






Complete Genome Sequence of *Bacillus velezensis* LABIM40, an Effective Antagonist of Fungal Plant Pathogens

Julia Pezarini Baptista,^a Paula Pinheiro Sanches,^a Gustavo Manoel Teixeira,^a Alexandre Tadachi Morey,^a Eliandro Reis Tavares,^a Sueli Fumie Yamada-Ogatta,^a Sérgio Paulo Dejato da Rocha,^a  Mariangela Hungria,^b Renan Augusto Ribeiro,^b Maria Isabel Balbi-Peña,^c Roberta Torres Chideroli,^d  Ulisses de Padua Pereira,^d  Admilton Gonçalves de Oliveira^{a,e}

^aDepartment of Microbiology, Universidade Estadual de Londrina (UEL), Londrina, Paraná, Brazil

^bEmbrapa Soja, Soil Biotechnology Laboratory, Londrina, Paraná, Brazil

^cDepartment of Agronomy, UEL, Londrina, Paraná, Brazil

^dDepartment of Preventive Veterinary Medicine, UEL, Londrina, Paraná, Brazil

^eLaboratory of Electron Microscopy and Microanalysis, UEL, Londrina, Paraná, Brazil

ABSTRACT *Bacillus velezensis* strain LABIM40 holds high potential for biological control of plant pathogens. Its complete genome contains one chromosome of 3,972,310 bp with 3,777 DNA coding sequences and displays 33 gene clusters potentially involved in the suppression of fungal pathogens.

The increasing demand for safe and sustainable food supplies requires an efficient control of the major plant diseases. Current management practices are largely based on the application of synthetic pesticides, but their massive use has caused serious environmental impact and human and animal health concerns. Therefore, there is need for new environmentally friendly technologies and products to replace partly or fully chemical-based pesticides, contributing to safer crop disease control (1).

The use of plant pathogen antagonists for biological control has increased significantly, particularly in the last decade. Aerobic endospore-forming bacteria, such as *Bacillus* spp., are present in soil microbial communities and are known to produce structurally diverse metabolites with various biological effects, including antimicrobial activity against plant pathogens and other plant growth-promoting properties (2). The metabolites produced by *Bacillus* spp. include aminoglycosides, polyketides, and several small ribosomally and nonribosomally synthesized peptides, such as bacteriocins, oligopeptides, and lipopeptides, all of which have important roles in suppressing fungal diseases (3–5).

In this study, we report the complete genome sequence of *Bacillus velezensis* LABIM40, isolated as an antagonist contaminant of *Fusarium oxysporum* *in vitro* in the city of Londrina, Brazil; the strain was deposited at the Microbial Collection of the Microbial Biotechnology Laboratory, Universidade Estadual de Londrina. The cell-free supernatant of the bacterium strongly inhibits the growth of the important fungal plant pathogens *Sclerotinia sclerotiorum*, *Rhizoctonia solani*, *Botrytis cinerea*, and *Macrophomina phaseolina* (6), confirming its high biotechnological potential for biological control.

The Genra Puregene genomic DNA kit (Qiagen, Brazil) was used for genomic DNA extraction. The *B. velezensis* LABIM40 genome was sequenced on the MiSeq platform, using a MiSeq version 3 reagent kit (600-cycle, Illumina, Brazil) at Embrapa Soja, Londrina. After we sequenced them, the reads were subjected to trimming and filtering using CLC Genomics Workbench version 10.0. Reads with an average Phred quality value of less than 30 and with one or more ambiguities were removed; reads smaller than 50 bp, as well as the last 10 nucleotides of the 3' end of each read, were also

Received 25 May 2018 Accepted 25 May 2018 Published 21 June 2018

Citation Baptista JP, Sanches PP, Teixeira GM, Morey AT, Tavares ER, Yamada-Ogatta SF, da Rocha SPD, Hungria M, Ribeiro RA, Balbi-Peña MI, Chideroli RT, Pereira UDP, de Oliveira AG. 2018. Complete genome sequence of *Bacillus velezensis* LABIM40, an effective antagonist of fungal plant pathogens. *Genome Announc* 6:e00595-18. <https://doi.org/10.1128/genomeA.00595-18>.

Copyright © 2018 Baptista et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Admilton Gonçalves de Oliveira, admilton@uel.br.

removed. Sequencing allowed a genome coverage of 212-fold, assembled into 19 contigs with CONTIGuator software (7). The genome was compared with other genomes of the same species, and the gaps were removed with recursive rounds of short reads mapped against the scaffold (8). The annotation was created using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP). The genome was estimated at 3,972,310 bp, with a G+C content of 46.5%, and the average nucleotide identity with its closest strain, *B. velezensis* S141 (9), was 99.1%. The genome of LABIM40 harbors 3,777 DNA coding sequences, 7 rRNA operons, 75 tRNAs, and 105 pseudogenes. Thirty-three putative gene clusters responsible for secondary metabolite biosynthesis were identified by using antiSMASH version 4.1.0 (10). Among them, we can highlight thiopeptides, polyketide synthase antibiotics, nonribosomal peptide synthetase antibiotics, bacteriocins, and terpenes, which may be responsible for the antifungal activity. The genome of *B. velezensis* LABIM40 may help to explore the metabolic pathways related to its antimicrobial activity, emphasizing its biotechnological potential.

Accession number(s). This whole-genome project was deposited at the DDBJ/EMBL/GenBank under the accession number [CP023748](https://doi.org/10.1093/bioinformatics/bty004) (BioProject PRJNA412668, BioSample SAMN07722662). The version described in this paper is the first version, CP023748.1.

ACKNOWLEDGMENTS

We acknowledge the financial support from the Brazilian National Council of Scientific and Technological Development (CNPq), the Fundação Araucária de Apoio ao Desenvolvimento Científico e Tecnológico do Estado do Paraná (FAPPR), and the Simbiose-Agro Company. S. F. Yamada-Ogatta and M. Hungria were supported by a research fellowship from CNPq. E. R. Tavares was supported by a postdoctoral fellowship from the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES).

REFERENCES

- Pérez-García A, Romero D, de Vicente A. 2011. Plant protection and growth stimulation by microorganisms: biotechnological applications of bacilli in agriculture. *Curr Opin Biotech* 22:187–193. <https://doi.org/10.1016/j.copbio.2010.12.003>.
- McSpadden Gardener BB. 2004. Ecology of *Bacillus* and *Paenibacillus* spp. in agricultural systems. *Phytopathology* 94:1252–1258. <https://doi.org/10.1094/PHYTO.2004.94.11.1252>.
- Finking R, Marahiel MA. 2004. Biosynthesis of nonribosomal peptides. *Annu Rev Microbiol* 58:453–488. <https://doi.org/10.1146/annurev.micro.58.030603.123615>.
- Tareq FS, Lee MA, Lee H-S, Lee Y-J, Lee JS, Hasan CM, Islam MT, Shin HJ. 2014. Non-cytotoxic antifungal agents: isolation and structures of gageopeptides A–D from a *Bacillus* strain 109GGC020. *J Agric Food Chem* 62:5565–5572. <https://doi.org/10.1021/jf502436r>.
- Walsh CT. 2008. The chemical versatility of natural-product assembly lines. *Acc Chem Res* 41:4–10. <https://doi.org/10.1021/ar7000414>.
- Baptista JP, Sanches PP, Higashi A, Gonçalves RM, Balbi-Peña MI, de Oliveira AG. 2017. Biocontrol potential of *Bacillus* sp. LABIM40 strain against plants pathogenic fungi, abstr 0542. Abstr 50th Brazilian Congress of Phytopathology, Uberlândia, Minas Gerais, Brazil, 21–23 August 2017.
- Galardini M, Biondi EG, Bazzicalupo M, Mengoni A. 2011. CONTIGuator: a bacterial genomes finishing tool for structural insights on draft genomes. *Source Code Biol Med* 6:11. <https://doi.org/10.1186/1751-0473-6-11>.
- Tsai IJ, Otto TD, Berriman M. 2010. Improving draft assemblies by iterative mapping and assembly of short reads to eliminate gaps. *Genome Biol* 11:R41. <https://doi.org/10.1186/gb-2010-11-4-r41>.
- Sibponkrung S, Kondo T, Tanaka K, Tittabutr P, Boonkerd N, Teaumroong N, Yoshida K. 2017. Genome sequence of *Bacillus velezensis* S141, a new strain of plant growth-promoting rhizobacterium isolated from soybean rhizosphere. *Genome Announc* 5(48):e01312-17. <https://doi.org/10.1128/genomeA.01312-17>.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <https://doi.org/10.1093/nar/gkv437>.