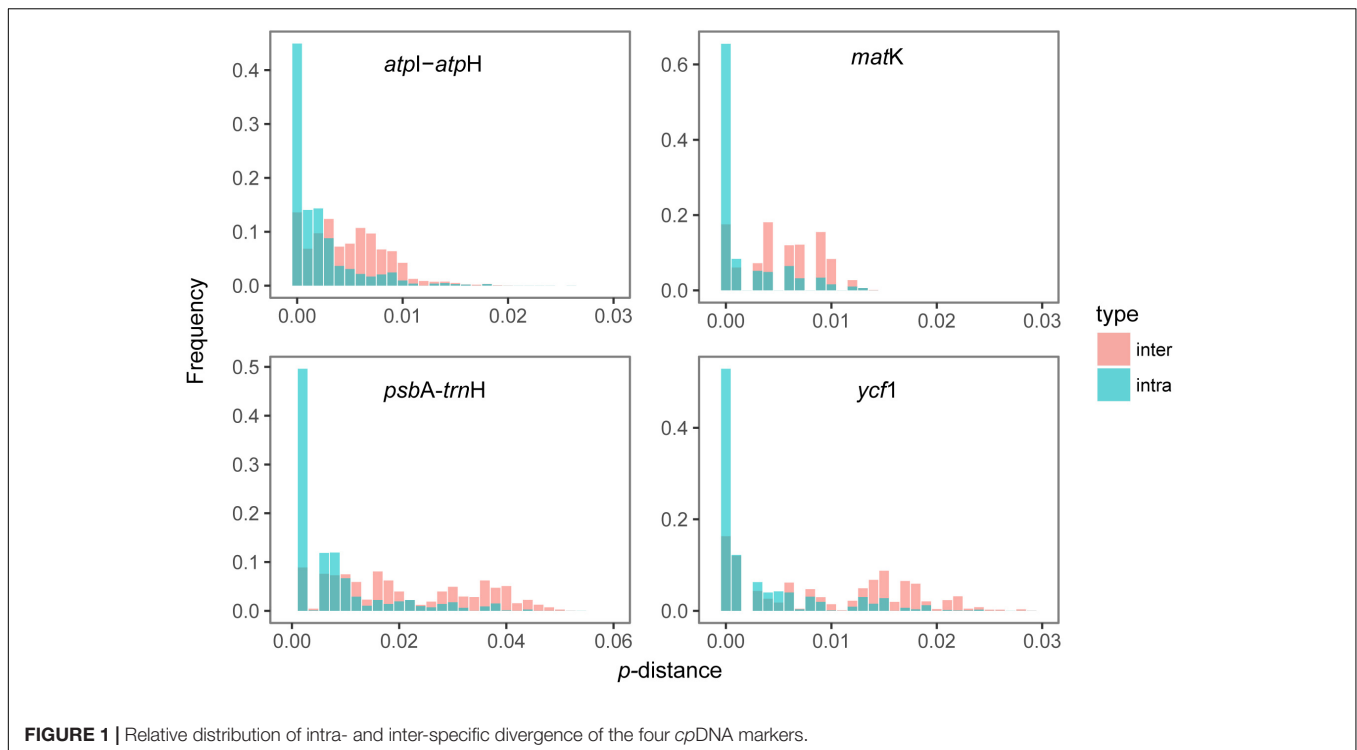
Subgenus *Quercus*, also known as the New World Clade that includes sections *Lobatae*, *Protobalanus*, and *Quercus*, formed a monophyletic clade (posterior probability = 1.00) (Figure 3). Section *Lobatae* was the only monophyletic clade (posterior probability = 1.00) inferred in this study. Two lineages were resolved in section *Quercus*, one with Eurasian species and the other one with the two American species (Figure 3). Section *Protobalanus* (*Q. chrysolepis*) was the sister group to the American lineage of section *Quercus* (Figure 3). However, we did

TABLE 1 | Characteristics of the four *cpDNA* markers used in this study.

Barcode	N	Ns	PCR	L	Vs		Indel	Pi	Hn	Hd
					Number	%				
<i>atpI-atpH</i>	345	72	92.2	1,187	128	10.78	99	0.00448	77	0.851
<i>matK</i>	374	74	100	691	64	9.26	3	0.00467	64	0.819
<i>psbA-trnH</i>	357	74	95.5	554	64	11.55	26	0.02012	68	0.905
<i>ycf1</i>	363	74	97.1	796	90	11.31	43	0.00954	66	0.827

N, number of samples; Ns, number of species; PCR success (%); L, aligned length; Vs, variable sites; Pi, nucleotide diversity; Hn, number of haplotypes; Hd, haplotype diversity.





**FIGURE 1** | Relative distribution of intra- and inter-specific divergence of the four cpDNA markers.

**TABLE 2** | Comparison of inter- and intraspecific  $p$ -distance of the four chloroplast markers and Wilcoxon rank sum test between inter- and intraspecific  $p$ -distance.

Barcode	Intraspecific distance (mean)	Theta (mean)	Coalescent depth (mean)	Interspecific distance (mean)	Wilcoxon rank sum test	
					W statistic	$p$ -value
<i>atpI-atpH</i>	0–0.0180 (0.0020)	0–0.0097 (0.0023)	0–0.0180 (0.0046)	0–0.0260 (0.0048)	15488000	<0.001
<i>matK</i>	0–0.0130 (0.0017)	0–0.0100 (0.0022)	0–0.0130 (0.0039)	0–0.0140 (0.0052)	20437000	<0.001
<i>psbA-trnH</i>	0–0.0430 (0.0069)	0–0.0380 (0.0082)	0–0.0430 (0.0145)	0–0.0530 (0.0201)	14029000	<0.001
<i>ycf1</i>	0–0.0240 (0.0030)	0–0.0153 (0.0036)	0–0.0240 (0.0072)	0–0.0300 (0.0095)	18544000	<0.001

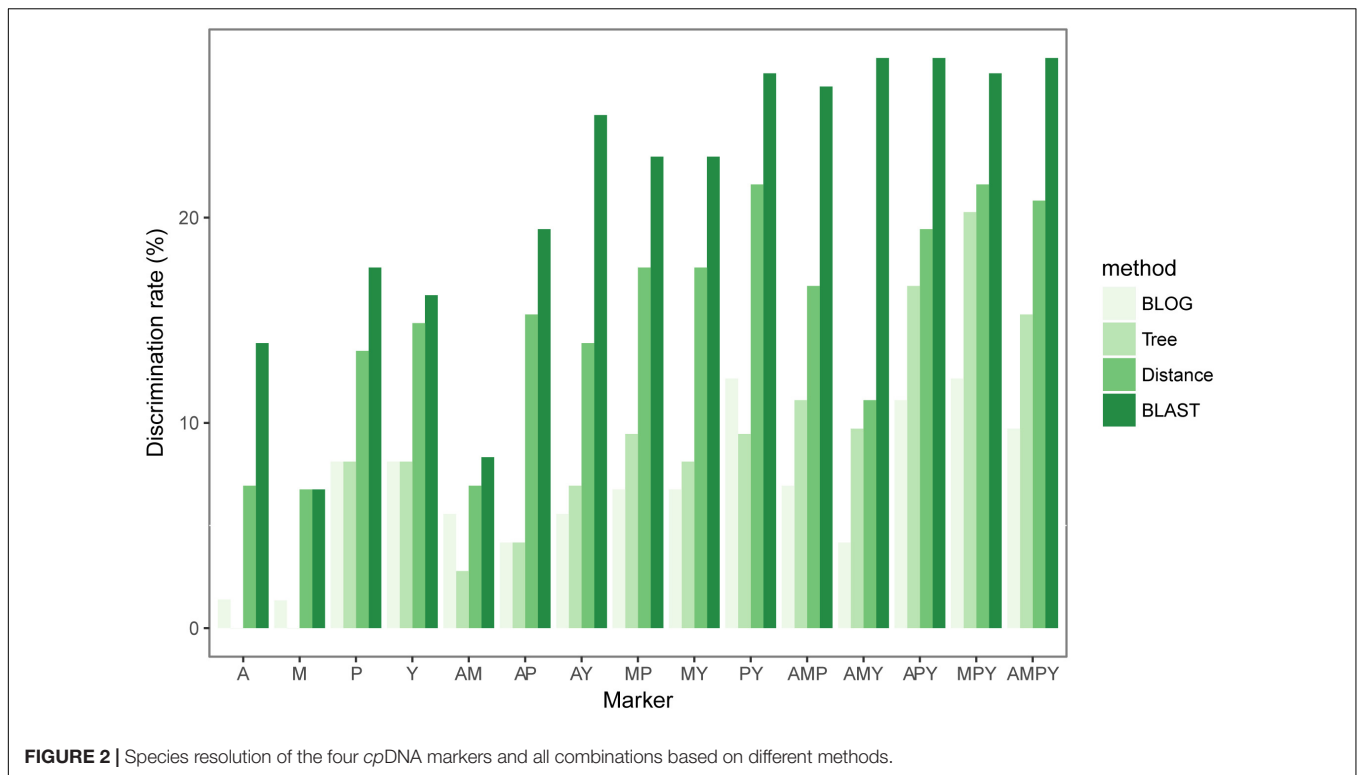
not find strong support for the monophyletic status of subgenus *Cerris* (the Old World Clade). Species in section *Cerris* almost formed a monophyletic lineage, except for two individuals. The Mediterranean *Q. suber* mixed with species of section *Ilex* and an individual of East Asian *Q. variabilis* nested in a sister lineage to a section *Ilex-Cyclobalanopsis* mixed lineage (Figure 3). We also inferred one main clade and 11 small clades in section *Cyclobalanopsis*. However, we did not find strong support for the monophyletic status of section *Cyclobalanopsis* (Figure 3). Species within the 11 small *Cyclobalanopsis* clades were mixed with section *Ilex* species. In turn, the cpDNA based phylogenetic reconstruction inferred that section *Ilex* was polyphyletic and its evolutionary history appeared rather complicated compared to the other sections in *Quercus* (Figure 3).

Similar to the phylogenetic reconstruction results, PCoA using genetic distance among all the six sections (Figure 4) showed that section *Cerris* is closely to a group of East Asian subtropical species, including *Q. acrodonta*, *Q. baronii*, *Q. phillyreoides*, *Q. dolicholepis*, and *Q. engleriana*. Species of section *Quercus* from Oregon showed close similarity to section *Lobatae*. The major clade of section *Cyclobalanopsis* was relatively isolated, with only

a few samples from Yunnan, SW China, are closely related to the individuals of section *Ilex* from the same region.

## Genetic Diversity Pattern

Section *Ilex* showed the highest diversity values for all the parameters (Table 3 and Figure 5), and formed three clusters in different quadrants on the PCoA plot (Figure 4). This suggests that section *Ilex* was the most diverse section in *Quercus*. Section *Quercus* was the second diverse section, with the second highest  $p$ -distance and haplotype diversity even only having a small number of samples (Table 3 and Figure 5). However, section *Cyclobalanopsis* with both high number of species and population sampling, yielded low genetic diversity levels, as the  $p$ -distance concentrated on low values (Figure 5) and the majority of individuals formed a small cluster in the PCoA analysis (Figure 5). Sections *Cerris* and *Lobatae* exhibited low genetic diversity as well (Table 3 and Figure 5). Nonetheless, we could not rule out the possibility that the low diversity might be caused by limited sampling, as we only covered four species and 18 individuals of section *Cerris* and five species (seven individuals) of section *Lobatae*.



## DISCUSSION

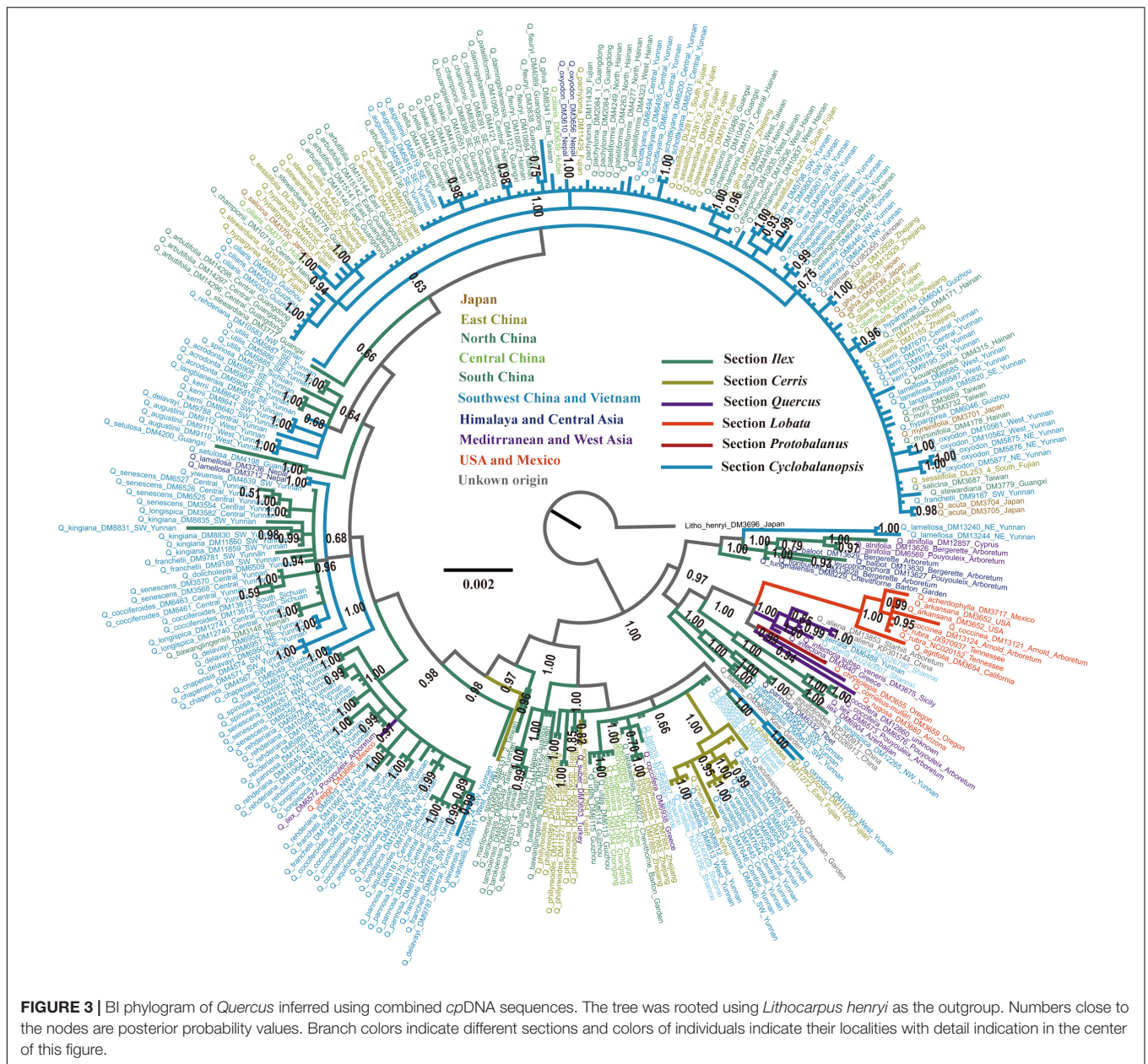
### The Application of cpDNA Markers on Oak Taxonomy and Systematics

We were able to successfully amplify and sequence the four cpDNA markers at high success rates (over 90%). There were differences in discrimination ability among each cpDNA marker. *PsbA-trnH* and *ycf1* had the highest discrimination ability whereas *matK* had the lowest. The chloroplast region *psbA-trnH* had the highest discriminatory power in other plant groups and community phylogeny (Kress et al., 2010; Hollingsworth et al., 2011). Three different combinations (*atpI-atpH* + *matK* + *psbA-trnH*, *atpI-atpH* + *matK* + *ycf1* and *atpI-atpH* + *matK* + *psbA-trnH* + *ycf1*) generated the highest species identification rate (27.78%) when used BLAST. Unfortunately, the overall identification success rates are not ideal and even the highest species resolution was below 30%. Meanwhile, none of the four chloroplast regions showed a barcode gap, which is an essential evaluation index for DNA barcodes (Hebert et al., 2003; Meier et al., 2006, 2008). The poor species resolution of universal chloroplast markers has been reported in East Asian oaks (Ohyama et al., 2001; Dong et al., 2015; Yang et al., 2017) and Euro-Mediterranean oaks (Piredda et al., 2011; Simeone et al., 2013), suggesting that these cpDNAs may not be ideal for resolving species identity in closely related oaks.

However, the universal cpDNA markers can provide information on resolving main infrageneric groups in oaks. First, two large clades of oaks, subgenera *Cerris* and *Quercus* could be distinguished when using the concatenated marker.

Second, section *Cerris* is almost recognizable among the East Asian oaks, because it formed a monophyletic lineage with only two exceptional individuals falling into different groups. Third, most individuals of section *Cyclobalanopsis* (84.4%) formed a large lineage that was easily distinguished from section *Ilex*. Few individuals of section *Ilex* showed close relationship with few species in section *Cyclobalanopsis* from SW China. In addition to the concentric rings and scales on the cupule wall, section *Cyclobalanopsis* and *Ilex* have distinct morphological traits, such as leaf architecture structures (Zhou et al., 1995; Luo and Zhou, 2002), leaf blade shape and margin teeth (Menitsky, 1984; Huang et al., 1999) that can easily be used to distinguish the two sections.

Chloroplast markers are effective barcodes in discriminating many plant species, such as Myristicaceae (Newmaster et al., 2008), *Pedicularis* (Yu et al., 2011), Asteraceae (Gao et al., 2010), *Carex* and *Kobresia* (Clerc-Blain et al., 2010). However, they have limitations when discriminating closely related species, which is quite common, not only in oaks but also in other plant taxa, such as *Salix* (Percy et al., 2014), *Rhododendron* (Yan et al., 2015), *Viburnum* (Clement and Donoghue, 2012), and *Curcuma* (Chen et al., 2015). In this study, various cpDNA haplotypes exist within a species and some of these haplotypes are shared interspecifically. Recent phylogeographic studies of the section *Ilex* from the Himalayas (Feng et al., 2016; Meng et al., 2017) and the pan-Mediterranean (Simeone et al., 2016; Vitelli et al., 2017) also showed similar scenario. Moreover, even using the complete plastid genome sequences for phylogenetic reconstruction, the monophyletic status of species was still rare in section *Quercus* (Pham et al., 2017). The

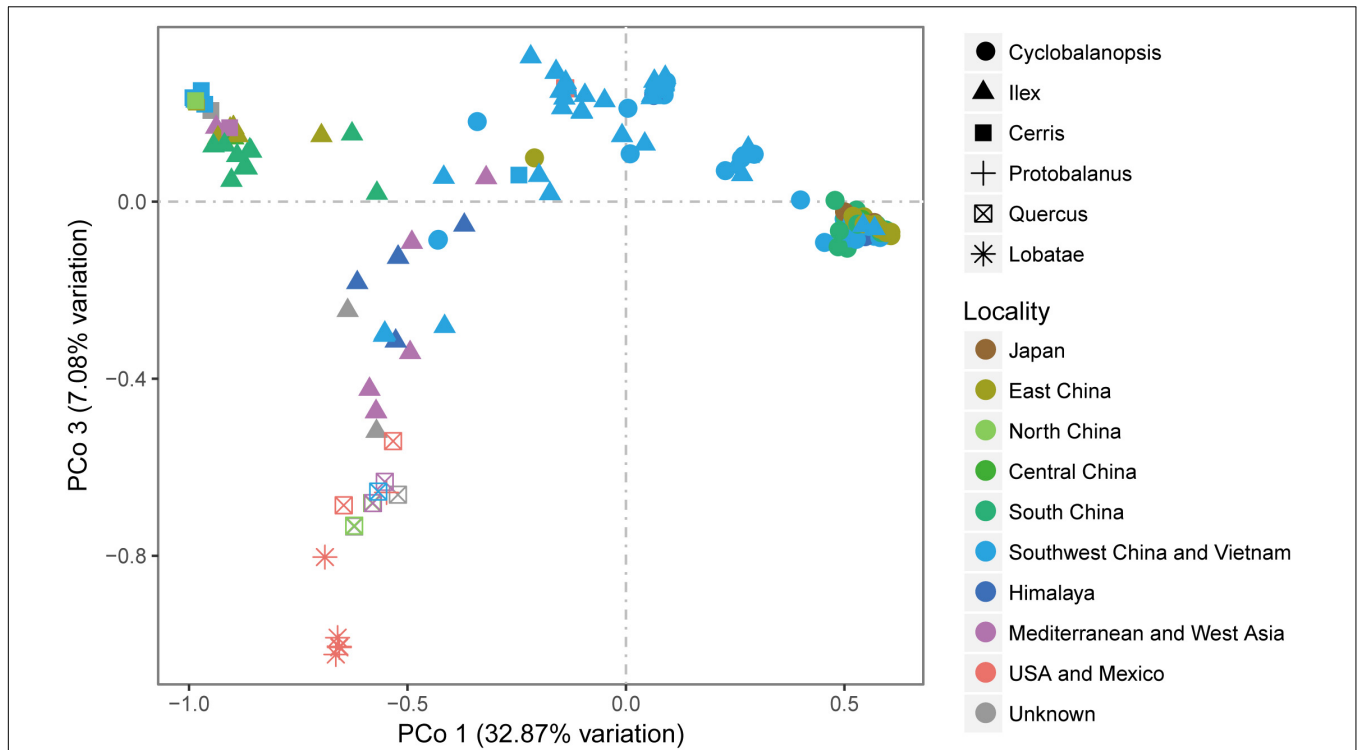


**FIGURE 3** | BI phylogram of *Quercus* inferred using combined *cpDNA* sequences. The tree was rooted using *Lithocarpus henryi* as the outgroup. Numbers close to the nodes are posterior probability values. Branch colors indicate different sections and colors of individuals indicate their localities with detail indication in the center of this figure.

chloroplast genome may not be an efficient marker for fine-scale DNA barcoding nor for inferring systematics of closely related oak species (Pham et al., 2017). Therefore, either subsets of chloroplast barcode sequences, or the complete chloroplast genome are unlikely to provide confident identification of the source species. All these facts restrict the application of *cpDNA* sequences on species identification in closely related oaks.

Aside from *cpDNA* markers, ITS (incl. ITS1, 5.8S, ITS2) has been used as a universal DNA barcode marker for species-level phylogenetic studies (Kress et al., 2005; Hughes et al., 2006). The *nrDNA* ITS and 5S-IGS have been applied to infer oak phylogeny, which roughly illustrate the phylogeny skeleton of the main lineages in *Quercus*, however, the deep

node phylogeny among sections are not resolved (Manos et al., 1999; Denk and Grimm, 2010). Indeed, orthologous sequences of *nrDNA* has better resolution than *cpDNA* at low taxonomical level to infer the phylogeny of oaks due to its biparental inheritance (Simeone et al., 2013). Nonetheless, *nrDNA* in oaks has paralogous copies and pseudogenes (Mayol and Rosselló, 2001; Bellarosa et al., 2005; Ma, 2006). Incorporating ITS paralogs in plant evolutionary studies is very risky because it can distort the phylogenetic signal (Mayol and Rosselló, 2001; Ma and Zhou, 2005; Feliner and Rosselló, 2007). To avoid incorporating ITS paralogs, PCR products need to be cloned into a vector for sequencing, which is time consuming and costly. Therefore, routine applicability of the ITS for barcoding is difficult.



**FIGURE 4 |** PCoA analysis of *Quercus* species based on the genetic distance obtained from combined *cpDNA* sequences.

Even single copy nuclear genes or a small batch of nuclear sequences may be not ideal barcoding makers for precise species identification in oaks, due to limited numbers of informative SNPs (Oh and Manos, 2008; Hubert et al., 2014). Recently, high-throughput markers, such as RAD-seq (Hipp et al., 2014; Cavender-Bares et al., 2015; Deng et al., 2017a,c) have demonstrated their power to successfully resolve the oak phylogeny with rich species and complex evolutionary history. Genome skimming also has great advantages for extending the plant barcode (Li et al., 2015; Coissac et al., 2016; Hollingsworth et al., 2016), allowing for the near-complete assembly of high-copy plastids, mitochondria and ribosomal DNA and possibly a fragmented nuclear genome assembly (Hollingsworth et al., 2016). These

advanced technologies offer a promising future for DNA barcoding in oaks and probably other taxonomical difficult groups.

### Performance of Different Barcoding Analysis Methods

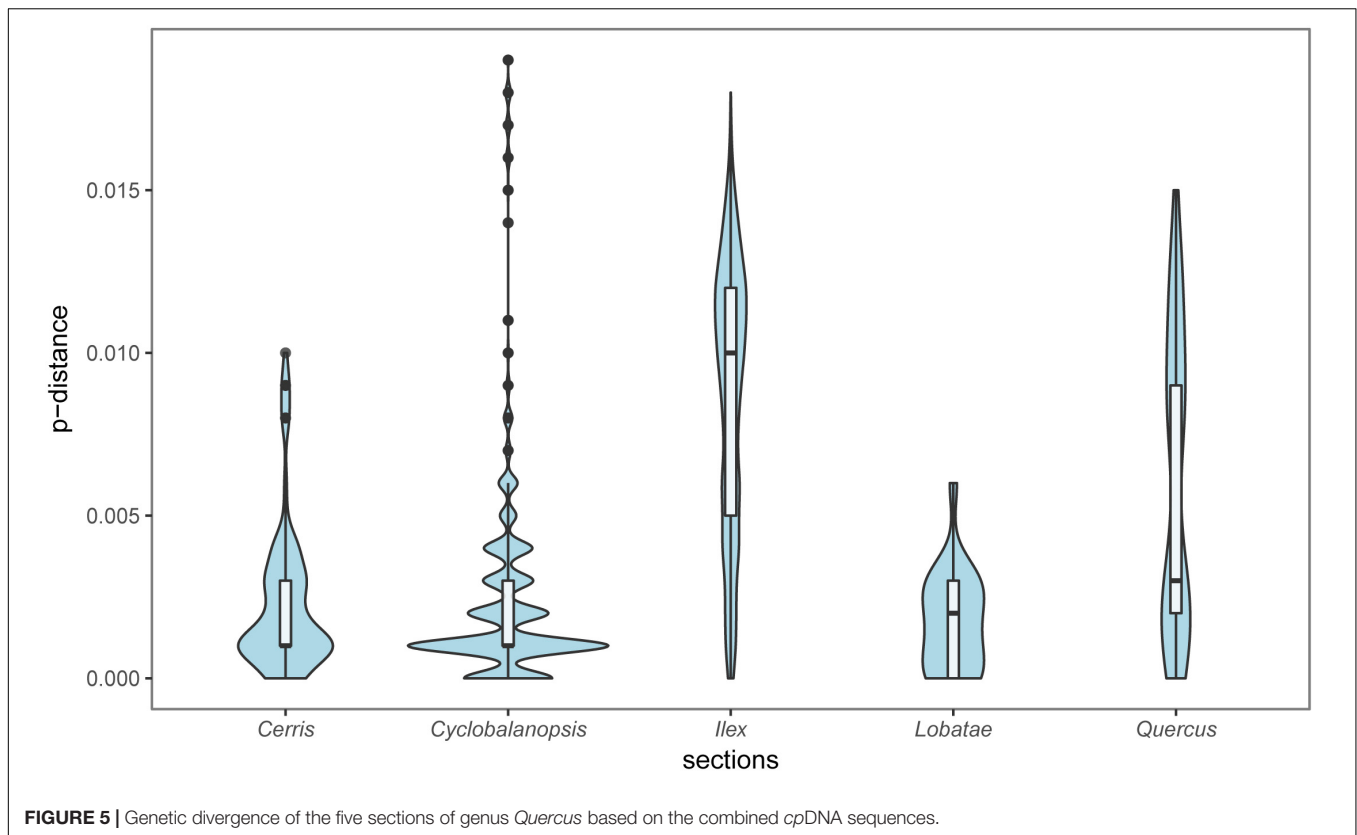
The different performances of the four methods may reflect the analytical theories implemented in each method. BLAST had the best performance, providing the highest species resolution for all single markers and combinations. BLAST calculates the similarity of sequences, and assume that individuals from a species will be more similar to those from other species (van Velzen et al., 2012). This method also yields higher identification rates in other barcoding studies in *Quercus*,

**TABLE 3 |** Genetic diversity obtained from the combined *cpDNA* sequences.

Dataset (section)	N	Ns	p-distance	Vs		Pi	Hn	Hd	Indel
				Number	%				
<i>Cyclobalanopsis</i>	147	29	0–0.019 (0.0020 ± 0.0031)	134	4.19	0.00205	54	0.837	55
<i>Ilex</i>	125	28	0–0.018 (0.0090 ± 0.0041)	220	6.88	0.00867	77	0.989	70
<i>Cerris</i>	18	4	0–0.011 (0.0025 ± 0.0026)	51	1.59	0.00257	9	0.895	32
<i>Quercus</i>	11	7	0–0.013 (0.0059 ± 0.0045)	67	2.10	0.00588	9	0.964	25
<i>Lobatae</i>	7	5	0–0.006 (0.0017 ± 0.0015)	17	0.53	0.00867	4	0.714	4
<i>Protobalanus</i>	1	1	–	–	–	–	–	–	–

N, number of samples; Ns, number of species; p-distance [min. – max. (mean ± SD)]; Vs, variable sites; Pi, nucleotide diversity; Hn, number of haplotypes; Hd, haplotype diversity.





*Curcuma*, *Rhododendron*, and bryophytes (Hassel et al., 2013; Chen et al., 2015; Yan et al., 2015; Yang et al., 2017). The distance-based method (ABGD) automatically finds the distance where the barcode gap is located to partition the data set into candidate species, and can be used even when the intra- and inter-species distances overlap (Puillandre et al., 2012). This explained though intra- and inter-species distances overlap in our dataset, which adds difficulty to discriminate species. Nevertheless, ABGD still provided a relatively high species discrimination rate. Conversely, the tree-based methods and diagnostic methods generate low species identification rate. The tree-based approach is depended on monophyletic status of species. However, we only found few species were resolved as monophyletic clades and successfully identified. The poor resolution of tree-based method is in concordance with other studies (Meier et al., 2006; Little and Stevenson, 2007; Virgilio et al., 2010; Little, 2011). It is worth noting that BLOG has been reported to generate the highest sequence identification success in oaks and other plants (van Velzen et al., 2012; Weitschek et al., 2013; Yang et al., 2017). Actually, in our study BLOG also provides the highest individual identification success (36.23%), but it demonstrated the lowest species identification success, which requires correct identification of all individuals of a species in both training and test sequences. BLOG identifies potential diagnostic nucleotide positions from training sequences and assigns species using logic formulas based on the species-specific (diagnostic) codes (Weitschek et al., 2013). However, the diagnostic codes based on training sequences may not always

cover the total variance of all sequences. Meanwhile, the total sequences are divided into training and test datasets in BLOG, as a result there are fewer samples (only one or two) in each dataset. Fewer samples may increase the discrimination rate and lead to a superficially high success rate. Hence, the high success rate produced by BLOG should be checked carefully, whether a species is correctly identified in both training and test dataset.

### The Factors That Might Contributed to the Low Resolution of *cpDNA* in East Asian Evergreen Oaks Conservation of the Chloroplast Genome

The poor species resolution of the four universal markers might be caused by multiple factors. Low mutation rates of chloroplast genome might lead to the poor resolution among different species. Actually, *cpDNA* has a low mutation rate, and much lower comparing to *nrDNA* (Lynch, 1997; Drouin et al., 2008). In theory, taxa with shorter generation time is likely to accumulate more mutations than long-lived species (Martin and Palumbi, 1993; Gaut et al., 1996). For woody plants with long reproductive cycle, their *cpDNA* mutation rates are much lower than the herbaceous species (Gaut et al., 1996). Generally, oaks have long generation time and oak seedlings need 3–45 years to produce their first acorns (Ducousso et al., 1993; Guyette et al., 2004; Kormanik et al., 2004). Consistent with the theory, the *cpDNA* substitution rates of oaks ( $0.19\text{--}0.96 \times 10^{-9}$  s/s/y) (Chen et al.,

2012; Xu et al., 2015, 2016; Feng et al., unpublished) are very low, below the average values reported for non-coding regions in other angiosperm lineages ( $1.2\text{--}1.7 \times 10^{-9}$  s/s/y) (Graur and Li, 2000). Similarly, the low mutation rate of chloroplast genome is also reported in other woody species (Qi et al., 2012; Guo et al., 2014). The low substitution rate of *cpDNA* of oaks resulted in fewer mutations between different individuals, which partly explains the low species resolution of *cpDNA* in oaks. However, the low resolution may also be due to selective sweeps in chloroplast genome and past genetic drift (Ellstrand and Elam, 1993; Muir and Filatov, 2007; Percy et al., 2014). Because of the lower effective population size compared to nuclear genome, chloroplast genome may be greatly affected by genetic drift, which reduces the genetic diversity (Birky, 1988; Ellstrand and Elam, 1993; Schaal et al., 1998). As the result, chloroplast genome demonstrates low genetic diversity.

### Complicated *cpDNA* Evolutionary History Involved Introgression and Incomplete Lineage Sorting

Chloroplast is maternally inherited in oaks (Dumolin et al., 1995). As a result, *cpDNA* is easy to transfer during the hybridization process, especially with asymmetric gene flow in the parental populations. Therefore, the *cpDNA* phylogeny only reflects the maternal evolutionary history. Oaks are notorious for interspecific hybridization, especially among the species in the same section (Curtu et al., 2007; Neophytou et al., 2011b; Moran et al., 2012; Leroy et al., 2017; McVay et al., 2017). Natural hybridization in East Asian evergreen oaks has been revealed and well demonstrated among sympatric species. In section *Cyclobalanopsis*, introgression between sympatric species (*Q. sessilifolia* and *Q. acuta* distributed in Korea and Japan; *Q. austrochinchinensis* and *Q. kerrii* distributed in Southwest China) were fully revealed using morphological traits and molecular markers (Tamaki and Okada, 2014; Song et al., 2015; An et al., 2017). The morphological intermediates have been frequently reported in section *Ilex* (Huang et al., 1999; Neophytou et al., 2007) and several hybridization zones with a series of morphological transitions have been discovered according to our field observation, indicating frequent interspecies hybridization. Meanwhile, the inconsistent evolutionary patterns between *cpDNA* and *nrDNA* were commonly detected in regional phylogeographic and population genetic analysis of section *Ilex*, which also implied introgression occurs in the Mediterranean and East Asia (Neophytou et al., 2011a,b; Meng et al., 2017; Vitelli et al., 2017). We found that the evolutionary history of the chloroplast genome is net-like rather than dichotomous, which hinders the discriminatory ability to infer the species tree using these maternal markers, especially in a frequent-hybridized taxa (Hollingsworth et al., 2011; Coissac et al., 2016). Moreover, driven by the peculiar paleogeographical histories of the studied regions, plastid genome may reveal geographic differentiation or distribution range of the ancestral lineage instead of the true species phylogeny (Petit et al., 2002; Gugger and Cavender-Bares, 2013; Simeone et al., 2016; Pham et al., 2017; Vitelli et al., 2017). Eurasian evergreen oaks also have strong *cpDNA* genetic differentiation across different geographical regions instead of species taxonomy. As we found in section *Ilex*, individuals from

Mediterranean-Himalaya, SW China and East China formed independent clusters, respectively. It is likely that active tectonic movements induced uplift of SE Himalaya fringes and adjacent region, which blocked the regional seed-mediated gene flow and resulted in the strong *cpDNA* geographic differentiation in these oaks.

Another explanation for haplotype sharing between species which located across long distance is the incomplete lineage sorting of the ancestral lineages (Piredda et al., 2011; Simeone et al., 2013). In addition, occasional introgression following the long-distance dispersal events might result in this pattern as well (Lumaret et al., 2002; Toumi and Lumaret, 2010). Therefore, the low *cpDNA* mutation rates, strong asymmetric interspecies gene flow and incomplete lineage sorting of ancestral polymorphism may have contributed to the low resolution of *cpDNA* for species delimitation for East Asian evergreen oaks.

### Different Genetic Diversity Patterns in Asian Evergreen Lineages

#### Why Is the *cpDNA* Phylogeny Non-monophyletic?

Within the subgenus *Cerris*, our data shows that section *Ilex* is closely related to both sections *Cyclobalanopsis* and *Cerris*. A similar unresolved complex phylogenetic relationship within subgenus *Cerris* was also found in other studies using *cpDNA* markers (Manos et al., 1999; Simeone et al., 2016). The chloroplast capture events within a genus are mostly due to hybridization (Fehrer et al., 2007; Acosta and Premoli, 2010). However, the hybridization between the extant sections in oaks is extremely rare in the wild (Costello et al., 2011). The non-monophyly of section *Ilex* plastomes and its close relationship with sections *Cyclobalanopsis* and *Cerris* may reflect an ancient introgression or incomplete lineage sorting of the chloroplast genome in the ancestor lineages of subgenus *Cerris*. In the future, this issue should be subjected to the further study using high-throughput molecular marker (e.g., GBS, RAD-seq) to resolve the species phylogeny. In addition, it needs to recruit multiple species populations of the three sections to reveal the ancient and/or contemporary gene flow among the lineages.

#### Why the Genetic Diversity in Sections of *Quercus* Is Different?

Theoretically, the deciduous lineages which are usually fast growing species, may have higher genetic variation, because they have shorter sexual maturation time comparing to evergreen lineages (Zuidema et al., 2009). Generally, it takes at least five or more years for seedlings of evergreen oaks to produce first acorns. But for the deciduous species, this cycle only needs 2–3 years (Wang and Gao, 2005; Wang et al., 2011), such as in *Q. serrata*, *Q. fabri*, and *Q. aliena*. However, the genetic variation patterns in oak sections may not match this theory well, as our study revealed that sections *Ilex* and *Quercus* have higher genetic variation, and other sections have lower variation. This result is slightly different to previous reports that sections *Ilex*, *Lobatae* and *Quercus* were the most variable lineages, and section *Cerris* was the least variable (Simeone et al., 2016). Currently, it is too early to conclude the overall genetic diversity patterns of the

genus *Quercus*, because of the uneven sampling of the main sections.

However, in the well-sampled Asian evergreen sections *Cyclobalanopsis* and *Ilex*, we found different patterns of genetic diversity. Trees of section *Ilex* grow at semi-arid habitats with slow biomass accumulation ratio and long sexual maturation cycle (Mediavilla and Escudero, 2003). All these biological traits of species of section *Ilex* tend to cause low *cpDNA* genetic diversity level. However, it was the most diverse lineage based on our study. Interestingly, section *Cyclobalanopsis* with high number of species and great morphological variation showed low genetic diversity. The different biogeographical histories and adaptabilities to the environment in the two sections may have driven their genetic diversity patterns. Section *Cyclobalanopsis* was an early derived (at the early Oligocene) section of subgenus *Cerris*, but their lineage diversification did not occur until the early Miocene and the highest diversification ratio achieved at the mid-late Miocene (Deng et al., 2017a). Instead, section *Ilex* was later derived at the early-middle Oligocene, but it shows a stepwise diversification pattern since the middle Oligocene onward (Deng et al., 2017c). Therefore, most species in section *Cyclobalanopsis* are probably younger than those in section *Ilex*. With longer evolutionary time, more *cpDNA* mutations may have accumulated, which may explain why the genetic diversity level of section *Ilex* is much higher than that of section *Cyclobalanopsis*. Moreover, section *Ilex* can grow in a wide geographic range with diversified habitats, aside from the prominent geographic barriers, the local adaptation to different environments may have also played a role in boosting the genetic diversity of section *Ilex* (Du et al., 2016; Feng et al., 2016; Meng et al., 2017; Feng et al., unpublished). In contrast, trees in section *Cyclobalanopsis* live mainly in warm and humid subtropical habitats (Huang et al., 1999; Deng et al., 2017a). As dominant trees, their large effective population size and the similar habitats may make them less influenced by topographic and climate changes. Shorter divergence time, long-term environment stability and strong gene flow among regional populations, may have altogether resulted in the low genetic diversity of the *cpDNA* in section *Cyclobalanopsis*.

## CONCLUSION AND PERSPECTIVE

This study reveals that the *cpDNA* markers have limited efficiency to identify the Asian evergreen oaks, but these *cpDNA* markers are still informative to infer the species placement to the main sections of *Quercus*. The different genetic diversity patterns

## REFERENCES

- Acosta, M. C., and Premoli, A. C. (2010). Evidence of chloroplast capture in South American *Nothofagus* (subgenus *Nothofagus*, Nothofagaceae). *Mol. Phylogenet. Evol.* 54, 235–242. doi: 10.1016/j.ympev.2009.08.008
- An, M., Deng, M., Zheng, S. S., Jiang, X. L., and Song, Y. G. (2017). Introgression threatens the genetic diversity of *Quercus austrocochinchinensis* (Fagaceae), an endangered oak: a case inferred by molecular markers. *Front. Plant Sci.* 8:229. doi: 10.3389/fpls.2017.00229

between the two evergreen oak sections *Ilex* and *Cyclobalanopsis* may have been shaped by their spatio-temporal histories and different adaptabilities to environments. The different evolutionary pattern of oaks revealed by the chloroplast and nuclear genomes is most likely due to the historical introgression in the ancestral lineages and recent or on-going gene flow between the closely related species. All these factors reduce the efficiency of *cpDNA* as species level barcodes. Even single copy nuclear genes or a small batch of nuclear sequences may not be ideal barcoding makers for precise species identification in oaks, due to limited number of informative SNPs. The advanced technologies, such as high-throughput markers (e.g., RAD-seq and GBS) and genome skimming offer a promising future for DNA barcoding in oaks and other taxonomic difficult groups.

## AUTHOR CONTRIBUTIONS

MD conceived and designed the experiments. YX, RL, and MY performed the experiments. MY and JS analyzed the data. MD was responsible for field collections and specimen identification. MY and MD wrote and revised the paper.

## FUNDING

This work was supported by grants from the Shanghai Municipal Administration of Forestation and City Appearances (G162405, G172406, and G162404), Southeast Asia Biodiversity Research Institute, Chinese Academy of Sciences (Y4ZK111B01), National Natural Science Foundation of China (31270267), Science and Technology Basic Work (S&T Basic Work) (2013FY112100), and International Partnership Program of Chinese Academy of Sciences (151111KYSB20170021).

## ACKNOWLEDGMENTS

We thank Xiaolong Jiang, Yigang Song, and Quanjian Li for their assistance with the sampling.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2018.00569/full#supplementary-material>

- Armstrong, K., and Ball, S. (2005). DNA barcodes for biosecurity: invasive species identification. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 360, 1813–1823. doi: 10.1098/rstb.2005.1713
- Ball, S. L., and Armstrong, K. F. (2006). DNA barcodes for insect pest identification: a test case with tussock moths (*Lepidoptera*: Lymantriidae). *Can. J. For. Res.* 36, 337–350. doi: 10.1098/rstb.2005.1713
- Bellarosa, R., Simeone, M. C., Papini, A., and Schirone, B. (2005). Utility of ITS sequence data for phylogenetic reconstruction of Italian *Quercus* spp. *Mol. Phylogenet. Evol.* 34, 355–370. doi: 10.1016/j.ympev.2004.10.014

- Bickford, D., Lohman, D. J., Sodhi, N. S., Ng, P. K., Meier, R., Winker, K., et al. (2007). Cryptic species as a window on diversity and conservation. *Trends Ecol. Evol.* 22, 148–155. doi: 10.1016/j.tree.2006.11.004
- Birky, C. W. (1988). “Evolution and variation in plant chloroplast and mitochondrial genomes,” in *Plant Evolutionary Biology*, eds L. D. Gottlieb and S. K. Jain (Dordrecht: Springer), 23–53.
- Bleeker, W., Klausmeyer, S., Peintinger, M., and Dienst, M. (2008). DNA sequences identify invasive alien *Cardamine* at Lake Constance. *Biol. Conserv.* 141, 692–698. doi: 10.1016/j.biocon.2007.12.015
- Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., et al. (2009). BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. doi: 10.1186/1471-2105-10-421
- Camus, A. (1936–1954). *Les Chênes: Monographie du Genre Quercus*. Paris: P. Lechevalier.
- Cao, M., and Zhang, J. (1997). Tree species diversity of tropical forest vegetation in Xishuangbanna, SW China. *Biodivers. Conserv.* 7, 995–1006. doi: 10.15517/rbt.v59i13212
- Cavender-Bares, J., González-Rodríguez, A., Eaton, D. A., Hipp, A. A., Beulke, A., and Manos, P. S. (2015). Phylogeny and biogeography of the American live oaks (*Quercus* subsection *Virentes*): a genomic and population genetics approach. *Mol. Ecol. Resour.* 24, 3668–3687. doi: 10.1111/mec.13269
- CBOL Plant Working Group (2009). A DNA barcode for land plants. *Proc. Natl. Acad. Sci. U.S.A.* 106, 12794–12797. doi: 10.1073/pnas.0905845106
- Chen, D. M., Zhang, X. X., Kang, H. H., Sun, X., Yin, S., Du, H. E., et al. (2012). Phylogeography of *Quercus variabilis* based on chloroplast DNA sequence in East Asia: multiple glacial refugia and mainland-migrated island populations. *PLoS One* 7:e47268. doi: 10.1371/journal.pone.0047268
- Chen, J., Zhao, J., Erickson, D. L., Xia, N., and Kress, W. J. (2015). Testing DNA barcodes in closely related species of *Curcuma* (Zingiberaceae) from Myanmar and China. *Mol. Ecol. Resour.* 15, 337–348. doi: 10.1111/1755-0998.12319
- Clement, W. L., and Donoghue, M. J. (2012). Barcoding success as a function of phylogenetic relatedness in *Viburnum*, a clade of woody angiosperms. *BMC Evol. Biol.* 12:73. doi: 10.1186/1471-2148-12-73
- Clerc-Blain, J. L., Starr, J. R., Bull, R. D., and Saarela, J. M. (2010). A regional approach to plant DNA barcoding provides high species resolution of sedges (*Carex* and *Kobresia*, Cyperaceae) in the Canadian Arctic Archipelago. *Mol. Ecol. Resour.* 10, 69–91. doi: 10.1111/j.1755-0998.2009.02725.x
- Coissac, E., Hollingsworth, P. M., Lavergne, S., and Taberlet, P. (2016). From barcodes to genomes: extending the concept of DNA barcoding. *Mol. Ecol.* 25, 1423–1428. doi: 10.1111/mec.13549
- Costello, L. R., Hagen, B. W., and Katherine, S. J. (2011). *Oaks in the Urban Landscape, Selection, Care and Preservation*. Richmond, VA: California University of California.
- Curtu, A. L., Gailing, O., and Finkeldey, R. (2007). Evidence for hybridization and introgression within a species-rich oak (*Quercus* spp.) community. *BMC Evol. Biol.* 7:218. doi: 10.1186/1471-2148-7-218
- Deng, M., Hipp, A., Song, Y. G., Li, Q. S., Coombes, A., and Cotton, A. (2014). Leaf epidermal features of *Quercus* subgenus *Cyclobalanopsis* (Fagaceae) and their systematic significance. *Bot. J. Linn. Soc.* 176, 224–259. doi: 10.1111/boj.12207
- Deng, M., Jiang, X. L., Hipp, A. L., Manos, P. S., and Hahn, M. (2017a). Phylogeny and biogeography of East Asian evergreen oaks (*Quercus* section *Cyclobalanopsis*; Fagaceae): insights into the Cenozoic history of evergreen broad-leaved forests in subtropical Asia. *Mol. Phylogenet. Evol.* 119, 170–181. doi: 10.1016/j.ympev.2017.11.003
- Deng, M., Jiang, X. L., Song, Y. G., Coombes, A., Yang, X. R., Xiong, Y. S., et al. (2017b). Leaf epidermal features of *Quercus* Group *Ilex* (Fagaceae) and their application to species identification. *Rev. Palaeobot. Palynol.* 237, 10–36. doi: 10.1016/j.revpalbo.2016.11.006
- Deng, M., Jiang, X. L., Su, T., Hipp, A., and Zhou, Z. K. (2017c). “Young dispersal from East Asia through the Himalya to the Mediterranean of the Tethys disjunction lineage-Biogeography of *Quercus* section *Ilex* (Fagaceae),” in *Proceedings of the IUFRO Genetics and Genomics of Fagaceae*, Shanghai.
- Deng, M., Li, Q. S., Yang, S. T., Xu, J., Song, Y. G., and Li, Q. J. (2013a). “Endangered Oaks in China and the conservation challenges,” in *Proceedings of the 5th Global Botanic Gardens Congress*, Guangzhou.
- Deng, M., Song, Y. G., Li, Q. J., and Li, Q. S. (2013b). Pollen morphology of *Quercus* subg. *Cyclobalanopsis* (Fagaceae) and its systematic implication. *Guihaia* 33, 368–375. doi: 10.5586/asbp.2012.005
- Denk, T., and Grimm, G. W. (2009). Significance of pollen characteristics for infrageneric classification and phylogeny in *Quercus* (Fagaceae). *Int. J. Plant Sci.* 170, 926–940.
- Denk, T., and Grimm, G. W. (2010). The oaks of western Eurasia: traditional classifications and evidence from two nuclear markers. *Taxon* 59, 351–366.
- Denk, T., Grimm, G. W., Manos, P. S., Deng, M., and Hipp, A. L. (2017). “An updated infrageneric classification of the oaks: review of previous taxonomic schemes and synthesis of evolutionary patterns,” in *Oaks Physiological Ecology. Exploring the Functional Diversity of Genus Quercus* L., eds E. Gil-Pelegrin, J. J. Peguero-Pina, and D. Sancho-Knapik (Berlin: Springer), 13–38.
- Denk, T., and Tekleva, M. V. (2014). Pollen morphology and ultrastructure of *Quercus* with focus on Group *Ilex* (= *Quercus* subgenus *Heterobalanus* (Oerst.) Menitsky): implications for oak systematics and evolution. *Grana* 53, 255–282. doi: 10.1080/00173134.2014918647
- Dong, W., Xu, C., Li, C., Sun, J., Zuo, Y., Shi, S., et al. (2015). ycf1, the most promising plastid DNA barcode of land plants. *Sci. Rep.* 5:8348. doi: 10.1038/srep08348
- Doyle, J. J., and Doyle, J. L. (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19, 11–15.
- Drouin, G., Daoud, H., and Xia, J. (2008). Relative rates of synonymous substitutions in the mitochondrial, chloroplast and nuclear genomes of seed plants. *Mol. Phylogenet. Evol.* 49, 827–831. doi: 10.1016/j.ympev.2008.09.009
- Du, F. K., Hou, M., Wang, W., Mao, K., and Hampe, A. (2016). Phylogeography of *Quercus aquifolioides* provides novel insights into the Neogene history of a major global hotspot of plant diversity in south-west China. *J. Biogeogr.* 44, 294–307. doi: 10.1111/jbi.12836
- Ducousso, A., Michaud, H., and Lumaret, R. (1993). Reproduction and gene flow in the genus *Quercus* L. *Ann. Sci. For.* 50(Suppl.), 91s–106s. doi: 10.1051/forest:19930708
- Dumolin, S., Demesure, B., and Petit, R. (1995). Inheritance of chloroplast and mitochondrial genomes in pedunculate oak investigated with an efficient PCR method. *Theor. Appl. Genet.* 91, 1253–1256. doi: 10.1007/BF00220937
- Edgar, R. C. (2004). MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 32, 1792–1797. doi: 10.1093/nar/gkh340
- Ellstrand, N. C., and Elam, D. R. (1993). Population genetic consequences of small population size: implications for plant conservation. *Annu. Rev. Ecol. Syst.* 24, 217–242. doi: 10.1146/annurev.es.24.110193.001245
- Fang, J. Y., and Yoda, K. (1991). Climate and vegetation in China V. Effect of climatic factors on the upper limit of distribution of evergreen broadleaf forest. *Ecol. Res.* 6, 113–125. doi: 10.1007/bf02353874
- Fehrer, J., Gemeinholzer, B., Chrtek, J., and Bräutigam, S. (2007). Incongruent plastid and nuclear DNA phylogenies reveal ancient intergeneric hybridization in *Pilosella* hawkweeds (*Hieracium*, Cichorieae, Asteraceae). *Mol. Phylogenet. Evol.* 42, 347–361. doi: 10.1016/j.ympev.2006.07.004
- Feliner, G. N., and Rosselló, J. A. (2007). Better the devil you know? Guidelines for insightful utilization of nrDNA ITS in species-level evolutionary studies in plants. *Mol. Phylogenet. Evol.* 44, 911–919. doi: 10.1016/j.ympev.2007.01.013
- Feng, L., Zheng, Q. J., Qian, Z. Q., Yang, J., Zhang, Y. P., Li, Z. H., et al. (2016). Genetic structure and evolutionary history of three Alpine Sclerophyllous oaks in East Himalaya-Hengduan Mountains and adjacent regions. *Front. Plant Sci.* 7:1688. doi: 10.3389/fpls.2016.01688
- Gao, T., Yao, H., Song, J. Y., Zhu, Y. J., Liu, C., and Chen, S. L. (2010). Evaluating the feasibility of using candidate DNA barcodes in discriminating species of the large Asteraceae family. *BMC Evol. Biol.* 10:324. doi: 10.1186/1471-2148-10-324
- Gaut, B. S., Morton, B. R., McCaig, B. C., and Clegg, M. T. (1996). Substitution rate comparisons between grasses and palms: synonymous rate differences at the nuclear gene *Adh* parallel rate differences at the plastid gene *rbcL*. *Proc. Natl. Acad. Sci. U.S.A.* 93, 10274–10279. doi: 10.1073/pnas.93.19.10274
- Goldstein, P. Z., and DeSalle, R. (2011). Integrating DNA barcode data and taxonomic practice: determination, discovery, and description. *Bioessays* 33, 135–147. doi: 10.1002/bies.201000036
- Gonzalez, M. A., Baraloto, C., Engel, J., Mori, S. A., Pétronelli, P., Riéra, B., et al. (2009). Identification of Amazonian trees with DNA barcodes. *PLoS One* 4:e7483. doi: 10.1371/journal.pone.0007483
- Graur, D., and Li, W. H. (2000). *Fundamentals of Molecular Evolution*, 2nd Edn. Sunderland, MA: Sinauer Associates Inc.



- Gugger, P. F., and Cavender-Bares, J. (2013). Molecular and morphological support for a Florida origin of the Cuban oak. *J. Biogeogr.* 40, 632–645. doi: 10.1111/j.1365-2699.2011.02610.x
- Guo, X. D., Wang, H. F., Bao, L., Wang, T. M., Bai, W. N., Ye, J. W., et al. (2014). Evolutionary history of a widespread tree species *Acer mono* in East Asia. *Ecol. Evol.* 4, 4332–4345. doi: 10.1002/ece3.1278
- Guyette, R. P., Muzika, R.-M., Kabrick, J., and Stambaugh, M. C. (2004). *A Perspective on Quercus Life History Characteristics and Forest Disturbance*. Gen. Tech. Rep. SRS-73. Asheville, NC: Department of Agriculture, 138–142.
- Hassel, K., Segreto, R., and Ekrem, T. (2013). Restricted variation in plant barcoding markers limits identification in closely related bryophyte species. *Mol. Ecol. Resour.* 13, 1047–1057. doi: 10.1111/1755-0998.12074
- Hebert, P. D., Cywinska, A., and Ball, S. L. (2003). Biological identifications through DNA barcodes. *Proc. R. Soc. B Biol. Sci.* 270, 313–321. doi: 10.1098/rspb.2002.2218
- Hipp, A. L., Eaton, D. A., Cavender-Bares, J., Fitzek, E., Nipper, R., and Manos, P. S. (2014). A framework phylogeny of the American oak clade based on sequenced RAD data. *PLoS One* 9:e93975. doi: 10.1371/journal.pone.0093975
- Hollingsworth, P. M. (2007). DNA barcoding: potential users. *Genomics Soc. Policy* 3, 44–47. doi: 10.1186/1746-5354-3-2-44
- Hollingsworth, P. M., Graham, S. W., and Little, D. P. (2011). Choosing and using a plant DNA barcode. *PLoS One* 6:e19254. doi: 10.1371/journal.pone.0019254
- Hollingsworth, P. M., Li, D. Z., van der Bank, M., and Twyford, A. D. (2016). Telling plant species apart with DNA: from barcodes to genomes. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 371:20150338. doi: 10.1098/rstb.2015.0338
- Huang, C., Zhang, Y., and Bartholomew, B. (1999). "Fagaceae," in *Flora of China 4 Cycadaceae through Fagaceae*, eds Z. Wu and P. Raven (Beijing: Science Press), 314–400.
- Hubert, F., Grimm, G. W., Jousselin, E., Berry, V., Franc, A., and Kremer, A. (2014). Multiple nuclear genes stabilize the phylogenetic backbone of the genus *Quercus*. *Syst. Biodivers.* 12, 405–423. doi: 10.1080/14772000.2014.941037
- Hughes, C. E., Eastwood, R. J., and Bailey, C. D. (2006). From famine to feast? Selecting nuclear DNA sequence loci for plant species-level phylogeny reconstruction. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 361, 211–225. doi: 10.1098/rstb.2005.1735
- Jiang, L., Xu, K., Fan, Q., and Peng, H. (2017). A new species of *Ilex* (Aquifoliaceae) from Jiangxi Province, China, based on morphological and molecular data. *Phytotaxa* 298, 147–157. doi: 10.11646/phytotaxa.298.2.4
- Jiang, X. L., An, M., Zheng, S. S., Deng, M., and Su, Z. H. (2018). Geographical isolation and environmental heterogeneity contribute to the spatial genetic patterns of *Quercus kerrii* (Fagaceae). *Heredity* 120, 219–233. doi: 10.1038/s41437-017-0012-7
- Jiang, X. L., Deng, M., and Li, Y. (2016). Evolutionary history of subtropical evergreen broad-leaved forest in Yunnan Plateau and adjacent areas: an insight from *Quercus schottkyana* (Fagaceae). *Tree Genet. Genomes* 12:104. doi: 10.1007/s11295-016-1063-2
- Kormanik, P. P., Sung, S.-J. S., Kormanik, T., Tibbs, T., and Zarnoch, S. J. (2004). *Northern Red Oak from Acorns to Acorns in 8 Years or Less*. Gen. Tech. Rep. SRS 71. Asheville, NC: Department of Agriculture Forest Service, 555–558.
- Kress, W. J., Erickson, D. L., Jones, F. A., Swenson, N. G., Perez, R., Sanjur, O., et al. (2009). Plant DNA barcodes and a community phylogeny of a tropical forest dynamics plot in Panama. *Proc. Natl. Acad. Sci. U.S.A.* 106, 18621–18626. doi: 10.1073/pnas.0909820106
- Kress, W. J., Erickson, D. L., Swenson, N. G., Thompson, J., Uriarte, M., and Zimmerman, J. K. (2010). Advances in the use of DNA barcodes to build a community phylogeny for tropical trees in a Puerto Rican forest dynamics plot. *PLoS One* 5:e15409. doi: 10.1371/journal.pone.0015409
- Kress, W. J., Wurdack, K. J., Zimmer, E. A., Weigt, L. A., and Janzen, D. H. (2005). Use of DNA barcodes to identify flowering plants. *Proc. Natl. Acad. Sci. U.S.A.* 102, 8369–8374. doi: 10.1073/pnas.0503123102
- Kumar, S., Stecher, G., and Tamura, K. (2016). MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol. Biol. Evol.* 33, 1870–1874. doi: 10.1093/molbev/msw054
- Lahaye, R., van der Bank, M., Bogarin, D., Warner, J., Pupulin, F., Gigot, G., et al. (2008). DNA barcoding the floras of biodiversity hotspots. *Proc. Natl. Acad. Sci. U.S.A.* 105, 2923–2928. doi: 10.1073/pnas.0709936105
- Laiou, A., Mandolini, L. A., Piredda, R., Bellarosa, R., and Simeone, M. C. (2013). DNA barcoding as a complementary tool for conservation and valorisation of forest resources. *Zookeys* 365, 197–213. doi: 10.3897/zookeys.365.5670
- Lee, W., Koh, S. H., Choi, W. I., Jung, C. S., Kim, I. K., Byun, B. K., et al. (2012). Barcoding forest insect pests in South Korea: constructing a basic endemic species dataset. *J. Asia Pac. Entomol.* 15, 363–368. doi: 10.1016/j.aspen.2012.01.008
- Leroy, T., Roux, C., Villate, L., Bodénès, C., Romiguier, J., Paiva, J. A., et al. (2017). Extensive recent secondary contacts between four European white oak species. *New Phytol.* 214, 865–878. doi: 10.1111/nph.14413
- Li, X. W., Yang, Y., Henry, R. J., Rossetto, M., Wang, Y. T., and Chen, S. L. (2015). Plant DNA barcoding: from gene to genome. *Bacteriol. Rev.* 90, 157–166. doi: 10.1111/brv.12104
- Liang, Y. Y., Liu, J., Huang, Y. S., and Lin, C. R. (2016). *Aspidistra erythrocephala* sp. nov. (Asparagaceae) from Guangxi, China. *Phytotaxa* 247, 295–298. doi: 10.1146/phytotaxa.247.4.9
- Librado, P., and Rozas, J. (2009). DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25, 1451–1452. doi: 10.1093/bioinformatics/btp187
- Little, D. P. (2011). DNA barcode sequence identification incorporating taxonomic hierarchy and within taxon variability. *PLoS One* 6:e20552. doi: 10.1371/journal.pone.002052
- Little, D. P., and Stevenson, D. W. (2007). A comparison of algorithms for the identification of specimens using DNA barcodes: examples from gymnosperms. *Cladistics* 23, 1–21. doi: 10.1111/j.1096-0031.2006.00126.x
- Liu, J., Moller, M., Gao, L. M., Zhang, D. Q., and Li, D. Z. (2011). DNA barcoding for the discrimination of Eurasian yews (*Taxus* L., Taxaceae) and the discovery of cryptic species. *Mol. Ecol. Resour.* 11, 89–100. doi: 10.1111/j.1755-0998.2010.02907.x
- Liu, J., Yan, H. F., Newmaster, S. G., Pei, N., Ragupathy, S., and Ge, X. J. (2015). The use of DNA barcoding as a tool for the conservation biogeography of subtropical forests in China. *Divers. Distrib.* 21, 188–199. doi: 10.1111/ddi.12276
- López-Pujol, J., Zhang, F. M., and Ge, S. (2006). Plant biodiversity in China: richly varied, endangered, and in need of conservation. *Biodivers. Conserv.* 15, 3983–4026. doi: 10.1007/s10531-005-3015-2
- Lumaret, R., Mir, C., Michaud, H., and Raynal, V. (2002). Phylogeographical variation of chloroplast DNA in holm oak (*Quercus ilex* L.). *Mol. Ecol.* 11, 2327–2336. doi: 10.1046/j.1365-294X.2002.01611.x
- Luo, Y., and Zhou, Z. (2000). Phytogeography of *Quercus* subg. *Cyclobalanopsis*. *Acta Bot. Yunnanica* 23, 1–16. doi: 10.3969/j.issn.2095-0845.2001.01.001
- Luo, Y., and Zhou, Z. K. (2002). Leaf architecture in *Quercus* subgenus *Cyclobalanopsis* (Fagaceae) from China. *Bot. J. Linn. Soc.* 140, 283–295. doi: 10.1046/j.1095-8339.2002.00097.x
- Lynch, M. (1997). Mutation accumulation in nuclear, organelle, and prokaryotic transfer RNA genes. *Mol. Biol. Evol.* 14, 914–925. doi: 10.1093/oxfordjournals.molbev.a025834
- Ma, C. L. (2006). *Phylogeny and Biogeography of Quercus Sect. Heterobalanus*. Ph.D. dissertation, Kunming Institute of Botany, Kunming.
- Ma, C. L., and Zhou, Z. K. (2005). Effect of ITS pseudogene on the phylogenetic study of *Quercus* (Fagaceae) and its revelation on the plant molecular phylogenetics. *Acta Bot. Yunnanica* 28, 127–132. doi: 10.3969/j.issn.2095-0845.2006.02.007
- Manos, P. S., Doyle, J. J., and Nixon, K. C. (1999). Phylogeny, biogeography, and processes of molecular differentiation in *Quercus* subgenus *Quercus* (Fagaceae). *Mol. Phylogenet. Evol.* 12, 333–349. doi: 10.1006/mpev.1999.0614
- Martin, A. P., and Palumbi, S. R. (1993). Body size, metabolic rate, generation time, and the molecular clock. *Proc. Natl. Acad. Sci. U.S.A.* 90, 4087–4091. doi: 10.1073/pnas.90.9.4087
- Mayol, M., and Rossello, J. A. (2001). Why nuclear ribosomal DNA spacers (ITS) tell different stories in *Quercus*. *Mol. Phylogenet. Evol.* 19, 167–176. doi: 10.1006/mpev.2001.0934
- McVay, J. D., Hipp, A. L., and Manos, P. S. (2017). A genetic legacy of introgression confounds phylogeny and biogeography in oaks. *Proc. R. Soc. B Biol. Sci.* 284:20170300. doi: 10.1098/rspb.2017.0300
- Mediavilla, S., and Escudero, A. (2003). Relative growth rate of leaf biomass and leaf nitrogen content in several Mediterranean woody species. *Plant Ecol.* 168, 321–332. doi: 10.1093/aob/mcl284

- Meier, R., Shiyang, K., Vaidya, G., and Ng, P. K. (2006). DNA barcoding and taxonomy in Diptera: a tale of high intraspecific variability and low identification success. *Syst. Biol.* 55, 715–728. doi: 10.1080/10635150600969864
- Meier, R., Zhang, G., and Ali, F. (2008). The use of mean instead of smallest interspecific distances exaggerates the size of the “barcoding gap” and leads to misidentification. *Syst. Biol.* 57, 809–813. doi: 10.1080/10635150802406343
- Meng, H. H., Su, T., Gao, X. Y., Li, J., Jiang, X. L., Sun, H., et al. (2017). Warm-cold colonization: response of oaks to uplift of the Himalaya-Hengduan Mountains. *Mol. Ecol.* 26, 3276–3294. doi: 10.1111/mec.14092
- Menitsky, L. L. (1984). *Oaks of Asia*. St. Petersburg: Leningosed Sciences.
- Meyer, C. P., and Paulay, G. (2005). DNA barcoding: error rates based on comprehensive sampling. *PLoS Biol.* 3:e422. doi: 10.1371/journal.pbio.0030422
- Miwa, H., Odrzykoski, I. J., Matsui, A., Hasegawa, M., Akiyama, H., Jia, Y., et al. (2009). Adaptive evolution of rbcL in *Conocephalum* (Hepaticae, bryophytes). *Gene* 441, 169–175. doi: 10.1016/j.gene.2008.11.020
- Montecchio, L., and Faccoli, M. (2014). First record of thousand cankers disease *Geosmithia morbida* and walnut twig beetle *Pityophthorus juglandis* on *Juglans nigra* in Europe. *Plant Dis.* 98, 696–696. doi: 10.1094/PDIS-10-13-1027-PDN
- Moran, E. V., Willis, J., and Clark, J. S. (2012). Genetic evidence for hybridization in red oaks (*Quercus* sect. *Lobatae*, Fagaceae). *Am. J. Bot.* 99, 92–100. doi: 10.3732/ajb.1100023
- Muir, G., and Filatov, D. (2007). A selective sweep in the chloroplast DNA of dioecious *Silene* (Section *Elisanthe*). *Genetics* 177, 1239–1247. doi: 10.1534/genetics.107.071969
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., Da Fonseca, G. A., and Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature* 403, 853–858. doi: 10.1038/35002501
- Nei, M., and Li, W. H. (1979). Mathematical model for studying genetic variation in terms of restriction endonucleases. *Proc. Natl. Acad. Sci. U.S.A.* 76, 5269–5273. doi: 10.1073/pnas.76.10.5269
- Neophytou, C., Aravanopoulos, F. A., Fink, S., and Dounavi, A. (2011a). Interfertile oaks in an island environment. II. Limited hybridization between *Quercus alnifolia* Poehc and *Q. coccifera* L. in a mixed stand. *Eur. J. For. Res.* 130, 623–635. doi: 10.1007/s10342-010-0454-4
- Neophytou, C., Dounavi, A., Fink, S., and Aravanopoulos, F. A. (2011b). Interfertile oaks in an island environment: I. High nuclear genetic differentiation and high degree of chloroplast DNA sharing between *Q. alnifolia* and *Q. coccifera* in Cyprus. A multipopulation study. *Eur. J. For. Res.* 130, 543–555. doi: 10.1007/s10342-010-0454-4
- Neophytou, C. H., Palli, G., Dounavi, A., and Aravanopoulos, F. A. (2007). Morphological differentiation and hybridization between *Quercus alnifolia* Poehc and *Quercus coccifera* L. (Fagaceae) in Cyprus. *Silvae Genet.* 56, 271–277. doi: 10.1515/sg-2007-0038
- Newmaster, S., Fazekas, A., Steeves, R., and Janovec, J. (2008). Testing candidate plant barcode regions in the Myricaceae. *Mol. Ecol. Resour.* 8, 480–490. doi: 10.1111/j.1471-8286.2007.02002.x
- Newmaster, S. G., and Ragupathy, S. (2009). Testing plant barcoding in a sister species complex of pantropical *Acacia* (Mimosoideae, Fabaceae). *Mol. Ecol. Resour.* 9(Suppl. 1), 172–180. doi: 10.1111/j.1755-0998.2009.02642.x
- Nixon, K. (2006). “Globe and neotropical distribution and diversity of oak (genus *Quercus*) and oak forests,” in *Ecology and Conservation of Neotropical Montane Oak Forests*, ed. M. Kappelle (Berlin: Springer), 3–13.
- Nixon, K. C. (1993). Infrageneric classification of *Quercus* (Fagaceae) and typification of sectional names. *Ann. Sci. For.* 50(Suppl. 1), 25s–34s. doi: 10.1051/forest:19930701
- Oh, S.-H., and Manos, P. S. (2008). Molecular phylogenetics and cupule evolution in Fagaceae as inferred from nuclear CRABS CLAW sequences. *Taxon* 57, 434–451.
- Ohya, M., Baba, K. I., and Itoh, T. (2001). Wood identification of Japanese *Cyclobalanopsis* species (Fagaceae) based on DNA polymorphism of the intergenic spacer between trnT and trnL 5' exon. *J. Wood Sci.* 47, 81–86. doi: 10.1007/bf00780554
- Peakall, R., and Smouse, P. E. (2012). GenALEX 6.5: genetic analysis in excel. Population genetic software for teaching and research—an update. *Mol. Ecol. Resour.* 28, 2537–2539. doi: 10.1111/j.1471-8286.2005.01155.x
- Percy, D. M., Argus, G. W., Cronk, Q. C., Fazekas, A. J., Kesanakurti, P. R., Burgess, K. S., et al. (2014). Understanding the spectacular failure of DNA barcoding in willows (*Salix*): does this result from a trans-specific selective sweep? *Mol. Ecol.* 23, 4737–4756. doi: 10.1111/mec.12837
- Petit, R. J., Carlson, J., Curtu, A. L., Loustau, M. L., Plomion, C., González-Rodríguez, A., et al. (2013). Fagaceae trees as models to integrate ecology, evolution and genomics. *New Phytol.* 197, 369–371. doi: 10.1111/nph.12089
- Petit, R. J., Csaikl, U. M., Bordács, S., Burg, K., Coart, E., Cottrell, J., et al. (2002). Chloroplast DNA variation in European white oaks: phylogeography and patterns of diversity based on data from over 2600 populations. *For. Ecol. Manage.* 156, 5–26. doi: 10.1016/S0378-1127(01)00645-4
- Pham, K. K., Hipp, A. L., Manos, P. S., and Cronn, R. C. (2017). A time and a place for everything: phylogenetic history and geography as joint predictors of oak plastome phylogeny. *Genome* 60, 720–732. doi: 10.1139/gen-2016-0191
- Phengkklai, C., Santisuk, T., and Larsen, K. (2008). “Fagaceae,” in *Flora of Thailand*, eds T. Santisuk and K. Larsen (Bangkok: The Forest Herbarium), 179–410.
- Piredda, R., Simeone, M. C., Attimonelli, M., Bellarosa, R., and Schirone, B. (2011). Prospects of barcoding the Italian wild dendroflora: oaks reveal severe limitations to tracking species identity. *Mol. Ecol. Resour.* 11, 72–83. doi: 10.1111/j.1755-0998.2010.02900.x
- Pitman, N. C., Mogollón, H., Dávila, N., Ríos, M., García-Villacorta, R., Guevara, J., et al. (2008). Tree community change across 700 km of lowland Amazonian forest from the Andean foothills to Brazil. *Biotropica* 40, 525–535. doi: 10.1111/j.1744-7429.2008.00424.x
- Posada, D., and Crandall, K. A. (1998). Modeltest: testing the model of DNA substitution. *Bioinformatics* 14, 817–818. doi: 10.1093/bioinformatics/14.9.817
- Puillandre, N., Lambert, A., Brouillet, S., and Achaz, G. (2012). ABGD, automatic barcode gap discovery for primary species delimitation. *Mol. Ecol. Resour.* 21, 1864–1877. doi: 10.1111/j.1365-294X.2011.05239.x
- Qi, X. S., Chen, C., Comes, H. P., Sakaguchi, S., Liu, Y. H., Tanaka, N., et al. (2012). Molecular data and ecological niche modelling reveal a highly dynamic evolutionary history of the East Asian Tertiary relict *Cercidiphyllum* (Cercidiphyllaceae). *New Phytol.* 196, 617–630. doi: 10.1111/j.1469-8137.2012.04242.x
- R Core Team (2015). *R: A Language and Environment for Statistical Computing*. Vienna: R Foundation for Statistical Computing.
- Ronquist, F., Teslenko, M., Van Der Mark, P., Ayres, D. L., Darling, A., Höhna, S., et al. (2012). MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst. Biol.* 61, 539–542. doi: 10.1093/sysbio/sys029
- Schaal, B., Hayworth, D., Olsen, K. M., Rauscher, J., and Smith, W. (1998). Phylogeographic studies in plants: problems and prospects. *Mol. Ecol.* 7, 465–474.
- Shapcott, A., Forster, P. I., Guymer, G. P., McDonald, W. J., Faith, D. P., Erickson, D., et al. (2015). Mapping biodiversity and setting conservation priorities for SE Queensland’s rainforests using DNA barcoding. *PLoS One* 10:e0122164. doi: 10.1371/journal.pone.0122164
- Simeone, M. C., Grimm, G. W., Papini, A., Vessella, F., Cardoni, S., Tordoni, E., et al. (2016). Plastome data reveal multiple geographic origins of *Quercus* Group *Ilex*. *PeerJ* 4:e1897. doi: 10.7287/peerj.preprints.1615
- Simeone, M. C., Piredda, R., Papini, A., Vessella, F., and Schirone, B. (2013). Application of plastid and nuclear markers to DNA barcoding of Euro-Mediterranean oaks (*Quercus*, Fagaceae): problems, prospects and phylogenetic implications. *Bot. J. Linn. Soc.* 172, 478–499. doi: 10.1111/boj.12059
- Song, Y., Deng, M., Hipp, A. L., and Li, Q. (2015). Leaf morphological evidence of natural hybridization between two oak species (*Quercus austrocochinensis* and *Q. kerrii*) and its implications for conservation management. *Eur. J. For. Res.* 134, 139–151. doi: 10.1007/s10342-014-0839-x
- Tamaki, I., and Okada, M. (2014). Genetic admixing of two evergreen oaks, *Quercus acuta* and *Q. sessilifolia* (subgenus *Cyclobalanopsis*), is the result of interspecific introgressive hybridization. *Tree Genet. Genomes* 10, 989–999. doi: 10.1007/s11295-014-0737-x
- Tang, C. Q. (2010). Subtropical montane evergreen broad-leaved forests of Yunnan, China: diversity, succession dynamics, human influence. *Front. Earth Sci. China* 4, 22–32. doi: 10.1007/s11707-009-0057-x
- Tao, T. (2010). *BLAST® Help. Standalone BLAST Setup for Unix*. Bethesda, MD: National Center for Biotechnology Information.
- Toumi, L., and Lumaret, R. (2010). Genetic variation and evolutionary history of holly oak: a circum-Mediterranean species-complex [*Quercus coccifera*

- L./*Q. calliprinos* (Webb) Holmboe, Fagaceae]. *Plant Syst. Evol.* 290, 159–171. doi: 10.1007/s00606-010-0358-2
- Valentini, A., Pompanon, F., and Taberlet, P. (2009). DNA barcoding for ecologists. *Trends Ecol. Evol.* 24, 110–117. doi: 10.1016/j.tree.2008.09.011
- Van De Wiel, C., Van Der Schoot, J., Van Valkenburg, J., Duistermaat, H., and Smulders, M. (2009). DNA barcoding discriminates the noxious invasive plant species, floating pennywort (*Hydrocotyle ranunculoides* Lf), from non-invasive relatives. *Mol. Ecol. Resour.* 9, 1086–1091. doi: 10.1111/j.1755-0998.2009.02547.x
- van Velzen, R., Weitschek, E., Felici, G., and Bakker, F. T. (2012). DNA barcoding of recently diverged species: relative performance of matching methods. *PLoS One* 7:e30490. doi: 10.1371/journal.pone.0030490
- Veldman, S., Otieno, J., Gravendeel, B., van Andel, T., and de Boer, H. (2014). “Conservation of endangered wild harvested medicinal plants: use of DNA barcoding,” in *Novel Plant Bioresources*, ed. A. Gurib-Fakim (Chichester: John Wiley & Sons), 81–88. doi: 10.1002/9781118460566.ch6
- Vilgalys, R. (2003). Taxonomic misidentification in public DNA databases. *New Phytol.* 160, 4–5. doi: 10.1046/j.1469-8137.2003.00894.x
- Virgilio, M., Backeljau, T., Nevado, B., and De Meyer, M. (2010). Comparative performances of DNA barcoding across insect orders. *BMC Bioinformatics* 11:206. doi: 10.1186/1471-2105-11-206
- Vitelli, M., Vessella, F., Cardoni, S., Pollegioni, P., Denk, T., Grimm, G. W., et al. (2017). Phylogeographic structuring of plastome diversity in Mediterranean oaks (*Quercus* Group Ilex, Fagaceae). *Tree Genet. Genomes* 13:3. doi: 10.1007/s11295-016-1086-8
- von Cräutlein, M., Korpelainen, H., Pietiläinen, M., and Rikkinen, J. (2011). DNA barcoding: a tool for improved taxon identification and detection of species diversity. *Biodivers. Conserv.* 20, 373–389. doi: 10.1007/s10531-010-9964-0
- Wang, C. W., Yang, B. Y., and Jin, X. H. (2017). *Herminium motuoensis* sp. nov. (Orchidaceae, Orchidoideae), a new species from Tibet, China. *Phytotaxa* 329, 197–200. doi: 10.11646/phytotaxa.329.2.14
- Wang, D. Y., Wang, Q., Wang, Y. L., Xiang, X. G., Huang, L. Q., and Jin, X. H. (2017). Evaluation of DNA barcodes in *Codonopsis* (Campanulaceae) and in some large angiosperm plant genera. *PLoS One* 12:e0170286. doi: 10.1371/journal.pone.0170286
- Wang, J. X., Liang, S. C., Wei, F., and Li, J. W. (2011). Spatial Pattern of *Quercus fabri* population in Karst Hills of Kaili in Guizhou. *J. Anhui Agric. Sci.* 39, 9639–9642. doi: 10.3969/j.issn.0517-6611.2011.16.068
- Wang, Z. L., and Gao, X. M. (2005). The regeneration of *Quercus aliena* var. *acuteserrata*: acorn status, seedling pool and size structure. *J. Northwest A & F Univ.* 25, 986–993. doi: 10.4028/www.scientific.net/amm.52-54.949
- Weitschek, E., Velzen, R., Felici, G., and Bertolazzi, P. (2013). BLOG 2.0: a software system for character-based species classification with DNA Barcode sequences. What it does, how to use it. *Mol. Ecol. Resour.* 13, 1043–1046. doi: 10.1111/1755-0998.12073
- Xiao, Y., Li, C., Hsieh, T. Y., Tian, D. K., Zhou, J. J., Zhang, D. G., et al. (2015). *Eutrema bulbiferum* (Brassicaceae), a new species with bulbils from Hunan, China. *Phytotaxa* 219, 233–242. doi: 10.11646/phytotaxa.219.3.3
- Xu, J., Deng, M., Jiang, X. L., Westwood, M., Song, Y. G., and Turkington, R. (2015). Phylogeography of *Quercus glauca* (Fagaceae), a dominant tree of East Asian subtropical evergreen forests, based on three chloroplast DNA interspace sequences. *Tree Genet. Genomes* 11:805. doi: 10.1007/s11295-014-0805-2
- Xu, J., Jiang, X. L., Deng, M., Westwood, M., Song, Y. G., and Zheng, S. S. (2016). Conservation genetics of rare trees restricted to subtropical montane cloud forests in southern China: a case study from *Quercus arbutifolia* (Fagaceae). *Tree Genet. Genomes* 12:90. doi: 10.1007/s11295-016-1048-1
- Yan, L. J., Liu, J., Moller, M., Zhang, L., Zhang, X. M., Li, D. Z., et al. (2015). DNA barcoding of *Rhododendron* (Ericaceae), the largest Chinese plant genus in biodiversity hotspots of the Himalaya-Hengduan Mountains. *Mol. Ecol. Resour.* 15, 932–944. doi: 10.1111/1755-0998.12353
- Yang, J., Vázquez, L., Chen, X., Li, H., Zhang, H., Liu, Z., et al. (2017). Development of chloroplast and nuclear DNA markers for Chinese oaks (*Quercus* subgenus *Quercus*) and assessment of their utility as DNA barcodes. *Front. Plant Sci.* 8:816. doi: 10.3389/fpls.2017.00816
- Yang, Y. M., Tian, K., Hao, J. M., Pei, S. J., and Yang, Y. X. (2004). Biodiversity and biodiversity conservation in Yunnan, China. *Biodivers. Conserv.* 13, 813–826. doi: 10.1023/b:bioc.0000014464.80847.02
- Yu, W. B., Huang, P. H., Ree, R. H., Liu, M. L., Li, D. Z., and Wang, H. (2011). DNA barcoding of *Pedicularis* L. (Orobanchaceae): evaluating four universal barcode loci in a large and hemiparasitic genus. *J. Syst. Evol.* 49, 425–437. doi: 10.1111/j.1759-6831.2011.00154.x
- Zhao, J. F., Feng, D. J., and Lei, Y. F. (2007). Wood study on *Quercus* and *Cyclobalanopsis* of Fagaceae in Shaanxi Province. *J. Northwest A & F Univ.* 35, 196–202. doi: 10.3321/j.issn:1671-9387.2007.10.038
- Zhou, Z. K., Wilkinson, H., and Wu, C. Y. (1995). Taxonomical and evolutionary implications of the leaf anatomy and architecture of *Quercus* L. subgenus *Quercus* from China. *Cathaya* 7, 1–34. doi: 10.1046/j.1095-8339.2002.00097.x
- Zuidema, P. A., Brienens, R. J., During, H. J., and Güneralp, B. (2009). Do persistently fast-growing juveniles contribute disproportionately to population growth? A new analysis tool for matrix models and its application to rainforest trees. *Am. Nat.* 174, 709–719.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Yan, Xiong, Liu, Deng and Song. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.