

Review

Bone Marrow Defects and Platelet Function: A Focus on MDS and CLL

Sarah Luu ¹ , Elizabeth E. Gardiner ² and Robert K. Andrews ^{1,*} 

¹ Australian Centre for Blood Diseases, Monash University, Melbourne, VIC 3004, Australia; sarah.luu@monash.edu

² ACRF Department of Cancer Biology and Therapeutics, The John Curtin School of Medical Research, The Australian National University, Canberra, ACT 2600, Australia; elizabeth.gardiner@anu.edu.au

* Correspondence: rob.andrews@monash.edu; Tel.: +61-03-9903-0136

Received: 20 April 2018; Accepted: 16 May 2018; Published: 18 May 2018



Abstract: The bloodstream typically contains >500 billion anucleate circulating platelets, derived from megakaryocytes in the bone marrow. This review will focus on two interesting aspects of bone marrow dysfunction and how this impacts on the quality of circulating platelets. In this regard, although megakaryocytes are from the myeloid lineage leading to granulocytes (including neutrophils), erythrocytes, and megakaryocytes/platelets, recent evidence has shown that defects in the lymphoid lineage leading to B cells, T cells, and natural killer (NK) cells also result in abnormal circulating platelets. Current evidence is limited regarding whether this latter phenomenon might potentially arise from (a) some form of as-yet-undetected defect common to both lineages; (b) adverse interactions occurring between cells of different lineages within the bone marrow environment; and/or (c) unknown disease-related factor(s) affecting circulating platelet receptor expression/function after their release from megakaryocytes. Understanding the mechanisms underlying how both myeloid and lymphoid lineage bone marrow defects lead to dysfunction of circulating platelets is significant because of the potential diagnostic and predictive value of peripheral platelet analysis for bone marrow disease progression, the additional potential effects of new anti-cancer drugs on platelet function, and the critical role platelets play in regulation of bleeding risk, inflammation, and innate immunity.

Keywords: platelets; bone marrow defects; glycoprotein Ib α ; glycoprotein VI

1. Introduction

The bloodstream typically contains >500 billion circulating anucleate platelets ($\sim 150\text{--}400 \times 10^9/\text{L}$ in $\sim 4\text{--}5$ L of blood) derived from megakaryocytes primarily situated in the bone marrow. In simple terms, megakaryocytes are within the myeloid lineage leading to granulocytes (including neutrophils), erythrocytes, and platelets, whereas the lymphoid lineage, derived from a common precursor, leads to B cells, T cells, and natural killer (NK) cells (Figure 1). Together, these different types of cells released into the vasculature play a crucial role in the transfer of oxygen, nutrients, and waste products throughout the body, in preventing blood loss following injury and promoting wound healing, and in fighting infection through innate and adaptive immune defense strategies [1–3]. On the other hand, dysfunction or deficiency of one or more of these cells due to bone marrow defects or other causes can result in bleeding, thrombotic disease, coagulopathy, inflammatory disease, increased infection risk, and/or (auto)immune disorders, with various cells typically playing multiple roles in initiating or regulating these interrelated health/disease processes.

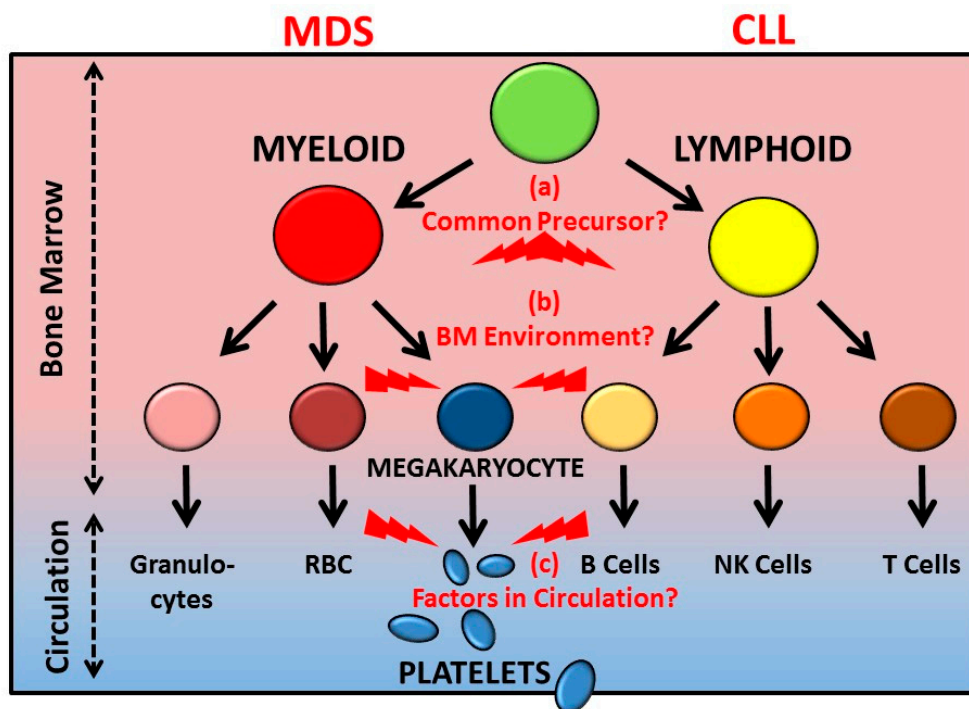


Figure 1. Haematopoietic cells are derived from the myeloid lineage leading to generation of granulocytes (including neutrophils), erythrocytes (red blood cells; RBC), and megakaryocytes that generate platelets, or from the lymphoid lineage producing immune B cells, T cells, and natural killer (NK) cells. Defects in the lymphoid lineage pathways surprisingly also lead to abnormal circulating platelets. Dysfunction of circulating platelets may be associated with myeloid lineage bone marrow defects such as myelodysplastic syndrome (MDS) or lymphoid lineage defects such as chronic lymphocytic leukemia (CLL), by unknown mechanisms potentially involving (a) some form of defect common to both lineages; (b) adverse interactions occurring between cells of different lineages within the bone marrow environment; and/or (c) unknown disease-related factor(s) affecting circulating platelet receptor expression/function after their release from megakaryocytes. Understanding mechanisms may be significant for understanding increased risks of bleeding or infection. See the text for references.

In this review, we will briefly highlight the role of human blood platelets in haemostasis and thrombosis, procoagulant and pro-inflammatory responses, innate immunity, and cancer. In this respect, although individuals may be thrombocytopenic, with platelet count $<150 \times 10^9/L$, in the absence of other causes, bleeding is generally not a major risk until platelets are $<5\text{--}10 \times 10^9/L$, suggesting there are far greater numbers of platelets in the circulation than required for their primary haemostatic function, and consistent with their expanding roles in other areas of vascular biology. In the absence of thrombocytopenia, platelets may be dysfunctional due to a plethora of causes, either inherited or acquired. This review will then consider how bone marrow defects of the myeloid or lymphoid lineage may result in defects of circulating platelets, using myelodysplastic syndrome (MDS) and chronic lymphocytic leukemia (CLL), respectively, as cases in point.

2. Platelet Function and Key Receptors, Including Roles in Cancer/Metastasis

In haemostasis, circulating platelets in flowing blood rapidly adhere at sites of vascular injury, become activated, secrete platelet agonists, procoagulant and pro-inflammatory factors from storage granules, release microparticles, and activate $\alpha\text{IIb}\beta_3$ and other integrins that mediate platelet aggregation or thrombus formation. The activated platelet surface through exposure of phosphatidyl serine and other pathways promotes the generation of thrombin, a potent human platelet agonist

that engages protease-activated receptors (PAR)-1 and 4, and thrombin-dependent fibrin formation in coagulation. The key platelet-specific receptors glycoprotein (GP)Ib α of the GPIb-IX-V complex and GPVI are critical for initiating platelet adhesion and activation particularly at elevated shear rates in flowing blood [4,5]. GPIb α of the leucine-rich repeat protein family binds von Willebrand factor (VWF), is a high-affinity binding site for thrombin, and also binds other coagulation factors (XII, XI, high molecular weight kininogen) and receptors P-selectin expressed on activated platelets or activated endothelial cells and α M β 2 on activated neutrophils; P-selectin expressed on the surface of activated platelets also adheres to P-selectin glycoprotein ligand-1 (PSGL-1) on neutrophils [6]. GPIb α forms a disulfide-linked complex with GPIb β , and noncovalently associates with GPIX and GPV, all members of the leucine-rich repeat family homologous to mammalian innate immune toll-like receptor (TLR) proteins [7] and to primitive defensive proteins (variable lymphocyte receptors) expressed in the jawless vertebrates, lampreys and hagfish [8]. GPV is involved in adhesion of the complex to collagen [9]. In addition, GPIb α also forms a noncovalent complex with GPVI on the human platelet surface [10]. GPVI of the immunoglobulin-like family is co-associated with the Fc receptor (FcR) γ , required for GPVI surface expression. FcR γ contains an immunoreceptor tyrosine-based activation motif (ITAM) linking engagement of GPVI/FcR γ by physiological ligands collagen and fibrin or non-physiological ligands such as cross-linked collagen-related peptide (CRP) or convulxin, to Src family kinase signalling pathways [4,5]. Conserved cytoplasmic sequences of GPIb-IX-V/GPVI are directly linked to various intracellular signalling/cytoskeletal proteins, including tumour necrosis factor (TNF) receptor-associated factor-4 (TRAF-4)/p47^{phox} of the nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (Nox) complexes generating intracellular reactive oxygen species (ROS) following engagement of GPIb α or GPVI [11,12], and the p85 subunit of phosphatidylinositol-3 (PI-3) kinase [13,14]. The major signalling pathways downstream of both GPIb-IX-V and GPVI engagement include Src, PI3-kinase, and spleen tyrosine kinase (Syk), leading to activation of phospholipase C γ (PLC γ) and mobilization of intracellular Ca²⁺, and platelet aggregation in response to selective engagement of GPIb α is blocked by inhibitors of Src, PI3-kinase, and Syk [15]. GPVI forms dimers on the platelet surface, and ligand-induced cross-linking or clustering of GPVI/FcR γ also leads to rapid Src kinase-dependent ITAM-mediated Syk phosphorylation, and activation of PLC γ leading to Ca²⁺ mobilization [16–19].

Importantly, cytoplasmic sequences of GPIb-IX-V/GPVI are also directly linked to calmodulin [20–22] which regulates a disintegrin and metalloproteinase (ADAM)-dependent extracellular shedding of ligand-binding domains of GPIb α (ADAM17), GPV (ADAM10/17), and GPVI (ADAM10) [23]. Plasma soluble GPVI (sGPVI) as well as soluble GPIb α and GPV, constitute unique platelet-specific biomarkers of platelet activation or dysfunction, with sGPVI levels in particular linked to specific bleeding risk [24,25]. Such regulation of platelet surface receptor density is likely to be increasingly studied with respect to adhesive function, signalling functions, interaction with endothelial cells or leukocytes, and clearance of platelets by macrophage adhesion in the liver [26,27]. Compared to existing antiplatelet drugs [28–30], all of which are limited to some extent by low efficacy or high bleeding risk, new targets including GPIb α and GPVI are emerging and antiplatelet therapy may have wider application in other human disease.

During inflammation, platelets adhere to neutrophils via GPIb α / α M β 2 and P-selectin/PSGL-1 interactions, and regulate angiogenesis and vascular integrity via GPVI, α IIb β 3 and the platelet C-type lectin-like receptor 2 (CLEC-2) [30–33]. Platelets also interact with bacteria and other pathogens via GPIb α , GPVI, α IIb β 3, and other mechanisms [34–36]. In addition to pro-inflammatory cytokines, activated platelets also release growth factors and microparticles which could all potentially play an important role in cancer, albeit the association between abnormal platelet function related to bleeding or thrombotic/coagulopathy risk, immunosuppression and cancer survival, growth and/or metastasis is complex [37–51]. Platelets may interact with metastatic cancer cells entering the blood stream via receptor-mediated adhesion and/or release of soluble factors, contribute to suppression of immune response towards cancer cells [52], and/or regulate localization within the microvasculature to enable

redistribution and enhance survival. Coagulopathy, including contact or tissue factor pathways, activated platelets, microparticles, thrombin generation, and platelet thrombin PAR receptors, may also play a role in cancer-related thrombosis [53–55]. The cause or effect of changes in platelet profiles and cancer is not always clear [46], however platelets may ultimately offer new anti-tumour therapeutic approaches [38,48–51,56,57]. Circulating platelets could therefore be informative regarding mechanisms, constitute therapeutic targets, and/or be useful diagnostically on progression or response to treatment. In this regard, platelet RNA is also emerging as an interesting new diagnostic parameter [58,59]. Here, we will consider two examples where acquired changes in platelet receptor expression or function have been reported, in myeloid or lymphoid blood cancers.

3. Myelodysplastic Syndrome (MDS) and Platelet Function

MDS is characterized by ineffective production (dysplasia) of myeloid blood cells (Figure 1), with accompanying risk of transformation to acute myelogenous leukaemia (AML). The underlying causes of MDS affect adults and less frequently children. These causes are heterogeneous, with multiple clonal haematopoietic defects identified and including germ line inheritable forms, in addition to a number of predisposing factors, and currently the main therapy involves haematopoietic stem cell transplantation [60,61]. There may be a variable extent of thrombocytopenia and other platelet/haemostatic defects associated with MDS, and there may be bleeding episodes of varying severity.

An acquired platelet GPVI defect associated with MDS has been reported, with essentially normal platelet counts and GPVI expression, but with no aggregation to platelet GPVI ligands (collagen, CRP, or convulxin), and with ultimate progression to AML [62]. Subsequent studies of 26 patients with MDS also showed platelet-related defects unrelated to thrombocytopenia, including high prevalence (>80%) of defective aggregation in response to collagen or other agonists (adrenaline, adenosine diphosphate (ADP), ristocetin) [63]. Defective platelet aggregation also strongly correlated with poor prognosis, although the underlying mechanism is not currently known. A further case of confirmed MDS-low risk with normal platelet count and increased bleeding tendency (recurring episodes over the previous 18 months), also showed defective GPVI-dependent aggregation (collagen, CRP), abnormally low GPVI surface expression on platelets and a signalling defect associated as a molecular abnormality in proteolytic processing of phosphorylated Syk [64]. Further, it was found that GPVI ligation induced intracellular ROS generation, independent of Syk activation or pathways leading to platelet aggregation or ectodomain shedding. There was a gradual decline in platelet count over several months, and also progression to AML [64]. Another study of 75 MDS cases with varying degrees of thrombocytopenia showed defective platelet activation, as well as reduced circulating immature platelet fraction and an increase of apoptotic markers in platelets consistent with defective platelet production [65].

While platelet GPVI-related dysfunction is associated with a significant proportion of MDS cases, further studies are no doubt required to fully evaluate these changes temporally and in response to treatment in order to discover the molecular link between specific defects and platelet function, and to establish any prognostic value in future. While the underlying mechanisms could speculatively involve either direct or indirect targeting of particular signalling or cytoskeletal proteins involved in GPVI expression or function, it is clear that more detailed experimental analysis perhaps involving megakaryocyte development and platelet production together with advanced screening of MDS is required to fully understand the precise causes and effects related to the altered platelet phenotype.

4. Chronic Lymphocytic Leukemia (CLL) and Platelet Function

CLL is characterized by defects in development of B-lymphocytes (Figure 1), but can also be associated with platelet/haemostatic dysfunction [66–69]. Ibrutinib, one of a number of recent drugs developed for the treatment of CLL, targets Bruton's tyrosine kinase (Btk), a signalling molecule that plays an essential role in B-cell development [66,69,70]. While reasonably well tolerated in clinical trials, its expanding use has corresponded with an increased bleeding risk, as well as other cardiovascular

complications and infections [66]. Ibrutinib treatment may also result in off-target inhibition of additional Src family kinase members [71]. Btk has previously been linked to signalling pathways downstream of platelet GPIb α and regulation of GPIb α -dependent thrombus formation [72], consistent with subsequent studies reporting an inhibitory effect of Ibrutinib on GPIb α /GPVI-dependent platelet function and potential exacerbation of bleeding risk [73–75]. Interestingly, circulating platelets from a small cohort of untreated refractory CLL patients showed significantly decreased surface expression of GPIb α and GPVI, whereas surface expression of α Ib β 3 and CD9 (control) were essentially normal [76]. Together, this suggests that Ibrutinib or other signalling pathway inhibitors [70] could exacerbate bleeding disorders in platelets already compromised by abnormally low receptor expression [76]. That is, while evidence for platelet dysfunction associated with CLL is more limited than is the case for MDS discussed above, and while spontaneous or clinically significant bleeding is not typically reported as being associated with CLL, significantly decreased levels of expression of GPIb α and GPVI, together with mildly reduced platelet count and/or the increased susceptibility to bleeding in the presence of anti-cancer drugs that impact upon key platelet signalling pathways, may ultimately have consequences in terms of monitoring bleeding risk or use of antiplatelet agents. Also, given the emerging non-haemostatic role for platelets mentioned above [1–3,30–36], defects in expression of GPIb α /GPVI could potentially be related to changes in inflammatory/infection risk or immune dysfunction. These platelet abnormalities prior to treatment could also be relevant to bleeding associated with use of other CLL drugs, such as those targeting B cell CLL/lymphoma 2 (BCL-2) proteins that regulate apoptosis/survival. Whereas a subsequent BCL-2-selective drug (ABT-199) had less impact on platelet function [77], pan-BCL-2 inhibitory drugs that target both BCL-2 and BCL-2-like protein 1 (ABT-737, ABT-263) resulted in pronounced transient thrombocytopenia and functional impairment of α Ib β 3 together with enhanced shedding of GPIb α and GPVI, resulting in markedly defective thrombus formation [78]. Further, unlike immune-related thrombocytopenia where anti-platelet antibodies acting at the Fc γ RIIa receptor markedly increase GPVI shedding resulting in lower surface expression and increased levels of sGPVI in plasma [76,79,80], in untreated CLL the plasma sGPVI levels were in the normal range, suggesting observed low platelet GPVI was unlikely to be the result of increased shedding.

Finally, current evidence is limited regarding whether the effect of CLL-related defects on circulating platelet profile might potentially result from (a) some form of as-yet-undetected defect common to both lineages; (b) adverse interactions occurring between cells of different lineages within the bone marrow environment; and/or (c) unknown disease-related factor(s) affecting circulating platelet receptor expression/function after their release from megakaryocytes. As CLL is not associated with either a unique cytogenetic or a molecular defect, mouse models of CLL generally recapitulate only some aspects of the disease, however it would be of interest to examine platelet and megakaryocyte function in one or more mouse models of progressive CLL [81]. As for MDS discussed above, further studies are now required to evaluate temporal changes in platelet receptor expression in early and late stage CLL, in the absence or presence of drugs or other treatment(s), to determine the relationship to platelet function and bleeding risk, as well as to establish any future clinical value in such analysis.

5. Conclusions and Future Directions

The heterogeneous nature of MDS and CLL is unlikely to result in a simple explanation for changes in platelet receptor expression and function associated with both diseases (Figure 1). In both cases, a platelet production defect could explain variation in platelet count as well as altered receptor density, although changes in circulating red blood cells, neutrophils or B cells could conceivably affect platelet receptor expression by increased platelet activation status, enhanced receptor shedding or other mechanisms. Cell co-culture experiments involving platelet production *in vitro* in the presence or absence of other mutant bone marrow cells could provide some useful information on the potential mechanisms involved. More extensive temporal analyses in human MDS or CLL, or the corresponding animal models, could also be informative. Finally, the extensive genetic defects underlying MDS or

CLL will ultimately be interpretable as genetic abnormalities associated with congenital and acquired platelet defects are increasingly understood. In any case, these studies of platelet-specific receptors highlight the scientific and potential clinical interest in such changes in human disease.

In conclusion, understanding the mechanisms underlying how both myeloid and lymphoid lineage bone marrow defects lead to dysfunction of circulating platelets is significant because of the potential diagnostic and predictive value of peripheral platelet analysis for bone marrow disease progression, the additional potential effects of new anti-cancer drugs on platelet function, and the critical role platelets play in regulation of bleeding risk, inflammation, and innate immunity. These findings ultimately may also be relevant to increased understanding of other human diseases (e.g., congenital platelet disorders, cardiovascular disease, stroke, autoimmune thrombocytopenia) and/or surgical procedures (e.g., circulatory support devices, splenectomy) where expression/function of platelet-specific receptors GPIb α /GPVI is compromised and there is concomitant altered haemostatic, thrombotic, inflammatory, and/or immune risk.

Author Contributions: S.L., E.E.G., and R.K.A. wrote the paper.

Acknowledgments: The authors acknowledge their research colleagues for helpful discussions and the National Health and Medical Research Council of Australia for financial support.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Li, J.L.; Zarbock, A.; Hidalgo, A. Platelets as autonomous drones for hemostatic and immune surveillance. *J. Exp. Med.* **2017**, *214*, 2193–2204. [[CrossRef](#)] [[PubMed](#)]
- Jenne, C.N.; Kubes, P. Platelets in inflammation and infection. *Platelets* **2015**, *26*, 286–292. [[CrossRef](#)] [[PubMed](#)]
- Koupenova, M.; Clancy, L.; Corkrey, H.A.; Freedman, J.E. Circulating platelets as mediators of immunity, inflammation, and thrombosis. *Circ. Res.* **2018**, *122*, 337–351. [[CrossRef](#)] [[PubMed](#)]
- Andrews, R.K.; Gardiner, E.E. Basic mechanisms of platelet receptor shedding. *Platelets* **2017**, *28*, 319–324. [[CrossRef](#)] [[PubMed](#)]
- Gardiner, E.E.; Andrews, R.K. Platelet receptor expression and shedding: Glycoprotein Ib-IX-V and glycoprotein VI. *Transfus. Med. Rev.* **2014**, *28*, 56–60. [[CrossRef](#)] [[PubMed](#)]
- Gardiner, E.E.; Andrews, R.K. Structure and function of platelet receptors initiating blood clotting. In *A Systems Biology Approach to Hematology*; Corey, S., Kimmel, M., Eds.; Springer: New York, NY, USA, 2014; pp. 263–275.
- Hamzeh-Cognasse, H.; Berthelot, P.; Tardy, B.; Pozzetto, B.; Bourlet, T.; Laradi, S.; Garraud, O.; Cognasse, F. Platelet toll-like receptors are crucial sensors of infectious danger moieties. *Platelets* **2018**. [[CrossRef](#)] [[PubMed](#)]
- Han, B.W.; Herrin, B.R.; Cooper, M.D.; Wilson, I.A. Antigen recognition by variable lymphocyte receptors. *Science* **2008**, *321*, 1834–1837. [[CrossRef](#)] [[PubMed](#)]
- Moog, S.; Mangin, P.; Lenain, N.; Strassel, C.; Ravanat, C.; Schuhler, S.; Freund, M.; Santer, M.; Kahn, M.; Nieswandt, B.; et al. Platelet glycoprotein V binds to collagen and participates in platelet adhesion and aggregation. *Blood* **2001**, *98*, 1038–1046. [[CrossRef](#)] [[PubMed](#)]
- Arthur, J.F.; Matzaris, M.; Gardiner, E.E.; Taylor, S.G.; Wijeyewickrema, L.C.; Ozaki, Y.; Kahn, M.L.; Andrews, R.K.; Berndt, M.C. Glycoprotein (GP)VI is associated with GPIb-IX-V on the membrane of resting and activated platelets. *Thromb. Haemost.* **2005**, *93*, 716–723. [[PubMed](#)]
- Arthur, J.F.; Shen, Y.; Gardiner, E.E.; Coleman, L.; Kenny, D.; Andrews, R.K.; Berndt, M.C. TNF Receptor-Associated Factor 4 (TRAF4) is a novel binding partner of glycoprotein Ib and glycoprotein VI in human platelets. *J. Thromb. Haemost.* **2011**, *9*, 163–172. [[CrossRef](#)] [[PubMed](#)]
- Arthur, J.F.; Gardiner, E.E.; Kenny, D.; Andrews, R.K.; Berndt, M.C. Platelet receptor redox regulation. *Platelets* **2008**, *19*, 1–8. [[CrossRef](#)] [[PubMed](#)]

13. Mu, F.T.; Andrews, R.K.; Arthur, J.F.; Munday, A.M.; Cranmer, S.L.; Jackson, S.P.; Stomski, F.; Lopez, A.F.; Berndt, M.C. A functional 14-3-3 ζ -independent association of PI3-kinase with glycoprotein Ib α , the major ligand-binding subunit of the platelet glycoprotein Ib-IX-V complex. *Blood* **2008**, *111*, 4580–4587. [[CrossRef](#)] [[PubMed](#)]
14. Mu, F.T.; Cranmer, S.L.; Andrews, R.K.; Berndt, M.C. Functional association of PI3-kinase with platelet glycoprotein Ib α , the major ligand-binding subunit of the glycoprotein Ib-IX-V complex. *J. Thromb. Haemost.* **2010**, *8*, 324–330. [[CrossRef](#)] [[PubMed](#)]
15. Gardiner, E.E.; Arthur, J.F.; Shen, Y.; Karunakaran, D.; Moore, L.A.; Schulte Am Esch, J., II; Andrews, R.K.; Berndt, M.C. GPIb α -selective activation of platelets induces platelet signalling events comparable to GPVI activation events. *Platelets* **2010**, *21*, 244–252. [[CrossRef](#)] [[PubMed](#)]
16. Jung, S.M.; Tsuji, K.; Moroi, M. Glycoprotein (GP) VI dimer as a major collagen-binding site of native platelets: Direct evidence obtained with dimeric GPVI-specific Fabs. *J. Thromb. Haemost.* **2009**, *7*, 1347–1355. [[CrossRef](#)] [[PubMed](#)]
17. Loyau, S.; Dumont, B.; Ollivier, V.; Boulaftali, Y.; Feldman, L.; Ajzenberg, N.; Jandrot-Perrus, M. Platelet glycoprotein VI dimerization, an active process inducing receptor competence, is an indicator of platelet reactivity. *Arterioscler. Thromb. Vasc. Biol.* **2012**, *32*, 778–785. [[CrossRef](#)] [[PubMed](#)]
18. Jung, S.M.; Moroi, M.; Soejima, K.; Nakagaki, T.; Miura, Y.; Berndt, M.C.; Gardiner, E.E.; Howes, J.M.; Pugh, N.; Bihan, D.; et al. Constitutive dimerization of glycoprotein VI (GPVI) in resting platelets is essential for binding to collagen and activation in flowing blood. *J. Biol. Chem.* **2012**, *287*, 30000–30013. [[CrossRef](#)] [[PubMed](#)]
19. Poulter, N.S.; Pollitt, A.Y.; Owen, D.M.; Gardiner, E.E.; Andrews, R.K.; Shimizu, H.; Ishikawa, D.; Bihan, D.; Farndale, R.W.; Moroi, M.; et al. Clustering of glycoprotein VI (GPVI) dimers upon adhesion to collagen as a mechanism to regulate GPVI signaling in platelets. *J. Thromb. Haemost.* **2017**, *15*, 549–564. [[CrossRef](#)] [[PubMed](#)]
20. Andrews, R.K.; Munday, A.D.; Mitchell, C.A.; Berndt, M.C. Interaction of calmodulin with the cytoplasmic domain of the glycoprotein Ib-IX-V complex. *Blood* **2001**, *98*, 681–687. [[CrossRef](#)] [[PubMed](#)]
21. Andrews, R.K.; Suzuki-Inoue, K.; Shen, Y.; Tulasne, D.; Watson, S.P.; Berndt, M.C. Interaction of calmodulin with the cytoplasmic domain of platelet glycoprotein VI. *Blood* **2002**, *99*, 4219–4221. [[CrossRef](#)] [[PubMed](#)]
22. Gardiner, E.E.; Arthur, J.F.; Berndt, M.C.; Andrews, R.K. Role of calmodulin in platelet receptor function. *Curr. Med. Chem. Cardiovasc. Hematol. Agents* **2005**, *3*, 173–179. [[CrossRef](#)]
23. Gardiner, E.E.; Karunakaran, D.; Shen, Y.; Andrews, R.K.; Berndt, M.C. Controlled shedding of platelet glycoprotein (GP)VI and GPIb-IX-V by ADAM family metalloproteinases. *J. Thromb. Haemost.* **2007**, *5*, 1530–1537. [[CrossRef](#)] [[PubMed](#)]
24. Gardiner, E.E.; Andrews, R.K. Plasma sGPVI: Changing levels in human disease. *Thromb. Res.* **2014**, *133*, 306–307. [[CrossRef](#)] [[PubMed](#)]
25. Muthiah, K.; Connor, D.; Ly, K.; Gardiner, E.E.; Andrews, R.K.; Qiao, J.; Rutgers, D.; Robson, D.; Low, J.; Jarvis, S.; et al. Longitudinal changes in haemostatic parameters and reduced pulsatility contribute to non-surgical bleeding in patients with centrifugal continuous flow left ventricular assist devices. *J. Heart Lung Transplant.* **2016**, *35*, 743–751. [[CrossRef](#)] [[PubMed](#)]
26. Gardiner, E.E.; Andrews, R.K. Metalloproteolytic receptor shedding . . . platelets “acting their age”. *Platelets* **2016**, *27*, 512–518.
27. Quach, M.E.; Chen, W.; Li, R. Mechanisms of platelet clearance and translation to improve platelet storage. *Blood* **2018**, *131*, 1512–1521. [[CrossRef](#)] [[PubMed](#)]
28. Metharom, P.; Berndt, M.C.; Baker, R.I.; Andrews, R.K. Current state and novel approaches of antiplatelet therapy. *Arterioscler. Thromb. Vasc. Biol.* **2015**, *35*, 1327–1338. [[CrossRef](#)] [[PubMed](#)]
29. McFadyen, J.D.; Schaff, M.; Peter, K. Current and future antiplatelet therapies: Emphasis on preserving haemostasis. *Nat. Rev. Cardiol.* **2018**, *15*, 181–191. [[CrossRef](#)] [[PubMed](#)]
30. Plow, E.F.; Wang, Y.; Simon, D.I. The search for new antithrombotic mechanisms and therapies that may spare hemostasis. *Blood* **2018**, *131*, 1899–1902. [[CrossRef](#)] [[PubMed](#)]
31. Walsh, T.G.; Metharom, P.; Berndt, M.C. The functional role of platelets in the regulation of angiogenesis. *Platelets* **2015**, *26*, 199–211. [[CrossRef](#)] [[PubMed](#)]
32. Deppermann, C. Platelets and vascular integrity. *Platelets* **2018**. [[CrossRef](#)] [[PubMed](#)]

33. Suzuki-Inoue, K.; Tsukiji, N.; Shirai, T.; Osada, M.; Inoue, O.; Ozaki, Y. Platelet CLEC-2: Roles beyond hemostasis. *Semin. Thromb. Hemost.* **2018**, *44*, 126–134. [[PubMed](#)]
34. Cox, D.; Kerrigan, S.W.; Watson, S.P. Platelets and the innate immune system: Mechanisms of bacterial-induced platelet activation. *J. Thromb. Haemost.* **2011**, *9*, 1097–1107. [[CrossRef](#)] [[PubMed](#)]
35. Andrews, R.K.; Arthur, J.A.; Gardiner, E.E. Neutrophil extracellular traps (NETs) and the role of platelets in infection. *Thromb. Haemost.* **2014**, *112*, 659–665. [[CrossRef](#)] [[PubMed](#)]
36. Hamzeh-Cognasse, H.; Damien, P.; Chabert, A.; Pozzetto, B.; Cognasse, F.; Garraud, O. Platelets and infections—Complex interactions with bacteria. *Front. Immunol.* **2015**, *6*, 82. [[CrossRef](#)] [[PubMed](#)]
37. Erpenbeck, L.; Schon, M.P. Deadly allies: The fatal interplay between platelets and metastasizing cancer cells. *Blood* **2010**, *115*, 3427–3436. [[CrossRef](#)] [[PubMed](#)]
38. Elaskalani, O.; Berndt, M.C.; Falasca, M.; Metharom, P. Targeting platelets for the treatment of cancer. *Cancers* **2017**, *9*, 94. [[CrossRef](#)] [[PubMed](#)]
39. Au, A.E.; Josefsson, E.C. Regulation of platelet membrane protein shedding in health and disease. *Platelets* **2017**, *28*, 342–353. [[CrossRef](#)] [[PubMed](#)]
40. Lowe, K.L.; Navarro-Nunez, L.; Watson, S.P. Platelet CLEC-2 and podoplanin in cancer metastasis. *Thromb. Res.* **2012**, *129* (Suppl. 1), S30–S37. [[CrossRef](#)]
41. Chang, Y.W.; Hsieh, P.W.; Chang, Y.T.; Lu, M.H.; Huang, T.F.; Chong, K.Y.; Liao, H.R.; Cheng, J.C.; Tseng, C.P. Identification of a novel platelet antagonist that binds to CLEC-2 and suppresses podoplanin-induced platelet aggregation and cancer metastasis. *Oncotarget* **2015**, *6*, 42733–42748. [[CrossRef](#)] [[PubMed](#)]
42. Lazar, S.; Goldfinger, L.E. Platelet microparticles and miRNA transfer in cancer progression: Many targets, modes of action, and effects across cancer stages. *Front. Cardiovasc. Med.* **2018**, *5*, 13. [[CrossRef](#)] [[PubMed](#)]
43. Riedl, J.; Pabinger, I.; Ay, C. Platelets in cancer and thrombosis. *Hamostaseologie* **2014**, *34*, 54–62. [[CrossRef](#)] [[PubMed](#)]
44. Franco, A.T.; Corken, A.; Ware, J. Platelets at the interface of thrombosis, inflammation, and cancer. *Blood* **2015**, *126*, 582–588. [[CrossRef](#)] [[PubMed](#)]
45. Coupland, L.A.; Hindmarsh, E.J.; Gardiner, E.E.; Parish, C.R. The influence of platelet membranes on tumour cell behaviour. *Cancer Metastasis Rev.* **2017**, *36*, 215–224. [[CrossRef](#)] [[PubMed](#)]
46. Menter, D.G.; Tucker, S.C.; Kopetz, S.; Sood, A.K.; Crissman, J.D.; Honn, K.V. Platelets and cancer: A casual or causal relationship: Revisited. *Cancer Metastasis Rev.* **2014**, *33*, 231–269. [[CrossRef](#)] [[PubMed](#)]
47. Sierko, E.; Wojtukiewicz, M.Z. Platelets and angiogenesis in malignancy. *Semin. Thromb. Hemost.* **2004**, *30*, 95–108. [[PubMed](#)]
48. Olsson, A.K.; Cedervall, J. The pro-inflammatory role of platelets in cancer. *Platelets* **2018**. [[CrossRef](#)] [[PubMed](#)]
49. Plantureux, L.; Crescence, L.; Dignat-George, F.; Panicot-Dubois, L.; Dubois, C. Effects of platelets on cancer progression. *Thromb. Res.* **2018**, *164* (Suppl. 1), S40–S47. [[CrossRef](#)] [[PubMed](#)]
50. Haemmerle, M.; Stone, R.L.; Menter, D.G.; Afshar-Kharghan, V.; Sood, A.K. The platelet lifeline to cancer: Challenges and opportunities. *Cancer Cell* **2018**. [[CrossRef](#)] [[PubMed](#)]
51. Gresele, P.; Malvestiti, M.; Momi, S. Anti-platelet treatments in cancer: Basic and clinical research. *Thromb. Res.* **2018**, *164* (Suppl. 1), S106–S111. [[CrossRef](#)] [[PubMed](#)]
52. Servais, L.; Wéra, O.; Dibato Epoh, J.; Delierneux, C.; Bouznad, N.; Rahmouni, S.; Mazzucchelli, G.; Baiwir, D.; Delvenne, P.; Lancellotti, P.; et al. Platelets contribute to the initiation of colitis-associated cancer by promoting immunosuppression. *J. Thromb. Haemost.* **2018**, *16*, 762–777. [[CrossRef](#)] [[PubMed](#)]
53. Wojtukiewicz, M.Z.; Hempel, D.; Sierko, E.; Tucker, S.C.; Honn, K.V. Protease-activated receptors (PARs)—Biology and role in cancer invasion and metastasis. *Cancer Metastasis Rev.* **2015**, *34*, 775–796. [[CrossRef](#)] [[PubMed](#)]
54. Date, K.; Ettlai, C.; Maraveyas, A. Tissue factor-bearing microparticles and inflammation: A potential mechanism for the development of venous thromboembolism in cancer. *J. Thromb. Haemost.* **2017**, *15*, 2289–2299. [[CrossRef](#)] [[PubMed](#)]
55. Campello, E.; Henderson, M.W.; Noubouossie, D.F.; Simioni, P.; Key, N.S. Contact system activation and cancer: New insights in the pathophysiology of cancer-associated thrombosis. *Thromb. Haemost.* **2018**, *118*, 251–265. [[CrossRef](#)] [[PubMed](#)]
56. Hyslop, S.R.; Josefsson, E.C. Undercover agents: Targeting tumours with modified platelets. *Trends Cancer* **2017**, *3*, 235–246. [[CrossRef](#)] [[PubMed](#)]

57. Xu, X.R.; Yousef, G.M.; Ni, H. Cancer and platelet crosstalk: Opportunities and challenges for aspirin and other antiplatelet agents. *Blood* **2018**, *131*, 1777–1789. [[CrossRef](#)] [[PubMed](#)]
58. Best, M.G.; Vancura, A.; Wurdinger, T. Platelet RNA as a circulating biomarker trove for cancer diagnostics. *J. Thromb. Haemost.* **2017**, *15*, 1295–1306. [[CrossRef](#)] [[PubMed](#)]
59. Tjon-Kon-Fat, L.A.; Sol, N.; Wurdinger, T.; Nilsson, R.J.A. Platelet RNA in cancer diagnostics. *Semin. Thromb. Hemost.* **2018**, *44*, 135–141. [[CrossRef](#)] [[PubMed](#)]
60. Locatelli, F.; Strahm, B. How I treat myelodysplastic syndromes of childhood. *Blood* **2018**, *131*, 1406–1414. [[CrossRef](#)] [[PubMed](#)]
61. Theilgaard-Monch, K.; Boultonwood, J.; Ferrari, S.; Giannopoulos, K.; Hernandez-Rivas, J.M.; Kohlmann, A.; Morgan, M.; Porse, B.; Tagliafico, E.; Zwaan, C.M.; et al. Gene expression profiling in MDS and AML: Potential and future avenues. *Leukemia* **2011**, *25*, 909–920. [[CrossRef](#)] [[PubMed](#)]
62. Bellucci, S.; Huisse, M.G.; Boval, B.; Hainaud, P.; Robert, A.; Fauvel-Lafeve, F.; Jandrot-Perrus, M. Defective collagen-induced platelet activation in two patients with malignant haemopathies is related to a defect in the GPVI-coupled signalling pathway. *Thromb. Haemost.* **2005**, *93*, 130–138. [[CrossRef](#)] [[PubMed](#)]
63. Girtovitis, F.I.; Ntaios, G.; Papadopoulos, A.; Ioannidis, G.; Makris, P.E. Defective platelet aggregation in myelodysplastic syndromes. *Acta Haematol.* **2007**, *118*, 117–122. [[CrossRef](#)] [[PubMed](#)]
64. Qiao, J.; Arthur, J.F.; Collett, M.; Shen, Y.; Mu, F.T.; Berndt, M.C.; Davis, A.K.; Andrews, R.K.; Gardiner, E.E. An acquired defect associated with abnormal signaling of the platelet collagen receptor glycoprotein VI. *Acta Haematol.* **2012**, *128*, 233–241. [[CrossRef](#)] [[PubMed](#)]
65. Martín, M.; de Paz, R.; Jiménez-Yuste, V.; Fernández Bello, I.; García Arias Salgado, E.; Alvarez, M.T.; Butta, N.V. Platelet apoptosis and agonist-mediated activation in myelodysplastic syndromes. *Thromb. Haemost.* **2013**, *109*, 909–919. [[PubMed](#)]
66. Brown, J.R. How I treat CLL patients with ibrutinib. *Blood* **2018**, *131*, 379–386. [[CrossRef](#)] [[PubMed](#)]
67. Abdool, A.; Donahue, A.C.; Wohlgemuth, J.G.; Yeh, C.H. Detection, analysis and clinical validation of chromosomal aberrations by multiplex ligation-dependent probe amplification in chronic leukemia. *PLoS ONE* **2010**, *5*, e15407. [[CrossRef](#)] [[PubMed](#)]
68. Hallek, M.; Shanafelt, T.D.; Eichhorst, B. Chronic lymphocytic leukaemia. *Lancet* **2018**, *391*, 1524–1537. [[CrossRef](#)]
69. Hallek, M.; Cheson, B.D.; Catovsky, D.; Caligaris-Cappio, F.; Dighiero, G.; Döhner, H.; Hillmen, P.; Keating, M.; Montserrat, E.; Chiorazzi, N.; et al. Guidelines for diagnosis, indications for treatment, response assessment and supportive management of chronic lymphocytic leukemia. *Blood* **2018**. [[CrossRef](#)] [[PubMed](#)]
70. Levade, M.; Severin, S.; Gratacap, M.P.; Ysebaert, L.; Payrastra, B. Targeting kinases in cancer therapies: Adverse effects on blood platelets. *Curr. Pharm. Des.* **2016**, *22*, 2315–2322. [[CrossRef](#)] [[PubMed](#)]
71. Bye, A.P.; Unsworth, A.J.; Desborough, M.J.; Hildyard, C.A.T.; Appleby, N.; Bruce, D.; Kriek, N.; Nock, S.H.; Sage, T.; Hughes, C.E.; et al. Severe platelet dysfunction in NHL patients receiving ibrutinib is absent in patients receiving acalabrutinib. *Blood Adv.* **2017**, *1*, 2610–2623. [[PubMed](#)]
72. Liu, J.; Fitzgerald, M.E.; Berndt, M.C.; Jackson, C.W.; Gartner, T.K. Bruton tyrosine kinase is essential for botrocetin/VWF-induced signaling and GPIb-dependent thrombus formation in vivo. *Blood* **2006**, *108*, 2596–2603. [[CrossRef](#)] [[PubMed](#)]
73. Busygina, K.; Jamasbi, J.; Seiler, T.; Deckmyn, H.; Weber, C.; Brandl, R.; Lorenz, R.; Siess, W. Oral Bruton tyrosine kinase inhibitors selectively block atherosclerotic plaque-triggered thrombus formation. *Blood* **2018**. [[CrossRef](#)] [[PubMed](#)]
74. Kamel, S.; Horton, L.; Ysebaert, L.; Levade, M.; Burbury, K.; Tan, S.; Cole-Sinclair, M.; Reynolds, J.; Filshie, R.; Schischka, S.; et al. Ibrutinib inhibits collagen-mediated but not ADP-mediated platelet aggregation. *Leukemia* **2015**, *29*, 783–787. [[CrossRef](#)] [[PubMed](#)]
75. Levade, M.; David, E.; Garcia, C.; Laurent, P.A.; Cadot, S.; Michallet, A.S.; Bordet, J.C.; Tam, C.; Sié, P.; Ysebaert, L.; et al. Ibrutinib treatment affects collagen and von Willebrand factor-dependent platelet functions. *Blood* **2014**, *124*, 3991–3995. [[CrossRef](#)] [[PubMed](#)]
76. Qiao, J.; Schoenwaelder, S.M.; Mason, K.D.; Tran, H.; Davis, A.K.; Kaplan, Z.S.; Jackson, S.P.; Kile, B.T.; Andrews, R.K.; Roberts, A.W.; et al. Low adhesion receptor levels on circulating platelets in patients with lymphoproliferative diseases before receiving Navitoclax (ABT-263). *Blood* **2013**, *121*, 1479–1481. [[CrossRef](#)] [[PubMed](#)]

77. Souers, A.J.; Levenson, J.D.; Boghaert, E.R.; Ackler, S.L.; Catron, N.D.; Chen, J.; Dayton, B.D.; Ding, H.; Enschede, S.H.; Fairbrother, W.J.; et al. ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. *Nat. Med.* **2013**, *19*, 202–208. [[CrossRef](#)] [[PubMed](#)]
78. Schoenwaelder, S.M.; Jarman, K.E.; Gardiner, E.E.; Hua, M.; Qiao, J.; White, M.J.; Josefsson, E.C.; Alwis, I.; Ono, A.; Willcox, A.; et al. Bcl-xL-inhibitory BH3 mimetics can induce a transient thrombocytopenia that undermines the hemostatic function of platelets. *Blood* **2011**, *118*, 1663–1674. [[CrossRef](#)] [[PubMed](#)]
79. Gardiner, E.E.; Al-Tamimi, M.; Mu, F.T.; Karunakaran, D.; Thom, J.Y.; Moroi, M.; Andrews, R.K.; Berndt, M.C.; Baker, R.I. Compromised ITAM-based platelet receptor function in a patient with immune thrombocytopenic purpura. *J. Thromb. Haemost.* **2008**, *6*, 1175–1182. [[CrossRef](#)] [[PubMed](#)]
80. Qiao, J.; Al Tamimi, M.; Baker, R.; Andrews, R.K.; Gardiner, E.E. The platelet Fc receptor, FcγRIIa. *Immunol. Rev.* **2015**, *268*, 241–252. [[CrossRef](#)] [[PubMed](#)]
81. Bresin, A.; D'Abundo, L.; Narducci, M.G.; Fiorenza, M.T.; Croce, C.M.; Negrini, M.; Russo, G. TCL1 transgenic mouse model as a tool for the study of therapeutic targets and microenvironment in human B-cell chronic lymphocytic leukemia. *Cell Death Dis.* **2016**, *7*, e2071. [[CrossRef](#)] [[PubMed](#)]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).