

## Tumor heterogeneity of gastric cancer: From the perspective of tumor-initiating cell

Jian-Peng Gao, Wei Xu, Wen-Tao Liu, Min Yan, Zheng-Gang Zhu

Jian-Peng Gao, Wei Xu, Wen-Tao Liu, Min Yan, Zheng-Gang Zhu, Department of Surgery, Shanghai Institute of Digestive Surgery, Ruijin Hospital, Shanghai Jiao Tong University, Shanghai 200025, China

ORCID number: Jian-Peng Gao (0000-0002-4627-6278); Wei Xu (0000-0002-5289-0944); Wen-Tao Liu (0000-0002-2166-8259); Min Yan (0000-0002-7486-2944); Zheng-Gang Zhu (0000-0003-4769-5463).

**Author contributions:** Gao JP performed the literature research, data analysis, and wrote the manuscript; Xu W performed the literature research and data analysis; Liu WT contributed to the literature research and provided a critical revision of the manuscript for important intellectual content; Yan M and Zhu ZG supervised the study and provided a critical revision of the manuscript prior to submission.

Supported by Shanghai Jiao Tong University Medical Engineering Cross Research Fund, No.YG2014MS59.

**Conflict-of-interest statement:** No potential conflicts of interest.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Invited manuscript

**Correspondence to:** Wen-Tao Liu, MD, PhD, Associate Professor, Surgeon, Surgical Oncologist, Department of Surgery, Shanghai Institute of Digestive Surgery, Ruijin Hospital, Shanghai Jiao Tong University, 197 Ruijin Road II, Shanghai 200025, China. [wt\\_mygod@163.com](mailto:wt_mygod@163.com)  
Telephone: +86-21-64370045  
Fax: +86-21-64314781

Received: March 20, 2018

Peer-review started: March 21, 2018

First decision: April 24, 2018

Revised: April 30, 2018

Accepted: May 26, 2018

Article in press: May 26, 2018

Published online: June 28, 2018

### Abstract

Gastric cancer (GC) remains one of the most common and malignant types of cancer due to its rapid progression, distant metastasis, and resistance to conventional chemotherapy, although efforts have been made to understand the underlying mechanism of this resistance and to improve clinical outcome. It is well recognized that tumor heterogeneity, a fundamental feature of malignancy, plays an essential role in the cancer development and chemoresistance. The model of tumor-initiating cell (TIC) has been proposed to explain the genetic, histological, and phenotypical heterogeneity of GC. TIC accounts for a minor subpopulation of tumor cells with key characteristics including high tumorigenicity, maintenance of self-renewal potential, giving rise to both tumorigenic and non-tumorigenic cancer cells, and resistance to chemotherapy. Regarding tumor-initiating cell of GC (GATIC), substantial studies have been performed to (1) identify the putative specific cell markers for purification and functional validation of GATICS; (2) trace the origin of GATICS; and (3) decode the regulatory mechanism of GATICS. Furthermore, recent studies demonstrate the plasticity of GATIC and the interaction between GATIC and its surrounding factors (TIC niche or tumor micro-environment). All these investigations pave the way for the development of GATIC-targeted therapy, which is in the phase of preclinical studies and clinical trials. Here, we interpret the heterogeneity of GC from the perspectives of TIC by reviewing the above-mentioned fundamental and clinical studies of GATICS. Problems encountered during the GATIC investigations and the potential solutions are

also discussed.

**Key words:** Gastric cancer; Tumor heterogeneity; Tumor-initiating cell

© **The Author(s) 2018.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Gastric cancer (GC) remains a severe malignancy with high incidence and mortality rates. One major underlying mechanism of GC rapid progression, extensive spreading, and chemoresistance is tumor heterogeneity could be explained by the gastric tumor-initiating cell (GATIC). Since the initial identification of putative GATICS in 2007, substantial studies have been performed to investigate various aspects of GATICS. Here, we systemically discuss the tumor heterogeneity of GC from the view of GATICS by reviewing studies on the identification and validation of GATICS, origination of GATICS, plasticity of GATICS and its underlying mechanism, and current status of chemotherapeutic agents targeting GATICS.

Gao JP, Xu W, Liu WT, Yan M, Zhu ZG. Tumor heterogeneity of gastric cancer: From the perspective of tumor-initiating cell. *World J Gastroenterol* 2018; 24(24): 2567-2581 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v24/i24/2567.htm> DOI: <http://dx.doi.org/10.3748/wjg.v24.i24.2567>

## INTRODUCTION

Gastric cancer (GC), with gastric adenocarcinoma as the most common histological type, is the fifth most common malignancy and the third leading cause of cancer-related death worldwide<sup>[1]</sup>. Despite its declining incidence and mortality rate in several developed countries, GC remains a common and deadly disease with a poor prognosis globally, especially in northeastern Asia and South America<sup>[2]</sup>. Although significant progress has been made regarding the development of anti-cancer therapeutic agents, chemoresistance and recurrence of GC are major obstacles for improving the overall survival rate of advanced GC patients<sup>[3]</sup>. Combined chemotherapy, such as fluoropyrimidine plus platinum derivatives, serves as the first-line treatment for patients with locally advanced or metastatic GC. Unfortunately, response rate of these patients to these major regimens is only approximately 50%, with overall survival ranging from 7 to 11 mo. For patients with relapsed GC, second-line treatment confers limited benefits of less than 6 mo<sup>[4,5]</sup>. With the advance of new techniques, especially high-throughput sequencing of GC patients' primary tumor materials, even at the single cell level, investigators have revealed the essential roles of tumor heterogeneity underlying the mechanism of tumor recurrence and drug resistance<sup>[6]</sup>.

Tumor heterogeneity is universally recognized as one of most fundamental features of malignancies.

It is noteworthy that the inherent variations exist not only between patients with same type of malignancies (intertumor heterogeneity) but also within any individual tumor (intratumor heterogeneity)<sup>[7]</sup>. Intratumor heterogeneity implies the inherent temporal-spatial differences between distinctive subpopulations of tumor cells within the same tumor at both genetic and epigenetic levels<sup>[8]</sup>. In-depth sequencing technology enables the identification of different regions of a single tumor entity that harbor subclones with distinct genetic and epigenetic features<sup>[9]</sup>. Moreover, non-cancerous cells (for instance, stromal cells, infiltrating immune cells, extracellular matrix, vascular endothelia cells, etc.) interact with the surrounding cancerous subpopulations and form the regional microenvironments<sup>[10]</sup>. Consequently, tumoral and microenvironmental factors confer distinctive subpopulations of cancer cells with distinctive biological features and differential responses to chemotherapeutics. This adds to the complexity of cancer treatment and leads to lethal consequences, including tumor relapse and chemoresistance. Regarding gastric cancer, multiple studies have demonstrated its intratumor heterogeneity at molecular, histological, and phenotypic levels<sup>[7]</sup>. For instance, novel molecular-based classification launched by the Cancer Genome Atlas in 2014 and other genomic studies not only highlight the heterogeneity of GC but also imply its negative influence on tumor response to therapeutics agents targeting human epidermal growth factor receptor 2 (HER2), epidermal growth factor receptor (EGFR), c-MYC, etc<sup>[11-13]</sup>.

Two major models have been proposed to interpret intratumor heterogeneity. The clonal evolution (CE) model posits that somatic mutations stochastically occur on various clones within the tumor, leading to their differential growth patterns. Mutated clones with growth advantages will expand, whereas others with disadvantageous mutations will be outcompeted. As this scenario might be presented at different sites and different stages during cancer progression, the spatial-temporal tumor heterogeneity is consequently installed<sup>[14]</sup>. Alternatively, the cancer stem cell (CSC)/tumor-initiating cell (TIC) model was introduced based on the discovery that only a minor subpopulation of cancer cells generates tumor *in vivo* and maintains its self-renewal potential<sup>[15]</sup>. Both CSCs and TICs are widely used in the literature. However, the term of TIC highlights the capacity of these cells to (re)generate tumors during *in vivo* serial xenotransplantation, which is currently the gold standard for functionally validating and evaluating their tumorigenic capacity and self-renewal potential<sup>[16]</sup>. Indeed, key features of these distinctive subsets of cancer cells include: (1) Initiating and maintaining tumor growth; (2) preserving self-renewal potential; (3) giving rise to both tumorigenic and non-tumorigenic cancer cells; and (4) being highly resistant to chemotherapy<sup>[17]</sup>. Consequently, TICs establish intratumor heterogeneity by generating a cellular hierarchy, with very primitive TICs at the apex generating both daughter TICs and more

differentiated non-TICs downwards. Recent genetic and functional studies not only identify somatic mutations within certain TIC clones but also demonstrate that these mutations influence their phenotypic features, generating distinctive TIC subclones<sup>[18]</sup>. As CE and TIC models are not mutually exclusive, these two models could be integrated. Remarkably, well-differentiated cells are shown to regain TIC properties through the process of dedifferentiation<sup>[19]</sup>. Collectively, these studies indicate that TICs are in dynamic status with substantial plasticity that is subjected to the regulation of multiple intrinsic and extrinsic factors<sup>[20,21]</sup>. These findings contribute to a comprehensive interpretation of intratumor heterogeneity through evolving characterization of TICs.

GC is both genetically and phenotypically heterogeneous, which could be explained by gastric tumor-initiating cells (GATICs) that interact with genetic/epigenetic and microenvironmental factors<sup>[22,23]</sup>. Here we systemically review the GATICs from multiple perspectives including: (1) Identification and origination of GATICs; (2) plasticity of GATICs and their regulatory mechanisms; and (3) clinical implications of GATIC-targeted therapy.

#### Identification and validation of GATICs

Identification of GATICs is executed from three major aspects: Putative cell surface markers, efflux potential, and chemotherapeutics of GATICs<sup>[24]</sup>. Further functional validation of GATICs can be achieved with serial xenotransplantation of purified TIC subpopulation, which aims to evaluate its tumorigenicity and self-renewal capacity *in vivo*<sup>[25]</sup>.

#### Identification of GATICs by cell surface markers

GATICs account for a minor subpopulation of GC cells that are endowed with significantly enhanced tumorigenic and self-renewal capacities. Key features of GATICs are partially similar with the characteristics of normal gastric stem cells (GSCs). Therefore, one major strategy of GATIC identification is through the detection of specific cell surface markers expressed on GSCs<sup>[26]</sup>.

CD44 was the first putative cell surface marker identified for GATICs. As a universally recognized stem cell marker, this transmembrane glycoprotein mainly mediates cell-cell interaction, cell adhesion, and migration in healthy tissues<sup>[27]</sup>. Studies have shown that CD44(+) tumor cells regenerate heterogeneous tumors, demonstrate self-renewal capacity *in vivo*, and promote tumor progression, especially metastasis through oncogenic and stemness-related signaling pathways<sup>[28]</sup>. In 2009, Takaishi *et al.*<sup>[29]</sup> demonstrated that fluorescence activated cell sorting (FACS)-sorted CD44(+) cells from three common GC cell lines exhibited enhanced TIC properties, including sphere formation *in vitro* and tumorigenicity in immune-deficient mice during serial transplantation, whereas CD44 knockdown induced compromised TIC properties both *in vitro* and *in vivo*. More specifically, CD44v8-10, a predominant CD44 variant, was identified

as a specific GATIC marker, as exogenous overexpression of CD44v8-10, rather than standard CD44, improved the tumorigenicity *in vivo*<sup>[30]</sup>. Since combination of multiple cell surface markers improved the specificity of TIC identification, several studies have further demonstrated other putative GATIC subpopulations expressing both CD44 and other cell surface markers, including EpCam, CD24, *etc*<sup>[31,32]</sup>. Notably, CD44(+)CD54(+) circulating tumor cells captured from GC patients' peripheral blood enabled sphere formation even at the single cell level. Moreover, the rapidly developed CD44(+)CD54(+) cell-derived tumors histologically resembled the matching primary tumor, indicating the improved specificity of GATIC population co-expressing multiple putative cell surface markers<sup>[33]</sup>.

CD90 was recently identified as a GATIC specific cell surface marker. Indeed, it is recognized as a marker for a variety of normal and cancer stem cells<sup>[34]</sup>. Jiang *et al.*<sup>[35]</sup> identified a GATIC population in primary gastric tumors characterized by its CD90 phenotypic features. CD90(+) cells were enriched in TIC-enriched cell culture condition, and these cells enabled tumor initiation with the minimum of  $1 \times 10^3$  cells. Remarkably, the cellular hierarchy of GC could be restored by CD90(+) cells at a single cell level, strongly indicating their self-renewal capacity in GC. It was further shown that both tumorigenicity and tumor progression could be compromised by anti-ERBB2 therapeutic monoclonal antibody through inhibiting CD90(+) cell population in GC tumor mass<sup>[36]</sup>.

Leucine rich repeat containing G-protein coupled receptor 5 (Lgr5) is a well-investigated stem cell marker across the gastrointestinal tract. Lgr5(+) stem cells were shown to drive self-renewal in the stomach and establish long-lived gastric units<sup>[37]</sup>. Thus, the role of Lgr5(+) cells during the initiation and progression of gastric malignancies has also been studied. Simon *et al.*<sup>[38]</sup> reported that an elevated number of Lgr5(+) stem cells might be involved in gastric tumorigenesis. Moreover, Gong *et al.*<sup>[39]</sup> showed that therapeutic antibodies targeting Lgr5(+) cells significantly inhibited tumor growth and prevented tumor recurrence, and Wang *et al.*<sup>[40]</sup> demonstrated that GATIC-enriched tumor spheres highly expressed Lgr5(+).

Aldehyde dehydrogenase (ALDH) is another universal marker for multiple cancers, including GC<sup>[41]</sup>. It is a family of enzymes that catalyze the oxidation of aromatic aldehydes to carboxyl acids. Multiple studies have shown that ALDH(+) GC cells not only contribute to the generation of heterogeneous tumor entity and the maintenance of GATIC features but also facilitate chemoresistance and tumor relapse<sup>[42,43]</sup>. An extensive study that profiled the expression of putative GATIC cell markers even claims that ALDH is one of the most specific biomarkers for GATICs in highly tumorigenic and chemoresistant non-cardiac GC, regardless of histological type<sup>[44]</sup>.

Furthermore, it has been accepted that combination

**Table 1** Putative cell surface markers of gastric tumor-initiating cell

Putative marker	Tumor sphere formation ( <i>in vitro</i> )	Tumor formation during serial engraftment ( <i>in vivo</i> )	Limiting dilution assay	Chemo-resistance	Others	Ref.
CD44	+	+	Not conducted	+	(1) Capacity of cell differentiation (2) Giving rise to both CD44+ and CD44- GC cells (3) Independent prognostic factor of GC patients	Takaishi <i>et al</i> <sup>[29]</sup> Yoon <i>et al</i> <sup>[119]</sup>
CD44/CD24	+	+	+	Not conducted	Capacity of cell differentiation	Zhang <i>et al</i> <sup>[32]</sup>
CD44/CD54	+	+	+	Not conducted	(1) Capacity of cell differentiation (2) Detectable from peripheral blood	Chen <i>et al</i> <sup>[33]</sup>
CD44/CD26	+	+	Not conducted	+	/	Nishikawa <i>et al</i> <sup>[127]</sup>
CD44/EpCam	+	+	+	+	(1) Capacity of cell differentiation (2) Restoration of histological heterogeneity from single CD44/EpCam+ cell	Han <i>et al</i> <sup>[31]</sup>
CD44v810/EpCAM	+	+	+	+	CD44v8-10 but not CD44 standard increase the frequency of tumor initiation	Lau <i>et al</i> <sup>[30]</sup>
CD90	+	+	+	+	Restoration of cellular hierarchy from single CD90+ cell	Jiang <i>et al</i> <sup>[35]</sup>
CD133	+	+	Not conducted	+	Independent prognostic factor of GC patients	Zhang <i>et al</i> <sup>[126]</sup>
Lgr5	+	+	Not conducted	Not conducted	/	Wang <i>et al</i> <sup>[40]</sup>
ALDH	+	+	+	+	/	Nishikawa <i>et al</i> <sup>[42]</sup>
ALDH1/REG4						Katsuno <i>et al</i> <sup>[45]</sup>
Oct4/Sox2/Nanog	+	+	Not conducted	+	Independent prognostic factor of GC patients	Liu <i>et al</i> <sup>[129]</sup>

ALDH: Aldehyde dehydrogenase; GC: Gastric cancer.

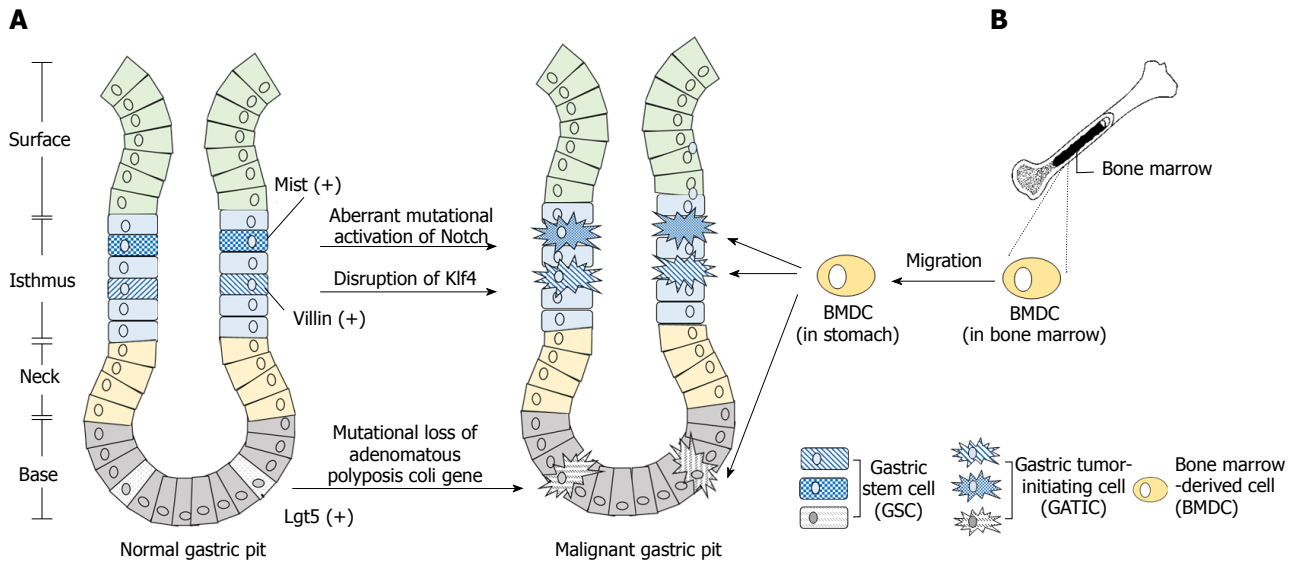
of multiple TIC markers could potentially/possibly increase the specificity of identified TIC subpopulations. In GC, the most common pattern of combined GATIC marker expression is CD44 plus another identified marker. All the putative GATIC markers, including GATICS expressing single and multiple cell surface markers, are summarized in Table 1.

Notably, several studies have demonstrated a significant correlation between reported GATIC markers and histological subtypes of GC. For instance, CD44 is more frequently and highly expressed in the intestinal subtype GC cells with moderate differentiation, whereas ALDH correlated with the diffuse subtype and epithelial-mesenchymal transition (EMT) phenotype<sup>[45,46]</sup>. Moreover, Lei *et al*<sup>[47]</sup> recently conducted novel classification of GC based on the analysis of gene expression patterns and claimed that the mesenchymal subtype had TIC properties such as high expression of putative GATIC markers, maintenance of an undifferentiated state, and enhanced activity of TIC-associated signaling pathways<sup>[48]</sup>. These studies further highlight tumor heterogeneity in terms of GATICS with specific markers differentially expressed by gastric malignancies.

#### Identification of GATICS by differential efflux potential

Although identification of TICs through putative cell surface markers has been extensively established, the specificity and accuracy of these markers are still under debate as the inconsistency of studies investigating the same markers has been observed<sup>[49]</sup>. An alternative to identify TICs is the differential efflux potential-based strategy. In principle, TICs highly express ATP-binding cassette (ABC) transporters, which facilitate the cells' ability to export foreign bodies, such as therapeutic compounds, and thereby endow TICs with key features of reduced chemosensitivity and enhanced multidrug resistance. Consequently, after exposing a whole cancer cell population to Hoechst 33342, a classic cell dye, a differential amount of residual Hoechst 33342 within non-TICs and TICs could be observed. Most cancer cells with much retained Hoechst 33342 were from the main population (MP), whereas the minor subpopulation of TICs with significantly eliminated Hoechst 33342 were defined as the side population (SP). As FACS enables the discrimination between MP and SP, this strategy has been widely applied to identify the TIC population<sup>[50]</sup>. In GC, Fukuda *et al*<sup>[51]</sup> and She *et al*<sup>[52]</sup> respectively





**Figure 1** Origin of gastric tumor-initiating cells. A: Evidence suggests that the gastric stem cell (GSC) is one major origin of the gastric tumor-initiating cell (GATIC). GSCs exist in the isthmus and bottom of the gastric pit. Certain genetic, epigenetic, and/or environmental factors potentially transform these GSCs into malignant GATICs. For instance, Mist(+) and Villin(+) GSCs in the isthmus and Lgr5(+) GSCs in the bottom could act as the origins of GATICs through multiple signaling pathways; B: GATICs are also presumed to originate partially from bone marrow-derived cells (BMDCs). The recruitment of BMDCs to stomach by chemokines and other factors is in parallel with the multi-step progression of gastric cancer (GC), which lays the basis for the presumption that BMDCs undergo the malignant transformation into GATICs and promote GC development, the underlying mechanism of which requires further investigation.

demonstrated that SP cells isolated from either GC cell lines or tumor samples exhibited improved sphere formation *in vitro* and tumorigenicity *in vivo*. Moreover, Tian *et al.*<sup>[53]</sup> isolated SP cells from multiple GC cell lines and showed that SP cells not only conferred an asymmetric cell division pattern but also exhibited elevated resistance to cisplatin and adriamycin, both of which are key characteristics of GATICs. Although these studies imply that SP cells possibly possess an enriched population of GATICs, a recent study suggested that not all SP cells contain GATICs. In this study, SP cells from BCG823 cell line showed equal tumorigenicity as MP cells, indicating that SP alone may not be potent enough to distinguish GATICs from normal GC cells<sup>[54]</sup>.

### Purification of GATICs through chemotherapeutic reagents

Apart from the above-mentioned strategies, chemotherapeutics screening can also be applied to obtain TIC populations that are inherently resistant to drugs. In fact, Xue *et al.*<sup>[55]</sup> reported a vincristine-preconditioning approach for GC cell line SGC7901 to obtain cells with increased GATIC markers, not only forming 3D structures resembling the differentiated gastric crypts but also displaying mesenchymal characteristics. A similar study conducted by Xu *et al.*<sup>[56]</sup> showed that preconditioning treatment of 5-fluorouracil to SGC7901 and AGS cell lines enriched the population with up-regulated GATIC markers, enhanced tumorigenicity and self-renewal *in vivo*, and enhanced toleration to the insults from chemotherapy.

## ORIGIN OF GATICS

As GC itself is highly heterogeneous, its initiation is also a

multi-stepwise process critically involving multiple factors. Genetic factors, infection of *H. pylori*, environmental elements, *etc.* contribute to the abnormal alteration of the gastric epithelium, especially metaplasia and dysplasia, which pave the basis for final development of gastric malignancies<sup>[57,58]</sup>. During this process, certain types of cells are abnormally transformed into GATICs, which install the cellular hierarchy with itself at the apex, giving rise to both self-renewal, tumorigenic GATICs and differentiated non-TICs. The major potential origins of GATICs are normal gastric stem cells and bone marrow-derived cells (Figure 1). Recent studies also preliminarily suggest that dedifferentiated gastric epithelial cells (GECs) could regain stemness features under certain circumstances, indicating its potential role as an origin of GATICs<sup>[59,60]</sup>. However, more rigorous and sufficient data are needed to support this proposal.

### GATICs derived from normal gastric stem cells

Gastric epithelial mucosa consists of four types of cells: chief cells, parietal cells, mucous cells, and entero-endocrine cells, all of which are derived from epithelial stem cells. These stem cells generate a cellular hierarchy of differentiation and proliferation and maintain the integrity of gastric mucosa<sup>[61]</sup>. Extensive studies of stem cells in the gastric epithelium identified a variety of GSCs with specific markers and restricted locations. For instance, Lgr5(+) cells at the bottom of adult pyloric glands<sup>[37]</sup>, Sox2(+) cells located slightly above the bottom of pyloric and fundic glands, Villinβ-gal/(+) and Mist1(+) cells in the isthmus *etc.* are all GSCs with distinctive functions during normal development of the gastric mucosa<sup>[62,63]</sup>.

It has long been postulated that resident tissue stem cells are the major resource of TICs, as malignancies could arise from the stem or progenitor cells *in situ*<sup>[26]</sup>.

In 2008, McDonald *et al.*<sup>[64]</sup> firstly proposed that the expansion and spread of mutated GSCs underlie the local progression of GC. Since then, multiple oncogenic mutations in GSCs have been identified. Adenoma is initiated when mutational loss of adenomatous polyposis coli gene induced by tamoxifen occurred in Lgr5(+) stem cells<sup>[37]</sup>. Under circumstance of chronic inflammation, aberrant activation of Notch and mutational alteration of E-cadherin in Mist(+) stem cells in the isthmus induced the development of intestinal and diffuse type of GC, respectively<sup>[63]</sup>. Moreover, mice with disrupted Krüppel-like factor 4 (Klf4) in Villin(+) antral mucosa cells develop GC more frequently than mice with normal Klf4 function, indicating dysfunctional mutation of a tumor suppressor gene in GSCs can also induce gastric carcinogenesis<sup>[65]</sup>. Taken together, it is speculated that genetic alterations and mutations transform normal GSCs into oncogenic GATICs, which then initiate and promote tumor development. In other words, normal GSCs are a major putative origin of GATICs (Figure 1A).

#### **GATICs derived from bone marrow-derived cells**

Bone marrow-derived cells (BMDCs) are recognized as the most primitive uncommitted stem cells in adult since they potentially possess substantial plasticity as well as exhibit motility, migrating towards the sites of inflammation or injury<sup>[66]</sup>. The potential role of BMDCs as a source of malignant cells was not been identified until 2004 when Houghton *et al.*<sup>[67]</sup> discovered that GC could originate from bone marrow-derived sources. In that study, *Helicobacter* C57BL/6 mouse model showed that *H. felis*-caused chronic inflammation and induced initiation and development of GC. This process was strikingly paralleled with the recruitment of BMDCs followed by dramatic population of bone marrow-derived glands in the abnormal gastric epithelial<sup>[67]</sup>. Varon *et al.*<sup>[68]</sup> further confirmed that the multi-step abnormal development of gastric epithelial induced by *H. pylori* was accompanied by significant accumulation of BMDCs. Notably, approximately 25% of the dysplasia lesions were bone-marrow derived. These discoveries strongly indicated that BMDCs, as a potential source of GATICs, could undergo abnormal transformation and contribute to GC progression, especially by migrating into the stem cell microenvironment of inflammatory tissues (Figure 1B)<sup>[69]</sup>. However, a recent study contradicted the claim and reported that BMDCs were only sporadically found in stroma and not the epithelium or glands of GC induced by carcinogens, including N-nitroso-N-methylurea and *H. felis*<sup>[70]</sup>. Therefore, more investigations are required to characterize further the process of aberrant transformation of BMDCs and to confirm them as a potential source of GATICs.

## **PLASTICITY OF GATICS AND ITS REGULATORY MECHANISMS**

### **Plasticity of GATICs**

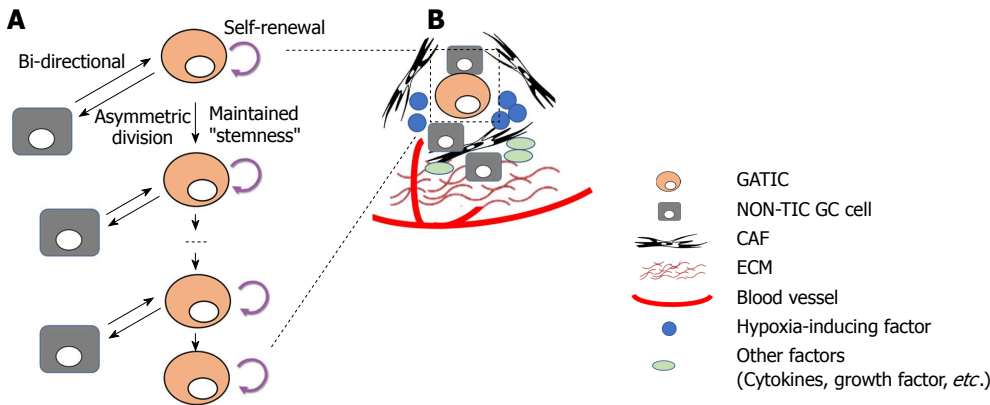
As briefly mentioned above, both the CE model and

TIC model are major proposals for the interpretation of intratumor heterogeneity. The CE model highlights the stochastic genetic/epigenetic alterations that occur in cancer cells at various sites and phases during cancer progression, resulting in the formation and expansion/extinction of different subpopulations of cancer cells with their own phenotypes<sup>[71]</sup>. In contrast, the TIC model stresses the essential role of highly tumorigenic and self-renewal TICs to generate daughter TICs with similar characteristics (symmetric division) and non-TICs with limited potential of self-renewal, tumorigenicity, and metastasis (asymmetric division)<sup>[72]</sup>. The co-existence of TICs and non-TICs with distinctive biological properties reflects the intratumor heterogeneity. Furthermore, it was repeatedly discovered that differentiated cancer cells could regain "stemness" under the regulation of multiple factors, demonstrating the existence of bidirectional conversion between TICs and non-TICs<sup>[73]</sup>. In light of this dynamic transition, TICs are more recognized as a phenotypic status or mode rather than a fixed subset of cancer cells, indicating that the plasticity of TICs plays an essential role in the intratumor heterogeneity<sup>[74]</sup>. Therefore, a unified model of CE and TICs was recently proposed to stress the plasticity of TICs: Primitive TICs with substantial capacity of tumorigenicity and self-renewal develop the cellular hierarchy. Meanwhile, they generate multiple subclones that subsequently acquire distinctive genetic mutations and/or epigenetics. Some subclones retain TIC features and continue to expand, whereas other subclones are in the non-TIC status. Nevertheless, a proportion of non-TIC subclones could reversely acquire TIC features under the regulation of genetic/epigenetic factors and/or tumor microenvironment (TME)<sup>[18]</sup>.

In GC, GATIC plasticity could also be identified. Bessède *et al.*<sup>[75]</sup> demonstrated that deletion of IQGAP1, a scaffold protein modulating cell plasticity and actin cytoskeleton, in the MKN74 GC cell line induced enhanced tumor sphere formation and increased expression of GATIC markers, both of which are major GATIC features *in vitro*. *In vivo* experiments further showed that *H. pylori*-induced gastric dysplasia was further enhanced by IQGAP1 deletion. Similarly, Yong *et al.*<sup>[76]</sup> reported that CagA-positive *H. pylori* induced the transformation of MKN45 and AGS GC cell lines into TIC-like cells as they manifested corresponding properties *in vitro*. A mechanism investigation showed that CagA upregulated Nanog and Oct4 *via* the Wnt/ $\beta$ -catenin pathway, which underlies the process of TIC status transition. Moreover, multiple studies have shown that the dedifferentiation of mature gastric epithelial cells can reacquire stemness features, including tumor-initiation, expression of TIC markers, *etc.*<sup>[60,77,78]</sup>. All these studies highlight TIC plasticity and demonstrate a dynamic conversion of cells with and without TIC properties when a variety of factors are involved in the process of regulation (Figure 2A).

### **Regulatory mechanisms of GATICs and their plasticity**

Both intrinsic factors (genetic and epigenetic alterations)



**Figure 2 Plasticity of gastric tumor-initiating cells.** A: Gastric tumor-initiating cells (GATICs) give rise to both daughter GATICs and non-TIC gastric cancer (GC) cells (asymmetric division) while maintain their self-renewal capacity and "stemness". Notably, recent studies demonstrate that differentiated non-TIC GC cells could undergo dedifferentiation and re-acquire the properties (or status) of GATICs. Thus, the bi-directional transition between TIC and non-TIC indicates the plasticity of GATICs, which is regulated by both genetic/epigenetic alterations and tumor microenvironmental factors; B: GATICs reside in the tumor-microenvironment, which consists of cancer cells (GATICs and non-TIC GC cells) as well as non-cancerous cells, such as cancer-associated fibroblast, extracellular matrix, blood supply, hypoxia (hypoxia-inducing factor), and other secreted factors, such as cytokines, growth factors. GATICs interact with these factors within the TIC niche, which exerts regulatory influence on the plasticity of GATICs through various signaling pathways.

and extrinsic factors (mainly tumor microenvironment) have been implicated in the regulation of GATICs and their plasticity (Figure 2).

**Genetic and epigenetic alterations:** Aberrantly dysregulated key effectors in several signaling pathways modulate gastric tumorigenesis and essential GATIC properties. Studies have shown that the activated hedgehog (HH) pathway significantly contributed to GC cell proliferation<sup>[79]</sup>. It was further demonstrated that sonic hedgehog (SHH) pathways are essential for maintaining the status of GATICs: Ptch and Gli1. Two SHH pathway key effectors were significantly overexpressed in GATIC-enriched sphere cultures. Significantly blocking this pathway not only decreased the expression of putative GATIC markers, including CD44 and CD24, but also reduced the self-renewal and tumorigenic capacity of GATICs, which are enriched in sphere cultures<sup>[80]</sup>. Nanog, a key transcription factor, not only maintained the self-renewal and pluripotency in embryonic stem cells but also contributed to the progression of multiple malignancies<sup>[81]</sup>. In GC, Nanog is aberrantly overexpressed in cancerous tissues. More importantly, it maintains the TIC features primarily through its interaction with multiple factors, especially the HH pathway and signal transducer and activator of transcription 3 (STAT3), which is another well-investigated regulating factor of GATICs<sup>[82]</sup>. Activated STAT3 has been observed in many types of cancers and is critically involved in cancer progression<sup>[83]</sup>. Jiang *et al.*<sup>[84]</sup> recently reported that interleukin (IL)-17 promoted the invasive transformation of quiescent GATICs through facilitating phosphorylation and subsequent activation of STAT3. Exposure of quiescent GATICs to an optimal duration and concentration of IL-17 led to an increase in N-cadherin and vimentin, which are epithelial-to-mesenchymal transition (EMT)-related makers, as well as in the invasive and clonal proliferative abilities of GATICs<sup>[84]</sup>.

Notably, epithelial cells that undergo the process of EMT acquire TIC phenotypes. Another EMT-contributing signaling pathway is the Wnt/ $\beta$ -catenin pathway, which reportedly maintains the self-renewal, tumorigenesis, and chemoresistance of TICs<sup>[85]</sup>. Oshima *et al.*<sup>[86]</sup> reported that the Wnt/ $\beta$ -catenin pathway maintained the undifferentiated status of gastric progenitor cells, and co-activation of both Wnt and prostaglandin E2 induced sequential metaplasia, dysplasia, and malignant transformation of the gastric epithelium. Other studies have shown that the Wnt/ $\beta$ -catenin pathway induced the acquirement and maintenance of TIC features in GC<sup>[87]</sup>. Investigations of other major stemness-associated factors, including TGF $\beta$ , Notch1 *etc.*, and their related signaling pathways hint their potential involvement in the maintenance of defined characteristics of TICs in certain types of cancers. However, more studies are required to validate their relationships with GATICs<sup>[88-90]</sup>.

Similarly, epigenetic alterations also critically regulate TICs and their plasticity. The malignant transformation of somatic cells, maintenance of TIC self-renewal capacity, and bidirectional transition between TICs and non-TICs are all under the regulatory influence of epigenetic factors<sup>[91]</sup>. Major epigenetic variations, including DNA hypermethylation, histone modification, and silencing of both microRNAs (miRNAs) and long non-coding RNAs (lncRNAs), have been shown to regulate GATICs. Activated oncogene ERAs not only drives the tumorigenic growth but is also involved in maintaining the TIC properties in GC<sup>[92,93]</sup>. Yashiro *et al.*<sup>[93]</sup> discovered that ERAs promoter methylation was found in six of seven GC cell lines without ERAs expression, which could be reversed by DNA methyltransferase inhibitor. Loss of methylation in the promoter of ERAs induced ERAs activation and further increased SP cells in which GATICs were enriched. Tomita *et al.*<sup>[94]</sup> reported that methylation and histone modification at the Tff1 promoter led to the extensive inactivation of the tumor suppressor gene Tff1 and fur-

ther facilitated the initiation of GC in MNU-treated wild-type mice. The epigenetic silencing in the TFF1 promoter could be partially reversed by gastrin, indicating the role of epigenetic silencing in the plasticity of GC development. Moreover, epigenetic alterations could indirectly regulate the status of GATICs through influencing key regulators in TIC-related signaling pathways. Yoda *et al.*<sup>[95]</sup> reported that aberrant methylation of DKK3, NKD1, and SFRP1, negative regulators of the WNT pathway, induced the activation of the Wnt/ $\beta$ -catenin pathway and further contributed to regulation of multiple aspects of GATIC functional activities. Wang *et al.*<sup>[96]</sup> reported that hypomethylation of the SHH promoter increased the aberrant expression of SHH ligands in GC cells, leading to further enhanced SHH pathway activity and dysregulated GATIC properties. Non-coding RNA can also exert regulatory effects on GATICs in a similar manner. For instance, microRNA-106b down-regulated Smad7 and subsequently activated the TGF- $\beta$ /Smad pathway, which in turn promoted the self-renewal and EMT characteristics of CD44(+) GC cells<sup>[97]</sup>. MicroRNA-483-5p was found to be overexpressed in GC spheroid cultures. Furthermore, this microRNA increased the expression of  $\beta$ -catenin and its downstream molecules, including cyclin D1, Bcl-2, and MMP2, which enhanced both invasiveness and self-renewal capacity of GC cells<sup>[98]</sup>.

**Tumor microenvironments (TIC niches):** Apart from intrinsic regulation of GATICs and their plasticity by genetic and epigenetic alterations, TIC niches are also key GATIC regulators within the tumor microenvironment (TME) (Figure 2B)<sup>[99]</sup>. In principal, TME is composed of fibroblastic stromal cells, endothelial and perivascular cells, immune cells, extracellular matrix (ECM), and networks of cytokines and growth factors, all of which play extensive roles in regulating GATICs<sup>[100]</sup>. It was reported that cancer-associated fibroblasts (CAFs) significantly increased the proportion of a side population (SP) of GC cells, with characteristics of cell stemness and the expression levels of GATICs markers in scirrhous gastric cancer cell lines. Mechanism investigation showed that this process was significantly influenced by the activity of TGF- $\beta$ <sup>[88]</sup>. Bone marrow-derived myofibroblasts (BMFs) are also major components of GATIC niches. The expansion and relocation of BMFs create a mesenchymal stem cell niche to facilitate tumor progression<sup>[101]</sup>. BMF-conditioned medium induced the formation of GC spheres expressing stem cell signatures and exhibiting features of self-renewal, and EMT transition through TGF- $\beta$  and Cxcr4/Cxcl12-dependent pathways<sup>[102]</sup>. Another fundamental characteristic of TIC niches is hypoxia. As tumor microenvironment is usually hypoxic, TICs are constantly under the pressure from hypoxia through the mediation of hypoxia-induced transcription factor 1 $\alpha$  and 2 $\alpha$  (HIF-1/2 $\alpha$ )<sup>[103]</sup>. In GC, it was shown that HIF-1 $\alpha$  induced EMT in GATIC-enriched spheroid cultures through the Snail signaling pathway<sup>[104]</sup>. Another study demonstrated that HIF-1 $\alpha$  down-regulated the

expression of CD133, a putative GATIC marker, in GC cell lines, suggesting that mTOR signaling is involved in the process<sup>[105]</sup>. Other major components in TIC niches, including tumor-associated macrophages, blood vessels, and soluble cytokines and growth factors, have been proven to regulate TIC activities and plasticity in multiples types of cancers. Therefore, it is reasonable to speculate that GATIC niche components exert extensive regulatory influences on GATICs and its plasticity.

## CLINICAL IMPLICATIONS OF TARGETING GATICS

Chemotherapy plays an essential role in the comprehensive treatment of GC. Unfortunately, development of tumor resistance to conventional chemotherapeutic agents poses a major obstacle in the eradication of GC. Intratumor heterogeneity of GC essentially underlies the chemoresistance. The bulk of proliferating progenitor cells or differentiated tumor cells could be effectively targeted by anti-cancer drugs, whereas a minor proportion of GATICs remain unaffected by cytotoxic agents<sup>[106]</sup>. GATICs survive from chemotherapy and re-generate heterogeneous subpopulations of GC cells, giving rise to tumor recurrence and leading to poor prognosis of GC patients.

### *Mechanisms of GATIC chemoresistance*

GATICs evade and/or tolerate the insults from chemotherapy through multiple routes. Firstly, GATICs highly express ABC transporters that function as efflux pumps of incoming reagents<sup>[107]</sup>. Consequently, GATICs constantly pump out anticancer drugs and avoid their cytotoxic effects<sup>[108]</sup>. Secondly, ALDH(+) GC cells are recognized as GATICs due to their substantially enhanced tumorigenicity and self-renewal capacity<sup>[109]</sup>. It was found that the expression of Notch1 and Shh was increased in this highly chemoresistant subpopulation, indicating that Notch1 and Shh signaling underlies the chemoresistance of GATICs<sup>[110]</sup>. Another well-recognized GATIC cell marker is CD44, which also confers GATICs with the capacity of drug resistance<sup>[111]</sup>. Chemoresistant GATICs are marked by increased glycolytic flux with activated pentose phosphate pathway (PPP)<sup>[112]</sup>. Tamada *et al.*<sup>[111]</sup> demonstrated that CD44 enhanced the glycolytic phenotype of GATICs by interacting with the pyruvate kinase M2, whereas CD44 ablation inhibited both glycolytic flux and PPP but increased intracellular level of reactive oxygen species, which are harmful to cancer cells, leading to enhanced effects of chemotherapy in hypoxia GC cells. All these mechanisms suggest that CD44 contributes to chemoresistance of GC cells through metabolic modulation. Other studies claim that stemness-related signaling pathways, including the PI3K/AKT signaling pathway and the WNT/ $\beta$ -catenin signaling pathway, also contribute to chemoresistance through their interaction with CD133 and ABCG2, respectively<sup>[110]</sup>. Moreover, the cancer cell-stroma interface of the TIC



niches within tumor microenvironment hinders the drug entrance, and thereby reduces the efficiency of chemotherapy<sup>[113]</sup>. These findings facilitate the design of therapeutic agents that specifically target GATICs and thereby improve the drug efficiency.

### **GATIC-Targeted therapies**

Based on the uncovered mechanisms of chemoresistance, two major strategies for elimination of the GATIC population have been developed: differentiation therapy and elimination therapy<sup>[114]</sup>. The former one implies treatment that induces GATIC differentiation to suppress their self-renewal capacity, and thereby making GC progression unsustainable in the long run. Han *et al.*<sup>[115]</sup> reported that ATOH1, a helix-loop-helix transcription factor, was induced during GATIC differentiation. They demonstrated that overexpression of ATOH1 in GATICs induced their differentiation and reduced their tumorigenicity both *in vitro* and *in vivo*, suggesting ATOH1 as a potential target for differentiation therapy targeting GATICs<sup>[115]</sup>. Similarly, lentiviral vector-based knockdown of PGK1, a metabolic enzyme that is involved in the dissemination of GC cells, induced the differentiation of the CD44(+) GATIC population and significantly inhibited both tumor growth and metastasis in immunodeficient mice<sup>[116]</sup>. These studies imply the feasibility of differentiation therapy to overcome chemoresistance of GC, although no therapeutic agent has yet been developed or entered clinical trials.

The other strategy to eliminate directly GATICs mainly focuses on self-renewal signaling pathways that are aberrantly overexpressed in GATICs. For instance, the SHH signaling pathway is abnormally dysregulated in GATICs. Ptch and Gli1 are two key SHH pathway genes targeted by cyclopamine<sup>[117]</sup>. Song *et al.*<sup>[80]</sup> reported that treatment of cyclopamine not only caused an enhanced reduction in self-renewal capacity but also improved the efficacy of oxaliplatin on GATIC-enriched tumor sphere cells. Vismodegib is another SHH pathway inhibitor that directly binds to SMO and subsequently inhibits the activation of downstream GLI family of transcription factors and their regulation on target genes<sup>[118]</sup>. A biomarker-based analysis of a phase 2 clinical trial of Vismodegib combined with FOLFOX vs FOLFOX demonstrated that Vismodegib could potentially reverse chemotherapy resistance in the population of patients with high CD44-expressing GC tumors<sup>[119]</sup>. Another featured pathway in GATICs is the Wnt/ $\beta$ -catenin signaling pathway, which is essentially involved in maintenance of TIC properties and induction of EMT. Gupta *et al.*<sup>[120]</sup> conducted a high-throughput screening to identify selective TIC inhibitors and discovered that salinomycin, a specific suppressor of Wnt/ $\beta$ -catenin pathway, potentially inhibited TICs in multiple cancer types. Zhi *et al.*<sup>[121]</sup> subsequently observed that chemoresistant GATICs highly expressing ALDH were relatively sensitive to salinomycin when compared to ALDH-low GC cells, indicating salinomycin as a selective therapy for GATIC

fraction. Similarly, Liu *et al.*<sup>[122]</sup> reported that ICG-001, a small molecule disrupting the co-activator of Wnt/ $\beta$ -catenin-mediated transcription, significantly suppressed GC cell growth, reduced their stemness properties, and enhanced their chemosensitivity to 5-Fu and cisplatin. Napabucasin is an orally administered small molecule that inhibits STAT3,  $\beta$ -catenin, and NANOG. Several studies have demonstrated its potent anti-stemness effect in various types of cancers<sup>[121]</sup>. A phase Ib/II clinical trial of Napabucasin combined with paclitaxel in advanced gastric and gastroesophageal junction (GEJ) adenocarcinoma not only demonstrated its safety but also observed its anti-cancer activity, leading to an ongoing phase III study of Napabucasin in combination with weekly paclitaxel as the second-line treatment for gastric/GEJ cancer<sup>[123]</sup>. Moreover, since overexpression of ABC transporters in GATICs leads to substantial efflux of therapeutic agents and chemoresistance, it is postulated that selectively inhibiting ABC transporters may be an alternative strategy to tackle chemoresistance. Indeed, multiple ABC transporter inhibitors (especially targeting ABCG2) have been recently developed to sensitize multidrug resistant (MDR) cancer cells<sup>[124,125]</sup>. Although some promising effects of improving chemosensitivity have been observed, significant side effects, such as cytotoxic effects on normal stem cells and blood-brain barrier, imply that substantial caution should be applied to obtain optimal outcomes<sup>[126]</sup>. So far, there are multiple developed therapeutic strategies targeting GATICs, either in the preclinical phase of experimental investigation or being tested in the clinical trials as developed chemotherapeutic agents. The most representative examples are shown in Table 2<sup>[127-137]</sup>.

## **CONCLUSION**

The complexity of GC remains largely unsolved due to its heterogeneity, especially intratumor heterogeneity. TIC model is proposed to interpret the heterogeneity of GC. Accumulating TIC investigations demonstrate that GATICs contribute to intratumor heterogeneity under the influence of genetic/epigenetic and microenvironmental factors. Recent studies show the bidirectional conversion between TIC and non-TIC status, indicating the plasticity of GATICs. Although the underlying mechanisms of this scenario have been studied to some extent, it remains unclear how GATICs are regulated and influenced by intrinsic and extrinsic factors. Technological advances in genomic, especially sequencing technique at the single cell level, could trace the developing route of individual GC cells and potentially better model the intertwined relationships between GATICs and their regulatory factors. With respect to the GATIC-target therapy, several *in vitro* and/or *in vivo* functional experiments have demonstrated that targeting GATICs reduced chemoresistance and thereby improved the outcomes of drug treatment. However, the disparity of drug effects between preclinical studies and clinical

**Table 2 Gastric tumor-initiating cell-targeted therapeutic strategies/agents**

Therapeutic target	Therapeutic agent	Investigation status	Underlying mechanism	Treatment	Result of treatment	Ref.
ATOH1	Lentiviral vector-based	Preclinical investigation	Overexpression of ATOH1 mediates its transcriptional activity to downstream genes and induces the differentiation of GATICs	Lentiviral vector-based overexpression of ATOH1	(1) Induction of CD44+/Lgr5+ GATICs differentiation (2) Reduced tumorigenicity of GATICs both <i>in vitro</i> and <i>in vivo</i>	Han <i>et al</i> <sup>[115]</sup>
PGK1	Lentiviral vector-based	Preclinical investigation	Knockdown of PGK1 alters the glycolytic metabolism of GATICs not only induces GATIC differentiation but also improve their chemosensitivity	Lentiviral vector-based knockdown of PGK1	(1) Induction of CD44+ GATICs differentiation (2) Inhibited tumor growth and metastasis <i>in vivo</i>	Zieker <i>et al</i> <sup>[116]</sup>
CD44v	Sulfasalazine	Phase I dose-escalation clinical study in EPOC1205	Targeting CD44v by inhibiting xCT which mainly interacts with CD44v and maintains high level of GSH	12 g/d, 4x/d with 2 wk as one cycle, oral administration	Reduced level of CD44v positive GATICs in some patients	Shitara <i>et al</i> <sup>[130]</sup>
EpCam	Catumaxomab	Phase II / III clinical trial of advanced gastric carcinoma_NCT00836654	Direct targeting CD3 and EpCam	Paracentesis +/- Catumaxomab	Clinical benefit (prolonged PFS and less symptoms of ascites) in GC patients with secondary malignant ascites	Heiss <i>et al</i> <sup>[131]</sup>
EpCam	Catumaxomab	Phase II clinical trial of advanced gastric carcinoma_NCT01784900	Direct targeting CD3 and EpCam	Surgical resection followed by Catumaxomab	Intra-/postoperative administration of catumaxomab within multimodal treatment is feasible and tolerable	Goéré <i>et al</i> <sup>[132]</sup>
c-MET	Rilotumumab	Phase III clinical trial of locally advanced or metastatic gastric and GEJ carcinoma_NCT01697072	Competitively targeting hepatocyte growth factor (HGF), ligand of c-MET receptor	ECX +/- Rilotumumab	Stopped early due to increased death risk	Doshi <i>et al</i> <sup>[133]</sup>
c-MET	Onartuzumab	Phase III clinical trial of metastatic HER2(-) and c-MET(+) Gastroesophageal Cancer_NCT01662869	Direct targeting c-MET as a MET antagonist	FOLFOX6 +/- Rilotumumab	Insignificant prolong of PFS (6.9 mo <i>vs</i> 5.7 mo) and OS (11.0 mo <i>vs</i> 9.7 mo)	Shah <i>et al</i> <sup>[134]</sup>
c-MET	Tivantinib	Phase I / II clinical trial of advanced and metastatic adenocarcinoma of distal esophagus, GEJ and stomach_NCT01611857	Inhibition of c-Met receptor tyrosine kinase	FOLFOX6 combined with Tivantinib	PFS: 6.1 mo and OS: 9.6 mo	Pant <i>et al</i> <sup>[135]</sup>
SHH signaling pathway	Cyclopamine	Preclinical investigation	Targeting overexpressed Ptch/Gli1 (key effectors in SHH pathway)	Direct addition of cyclopamine (5 $\mu$ mol/L <i>in vitro</i> and 10 $\mu$ mol/L <i>in vivo</i> )	(1) Reduced self-renewing capacity of GATIC-enriched tumor sphere (2) Enhanced efficacy of Oxaliplatin/Mitomycin inhibiting proliferation of tumor sphere	Song <i>et al</i> <sup>[80]</sup>
SHH signaling pathway	Vismodegib	Phase II clinical trial of advanced gastric and GEJ carcinoma_NCT00982592	Targeting Smoothed (SMO) and its downstream GLI family members	FOLFOX +/- Vismodegib	(1) No significant improvement of anti-tumor activity (2) Potentially reverse the chemotherapy resistance of patients with high CD44-expressing tumor cells	Cohen <i>et al</i> <sup>[136]</sup>
Wnt/ $\beta$ -catenin signaling pathway	Salinomycin	Preclinical investigation	Blocking and degrading LRP6 (Wnt co-receptor)	Direct addition of Salinomycin (ranging from 1 $\mu$ mol/L to 100 $\mu$ mol/L <i>in vitro</i> )	Effectively kill ALDH-high GATICs which are resistant to 5-FU and CDDP	Mao <i>et al</i> <sup>[87]</sup>
Wnt/ $\beta$ -catenin signaling pathway	ICG-001	Preclinical investigation	Inhibiting CBP (co-activator of Wnt/ $\beta$ -catenin pathway)	Direct addition of ICG-001 (50 mg/kg/d, <i>in vivo</i> )	(1) Suppressed GC cell growth and metastasis both <i>in vitro</i> and <i>in vivo</i> (2) Reduced self-renewal capacity and enhanced efficacy of 5-Fu/cisplatin	Liu <i>et al</i> <sup>[122]</sup>

STAT3 signaling pathway	Napabucasin	Phase I b/ II dose-escalation and extension study of advanced gastric and GEJ carcinoma_NCT01325441	Direct targeting Stat3, $\beta$ -catenin and NANOG	Paclitaxel +/- Napabucasin	(1) Well-tolerated by GC patients even receiving high doses of chemotherapy (2) Observed anti-tumor activity but still needs to be further confirmed in the on-going BRIGHTER phase III clinical trial	Shah <i>et al.</i> <sup>[137]</sup>
-------------------------	-------------	---	--	----------------------------	---	-------------------------------------

GATIC: Gastric tumor-initiating cell; SHH: Sonic hedgehog; GEJ: Gastroesophageal junction; GC: Gastric cancer; ALDH: Aldehyde dehydrogenase.

trials has also been repeatedly observed. One major explanation is that current *in vitro* and *in vivo* models failed to recapitulate the real TME that plays crucial roles in regulating GATIC phenotypes and plasticity. With the application of Matrigel® and other specific cell culture materials, three-dimensional spheroid and even organoid cultures of GC have been recently generated to enrich TIC subpopulation and mimic the real status of GC cells within the microenvironment. As stroma and ECM, key aspects of TME, are still missing in current cultivation system, new methods, such as co-culture of patient-derived cancer cells and stromal cells within the ECM-like “scaffolds”, will be developed in the near future to represent better the tumor heterogeneity.

## REFERENCES

- 1 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin* 2016; **66**: 7-30 [PMID: 26742998 DOI: 10.3322/caac.21332]
- 2 Saikawa Y, Fukuda K, Takahashi T, Nakamura R, Takeuchi H, Kitagawa Y. Gastric carcinogenesis and the cancer stem cell hypothesis. *Gastric Cancer* 2010; **13**: 11-24 [PMID: 20373071 DOI: 10.1007/s10120-009-0537-4]
- 3 Marin JJ, Al-Abdulla R, Lozano E, Briz O, Bujanda L, Banales JM, Macias RI. Mechanisms of Resistance to Chemotherapy in Gastric Cancer. *Anticancer Agents Med Chem* 2016; **16**: 318-334 [PMID: 26234359 DOI: 10.2174/1871520615666150803125121]
- 4 Thuss-Patience PC, Kretzschmar A, Bichev D, Deist T, Hinke A, Breithaupt K, Dogan Y, Gebauer B, Schumacher G, Reichardt P. Survival advantage for irinotecan versus best supportive care as second-line chemotherapy in gastric cancer—a randomised phase III study of the Arbeitsgemeinschaft Internistische Onkologie (AIO). *Eur J Cancer* 2011; **47**: 2306-2314 [PMID: 21742485 DOI: 10.1016/j.ejca.2011.06.002]
- 5 Kang JH, Lee SI, Lim DH, Park KW, Oh SY, Kwon HC, Hwang IG, Lee SC, Nam E, Shin DB, Lee J, Park JO, Park YS, Lim HY, Kang WK, Park SH. Salvage chemotherapy for pretreated gastric cancer: a randomized phase III trial comparing chemotherapy plus best supportive care with best supportive care alone. *J Clin Oncol* 2012; **30**: 1513-1518 [PMID: 22412140 DOI: 10.1200/JCO.2011.39.4585]
- 6 McGranahan N, Swanton C. Clonal Heterogeneity and Tumor Evolution: Past, Present, and the Future. *Cell* 2017; **168**: 613-628 [PMID: 28187284 DOI: 10.1016/j.cell.2017.01.018]
- 7 Gullo I, Carneiro F, Oliveira C, Almeida GM. Heterogeneity in Gastric Cancer: From Pure Morphology to Molecular Classifications. *Pathobiology* 2018; **85**: 50-63 [PMID: 28618420 DOI: 10.1159/000473881]
- 8 Welch DR. Tumor Heterogeneity--A ‘Contemporary Concept’ Founded on Historical Insights and Predictions. *Cancer Res* 2016; **76**: 4-6 [PMID: 26729788 DOI: 10.1158/0008-5472.CAN-15-3024]
- 9 Gerlinger M, Rowan AJ, Horswell S, Math M, Larkin J, Endesfelder D, Gronroos E, Martinez P, Matthews N, Stewart A, Tarpey P, Varela I, Phillimore B, Begum S, McDonald NQ, Butler A, Jones D, Raine K, Latimer C, Santos CR, Nohadani M, Eklund AC, Spencer-Dene B, Clark G, Pickering L, Stamp G, Gore M, Szallasi Z, Downward J, Futreal PA, Swanton C. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med* 2012; **366**: 883-892 [PMID: 22397650 DOI: 10.1056/NEJMoa1113205]
- 10 Junttila MR, de Sauvage FJ. Influence of tumour micro-environment heterogeneity on therapeutic response. *Nature* 2013; **501**: 346-354 [PMID: 24048067 DOI: 10.1038/nature12626]
- 11 Alsina M, Gullo I, Carneiro F. Intratumoral heterogeneity in gastric cancer: a new challenge to face. *Ann Oncol* 2017; **28**: 912-913 [PMID: 28368465 DOI: 10.1093/annonc/mdx134]
- 12 Cancer Genome Atlas Research Network. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature* 2014; **513**: 202-209 [PMID: 25079317 DOI: 10.1038/nature13480]
- 13 Böger C, Krüger S, Behrens HM, Bock S, Haag J, Kalthoff H, Röcken C. Epstein-Barr virus-associated gastric cancer reveals intratumoral heterogeneity of PIK3CA mutations. *Ann Oncol* 2017; **28**: 1005-1014 [PMID: 28453696 DOI: 10.1093/annonc/mdx047]
- 14 Anderson AR, Weaver AM, Cummings PT, Quaranta V. Tumor morphology and phenotypic evolution driven by selective pressure from the microenvironment. *Cell* 2006; **127**: 905-915 [PMID: 17129778 DOI: 10.1016/j.cell.2006.09.042]
- 15 Baccelli I, Trumpp A. The evolving concept of cancer and metastasis stem cells. *J Cell Biol* 2012; **198**: 281-293 [PMID: 22869594 DOI: 10.1083/jcb.201202014]
- 16 Ishizawa K, Rasheed ZA, Karisch R, Wang Q, Kowalski J, Susky E, Pereira K, Karamboulas C, Moghal N, Rajeshkumar NV, Hidalgo M, Tsao M, Ailles L, Waddell TK, Maitra A, Neel BG, Matsui W. Tumor-initiating cells are rare in many human tumors. *Cell Stem Cell* 2010; **7**: 279-282 [PMID: 20804964 DOI: 10.1016/j.stem.2010.08.009]
- 17 Fulawka L, Donizy P, Halon A. Cancer stem cells--the current status of an old concept: literature review and clinical approaches. *Biol Res* 2014; **47**: 66 [PMID: 25723910 DOI: 10.1186/0717-6287-47-66]
- 18 Kreso A, Dick JE. Evolution of the cancer stem cell model. *Cell Stem Cell* 2014; **14**: 275-291 [PMID: 24607403 DOI: 10.1016/j.stem.2014.02.006]
- 19 Medema JP. Cancer stem cells: the challenges ahead. *Nat Cell Biol* 2013; **15**: 338-344 [PMID: 23548926 DOI: 10.1038/ncb2717]
- 20 Shackleton M, Quintana E, Fearon ER, Morrison SJ. Heterogeneity in cancer: cancer stem cells versus clonal evolution. *Cell* 2009; **138**: 822-829 [PMID: 19737509 DOI: 10.1016/j.cell.2009.08.017]
- 21 Prasetyanti PR, Medema JP. Intra-tumor heterogeneity from a cancer stem cell perspective. *Mol Cancer* 2017; **16**: 41 [PMID: 28209166 DOI: 10.1186/s12943-017-0600-4]
- 22 Tatematsu M, Tsukamoto T, Inada K. Stem cells and gastric cancer: role of gastric and intestinal mixed intestinal metaplasia. *Cancer Sci* 2003; **94**: 135-141 [PMID: 12708487 DOI: 10.1111/j.1349-7006.2003.tb01409.x]
- 23 Li K, Dan Z, Nie YQ. Gastric cancer stem cells in gastric carcinogenesis, progression, prevention and treatment. *World J Gastroenterol* 2014; **20**: 5420-5426 [PMID: 24833872 DOI: 10.3748/wjg.v20.i18.5420]
- 24 Bekaii-Saab T, El-Rayes B. Identifying and targeting cancer stem cells in the treatment of gastric cancer. *Cancer* 2017; **123**: 1303-1312 [PMID: 28117883 DOI: 10.1002/cncr.30538]
- 25 Rycaj K, Tang DG. Cell-of-Origin of Cancer versus Cancer Stem Cells: Assays and Interpretations. *Cancer Res* 2015; **75**: 4003-4011 [PMID: 26292361 DOI: 10.1158/0008-5472.CAN-15-0798]
- 26 Takaishi S, Okumura T, Wang TC. Gastric cancer stem cells. *J*

- Clin Oncol* 2008; **26**: 2876-2882 [PMID: 18539967 DOI: 10.1200/JCO.2007.15.2603]
- 27 **Williams K**, Motiani K, Giridhar PV, Kasper S. CD44 integrates signaling in normal stem cell, cancer stem cell and (pre)metastatic niches. *Exp Biol Med* (Maywood) 2013; **238**: 324-338 [PMID: 23598979 DOI: 10.1177/1535370213480714]
- 28 **Naor D**, Nedvetzki S, Golan I, Melnik L, Faitelson Y. CD44 in cancer. *Crit Rev Clin Lab Sci* 2002; **39**: 527-579 [PMID: 12484499 DOI: 10.1080/10408360290795574]
- 29 **Takaishi S**, Okumura T, Tu S, Wang SS, Shibata W, Vigneshwaran R, Gordon SA, Shimada Y, Wang TC. Identification of gastric cancer stem cells using the cell surface marker CD44. *Stem Cells* 2009; **27**: 1006-1020 [PMID: 19415765 DOI: 10.1002/stem.30]
- 30 **Lau WM**, Teng E, Chong HS, Lopez KA, Tay AY, Salto-Tellez M, Shabbir A, So JB, Chan SL. CD44v8-10 is a cancer-specific marker for gastric cancer stem cells. *Cancer Res* 2014; **74**: 2630-2641 [PMID: 24618343 DOI: 10.1158/0008-5472.CAN-13-2309]
- 31 **Han ME**, Jeon TY, Hwang SH, Lee YS, Kim HJ, Shim HE, Yoon S, Baek SY, Kim BS, Kang CD, Oh SO. Cancer spheres from gastric cancer patients provide an ideal model system for cancer stem cell research. *Cell Mol Life Sci* 2011; **68**: 3589-3605 [PMID: 21448722 DOI: 10.1007/s00018-011-0672-z]
- 32 **Zhang C**, Li C, He F, Cai Y, Yang H. Identification of CD44+CD24+ gastric cancer stem cells. *J Cancer Res Clin Oncol* 2011; **137**: 1679-1686 [PMID: 21882047 DOI: 10.1007/s00432-011-1038-5]
- 33 **Chen T**, Yang K, Yu J, Meng W, Yuan D, Bi F, Liu F, Liu J, Dai B, Chen X, Wang F, Zeng F, Xu H, Hu J, Mo X. Identification and expansion of cancer stem cells in tumor tissues and peripheral blood derived from gastric adenocarcinoma patients. *Cell Res* 2012; **22**: 248-258 [PMID: 21727908 DOI: 10.1038/cr.2011.109]
- 34 **Shaikh MV**, Kala M, Nivsarkar M. CD90 a potential cancer stem cell marker and a therapeutic target. *Cancer Biomark* 2016; **16**: 301-307 [PMID: 27062695 DOI: 10.3233/CBM-160590]
- 35 **Jiang J**, Zhang Y, Chuai S, Wang Z, Zheng D, Xu F, Zhang Y, Li C, Liang Y, Chen Z. Trastuzumab (herceptin) targets gastric cancer stem cells characterized by CD90 phenotype. *Oncogene* 2012; **31**: 671-682 [PMID: 21743497 DOI: 10.1038/ncr.2011.282]
- 36 **Singh SR**. Gastric cancer stem cells: a novel therapeutic target. *Cancer Lett* 2013; **338**: 110-119 [PMID: 23583679 DOI: 10.1016/j.canlet.2013.03.035]
- 37 **Barker N**, Huch M, Kujala P, van de Wetering M, Snippert HJ, van Es JH, Sato T, Stange DE, Begthel H, van den Born M, Danenberg E, van den Brink S, Korving J, Abo A, Peters PJ, Wright N, Poulsom R, Clevers H. Lgr5(+ve) stem cells drive self-renewal in the stomach and build long-lived gastric units in vitro. *Cell Stem Cell* 2010; **6**: 25-36 [PMID: 20085740 DOI: 10.1016/j.stem.2009.11.013]
- 38 **Simon E**, Petke D, Böger C, Behrens HM, Warneke V, Ebert M, Röcken C. The spatial distribution of LGR5+ cells correlates with gastric cancer progression. *PLoS One* 2012; **7**: e35486 [PMID: 22530031 DOI: 10.1371/journal.pone.0035486]
- 39 **Gong X**, Azhdarinia A, Ghosh SC, Xiong W, An Z, Liu Q, Carmon KS. LGR5-Targeted Antibody-Drug Conjugate Eradicates Gastrointestinal Tumors and Prevents Recurrence. *Mol Cancer Ther* 2016; **15**: 1580-1590 [PMID: 27207778 DOI: 10.1158/1535-7163.MCT-16-0114]
- 40 **Wang Z**, Liu C. Lgr5-Positive Cells are Cancer-Stem-Cell-Like Cells in Gastric Cancer. *Cell Physiol Biochem* 2015; **36**: 2447-2455 [PMID: 26279446 DOI: 10.1159/000430205]
- 41 **Clark DW**, Palle K. Aldehyde dehydrogenases in cancer stem cells: potential as therapeutic targets. *Ann Transl Med* 2016; **4**: 518 [PMID: 28149880 DOI: 10.21037/atm.2016.11.82]
- 42 **Nishikawa S**, Konno M, Hamabe A, Hasegawa S, Kano Y, Ohta K, Fukusumi T, Sakai D, Kudo T, Haraguchi N, Satoh T, Takiguchi S, Mori M, Doki Y, Ishii H. Aldehyde dehydrogenase high gastric cancer stem cells are resistant to chemotherapy. *Int J Oncol* 2013; **42**: 1437-1442 [PMID: 23440340 DOI: 10.3892/ijo.2013.1837]
- 43 **Senel F**, Kökenek Unal TD, Karaman H, Inanç M, Aytekin A. Prognostic Value of Cancer Stem Cell Markers CD44 and ALDH1/2 in Gastric Cancer Cases *Asian Pac J Cancer Prev* 2017; **18**: 2527-2531 [PMID: 28952294 DOI: 10.22034/apjcp.2017.18.9.2527]
- 44 **Nguyen PH**, Giraud J, Chambonnier L, Dubus P, Wittkop L, Belleannée G, Collet D, Soubeyran I, Evrard S, Rousseau B, Senant-Dugot N, Mégraud F, Mazurier F, Varon C. Characterization of Biomarkers of Tumorigenic and Chemoresistant Cancer Stem Cells in Human Gastric Carcinoma. *Clin Cancer Res* 2017; **23**: 1586-1597 [PMID: 27620279 DOI: 10.1158/1078-0432.CCR-15-2157]
- 45 **Katsuno Y**, Ehata S, Yashiro M, Yanagihara K, Hirakawa K, Miyazono K. Coordinated expression of REG4 and aldehyde dehydrogenase 1 regulating tumorigenic capacity of diffuse-type gastric carcinoma-initiating cells is inhibited by TGF- $\beta$ . *J Pathol* 2012; **228**: 391-404 [PMID: 22430847 DOI: 10.1002/path.4020]
- 46 **Nosrati A**, Naghshvar F, Khanari S. Cancer Stem Cell Markers CD44, CD133 in Primary Gastric Adenocarcinoma. *Int J Mol Cell Med* 2014; **3**: 279-286 [PMID: 25635255]
- 47 **Lei Z**, Tan IB, Das K, Deng N, Zouridis H, Pattison S, Chua C, Feng Z, Guan YK, Ooi CH, Ivanova T, Zhang S, Lee M, Wu J, Ngo A, Manesh S, Tan E, Teh BT, So JB, Goh LK, Boussioutas A, Lim TK, Flotow H, Tan P, Rozen SG. Identification of molecular subtypes of gastric cancer with different responses to PI3-kinase inhibitors and 5-fluorouracil. *Gastroenterology* 2013; **145**: 554-565 [PMID: 23684942 DOI: 10.1053/j.gastro.2013.05.010]
- 48 **Turner ES**, Turner JR. Expanding the Lauren classification: a new gastric cancer subtype? *Gastroenterology* 2013; **145**: 505-508 [PMID: 23891604 DOI: 10.1053/j.gastro.2013.07.019]
- 49 **Miller TJ**, McCoy MJ, Hemmings C, Bulsara MK, Iacopetta B, Platell CF. Objective analysis of cancer stem cell marker expression using immunohistochemistry. *Pathology* 2017; **49**: 24-29 [PMID: 27913042 DOI: 10.1016/j.pathol.2016.09.063]
- 50 **Morii E**. Heterogeneity of tumor cells in terms of cancer-initiating cells. *J Toxicol Pathol* 2017; **30**: 1-6 [PMID: 28190919 DOI: 10.1293/tox.2016-0056]
- 51 **Fukuda K**, Saikawa Y, Ohashi M, Kumagai K, Kitajima M, Okano H, Matsuzaki Y, Kitagawa Y. Tumor initiating potential of side population cells in human gastric cancer. *Int J Oncol* 2009; **34**: 1201-1207 [PMID: 19360333]
- 52 **She JJ**, Zhang PG, Wang X, Che XM, Wang ZM. Side population cells isolated from KATO III human gastric cancer cell line have cancer stem cell-like characteristics. *World J Gastroenterol* 2012; **18**: 4610-4617 [PMID: 22969237 DOI: 10.3748/wjg.v18.i33.4610]
- 53 **Tian T**, Zhang Y, Wang S, Zhou J, Xu S. Sox2 enhances the tumorigenicity and chemoresistance of cancer stem-like cells derived from gastric cancer. *J Biomed Res* 2012; **26**: 336-345 [PMID: 23554769 DOI: 10.7555/JBR.26.20120045]
- 54 **Zhang H**, Xi H, Cai A, Xia Q, Wang XX, Lu C, Zhang Y, Song Z, Wang H, Li Q, Chen L, Guo Z. Not all side population cells contain cancer stem-like cells in human gastric cancer cell lines. *Dig Dis Sci* 2013; **58**: 132-139 [PMID: 22878917 DOI: 10.1007/s10620-012-2330-1]
- 55 **Xue Z**, Yan H, Li J, Liang S, Cai X, Chen X, Wu Q, Gao L, Wu K, Nie Y, Fan D. Identification of cancer stem cells in vincristine preconditioned SGC7901 gastric cancer cell line. *J Cell Biochem* 2012; **113**: 302-312 [PMID: 21913215 DOI: 10.1002/jcb.23356]
- 56 **Xu ZY**, Tang JN, Xie HX, Du YA, Huang L, Yu PF, Cheng XD. 5-Fluorouracil chemotherapy of gastric cancer generates residual cells with properties of cancer stem cells. *Int J Biol Sci* 2015; **11**: 284-294 [PMID: 25678847 DOI: 10.7150/ijbs.10248]
- 57 **Massarrat S**, Stolte M. Development of gastric cancer and its prevention. *Arch Iran Med* 2014; **17**: 514-520 [PMID: 24979566]
- 58 **Zavros Y**. Initiation and Maintenance of Gastric Cancer: A Focus on CD44 Variant Isoforms and Cancer Stem Cells. *Cell Mol Gastroenterol Hepatol* 2017; **4**: 55-63 [PMID: 28560289 DOI: 10.1016/j.jcmgh.2017.03.003]
- 59 **Aoi T**, Yae K, Nakagawa M, Ichisaka T, Okita K, Takahashi K, Chiba T, Yamanaka S. Generation of pluripotent stem cells from adult mouse liver and stomach cells. *Science* 2008; **321**: 699-702 [PMID: 18276851 DOI: 10.1126/science.1154884]
- 60 **Choi YJ**, Kim N, Chang H, Lee HS, Park SM, Park JH, Shin CM, Kim JM, Kim JS, Lee DH, Jung HC. Helicobacter pylori-induced epithelial-mesenchymal transition, a potential role of gastric cancer initiation and an emergence of stem cells. *Carcinogenesis* 2015; **36**:



- 553-563 [PMID: 25784376 DOI: 10.1093/carcin/bgv022]
- 61 **Wright NA.** Epithelial stem cell repertoire in the gut: clues to the origin of cell lineages, proliferative units and cancer. *Int J Exp Pathol* 2000; **81**: 117-143 [PMID: 10762441 DOI: 10.1046/j.1365-2613.2000.00146.x]
- 62 **Qiao XT, Ziel JW, McKimpton W, Madison BB, Todisco A, Merchant JL, Samuelson LC, Gumucio DL.** Prospective identification of a multilineage progenitor in murine stomach epithelium. *Gastroenterology* 2007; **133**: 1989-1998 [PMID: 18054570 DOI: 10.1053/j.gastro.2007.09.031]
- 63 **Hayakawa Y, Ariyama H, Stancikova J, Sakitani K, Asfaha S, Renz BW, Dubeykovskaya ZA, Shibata W, Wang H, Westphalen CB, Chen X, Takemoto Y, Kim W, Khurana SS, Taylor Y, Nagar K, Tomita H, Hara A, Sepulveda AR, Setlik W, Gershon MD, Saha S, Ding L, Shen Z, Fox JG, Friedman RA, Konieczny SF, Worthley DL, Korinek V, Wang TC.** Mist1 Expressing Gastric Stem Cells Maintain the Normal and Neoplastic Gastric Epithelium and Are Supported by a Perivascular Stem Cell Niche. *Cancer Cell* 2015; **28**: 800-814 [PMID: 26585400 DOI: 10.1016/j.ccell.2015.10.003]
- 64 **McDonald SA, Greaves LC, Gutierrez-Gonzalez L, Rodriguez-Justo M, Deheragoda M, Leedham SJ, Taylor RW, Lee CY, Preston SL, Lovell M, Hunt T, Elia G, Oukrif D, Harrison R, Novelli MR, Mitchell I, Stoker DL, Turnbull DM, Jankowski JA, Wright NA.** Mechanisms of field cancerization in the human stomach: the expansion and spread of mutated gastric stem cells. *Gastroenterology* 2008; **134**: 500-510 [PMID: 18242216 DOI: 10.1053/j.gastro.2007.11.035]
- 65 **Li Q, Jia Z, Wang L, Kong X, Li Q, Guo K, Tan D, Le X, Wei D, Huang S, Mishra L, Xie K.** Disruption of Klf4 in villin-positive gastric progenitor cells promotes formation and progression of tumors of the antrum in mice. *Gastroenterology* 2012; **142**: 531-542 [PMID: 22155367 DOI: 10.1053/j.gastro.2011.11.034]
- 66 **Krause DS, Theise ND, Collector MI, Henegariu O, Hwang S, Gardner R, Neutzel S, Sharkis SJ.** Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. *Cell* 2001; **105**: 369-377 [PMID: 11348593 DOI: 10.1016/S0092-8674(01)00328-2]
- 67 **Houghton J, Stoicov C, Nomura S, Rogers AB, Carlson J, Li H, Cai X, Fox JG, Goldenring JR, Wang TC.** Gastric cancer originating from bone marrow-derived cells. *Science* 2004; **306**: 1568-1571 [PMID: 15567866 DOI: 10.1126/science.1099513]
- 68 **Varon C, Dubus P, Mazurier F, Asencio C, Chambonnier L, Ferrand J, Giese A, Senant-Dugot N, Carloti M, Mégraud F.** Helicobacter pylori infection recruits bone marrow-derived cells that participate in gastric preneoplasia in mice. *Gastroenterology* 2012; **142**: 281-291 [PMID: 22062361 DOI: 10.1053/j.gastro.2011.10.036]
- 69 **McLarnon A.** Helicobacter pylori: Bone-marrow-derived cells could cause gastric preneoplasia in chronic Helicobacter pylori infection. *Nat Rev Gastroenterol Hepatol* 2011; **9**: 4 [PMID: 22158384 DOI: 10.1038/nrgastro.2011.226]
- 70 **Yang C, Gu L, Deng D.** Bone marrow-derived cells may not be the original cells for carcinogen-induced mouse gastrointestinal carcinomas. *PLoS One* 2013; **8**: e79615 [PMID: 24260263 DOI: 10.1371/journal.pone.0079615]
- 71 **Shlush LI, Hershkovitz D.** Clonal evolution models of tumor heterogeneity. *Am Soc Clin Oncol Educ Book* 2015; e662-e665 [PMID: 25993239 DOI: 10.14694/EdBook\_AM.2015.35.e662]
- 72 **Calderwood SK.** Tumor heterogeneity, clonal evolution, and therapy resistance: an opportunity for multitargeting therapy. *Discov Med* 2013; **15**: 188-194 [PMID: 23545047]
- 73 **Chow KH, Shin DM, Jenkins MH, Miller EE, Shih DJ, Choi S, Low BE, Philip V, Rybinski B, Bronson RT, Taylor MD, Yun K.** Epigenetic states of cells of origin and tumor evolution drive tumor-initiating cell phenotype and tumor heterogeneity. *Cancer Res* 2014; **74**: 4864-4874 [PMID: 25136069 DOI: 10.1158/0008-5472.CAN-13-3293]
- 74 **Cabrera MC, Hollingsworth RE, Hurt EM.** Cancer stem cell plasticity and tumor hierarchy. *World J Stem Cells* 2015; **7**: 27-36 [PMID: 25621103 DOI: 10.4252/wjsc.v7.i1.27]
- 75 **Bessède E, Molina S, Acuña-Amador L, Dubus P, Staedel C, Chambonnier L, Buissonnière A, Sifré E, Giese A, Bénégat L, Rousseau B, Costet P, Sacks DB, Mégraud F, Varon C.** Deletion of IQGAP1 promotes Helicobacter pylori-induced gastric dysplasia in mice and acquisition of cancer stem cell properties in vitro. *Oncotarget* 2016; **7**: 80688-80699 [PMID: 27729612 DOI: 10.18632/oncotarget.12486]
- 76 **Yong X, Tang B, Xiao YF, Xie R, Qin Y, Luo G, Hu CJ, Dong H, Yang SM.** Helicobacter pylori upregulates Nanog and Oct4 via Wnt/β-catenin signaling pathway to promote cancer stem cell-like properties in human gastric cancer. *Cancer Lett* 2016; **374**: 292-303 [PMID: 26940070 DOI: 10.1016/j.canlet.2016.02.032]
- 77 **Nam KT, Lee HJ, Sousa JF, Weis VG, O'Neal RL, Finke PE, Romero-Gallo J, Shi G, Mills JC, Peek RM Jr, Konieczny SF, Goldenring JR.** Mature chief cells are cryptic progenitors for metaplasia in the stomach. *Gastroenterology* 2010; **139**: 2028-2037.e9 [PMID: 20854822 DOI: 10.1053/j.gastro.2010.09.005]
- 78 **Fujii Y, Yoshihashi K, Suzuki H, Tsutsumi S, Mutoh H, Maeda S, Yamagata Y, Seto Y, Aburatani H, Hatakeyama M.** CDX1 confers intestinal phenotype on gastric epithelial cells via induction of stemness-associated reprogramming factors SALL4 and KLF5. *Proc Natl Acad Sci USA*; **109**: 20584-20589 [PMID: 23112162 DOI: 10.1073/pnas.1208651109]
- 79 **Wan J, Zhou J, Zhao H, Wang M, Wei Z, Gao H, Wang Y, Cui H.** Sonic hedgehog pathway contributes to gastric cancer cell growth and proliferation. *Biores Open Access* 2014; **3**: 53-59 [PMID: 24804165 DOI: 10.1089/biores.2014.0001]
- 80 **Song Z, Yue W, Wei B, Wang N, Li T, Guan L, Shi S, Zeng Q, Pei X, Chen L.** Sonic hedgehog pathway is essential for maintenance of cancer stem-like cells in human gastric cancer. *PLoS One* 2011; **6**: e17687 [PMID: 21394208 DOI: 10.1371/journal.pone.0017687]
- 81 **Zhao X, Wang F, Hou M.** Expression of stem cell markers nanog and PSCA in gastric cancer and its significance. *Oncol Lett* 2016; **11**: 442-448 [PMID: 26870231 DOI: 10.3892/ol.2015.3884]
- 82 **Gong S, Li Q, Jeter CR, Fan Q, Tang DG, Liu B.** Regulation of NANOG in cancer cells. *Mol Carcinog* 2015; **54**: 679-687 [PMID: 26013997 DOI: 10.1002/mc.22340]
- 83 **Aggarwal BB, Kunnumakara AB, Harikumar KB, Gupta SR, Tharakan ST, Koca C, Dey S, Sung B.** Signal transducer and activator of transcription-3, inflammation, and cancer: how intimate is the relationship? *Ann N Y Acad Sci* 2009; **1171**: 59-76 [PMID: 19723038 DOI: 10.1111/j.1749-6632.2009.04911.x]
- 84 **Jiang YX, Yang SW, Li PA, Luo X, Li ZY, Hao YX, Yu PW.** The promotion of the transformation of quiescent gastric cancer stem cells by IL-17 and the underlying mechanisms. *Oncogene* 2017; **36**: 1256-1264 [PMID: 27524415 DOI: 10.1038/ncr.2016.291]
- 85 **Mohammed MK, Shao C, Wang J, Wei Q, Wang X, Collier Z, Tang S, Liu H, Zhang F, Huang J, Guo D, Lu M, Liu F, Liu J, Ma C, Shi LL, Athiviraham A, He TC, Lee MJ.** Wnt/β-catenin signaling plays an ever-expanding role in stem cell self-renewal, tumorigenesis and cancer chemoresistance. *Genes Dis* 2016; **3**: 11-40 [PMID: 27077077 DOI: 10.1016/j.gendis.2015.12.004]
- 86 **Oshima H, Matsunaga A, Fujimura T, Tsukamoto T, Taketo MM, Oshima M.** Carcinogenesis in mouse stomach by simultaneous activation of the Wnt signaling and prostaglandin E2 pathway. *Gastroenterology* 2006; **131**: 1086-1095 [PMID: 17030179 DOI: 10.1053/j.gastro.2006.07.014]
- 87 **Mao J, Fan S, Ma W, Fan P, Wang B, Zhang J, Wang H, Tang B, Zhang Q, Yu X, Wang L, Song B, Li L.** Roles of Wnt/β-catenin signaling in the gastric cancer stem cells proliferation and salinomycin treatment. *Cell Death Dis* 2014; **5**: e1039 [PMID: 24481453 DOI: 10.1038/cddis.2013.515]
- 88 **Hasegawa T, Yashiro M, Nishii T, Matsuoka J, Fuyuhiko Y, Morisaki T, Fukuoka T, Shimizu K, Shimizu T, Miwa A, Hirakawa K.** Cancer-associated fibroblasts might sustain the stemness of scirrhous gastric cancer cells via transforming growth factor-β signaling. *Int J Cancer* 2014; **134**: 1785-1795 [PMID: 24155219 DOI: 10.1002/ijc.28520]
- 89 **Fu Y, Li H, Hao X.** The self-renewal signaling pathways utilized by gastric cancer stem cells. *Tumour Biol* 2017; **39**: 1010428317697577 [PMID: 28378630 DOI: 10.1177/1010428317697577]
- 90 **Demitrack ES, Samuelson LC.** Notch as a Driver of Gastric Epithelial Cell Proliferation. *Cell Mol Gastroenterol Hepatol* 2017; **3**:

- 323-330 [PMID: 28462374 DOI: 10.1016/j.jcmgh.2017.01.012]
- 91 **Toh TB**, Lim JJ, Chow EK. Epigenetics in cancer stem cells. *Mol Cancer* 2017; **16**: 29 [PMID: 28148257 DOI: 10.1186/s12943-017-0596-9]
- 92 **Kubota E**, Kataoka H, Aoyama M, Mizoshita T, Mori Y, Shimura T, Tanaka M, Sasaki M, Takahashi S, Asai K, Joh T. Role of ES cell-expressed Ras (ERas) in tumorigenicity of gastric cancer. *Am J Pathol* 2010; **177**: 955-963 [PMID: 20566745 DOI: 10.2353/ajpath.2010.091056]
- 93 **Yashiro M**, Yasuda K, Nishii T, Kaizaki R, Sawada T, Ohira M, Hirakawa K. Epigenetic regulation of the embryonic oncogene ERas in gastric cancer cells. *Int J Oncol* 2009; **35**: 997-1003 [PMID: 19787253 DOI: 10.3892/ijo.00000414]
- 94 **Tomita H**, Takaishi S, Menhennott TR, Yang X, Shibata W, Jin G, Betz KS, Kawakami K, Minamoto T, Tomasetto C, Rio MC, Lerkowit N, Varro A, Giraud AS, Wang TC. Inhibition of gastric carcinogenesis by the hormone gastrin is mediated by suppression of TFF1 epigenetic silencing. *Gastroenterology* 2011; **140**: 879-891 [PMID: 21111741 DOI: 10.1053/j.gastro.2010.11.037]
- 95 **Yoda Y**, Takeshima H, Niwa T, Kim JG, Ando T, Kushima R, Sugiyama T, Katai H, Noshiro H, Ushijima T. Integrated analysis of cancer-related pathways affected by genetic and epigenetic alterations in gastric cancer. *Gastric Cancer* 2015; **18**: 65-76 [PMID: 24510342 DOI: 10.1007/s10120-014-0348-0]
- 96 **Wang LH**, Choi YL, Hua XY, Shin YK, Song YJ, Youn SJ, Yun HY, Park SM, Kim WJ, Kim HJ, Choi JS, Kim SH. Increased expression of sonic hedgehog and altered methylation of its promoter region in gastric cancer and its related lesions. *Mod Pathol* 2006; **19**: 675-683 [PMID: 16528374 DOI: 10.1038/modpathol.3800573]
- 97 **Yu D**, Shin HS, Lee YS, Lee YC. miR-106b modulates cancer stem cell characteristics through TGF- $\beta$ /Smad signaling in CD44-positive gastric cancer cells. *Lab Invest* 2014; **94**: 1370-1381 [PMID: 25286029 DOI: 10.1038/labinvest.2014.125]
- 98 **Wu K**, Ma L, Zhu J. miR-483-5p promotes growth, invasion and self-renewal of gastric cancer stem cells by Wnt/ $\beta$ -catenin signaling. *Mol Med Rep* 2016; **14**: 3421-3428 [PMID: 27511210 DOI: 10.3892/mmr.2016.5603]
- 99 **Ye J**, Wu D, Wu P, Chen Z, Huang J. The cancer stem cell niche: cross talk between cancer stem cells and their microenvironment. *Tumour Biol* 2014; **35**: 3945-3951 [PMID: 24420150 DOI: 10.1007/s13277-013-1561-x]
- 100 **Korkaya H**, Liu S, Wicha MS. Breast cancer stem cells, cytokine networks, and the tumor microenvironment. *J Clin Invest* 2011; **121**: 3804-3809 [PMID: 21965337 DOI: 10.1172/JCI57099]
- 101 **Quante M**, Tu SP, Tomita H, Gonda T, Wang SS, Takashi S, Baik GH, Shibata W, Diprete B, Betz KS, Friedman R, Varro A, Tycko B, Wang TC. Bone marrow-derived myofibroblasts contribute to the mesenchymal stem cell niche and promote tumor growth. *Cancer Cell* 2011; **19**: 257-272 [PMID: 21316604 DOI: 10.1016/j.ccr.2011.01.020]
- 102 **Zhu L**, Cheng X, Shi J, Jiacheng L, Chen G, Jin H, Liu AB, Pyo H, Ye J, Zhu Y, Wang H, Chen H, Fang J, Cai L, Wang TC, Yang CS, Tu SP. Crosstalk between bone marrow-derived myofibroblasts and gastric cancer cells regulates cancer stemness and promotes tumorigenesis. *Oncogene* 2016; **35**: 5388-5399 [PMID: 27109105 DOI: 10.1038/ncr.2016.76]
- 103 **Li Z**, Rich JN. Hypoxia and hypoxia inducible factors in cancer stem cell maintenance. *Curr Top Microbiol Immunol* 2010; **345**: 21-30 [PMID: 20582533 DOI: 10.1007/82\_2010\_75]
- 104 **Yang SW**, Zhang ZG, Hao YX, Zhao YL, Qian F, Shi Y, Li PA, Liu CY, Yu PW. HIF-1 $\alpha$  induces the epithelial-mesenchymal transition in gastric cancer stem cells through the Snail pathway. *Oncotarget* 2017; **8**: 9535-9545 [PMID: 28076840 DOI: 10.18632/oncotarget.14484]
- 105 **Matsumoto K**, Arai T, Tanaka K, Kaneda H, Kudo K, Fujita Y, Tamura D, Aomatsu K, Tamura T, Yamada Y, Saijo N, Nishio K. mTOR signal and hypoxia-inducible factor-1  $\alpha$  regulate CD133 expression in cancer cells. *Cancer Res* 2009; **69**: 7160-7164 [PMID: 19738050 DOI: 10.1158/0008-5472.CAN-09-1289]
- 106 **Abdullah LN**, Chow EK. Mechanisms of chemoresistance in cancer stem cells. *Clin Transl Med* 2013; **2**: 3 [PMID: 23369605 DOI: 10.1186/2001-1326-2-3]
- 107 **Choi CH**. ABC transporters as multidrug resistance mechanisms and the development of chemosensitizers for their reversal. *Cancer Cell Int* 2005; **5**: 30 [PMID: 16202168 DOI: 10.1186/1475-2867-5-30]
- 108 **Saini V**, Shoemaker RH. Potential for therapeutic targeting of tumor stem cells. *Cancer Sci* 2010; **101**: 16-21 [PMID: 19860850 DOI: 10.1111/j.1349-7006.2009.01371.x]
- 109 **Shang Z**, Xu Y, Liang W, Liang K, Hu X, Wang L, Zou Z, Ma Y. Isolation of cancer progenitor cells from cancer stem cells in gastric cancer. *Mol Med Rep* 2017; **15**: 3637-3643 [PMID: 28393208 DOI: 10.3892/mmr.2017.6423]
- 110 **Katoh M**. Networking of WNT, FGF, Notch, BMP, and Hedgehog signaling pathways during carcinogenesis. *Stem Cell Rev* 2007; **3**: 30-38 [PMID: 17873379 DOI: 10.1007/s12015-007-0006-6]
- 111 **Tamada M**, Nagano O, Tateyama S, Ohmura M, Yae T, Ishimoto T, Sugihara E, Onishi N, Yamamoto T, Yanagawa H, Suematsu M, Saya H. Modulation of glucose metabolism by CD44 contributes to antioxidant status and drug resistance in cancer cells. *Cancer Res* 2012; **72**: 1438-1448 [PMID: 22293754 DOI: 10.1158/0008-5472.CAN-11-3024]
- 112 **Ishimoto T**, Sugihara H, Watanabe M, Sawayama H, Iwatsuki M, Baba Y, Okabe H, Hidaka K, Yokoyama N, Miyake K, Yoshikawa M, Nagano O, Komohara Y, Takeya M, Saya H, Baba H. Macrophage-derived reactive oxygen species suppress miR-328 targeting CD44 in cancer cells and promote redox adaptation. *Carcinogenesis* 2014; **35**: 1003-1011 [PMID: 24318997 DOI: 10.1093/carcin/bgt402]
- 113 **Bissell MJ**, Labarge MA. Context, tissue plasticity, and cancer: are tumor stem cells also regulated by the microenvironment? *Cancer Cell* 2005; **7**: 17-23 [PMID: 15652746 DOI: 10.1016/j.ccr.2004.12.013]
- 114 **Stojnev S**, Krstic M, Ristic-Petrovic A, Stefanovic V, Hattori T. Gastric cancer stem cells: therapeutic targets. *Gastric Cancer* 2014; **17**: 13-25 [PMID: 23563919 DOI: 10.1007/s10120-013-0254-x]
- 115 **Han ME**, Baek SJ, Kim SY, Kang CD, Oh SO. ATOH1 Can Regulate the Tumorigenicity of Gastric Cancer Cells by Inducing the Differentiation of Cancer Stem Cells. *PLoS One* 2015; **10**: e0126085 [PMID: 25950549 DOI: 10.1371/journal.pone.0126085]
- 116 **Zieker D**, Bühler S, Ustündag S, Königsrainer J, Manncke S, Bajaeifer K, Vollmer J, Fend F, Northoff H, Königsrainer A, Glatzle J. Induction of tumor stem cell differentiation--novel strategy to overcome therapy resistance in gastric cancer. *Langenbecks Arch Surg* 2013; **398**: 603-608 [PMID: 23412594 DOI: 10.1007/s00423-013-1058-5]
- 117 **Chen JK**, Taipale J, Cooper MK, Beachy PA. Inhibition of Hedgehog signaling by direct binding of cyclopamine to Smoothened. *Genes Dev* 2002; **16**: 2743-2748 [PMID: 12414725 DOI: 10.1101/gad.1025302]
- 118 **Brinkhuizen T**, Reinders MG, van Geel M, Hendriksen AJ, Paulussen AD, Winnepenninckx VJ, Keymeulen KB, Soetekouw PM, van Steensel MA, Mosterd K. Acquired resistance to the Hedgehog pathway inhibitor vismodegib due to smoothened mutations in treatment of locally advanced basal cell carcinoma. *J Am Acad Dermatol* 2014; **71**: 1005-1008 [PMID: 25199678 DOI: 10.1016/j.jaad.2014.08.001]
- 119 **Yoon C**, Park DJ, Schmidt B, Thomas NJ, Lee HJ, Kim TS, Janjigian YY, Cohen DJ, Yoon SS. CD44 expression denotes a subpopulation of gastric cancer cells in which Hedgehog signaling promotes chemotherapy resistance. *Clin Cancer Res* 2014; **20**: 3974-3988 [PMID: 24947926 DOI: 10.1158/1078-0432.CCR-14-0011]
- 120 **Gupta PB**, Onder TT, Jiang G, Tao K, Kuperwasser C, Weinberg RA, Lander ES. Identification of selective inhibitors of cancer stem cells by high-throughput screening. *Cell* 2009; **138**: 645-659 [PMID: 19682730 DOI: 10.1016/j.cell.2009.06.034]
- 121 **Zhi QM**, Chen XH, Ji J, Zhang JN, Li JF, Cai Q, Liu BY, Gu QL, Zhu ZG, Yu YY. Salinomycin can effectively kill ALDH(high) stem-like cells on gastric cancer. *Biomed Pharmacother* 2011; **65**: 509-515 [PMID: 21996439 DOI: 10.1016/j.biopha.2011.06.006]
- 122 **Liu Y**, Chen H, Zheng P, Zheng Y, Luo Q, Xie G, Ma Y, Shen L. ICG-001 suppresses growth of gastric cancer cells and reduces chemoresistance of cancer stem cell-like population. *J Exp Clin Cancer Res* 2017; **36**: 125 [PMID: 28893318 DOI: 10.1186/

- s13046-017-0595-0]
- 123 **Sonbol MB**, Bekaii-Saab T. A clinical trial protocol paper discussing the BRIGHTER study. *Future Oncol* 2018; **14**: 901-906 [PMID: 29297698 DOI: 10.2217/fon-2017-0406]
  - 124 **Peng H**, Dong Z, Qi J, Yang Y, Liu Y, Li Z, Xu J, Zhang JT. A novel two mode-acting inhibitor of ABCG2-mediated multidrug transport and resistance in cancer chemotherapy. *PLoS One* 2009; **4**: e5676 [PMID: 19479068 DOI: 10.1371/journal.pone.0005676]
  - 125 **Peng H**, Qi J, Dong Z, Zhang JT. Dynamic vs static ABCG2 inhibitors to sensitize drug resistant cancer cells. *PLoS One* 2010; **5**: e15276 [PMID: 21151870 DOI: 10.1371/journal.pone.0015276]
  - 126 **Lou H**, Dean M. Targeted therapy for cancer stem cells: the patched pathway and ABC transporters. *Oncogene* 2007; **26**: 1357-1360 [PMID: 17322922 DOI: 10.1038/sj.onc.1210200]
  - 127 **Nishikawa S**, Konno M, Hamabe A, Hasegawa S, Kano Y, Fukusumi T, Satoh T, Takiguchi S, Mori M, Doki Y, Ishii H. Surgically resected human tumors reveal the biological significance of the gastric cancer stem cell markers CD44 and CD26. *Oncol Lett* 2015; **9**: 2361-2367 [PMID: 26137071 DOI: 10.3892/ol.2015.3063]
  - 128 **Zhang X**, Hua R, Wang X, Huang M, Gan L, Wu Z, Zhang J, Wang H, Cheng Y, Li J, Guo W. Identification of stem-like cells and clinical significance of candidate stem cell markers in gastric cancer. *Oncotarget* 2016; **7**: 9815-9831 [PMID: 26769843 DOI: 10.18632/oncotarget.6890]
  - 129 **Liu J**, Ma L, Xu J, Liu C, Zhang J, Liu J, Chen R, Zhou Y. Spheroid body-forming cells in the human gastric cancer cell line MKN-45 possess cancer stem cell properties. *Int J Oncol* 2013; **42**: 453-459 [PMID: 23229446 DOI: 10.3892/ijo.2012.1720]
  - 130 **Shitara K**, Doi T, Nagano O, Imamura CK, Ozeki T, Ishii Y, Tsuchihashi K, Takahashi S, Nakajima TE, Hironaka S, Fukutani M, Hasegawa H, Nomura S, Sato A, Einaga Y, Kuwata T, Saya H, Ohtsu A. Dose-escalation study for the targeting of CD44v+ cancer stem cells by sulfasalazine in patients with advanced gastric cancer (EPOC1205). *Gastric Cancer* 2017; **20**: 341-349 [PMID: 27055559 DOI: 10.1007/s10120-016-0610-8]
  - 131 **Heiss MM**, Murawa P, Koralewski P, Kutarska E, Kolesnik OO, Ivanchenko VV, Dudnichenko AS, Aleknaviciene B, Razbadauskas A, Gore M, Ganea-Motan E, Ciuleanu T, Wimberger P, Schmittl A, Schmalfeldt B, Burges A, Bokemeyer C, Lindhofer H, Lahr A, Parsons SL. The trifunctional antibody catumaxomab for the treatment of malignant ascites due to epithelial cancer: Results of a prospective randomized phase II/III trial. *Int J Cancer* 2010; **127**: 2209-2221 [PMID: 20473913 DOI: 10.1002/ijc.25423]
  - 132 **Goéré D**, Gras-Chaput N, Aupérin A, Flament C, Mariette C, Glehen O, Zitvogel L, Elias D. Treatment of gastric peritoneal carcinomatosis by combining complete surgical resection of lesions and intraperitoneal immunotherapy using catumaxomab. *BMC Cancer* 2014; **14**: 148 [PMID: 24589307 DOI: 10.1186/1471-2407-14-148]
  - 133 **Doshi S**, Gisleskog PO, Zhang Y, Zhu M, Oliner KS, Loh E, Perez Ruixo JJ. Rilotumumab exposure-response relationship in patients with advanced or metastatic gastric cancer. *Clin Cancer Res* 2015; **21**: 2453-2461 [PMID: 25712685 DOI: 10.1158/1078-0432.CCR-14-1661]
  - 134 **Shah MA**, Bang YJ, Lordick F, Alsina M, Chen M, Hack SP, Bruey JM, Smith D, McCaffery I, Shames DS, Phan S, Cunningham D. Effect of Fluorouracil, Leucovorin, and Oxaliplatin With or Without Onartuzumab in HER2-Negative, MET-Positive Gastroesophageal Adenocarcinoma: The METGastric Randomized Clinical Trial. *JAMA Oncol* 2017; **3**: 620-627 [PMID: 27918764 DOI: 10.1001/jamaoncol.2016.5580]
  - 135 **Pant S**, Patel MR, Kurkjian C, Hemphill MB, Flores MRC, Thompson DS. A phase II study of the c-Met inhibitor tivantinib (tiv) in combination with FOLFOX for the treatment of patients (pts) with previously untreated metastatic adenocarcinoma of the distal esophagus, gastroesophageal (GE) junction, or stomach. *J Clin Oncol* 2015; **33**: 4065
  - 136 **Cohen DJ**, Christos PJ, Kindler HL, Catenacci DVT, Bekaii-Saab TB, Tahiri S. Vismodegib (V), a hedgehog (HH) pathway inhibitor, combined with FOLFOX for first-line therapy of patients (pts) with advanced gastric and gastroesophageal junction (GEJ) carcinoma: A New York Cancer Consortium led phase II randomized study. *J Clin Oncol* 2013; **31**: 4011 [DOI: 10.1200/jco.2013.31.4\_suppl.67]
  - 137 **Shah MA**, Muro K, Shitara K, Tebbutt NC, Bang Y-J, Lordick F. The BRIGHTER trial: A phase III randomized double-blind study of BBI608 + weekly paclitaxel versus placebo (PBO) + weekly paclitaxel in patients (pts) with pretreated advanced gastric and gastro-esophageal junction (GEJ) adenocarcinoma. *J Clin Oncol* 2015; **33**: TPS4139

**P- Reviewer:** Chueh PJ, Kim SY **S- Editor:** Wang XJ  
**L- Editor:** Filipodia **E- Editor:** Yin SY





Published by **Baishideng Publishing Group Inc**  
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA  
Telephone: +1-925-223-8242  
Fax: +1-925-223-8243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>



ISSN 1007-9327

