

## Purification of Total RNA from DSS-treated Murine Tissue via Lithium Chloride Precipitation

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**[Abstract]** We have developed a protocol to purify RNA from DSS (Dextran Sulfate Sodium)-treated mouse tissues. This method, which prevents downstream inhibition of q-RT-PCR observed in DSS-treated tissues, relies on successive precipitations with lithium chloride.

**Keywords:** Dextran sodium sulfate, Colitis, Lithium chloride, RNA, q-RT-PCR

**[Background]** Dextran Sulfate Sodium (DSS) is very commonly used in laboratories to induce colitis in rodents. Specifically, it mimics the clinical and histological features of human Inflammatory Bowel Disease (IBD) with Ulcerative Colitis (UC) characteristics. DSS is diluted in the drinking water and penetrates tissues. We have observed that contamination of RNA extracts with DSS prevented successful subsequent amplification processes from the colon and small intestine, but also blood and other tissues obtained from DSS-treated animals. We had previously shown that the presence of DSS in the samples inhibited reverse transcription and polymerase chain reaction amplification (Viennois *et al.*, 2013). This inhibitory effect was observed in a dose depended manner by Kerr *et al.* and they suggested a poly-A-purification based technique to remove DSS from total RNA extract (Kerr *et al.*, 2012). We hereby propose another efficient and economical method for purifying total RNA extracts from DSS traces based on lithium chloride (LiCl) precipitations. This method has been extensively used in our laboratory as well as in others (Chassaing *et al.*, 2012, Li *et al.*, 2016); however, no attempt has been taken to document the procedure in detail. Therefore, we provide a detailed description of LiCl purification procedure of total RNA primarily isolated from DSS-treated murine tissue with another method (Trizol, Spin column-based nucleic acid purification...).

### Materials and Reagents

1. Pipette tips (0.1-10 µl, 1-200 µl, 100-1,000 µl)
2. Eppendorf Safe-Lock Tubes, 1.5 ml (Eppendorf, catalog number: 022363204)
3. 8 M lithium chloride (LiCl) (SIGMA Lithium Chloride Solution, 8 M Solution, Sigma-Aldrich, catalog number: L7026-100ML, 090M8728)
4. RT-PCR Grade Water (Thermo Fisher Scientific, Ambion™, catalog number: AM9935)
5. Pure 100% ethanol (Decon Labs, catalog number: 2716)
6. 3 M sodium acetate, pH 5.2 (see Recipes)
7. 70% ethanol (see Recipes)

## Equipment

1. Pipettes 0.5-10  $\mu$ l, 10-100  $\mu$ l and 100-1,000  $\mu$ l (Eppendorf, model: Research<sup>®</sup> plus, Variable Adjustable Volume Pipettes)
2. Refrigerated centrifuge (Thermo Fisher Scientific, Thermo Scientific<sup>™</sup>, model: Sorvall<sup>™</sup> Legend<sup>™</sup> Micro 21R, or equivalent)
3. Multi-Mode Microplate Reader (BioTek Instruments, model: BioTek<sup>™</sup> Synergy<sup>™</sup> 2)
4. -20 °C freezer

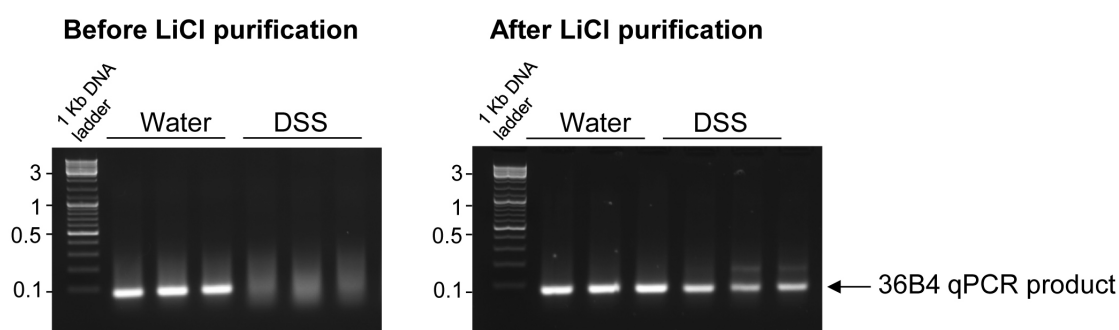
## Procedure

1. RNAs are in solution in RNase-free water after previous isolation with any method (Trizol, Spin column-based nucleic acid purification).
2. Add 0.1 volume of 8 M LiCl solution to 1 volume of RNA solution (*i.e.*, 5  $\mu$ l of 8 M LiCl for 50  $\mu$ l RNA solution).
3. Mix well by pipetting up and down and incubate on ice for 2 h.
4. After the incubation, centrifuge at 14,000  $\times g$  for 30 min, at 4 °C.
5. Discard the supernatant and dissolve the pellet (that might be almost invisible depending on the initial quantity of RNA) in 200  $\mu$ l RNase-free water.
6. Repeat steps from 1 to 5. Briefly, perform a second precipitation with 0.1 volume of LiCl (20  $\mu$ l), mix by pipetting and incubate the solution for 2 h on ice, centrifuge and dissolve the pellet in 200  $\mu$ l of water.
7. Add 0.1 volume (*i.e.*, 20  $\mu$ l) of 3 M sodium acetate (pH = 5.2) and 2.0 volumes (*i.e.*, 400  $\mu$ l) of -20 °C prechilled 100% ethanol to 1 volume of RNA solution. Incubate at -20 °C for 30 min.
8. Centrifuge at 14,000  $\times g$  for 30 min at 4 °C.
9. Discard the supernatant and wash the pellet with 100  $\mu$ l of -20 °C prechilled 70% ethanol.
10. Centrifuge at 14,000  $\times g$  for 10 min at 4 °C.
11. Remove the supernatant carefully with a pipet without disturbing the pellet which might be invisible at this step.  
*Note: The supernatant should be removed carefully. The expected location of the pellet (which is determined by the position of the tube in the centrifuge) must be considered for pipetting up the supernatant.*
12. Let the pellet air dry at room temperature for 5-10 min.
13. Dissolve the pellet in RNase free water in a volume smaller or equal to the initial volume of RNA solution in Step 1.
14. Determine RNA yield by measuring the absorbance peak at 260 nm with a multi-mode microplate reader. The absorbance ratio at 260/280 nm is also determined to evaluate the purity of RNA with an acceptable ratio from 2 to 2.2.

15. RNAs from DSS-treated tissues can now be used for cDNA synthesis and qPCR according to standard protocols.

### Data analysis

Agarose gel electrophoresis demonstrates that, before LiCl precipitation, housekeeping gene 36B4 cDNA amplification products are obtained from non-DSS-treated mice as indicated by a clear band while amplification is inhibited in DSS-treated mice as indicated by the smear (Figure 1). After LiCl precipitation, cDNA obtained from non-DSS and DSS treated samples were both successfully amplified, as attested by a distinct band (Figure 1).



**Figure 1. LiCl purification allows amplification of cDNA from DSS-treated tissues.** Total RNAs were extracted from colonic tissues obtained from mice treated with DSS diluted at 3% in the drinking water or water controls. After cDNA synthesis, qPCR for the housekeeping gene 36B4 was performed and the product of amplification was visualized by electrophoresis. The presence of a smear indicates that the amplification of 36B4 cDNA was inhibited in DSS-treated mice before LiCl purification. After purification of the DSS- and non-DSS RNA samples with LiCl, clear bands indicate that 36B4 cDNA was successfully amplified from both DSS- and non-DSS samples.

### Notes

1. This protocol might be associated with a decrease of RNA yield. The volume of re-suspension in Step 13 might be adjusted according to the initial volume and should be the same or smaller (if the RNA should be at a similar or higher concentration than before the LiCl purification).
2. Due to the partial loss of RNA that might occur during the successive precipitations, the protocol should be performed on RNA samples with an initial concentration of no less than 200-300 ng/ $\mu$ l. The best results are obtained on samples from 500 ng/ $\mu$ l initial RNA concentration and above.
3. Within the same experiment, all groups of samples, whether they were obtained from mice administered with DSS or from the water control group, should be equally subjected to the LiCl purification process.

## **Recipes**

1. 3 M sodium acetate, pH 5.2 (for 100 ml)  
24.61 g of sodium acetate  
100 ml of Milli-Q® ultra-pure water  
Adjust pH to 5.2 with HCl
2. 70% ethanol  
70% of 100% ethanol  
30% of Milli-Q® ultra-pure water

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