

# The upregulated expression of vascular endothelial growth factor in surgically treated patients with recurrent/radioresistant cervical cancer of the uterus

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**Abstract.** Vascular endothelial growth factor (VEGF) inhibitors have been utilized for the treatment against advanced or recurrent cervical carcinoma as a novel therapeutic modality. However, the expression level of VEGF in post-radiotherapy relapsed/persistent cervical cancer remains to be elucidated. The aim of the present study was to investigate the expression of VEGF and associated molecules using tumor samples from patients with post-radiotherapy relapsed/persistent cervical cancer. From a database of 826 patients who were treated at our institution between 2003 and 2015, eight patients with post-radiotherapy relapsed/persistent cervical cancer were identified, and 20 patients who underwent initial surgery alone were used as a control. Using samples from these patients, the expression levels of VEGF-A, VEGF receptor-1 (VEGFR-1) and hypoxia inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) were immunohistochemically categorized as negative or weakly, moderately, or strongly positive according to the size of the staining area, and intensity. In carcinoma cells, the expression levels of VEGF-A, VEGFR-1 and HIF-1 $\alpha$  were significantly higher in post-radiotherapy relapsed/persistent cervical cancer compared with control patients ( $P=0.0003$ ,  $0.0003$ , and  $0.0001$ , respectively). In stroma cells, similar tendencies with statistical significance were observed ( $P=0.0014$  and  $P<0.0001$ , respectively). In addition, the expression levels of VEGF-A and VEGFR-1 in carcinoma cells were significantly correlated with each other

( $P<0.0001$ ). A significantly higher expression of VEGF was identified in post-radiotherapy relapsed/persistent cervical cancer compared with typical specimens from cervical cancer. The findings provide a novel insight into the clinical treatment for recurrent/persistent cervical cancer using a VEGF antagonist.

## Introduction

Cervical cancer is one of the most common malignancies in females worldwide. Mortality rates associated with uterine cervical cancer have declined due to the widespread use of cancer screening for the prevention and early detection of cervical cancer (1). Moreover, since concurrent chemoradiotherapy (CCRT) has been established as a standard treatment, the prognosis of those patients has improved (2,3). However, about one third of patients experience recurrence within five years (4), with a median survival after recurrence of 15 months (5), and less than 5% of them survive for 5 years (6). Thus, the oncologic outcome is far from satisfactory. Especially, the prognosis of patients with recurrent disease within a previously irradiated field is unfavorable (7). In addition, earlier studies indicated that response rates to chemotherapy in those patients were poorer than that of those with out-of-field recurrence (8,9). Therefore, the oncologic outcome of patients with post-radiotherapy relapsed/persistent cervical cancer (PRRCC) is still poor. Recently, bevacizumab, a humanized monoclonal antibody targeting vascular endothelial growth factor A (VEGF-A), has been approved for this tumor, and moreover immunotherapy is under investigation (10).

VEGF-A is a multifunctional and an important molecule in endothelial signaling and angiogenesis. VEGF-A binds to its receptor VEGFR-1, and the downstream signaling is thought to be involved in cancer proliferation and invasion (11). There have been several reports showing that the overexpression of VEGF in cancer cells or serum is correlated with radioresistance and poor disease-free survival (12-16), and a meta-analysis suggested that high expressions of VEGF was significantly associated with poor survival outcome (17). Although VEGF inhibitors such as bevacizumab are widely used against several

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*Abbreviation:* PRRCC, post-radiotherapy relapsed/persistent cervical cancer

*Key words:* uterine cervical cancer, radioresistance, VEGF, VEGFR-1, HIF-1 $\alpha$

solid cancers, evidence regarding their efficacy against cervical cancer is not satisfactory, especially PRRCC. Particularly, to our best knowledge, there have been no reports on the expression level of VEGF in PRRCC based on the fact that surgical treatment is rarely performed as a salvage therapy for those patients (6). In sophisticated randomized clinical trials, the addition of a VEGF inhibitor such as bevacizumab to combination chemotherapy led to a significant improvement of the oncologic outcome of patients with recurrent, persistent, and highly metastatic cervical cancer (18,19). Accordingly, bevacizumab has been applied in actual clinical practice for this tumor. These results led us to hypothesize that the expression of VEGF may be upregulated in those patients.

Reviewing 826 clinical records of cervical cancer patients in our institute from 2003 to 2015, we identified eight patients with PRRCC who underwent debulking surgery, and evaluated the expression of VEGF immunohistochemically. In the present study, we further clarify the upregulation of VEGF in radioresistant cervical cancer by evaluating the expressions of VEGFR-1 and hypoxia inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ).

## Materials and methods

**Patients.** We retrospectively reviewed all the records of 826 patients with cervical cancer who were initially treated in our hospital from 2003 to 2015. Written informed consent was acquired from all patients. This study was approved by the Ethics Committee of our institute (Approval no. 2013-0078). Treatment strategies for each patient were determined by several gynecologic oncologists in our hospital depending on their age, performance status (PS), and spread of the disease. For example, as primary treatments, patients who were in the early stage and had a good PS were indicated for radical hysterectomy, and the other patients were treated with CCRT or radiotherapy alone. As treatments for recurrence, most patients were treated with chemotherapy, and only a few patients with localized disease were selected for surgical resection.

Ninety-seven patients had PRRCC, and 14 of them underwent surgery for a recurrent lesion. After excluding six patients because of lymph node metastasis or small residual tumors, eight patients with uterine or vaginal stump recurrence were investigated. Twenty patients who underwent radical surgery without neoadjuvant therapy were extracted as a control (Fig. 1).

**Immunohistochemical (IHC) staining and its evaluation.** Archival formalin-fixed, paraffin-embedded tumor tissue obtained at surgery was used in this study. Sections of 4- $\mu$ m thickness were prepared using a microtome. The sections were deparaffinized and rehydrated, subjected to antigen retrieval in 10 mM sodium citrate (pH 6.0) for 20 min at 95°C in a microwave, and treated with 0.3% hydrogen peroxide in methanol for 20 min after being washed with phosphate-buffered saline (PBS). Then, the sections were blocked with appropriate serum using Histofine SAB-PO(R) kit or Histofine SAB-PO(M) kit according to the manufacturer's protocol (Nichirei, Tokyo, Japan), and incubated with an appropriate first antibody diluted by PBS at 4°C overnight. After rinsing with PBS, the sections were incubated with an appropriate second antibody, and then peroxidase labeled streptavidin using the kit. Then, the sections

were rinsed with PBS and developed by the 3, 3'-diaminobenzidine (DAB) substrate-chromogen. After rinsing in water, the sections were incubated with hematoxylin, dehydrated, and mounted. Details about the reagents are presented in Table I.

Based on the IHC activity, a four-tiered semi-quantitative score was assigned according to the intensity and area of stained cells, as follows: For the evaluation of IHC expression, the staining intensity was scored as: 0, negative; 1, weak; 2, medium; or 3, strong. The percentage of the staining area was scored as 'focal' (1-10%), 'sporadic' (11-50%), and diffuse (>51%) relative to the total tumor area. Carcinoma cells and stroma were separately evaluated by two researchers, and the final score was decided according to Table II ('negative', 'weak', 'moderate', and 'strong', respectively).

All photographs were taken using Zeiss Axio Imager.A1 (Carl Zeiss, Tokyo, Japan).

**Statistics.** All statistical analyses were performed with EZR (Saitama Medical Center, Jichi Medical University, Saitama, Japan), which is a graphical user interface for R (The R Foundation for Statistical Computing, Vienna, Austria). More precisely, it is a modified version of R commander designed to add statistical functions frequently used in biostatistics. Differences between recurrent cancer and control patients were assessed by the Mann-Whitney U test and t-test, and the correlation of each expression was assessed by Spearman's correlation coefficient. Differences at  $P < 0.05$  were considered significant.

## Results

We first compared clinical backgrounds of eight patients with PRRCC and those of 20 patients with primary uterine cervical cancer. Distributions of the age, tumor size, and lymphovascular space invasion between the two groups were not significantly different. All patients had squamous cell carcinoma (SCC), and the serum SCC level at surgery was not significantly different (Table III). Detailed characteristics of PRRCC patients are presented in Table IV. All of the patients had previously received radiotherapy, and six of them had received CCRT. Salvage hysterectomy was performed for seven patients, and pelvic exenteration was performed for case 2, who had vaginal stump recurrence after vaginal total hysterectomy for carcinoma *in situ* (CIS).

Representative images of IHC are shown in Fig. 2. In both carcinoma and stroma cells, the expressions of VEGF-A were significantly higher in the PRRCC group than in controls [PRRCC vs. control:  $P = 0.0003$  (carcinoma) and  $P = 0.0014$  (stroma), respectively] (Fig. 2A and Table VA). Similarly, the expressions of VEGFR-1 were also significantly stronger [PRRCC vs. control:  $P = 0.0003$  (carcinoma) and  $P < 0.0001$  (stroma), respectively] (Fig. 2B and Table VB). Of note, the expressions of VEGF-A and VEGFR-1 in carcinoma cells were significantly correlated with each other (Spearman's correlation coefficient: 0.856;  $P < 0.0001$ ) (Fig. 3A). In addition, the correlation of the VEGF-A expression in carcinoma cells and the VEGFR-1 expression in stroma cells was moderate (Spearman's correlation coefficient: 0.484;  $P = 0.0090$ ) (Fig. 3B), and that of the VEGF-A expression in stroma cells and the VEGFR-1 expression in carcinoma

Table I. Details about antibodies and immunohistochemistry kits.

A, Primary antibodies				
Antibody name	Manufacturer	Product no.	Host	Dilution rate
Anti-VEGF antibody	Abcam	ab46154	Rabbit	1:100
Anti-VEGF receptor 1 antibody	Abcam	ab2350	Rabbit	1:100
Anti-HIF-1 $\alpha$ antibody	Abcam	ab1Rabbit	1:100	
B, Immunohistochemistry kits				
Kit name	Manufacturer	Product no.		
Histofine SAB-PO (R) kit	Nichirei	424032		
10% Normal goat serum				
Biotin labeled anti-rabbit IgG antibody				
Peroxidase labeled streptavidin				
Histofine SAB-PO (M) kit	Nichirei	424022		
10% Normal rabbit serum				
Biotin labeled anti-mouse IgG + IgA + IgM antibody				
Peroxidase labeled streptavidin				
Liquid DAB+ Substrate Chromogen system	Dako	K3468		

VEGF, vascular endothelial growth factor; HIF-1 $\alpha$ , hypoxia inducible factor-1 $\alpha$ .

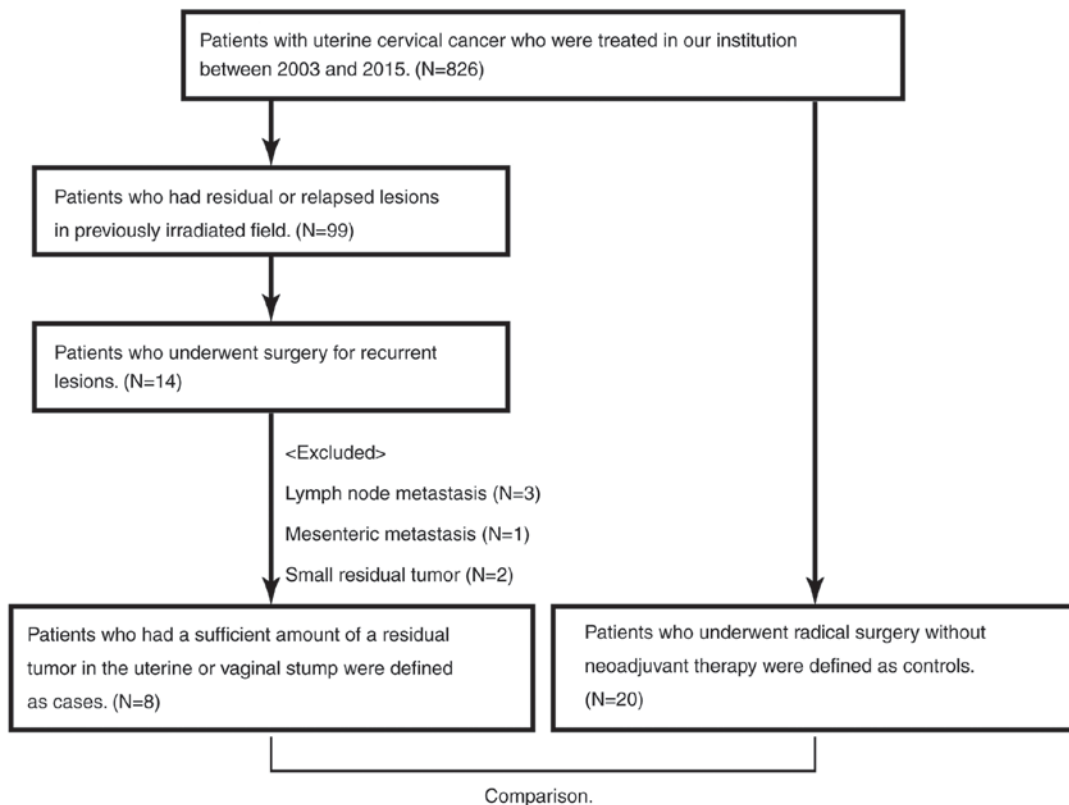


Figure 1. Flowchart of patient inclusion.

cells was weak (Spearman's correlation coefficient: 0.258;  $P=0.185$ ) (Fig. 3C).

The expression of HIF-1 $\alpha$  in carcinoma cells was also significantly stronger in the PRCC group than control group,

Table II. The evaluation of immunohistochemistry.

A, Cancer area				
Cancer area	Intensity			
	Negative	Weak	Medium	Strong
Focal	0	1	1	2
Sporadic	0	1	2	3
Diffuse	0	2	2	3
B, Stroma area				
Stroma area	Intensity			
	Negative	Weak	Medium	Strong
Focal	0	1	1	
Sporadic	0	1	2	
Diffuse	0	2	3	

although that in stroma cells was weak and showed no significant difference ( $P=0.343$ ) (Fig. 2C and Table VC). Moreover, the expression of HIF-1 $\alpha$  was significantly correlated with that of VEGF-A in carcinoma cells, but not in stroma cells [Spearman's correlation coefficient: 0.797;  $P<0.0001$  (carcinoma);  $P=0.343$  (stroma)] (Fig. 3D).

## Discussion

VEGF is an important factor for tumor angiogenesis, and there have been a number of reports evaluating the VEGF expression of primary surgery specimens or that of serum in uterine cervical cancer (12-16). However, to the best of our knowledge, there have been no reports concerning VEGF expression in PRRCC. In this study, we investigated the expression of VEGF and related molecules using tumor samples from patients with PRRCC. The expressions of both VEGF-A and VEGFR-1 were significantly higher in PRRCC sections than in controls. These results led us to hypothesize two possible mechanisms: 'natural selection' and 'evolution'. Intra-tumor genetic heterogeneity is also known in cervical cancer, and subpopulations of each tumor showed differential responses to chemoradiotherapy (20). In addition, patients with high VEGF expression in cancer tissue or serum were associated with a poor response to radiotherapy and poor survival (12-15). Therefore, the 'natural selection' hypothesis suggests that subpopulations of cervical cancer with high VEGF expression survive through chemoradiation, and then these selected subpopulations develop, leading to recurrence. On the other hand, in addition to its therapeutic effects, ionizing radiation is known to promote the malignant behaviors of surviving cancer cells. Radiation induced HIF-1 $\alpha$  and VEGF, and those factors were related to radioresistance (21,22). Therefore, the 'evolution' hypothesis suggests that some cervical cancer cells are evolutionarily induced to acquire VEGF expression by radiation while most of them are killed, and then cancer with acquired

Table III. Patients' characteristics.

Characteristic	Cases (N=8)		Controls (N=20)	P-value
	#1	#2		
Age				0.321
Median	43	44	39	
(Range)	(34-73)	(34-74)	(20-68)	
Stage				0.795
CIS	1		0	
Stage I	3		10	
Stage II	4		10	
Tumor size				0.100
<4 cm	5	8	18	
$\geq$ 4 cm	3	0	2	
LVSI				0.591
Yes		6	15	
No		1	5	
Unknown		1	0	
Nodal metastasis				0.576
Yes	3	1	10	
No	5	0	10	
Unknown		7		
Serum SCC level				0.660
<2.0 ng/ml	3	4	8	
$\geq$ 2.0 ng/ml	4	4	12	
Unknown	1			
Previous treatment				<0.0001
CCRT		5	0	
RT		3	0	

#1, at diagnosis of uterine cervical cancer; #2, at recurrent diagnosis; CIS, carcinoma *in situ*; LVSI, lymphovascular space invasion; SCC, squamous cell carcinoma; CCRT, concurrent chemoradiotherapy; RT, radiotherapy.

resistance develops, leading to recurrence. Regardless of the two independent hypotheses, if VEGF and its receptor are upregulated in PRRCC, VEGF-targeting therapy is expected as an effective therapeutic strategy for this tumor.

Hypoxia is an important cancer microenvironment, and most solid human cancers including cervical cancer are known to induce such an environment (23). It is possible that PRRCC tissue is exposed to hypoxic conditions by tissue fibrosis after radiotherapy. HIF-1 $\alpha$  has been reported to mediate essential homeostatic responses by activating the transcription of multiple genes including VEGF (24). Indeed, in the present study, we showed that the expression of HIF-1 $\alpha$  in carcinoma cells was also significantly higher in the PRRCC than control group consistent with our current findings. According to earlier studies, hypoxic conditions enhanced the radiation resistance dependent on HIF-1 $\alpha$  by elevating the expression of VEGF and inhibiting the expression of p53 (25). In addition, high-level expression of HIF-1 $\alpha$  is associated with treatment-resistance, and, conversely, the inhibition of HIF-1 $\alpha$  transactivation



Table IV. Characteristics and oncologic outcomes of patients with recurrence.

No.	Age <sup>a</sup>	TNM	Age <sup>b</sup>	PFS (months)	Previous treatments	OS (months)	Outcome
1	34	cT1b2N1M0	34	3	CCRT (PFx4 kur + WP 56.4 Gy, RALS 15 Gy)	20	DOD
2	35	pTisN0M0	38	16	VTH → CCRT (PF x5 kur + WP 50.4 Gy, RALS 13 Gy)	47	NED
3	37	cT2bN0M0	40	4	CCRT (PFx5 kur + WP 50.4 Gy, RALS 16 Gy) → TC x6 kur → CPT-11 x3 kur	44	NED
4	42	cT1b2N0M0	43	7	CCRT (PFx2 kur + WP 50.4 Gy, RALS 24 Gy)	13	DOD
5	44	cT2bN1M0	45	16	CCRT (PFx1 kur, CBDCA x1 kur, NDP x3 kur + WP 50.4 Gy, RALS 18 Gy)	41	NED
6	53	cT1bN0M0	54	5	RT (WP 50.4 Gy, RALS 24 Gy) → PF x2 kur → TP x4 kur	24	DOD
7	73	cT2bN0M0	74	5	RT (WP 50.4 Gy, RALS 13 Gy)	35	DOD
8	73	cT2aN0M0	74	10	RT (WP 50.4 Gy, RALS 9 Gy)	14	NED

<sup>a</sup>At diagnosis of uterine cervical cancer, <sup>b</sup>At recurrent diagnosis. PFS, progression free survival; OS, overall survival; VTH, vaginal total hysterectomy; CCRT, concurrent chemoradiotherapy; RT, radiotherapy; WP, whole pelvis; RALS, remote after loading system; PF, cisplatin and 5-FU; TC, paclitaxel and carboplatin; CPT-11, irinotecan; CBDCA, carboplatin; NDP, nedaplatin; TP, paclitaxel and cisplatin; DOD, died of disease; NED, no evidence of disease.

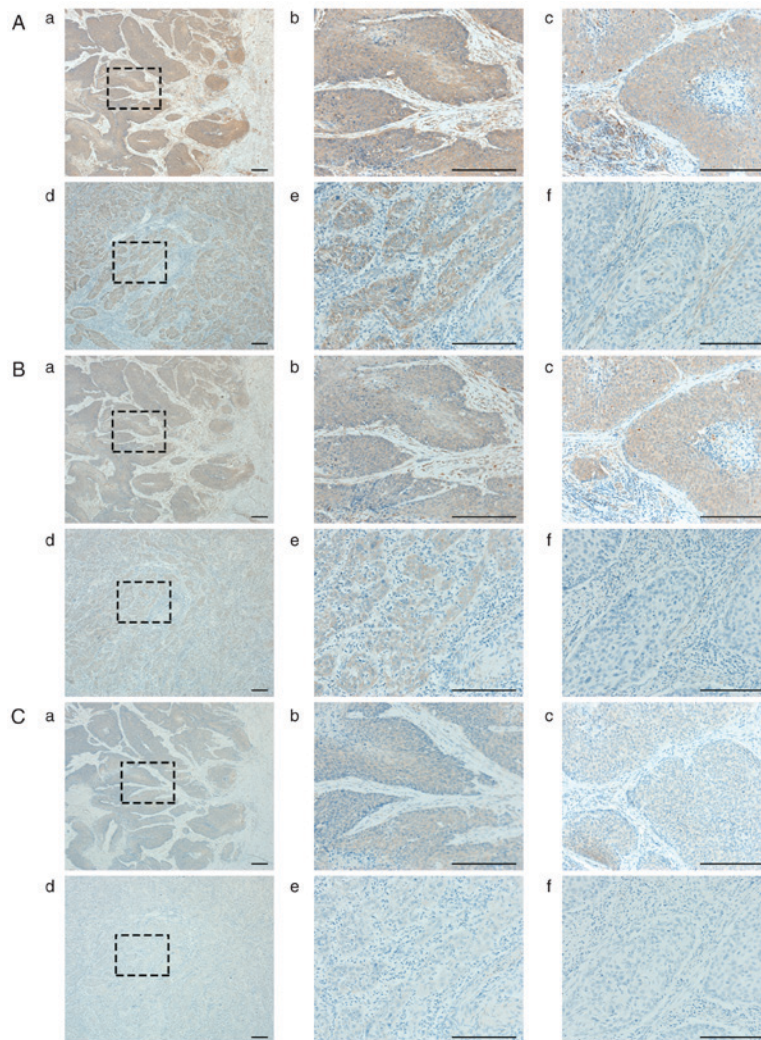


Figure 2. Representative images of immunohistochemistry. (A, B, and C) Representative images of VEGF-A, VEGFR-1, and HIF-1 $\alpha$ , respectively. a and b show those of case 3, c shows that of case 2, d and e show those of control 1, and f shows that of control 2. All scale bars, 200  $\mu$ m. A-b shows strong expression in cancer (cancer-3), A-c: cancer-3, A-e: cancer-2, and A-f: cancer-0. A-b shows strong expression in stroma (stroma-3), A-c: stroma-3, A-e: stroma-0, and A-f: stroma-1. B-b: cancer-3, B-c: cancer-3, B-e: cancer-2, and A-f: cancer-0. B-b: stroma-3, B-c: stroma-2, B-e: stroma-0, and A-f: stroma-1. C-b: cancer-2, C-c: cancer-2, C-e: cancer-1, and A-f: cancer-0. C-b: stroma-1, C-c: stroma-1, C-e: stroma-0, and A-f: stroma-0.

Table V. The expressions of VEGF-A, VEGFR-1, and HIF-1 $\alpha$ .

A, VEGF-A expression					
VEGF-A	Negative	Weak	Moderate	Strong	P-value
<b>Cancer</b>					
Cases			2	6	0.0003
Controls	6	8	5	1	
<b>Stroma</b>					
Cases	2	1		5	0.0014
Controls	16	4			
B, VEGFR-1 expression					
VEGFR-1	Negative	Weak	Moderate	Strong	P-value
<b>Cancer</b>					
Cases			2	6	0.0003
Controls	4	9	6	1	
<b>Stroma</b>					
Cases			5	3	<0.0001
Controls	13	7			
C, HIF-1 $\alpha$ expression					
HIF-1 $\alpha$	Negative	Weak	Moderate	Strong	P-value
<b>Cancer</b>					
Cases	1	3	4		0.0001
Controls	18	1	1		
<b>Stroma</b>					
Cases	4	4			0.343
Controls	14	6			

enhances radiotherapy responses (23,26). Burri *et al* reported that multivariate analyses revealed HIF-1 $\alpha$  expression to be an independent factor for overall survival based on an immunohistochemical analysis of 78 patients with uterine cervix carcinoma treated with external beam radiotherapy (27). This evidence prompted us to hypothesize that HIF-1 $\alpha$  plays a crucial role in VEGF upregulation and the treatment refractoriness of PRRCC.

In the current study, high-level expressions of VEGF-A and VEGFR-1 were observed in the stroma as well as in carcinoma cells. Cancer-associated fibroblasts (CAFs) are major components of the tumor stroma and involved in tumor progression. A previous report demonstrated the effects to protect against radiation of CAF-cancer cell crosstalk through multiple growth factors including VEGF *in vitro* (28). In order to inhibit VEGF-VEGFR interactions between carcinoma and stroma cells, VEGF inhibitors such as bevacizumab have been widely used. In cervical cancer, VEGF inhibitors also showed clinical benefits for patients with advanced, persistent, or recurrent lesions (18,19,29). Especially, according to Tewari's sub-group analysis, bevacizumab was more favorable

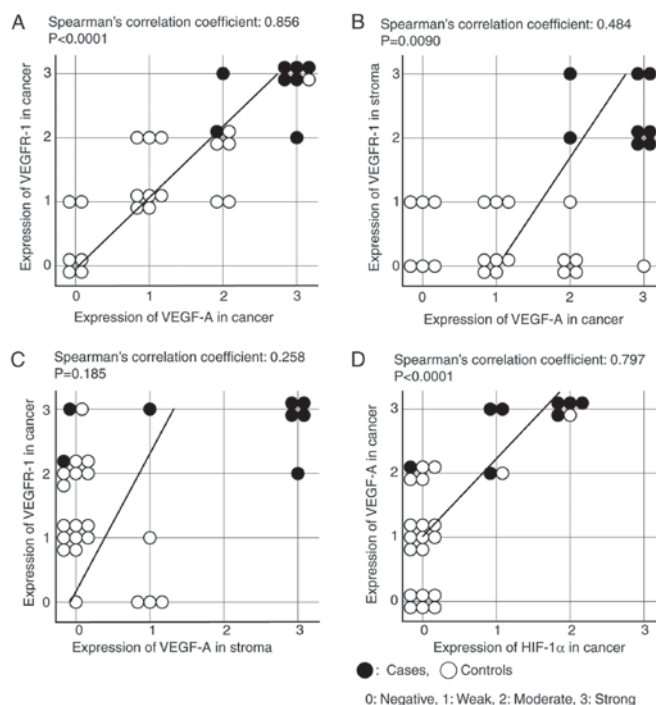


Figure 3. The correlations among the expressions of VEGF-A, VEGFR-1, and HIF-1 $\alpha$ . (A) The correlation between the expression of VEGF-A in cancer and that of VEGFR-1 in cancer. (B) The correlation between the expression of VEGF-A in cancer and that of VEGFR-1 in the stroma. (C) The correlation between the expression of VEGF-A in the stroma and that of VEGFR-1 in cancer. (D) The correlation between the expression of HIF-1 $\alpha$  in cancer and that of VEGF-A in cancer.

in patients with recurrent or persistent lesions than those with advanced lesions, and also in those who previously received chemoradiotherapy (18). These results suggest that recurrent or persistent cancer after chemoradiotherapy expressed VEGF-A and VEGFR-1 more strongly than advanced cancer, being consistent with our results.

The main limitation of this study was the fact that only eight patients with PRRCC were available despite the enrollment of over 800 patients with cervical cancer. This limited patient number is consistent with the actual clinical situation whereby the selection of surgical treatment for PRRCC is extremely rare. Second, we did not evaluate the association between the VEGF expression and efficacy of VEGF inhibitors in patients with PRRCC. Moreover, we could not directly compare the immunohistochemical expressions between pre- and post-treatment sample sets in the same patient. Actually, it was difficult to obtain enough specimens from patients with primary CCRT before treatment. As a result, we used specimens of primary surgery as a control. An additional large-scale study to confirm our current findings is desirable by accumulating more patients with PRRCC from multiple institutions. Therefore, in the present study, we could not verify the direct effect of the radiation-induced expression of VEGF in tumor tissues of patients with PRRCC. We would like to verify the radiation-induced upregulation of VEGF expression effects *in vitro* and using an animal model in a future study.

In conclusion, the expressions of VEGF-A and VEGFR-1 were significantly upregulated in PRRCC. These results are important and valuable because there has been no evidence

of VEGF expression in PRRCC. For further evaluation, a large-scale study of VEGF in advanced, residual, and recurrent cervical cancer is desired and the efficacy of VEGF inhibitors must be investigated. The prognoses of these patients are expected to improve in the future. We believe that our results will help clarify the efficacy of bevacizumab.

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