CASE REPORT Open Access



A de novo *KMT2D* mutation in a girl with Kabuki syndrome associated with endocrine symptoms: a case report

Jung-Eun Moon, Su-Jeong Lee and Cheol Woo Ko*

Abstract

Background: Kabuki syndrome is characterized by distinctive facial features and varying degrees of growth retardation. It leads to malformations in skeletal, urogenital and cardiac structures; moreover, endocrine conditions such as premature thelarche, precocious puberty, growth hormone deficiency, diabetes insipidus, thyroid dysfunction and obesity have been reported. Kabuki syndrome is caused by a heterozygous mutation in the *KMT2D* or *KDM6A* genes.

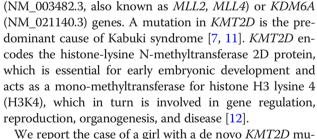
Case presentation: An 11-year-old girl with the typical facial features of Kabuki syndrome visited our hospital due to her short stature. She was found to have the de novo heterozygous mutation of c.8200C > T, p(Arg2734*) in exon 32 of the *KMT2D* gene and was diagnosed with Kabuki syndrome. The patient also exhibited endocrine abnormalities such as a constitutional delay of puberty, transiently congenial hypothyroidism, obesity and growth hormone deficiency.

Conclusions: This is a case of a mutation in the *KMT2D* gene in a girl with Kabuki syndrome who presented with endocrine symptoms (constitutional delay of puberty, hypothyroidism, obesity and growth hormone deficiency).

Keywords: Kabuki syndrome, KMT2D gene, Growth hormone deficiency, Constitutional delay of puberty

Background

Kabuki syndrome (OMIM#147920), which was first reported in Japan in 1981, is a rare disease with an incidence of 1:32,000 in Japan [1, 2]. At least 589 patients with Kabuki syndrome due to KDM6A mutations have been reported [3]. Kabuki syndrome is characterized by distinctive facial features, including long palpebral fissures with eversion of the lateral third of the lower eyelids, arched eyebrows and a short nasal columella with a broad and depressed nasal tip; it also manifests as varying degrees of growth retardation [4]. Kabuki syndrome is a multiple malformation syndrome that is accompanied by skeletal conditions such as cleft palate and cranial abnormalities (including coronal and metopic synostosis), as well as urogenital and cardiac abnormalities [5–7]. Patients with this syndrome also exhibit endocrine system-related conditions such as premature thelarche, precocious puberty, diabetes insipidus, thyroid dysfunction, obesity and growth hormone (GH) deficiency [8-10].



Kabuki syndrome is caused by mutations in the KMT2D

We report the case of a girl with a de novo *KMT2D* mutation who had the typical Kabuki syndrome features as well as endocrine-related symptoms, including a constitutional delay of puberty (CDP), obesity and GH deficiency.

Case presentation

An 11-year, 7-month-old girl visited the Kyungpook National Children's Hospital because of a short stature. The patient had been born via vaginal delivery at a gestational age of 38 weeks; she weighed 2.6 kg at birth, and there were no perinatal problems. However, she had an incomplete cleft palate and craniosynostosis at birth. Hearing loss in both ears was detected at the age of 1 month by brainstem-evoked response audiometry during an



^{*} Correspondence: cwko@knu.ac.kr; https://orcid.org/0000-0002-0643-7233 Department of Pediatrics, Kyungpook National University School of Medicine, Kyungpook National University Children's Hospital, 807, Hoguk-ro, Buk-gu, Daegu 41404, Republic of Korea

initial screening test. An 'inborn errors of metabolism' workup performed after birth revealed congenital hypothyroidism, for which the patient received levothyroxine at the local hospital until she was 3 years old; normal thyroid function was confirmed after discontinuing the medication.

When she was 1 year old, she was transferred to our hospital because of a developmental delay and for a repair of her incomplete cleft palate and craniosynostosis. Brain computed tomography revealed an auditory defect, and she began wearing hearing aids, as prescribed by an otolaryngologist. The patient underwent skull reconstruction and incomplete cleft palate repair at the Departments of Plastic Surgery and Neurosurgery, respectively. She was scheduled for follow-up at the Department of Pediatric Neurology due to her developmental language delay, but she was lost to follow-up.

When the patient re-visited our hospital at the age of 11 years and 7 months, she had a height of 124.8 cm (standard deviation score [SDS]: – 3.6), weight of 46 kg (SDS: 0.65 kg), and body mass index (BMI) of 29.53 kg/m² (Z score: 1.89). She had distinctive facial features, including abnormally long openings between the eyelids, arch-shaped eyebrows, a thin upper lip, and large ears (Fig. 1). Moreover, she showed postnatal growth retardation and skeletal anomalies, including an incomplete cleft palate, craniosynostosis, and brachydactyly (Fig. 1). These dysmorphic features and her developmental delay



Fig. 1 The patient shows the typical facial features of Kabuki syndrome, such as long palpebral fissures, everted lower eyelids, arched eyebrows, a broad nasal root, thin upper and full lower lips, and brachydactyly in both hands and feet

were considered suggestive of Kabuki syndrome. Her karyotype was 46,XX. Her bone age was 8 years and 10 months, which was considered delayed according to the Greulich and Pyle atlas [13]. Based on these characteristic clinical features, mutational analyses for the *KMT2D* and *KDM6A* genes were performed.

The CARE guidelines were followed in reporting this case, and the molecular analyses were approved by the Institutional Review Board of the Kyungpook National University Chilgok Hospital after obtaining informed consent from the patient's parents. Genomic DNA was extracted from peripheral blood, and library preparations were performed with the TruSight One Sequencing Panel (Illumina, Inc., San Diego, CA, USA), which enriches a 12 Mb region spanning 4813 genes of clinical relevance. Large-scale parallel sequencing was performed with the Illumina NextSeq platform. Sequence reads were aligned with the hg19 reference sequence using the Burrow-Wheeler Aligner (version 0.7.12, MEM algorithm). Duplicate reads were removed using Picard Tools (version 1.96). Local realignment and base quality recalibration were performed with the Genome Analysis Toolkit (GATK, version 3.5), and variant calling was performed using the GATK HaplotypeCaller. Variants were annotated with the Variant Effect Predictor and Anno-Var. Common variants with minor allele frequencies (≥1%) were filtered out using public databases (1000 Genomes Project, Exome Variant Server, and Exome Aggregation Consortium). Population-specific common variants were further filtered using the Korean Reference Genome Database [14]. The patient had the nonsense mutation of c.8200C > T, p(Arg2734*) in exon 32 in the KMT2D gene. As neither parent had this mutation, as determined by Sanger sequencing, the patient had a de novo heterozygous mutation in the *KMT2D* gene (Additional file 1: Figure S1).

We then performed additional examinations to detect other anomalies associated with Kabuki syndrome. Two-dimensional echocardiography, abdominal ultrasonography, and brain magnetic resonance imaging that had been performed at 1 year of age, showed no abnormalities. Ophthalmological findings were also normal.

The patient exhibited several endocrine problems (transiently congenital hypothyroidism, CDP, obesity and GH deficiency); additional endocrine tests were performed upon admission to our hospital (at the age of 11 years and 7 months).

The patient had transiently congenital hypothyroidism. When she was 7 days old, her free thyroxine (T4) was 1.09 ng/dL (reference range [RR], 0.8–2.0 ng/dL), triiodothyronine (T3) was 1.3 ng/mL (RR, 0.2–2.0 ng/mL), and thyroid-stimulating hormone (TSH) was 22.8 mIU/L (RR, 0.7–13.1 mIU/L). She received levothyroxine until she was 3 years old, and normal thyroid function was confirmed after discontinuing the medication. Upon admission to our hospital, she was 11 years and 7 month old her free

T4 was 1.6 ng/dL (RR, 0.8–1.8 ng/dL), T3 was 0.8 ng/mL (RR, 0.6–1.9 ng/mL), and TSH was 3.5 mIU/L (RR, 0.3–4.0 mIU/L), indicating normal thyroid function.

Regarding puberty, the patient showed Tanner stage I–II of breast development; her basal luteinizing hormone level was < 0.07 mIU/mL (RR, 5–20 mIU/mL), basal follicle-stimulating hormone level was 6.19 mIU/mL (RR, 0.3–10 mIU/mL), and estradiol level was < 11.8 pg/mL (RR, 0–16 pg/mL), suggesting a CDP.

She was obese and had a buffalo hump. Her cortisol levels while awake at 8 AM, 5 PM, and midnight were 6.93 μg/dL (RR, 3–22 μg/dL), 12.46 μg/dL (RR, 3–22 μg/ dL), and 14.87 μg/dL (RR, <7.5 μg/dL), respectively. Her corresponding adrenocorticotropic hormone levels were 34.2 pg/mL, 40 pg/mL, and 42.8 pg/mL, respectively (RR for all, 0–60 pg/mL). Her urine-free cortisol was $28.9 \mu g/dL$ (RR, 4.3-176 µg/dL). Even though the midnight cortisol level was elevated, a low-dose dexamethasone suppression test showed a cortisol level of 0.68 μ g/dL (RR, < 1.8 μ g/dL). A lipid profile showed a total cholesterol level of 150 mg/ dL (RR, 125-200 mg/dL), triglyceride level of 66 mg/dL (RR, < 200 mg/dL), and LDL-cholesterol level of 53 mg/dL (RR, < 130 mg/dL). Her aspartate aminotransferase and alanine aminotransferase levels were 24 U/L (RR, < 97 U/L) and 17 U/L (RR, < 41 U/L), respectively.

A GH stimulation test (spanning 2 days) was performed because of her short stature and delayed bone age. On the first day, the patient was administered levodopa (500 mg); the samples for the GH assessment were acquired at 0, 30, 60, 90, and 120 min. On the next day, the patient fasted for 8 h in the morning and was intravenously administered 0.1 U/kg of rapid-acting insulin diluted in 5 mL of normal saline for over 1 min, after which the samples for GH assessment were acquired at 0, 15, 30, 60, 90, and 120 min. The maximum GH concentrations in both tests were below 5 μ g/L (RR, > 7 μ g/ L); therefore, the patient was diagnosed with GH deficiency. Her insulin-like growth factor 1 (IGF-1) and insulin-like growth factor-binding protein 3 (IGFBP-3) levels were 130.5 ng/mL (RR, 99-537 ng/mL, SDS: -1.67) and 3760 ng/mL (RR, 2400-8400 ng/mL, SDS: 0.91), respectively. The patient started GH replacement therapy (Eutropin 0.03 mg/kg/day) when she was 11 years and 7 months old and is currently scheduled for regular follow-ups for growth and puberty changes.

Discussion and conclusions

In the present case, a de novo nonsense mutation in the *KMT2D* gene was confirmed in a girl who had the facial features distinctive of Kabuki syndrome and several endocrine conditions such as transiently congenital hypothyroidism, CDP, obesity, and GH deficiency.

Aimée et al. first reported patients with Kabuki syndrome due to the nonsense mutation c.8200C > T, p

(Arg2734 *) in exon 32 of the *KMT2D* gene in 2011 [15]. A mutation in the same position was reported by Dina et al. in patients with Kabuki syndrome and a short stature [16]. In our case, multiple endocrine conditions, including a short stature, were also observed.

KMT2D (MLL2) plays a critical role in regulating HOX genes and embryonic growth; it also interacts with nuclear receptors and is involved in the hormone-dependent regulation of genes that play crucial roles in reproduction and organogenesis [17]. In particular, HOXC6 is critical for mammary gland development and milk production and plays a critical role in breast carcinoma development. Moreover, in an in vivo study, HOXC6 was reported to regulate the expression of the bone morphogenic protein 7, fibroblast growth factor receptor 2, and IGFBP3 [18, 19]. Ansari et al. reported that KMT2D (MLL2) knockdown suppressed estradiol-medicated-HOXC6 regulation [20]. Thus, it can be speculated that the skull anomalies, cleft palate, and short stature that are characteristic of patients with a KMT2D mutation are the result of FGFR2 and IGFBP3 expression via downregulation of the HOXC6 gene. Approximately 41% of patients with Kabuki syndrome show premature thelarche, and some experience precocious puberty; these manifestations could be secondary to a low hypothalamic sensitivity to the suppressive effects of sex hormones on gonadotropin secretion [21, 22]. Our patient exhibited a CDP; this may be attributable to HOXC6 downregulation because of the mutation in KMT2D, which hinders mammary gland development.

More than 50% of patients with Kabuki syndrome are overweight or obese, like our patient, during childhood or adolescence, although the exact cause is unknown [23]. The ER is associated with human disease, particularly with obesity [24]. In an in vivo study by Ohlsson et al., *ER* knockout mice had an obvious increase in total body fat [25]. Because KMT2D (MLL2) is involved in ER-dependent gene regulation, it is feasible that a mutation in *KMT2D* may be linked to obesity.

Patients with Kabuki syndrome also show postnatal growth retardation, but the cause is unclear. Schott et al. previously found that 27.8% of patients with Kabuki syndrome who were confirmed to have a *KMT2D* mutation had GH deficiency [26]. Considering that the prevalence of GH deficiency is only 1% in the general population, it is evidently very high in patients with Kabuki syndrome. The same group later showed that GH treatment produced a significant increase in height SDS after 1 year [27]. Our patient is currently receiving GH therapy because of her GH deficiency, and she is scheduled for follow-up visits to assess her growth progress.

In conclusion, a Korean girl was diagnosed with Kabuki syndrome caused by the de novo nonsense mutation of c.8200C > T, p(Arg2734*) in exon 32 in the *KMT2D* gene. The patient presented with the typical Kabuki syndrome

phenotypes and several endocrine symptoms (CDP, obesity, transiently congenital hypothyroidism, and GH deficiency). Further molecular studies of the *KMT2D* gene and *HOXC6*-mediated ER regulation are required to better understand Kabuki syndrome with multiple endocrine disorders.

Additional file

Additional file 1: Figure S1. Sanger sequencing confirmed a heterozygous mutation in *KMT2D*. The de novo heterozygous mutation c.8200C > T, p(Arg2734*) in exon 32 in *KMT2D* was identified by targeted exome sequencing and confirmed by Sanger sequencing analysis. (PDF 73 kb)

Abbreviations

BMI: Body mass index; CDP: Constitutional delay of puberty; ER: Estrogen receptor; GH: Growth hormone; H3K4: Histone H3 lysine 4; IGF-1: Insulin-like growth factor-1; IGFBP-3: Insulin-like growth factor-binding protein 3; KMT2D: Histone-lysine N-methyltransferase 2D; MLL: Mixed lineage leukemia; RR: Reference range; SDS: Standard deviation score; T3: Triiodothyronine; T4: Thyroxine; T5H: Thyroid-stimulating hormone

Availability of data and materials

Data from this study that do not pertain to identifiable patient information are freely available and provided as supplemental material and/or can be obtained by contacting the corresponding author.

Authors' contributions

JEM was responsible for the acquisition of the clinical information and the writing as well as reviewing of the manuscript and figures. She was also responsible for the collection of the clinical data and arranged the genetic studies for the patient and her family. SJL was the study coordinator and was involved in revising the manuscript. JEM, SJL, and CWK analyzed and interpreted the data and drafted the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

This study was approved by the Institutional Review Board of the Kyungpook National University Chilgok Hospital. Written informed consent for the molecular studies was obtained from the patient's parents. A copy of the consent is available for review by the editor of this journal.

Consent for publication

The patient's parents consented to the publication of the case and any accompanying images in writing.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 16 December 2017 Accepted: 15 May 2018 Published online: 18 June 2018

References

- Niikawa N, Matsuura N, Fukushima Y, Ohsawa T, Kajii T. Kabuki make-up syndrome: a syndrome of mental retardation, unusual facies, large and protruding ears, and postnatal growth deficiency. J Pediatr. 1981;99:565–9.
- Niikawa N, Kuroki Y, Kajii T, Matsuura N, Ishikiriyama S, Tonoki H, et al. Kabuki make-up (Niikawa Kuroki) syndrome: A study of 62 patients. Am J Med Genet. 1988;31:565–89.
- Chengqi X, Chun W, Yachen W, Jingyuan Z, Liang W, Liang W, et al. Identification of novel KMT2D mutations in two Chinese children with kabuki syndrome: a case report and systematic literature review. BMC Med Genet. 2018;19:31.

- Kawame H, Hannibal MC, Hudgins L, Pagon RA. Phenotypic spectrum and management issues in kabuki syndrome. J Pediatr. 1999;134:480–5.
- Ewart-Toland A, Enns GM, Cox VA, Mohan GC, Rosenthal P, Golabi M. Severe congenital anomalies requiring transplantation in children with kabuki syndrome. Am J Med Genet. 1998;80:362–7.
- Matsumoto N, Niikawa N. Kabuki make-up syndrome: a review. Am J Med Genet C Semin Med Genet. 2003;117C:57–65.
- Bögershausen N, Wollnik B. Unmasking Kabuki syndrome. Clin Genet. 2013; 83:201–11.
- Devriendt K, Lemli L, Craen M, de Zegher F. Growth hormone deficiency and premature thelarche in a female infant with kabuki makeup syndrome. Horm Res. 1995;43:303–6.
- Bereket A, Turan S, Alper G, Comu S, Alpay H, Akalin F. Two patients with kabuki syndrome presenting with endocrine problems. J Pediatr Endocrinol Metab. 2001;14:215–20.
- Tawa R, Kaino Y, Ito T, Goto Y, Kida K, Matsuda H. A case of kabuki make-up syndrome with central diabetes insipidus and growth hormone neurosecretory dysfunction. Acta Paediatr Jpn. 1993;36:412–5.
- Ng SB, Bigham AW, Buckingham KJ, Hannibal MC, McMillin MJ, Gildersleeve HI, et al. Exome sequencing identifies MLL2 mutations as a cause of kabuki syndrome. Nature Genet. 2010;42:790–3.
- 12. Froimchuk E, Jang Y, Ge K. Histone H3 lysine 4 methyltransferase KMT2D. Gene. 2017;627:337–42.
- Greulich WW, Pyle SI. Radiographic atlas of skeletal development of the hand and wrist. 2nd ed. Stanford: Stanford university press; 1959.
- Korean Reference Genome Database. http://152.99.75.168/KRGDB. Accessed 17 Apr 2018.
- Aimée DCP, Alexander PAS, Marinus JB, Demis T, Crool PV, Yvonne D, et al. MLL2 mutation spectrum in 45 patients with kabuki syndrome. Hum Mutat. 2011;32:F2018–25.
- Dina AS, Marinus JB, Willem JMG, Koenraad D, Luc JIZ, Constance TRMS. Growth pattern in kabuki syndrome with a KMT2D mutation. Am J Med Genet. 2016;170A:3172–9.
- Ansari KI, Mandal SS. Mixed lineage leukemia: roles in gene expression. hormone signaling and mRNA processing FEBS J. 2010;277:1790–804.
- Castronovo V, Kusaka M, Chariot A, Gielen J, Sobel M. Homeobox genes: potential candidates for the transcriptional control of the transformed and invasive phenotype. Biochem Pharmacol. 1994;47:137–43.
- Garcia-Gasca A, Spyropoulos DD. Differential mammary morphogenesis along the anteroposterior axis in Hoxc6 gene targeted mice. Dev Dyn. 2000; 219:261–76.
- Ansari KI, Hussain I, Shrestha B, Kasiri S, Mandal SS. HOXC6 is transcriptionally regulated via coordination of MLL histone methylase and estrogen receptor in an estrogen environment. J Mol Biol. 2011;411:334–49.
- 21. Banka S, Veeramachaneni R, Reardon W, Howard E, Bunstone S, Ragge N, et al. How genetically heterogeneous is kabuki syndrome?: MLL2 testing in 116 patients, review and analyses of mutation and phenotypic spectrum. Eur J Hum Genet. 2012;20:381–8.
- 22. Kuroki Y, Katsumata N, Eguchi T, Fukushima Y, Suwa S, Kajii T. Precocious puberty in kabuki makeup syndrome. J Pediatr. 1987;110:750–2.
- White SM, Thompson EM, Kidd A, Savarirayan R, Turner A, Amor D, et al. Growth, behavior, and clinical findings in 27 patients with kabuki (Niikawa–Kuroki) syndrome. Am J Med Genet A. 2004;127:118–27.
- Deroo BJ, Korach KS. Estrogen receptors and human disease. J Clin Invest. 2006;116:561–70.
- Ohlsson C, Hellberg N, Parini P, Vidal O, Bohlooly M, Rudling M, et al. Obesity and disturbed lipoprotein profile in estrogen receptor-α-deficient male mice. Biochem Biophys Res Commun. 2000;278:640–5.
- Schott DA, Gerver WJ, Stumpel CT. Growth hormone stimulation tests in children with kabuki syndrome. Horm Res Paediatr. 2016;86:319–24.
- Schott DA, Gerver WJ, Stumpel CTRM. Growth hormone therapy in children with kabuki syndrome: 1-year treatment results. Horm Res Paediatr. 2017;88:258–64.