

# The potential mechanism of mitochondrial dysfunction in septic cardiomyopathy

Journal of International Medical Research

2018, Vol. 46(6) 2157–2169

© The Author(s) 2018

Reprints and permissions:

[sagepub.co.uk/journalsPermissions.nav](http://sagepub.co.uk/journalsPermissions.nav)

DOI: 10.1177/0300060518765896

[journals.sagepub.com/home/imr](http://journals.sagepub.com/home/imr)**Pan Pan, Xiaoting Wang and Dawei Liu**

## Abstract

Septic cardiomyopathy is one of the most serious complications of sepsis or septic shock. Basic and clinical research has studied the mechanism of cardiac dysfunction for more than five decades. It has become clear that myocardial depression is not related to hypoperfusion. As the heart is highly dependent on abundant adenosine triphosphate (ATP) levels to maintain its contraction and diastolic function, impaired mitochondrial function is lethally detrimental to the heart. Research has shown that mitochondria play an important role in organ damage during sepsis. The mitochondria-related mechanisms in septic cardiomyopathy have been discussed in terms of restoring mitochondrial function. Mitochondrial uncoupling proteins located in the mitochondrial inner membrane can promote proton leakage across the mitochondrial inner membrane. Recent studies have demonstrated that proton leakage is the essential regulator of mitochondrial membrane potential and the generation of reactive oxygen species (ROS) and ATP. Other mechanisms involved in septic cardiomyopathy include mitochondrial ROS production and oxidative stress, mitochondria  $\text{Ca}^{2+}$  handling, mitochondrial DNA in sepsis, mitochondrial fission and fusion, mitochondrial biogenesis, mitochondrial gene regulation and mitochondria autophagy. This review will provide an overview of recent insights into the factors contributing to septic cardiomyopathy.

## Keywords

Sepsis, cardiomyopathy, mitochondrial, uncoupling proteins

Date received: 11 November 2017; accepted: 27 February 2018

---

Department of Critical Care Medicine, Peking Union Medical College Hospital, Peking Union Medical College & Chinese Academy of Medical Sciences, Beijing, China

---

## Corresponding author:

Dawei Liu, Department of Critical Care Medicine, Peking Union Medical College Hospital, 1 Shuaifuyuan Road, Dongcheng District, Beijing 100730, China.

Email: [dwliu98@163.com](mailto:dwliu98@163.com)

Xiaoting Wang, Department of Critical Care Medicine, Peking Union Medical College Hospital, 1 Shuaifuyuan Road, Dongcheng District, Beijing 100730, China.

Email: [icuting@163.com](mailto:icuting@163.com)



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<http://www.creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

## Introduction

Sepsis is defined as an immune and inflammatory response that is capable of inducing multi-organ dysfunction.<sup>1</sup> Recent data indicate that mortality rate from sepsis or septic shock is approximately 40% in intensive care units (ICU).<sup>2</sup> It is the main cause of death among patients in hospital and the mortality is as high as 28.7% despite progress having been made in its treatment.<sup>2,3</sup> In the US, the mortality rate due to sepsis is higher than that from prostate cancer, breast cancer and AIDS combined, and this figure is growing annually.<sup>4-6</sup> The high morbidity and mortality associated with sepsis and septic shock make them the 10th most common cause of death in the US.<sup>7</sup> An important organ system frequently involved in sepsis is the cardiovascular system, with septic cardiomyopathy being one of the most serious complications associated with sepsis or septic shock, which may progress to left and right heart systolic and diastolic failure.<sup>8</sup> Basic and clinical research has studied its mechanism of dysfunction for more than five decades.

In 1951, a hyperdynamic state was identified in patients with sepsis or septic shock, which was the first cardiovascular event shown to be caused by sepsis.<sup>9</sup> In subsequent famous studies, fluid therapy was attempted in these patients and appropriate and sufficient volume resuscitation was demonstrated to be one of the most effective therapeutic treatments for sepsis.<sup>10</sup> However, in the mid-1980s, many clinicians found that some septic patients had normal or even a slightly higher cardiac output with descended ejection fraction and stroke volume.<sup>11</sup> It was also shown that septic patients with cardiovascular dysfunction had a higher mortality rate than those without cardiovascular dysfunction.<sup>12</sup> From then on, researchers paid more and more attention to septic cardiomyopathy and

attempted to clarify the mechanism of this critical manifestation.

A number of studies identified that myocardial depression was not related to hypoperfusion as an adequate oxygen supply had already been proved in experiments on human and animals, while a circulating depressant factor in septic shock, which was first proposed fifty years ago,<sup>13</sup> must play an important role in heart dysfunction.<sup>14</sup> Other mechanisms like mitochondrial dysfunction/apoptosis, cellular damage, cell signalling, autonomic dysfunction, decreased coronary blood flow, increased heat shock protein or adhesion molecules and myocardial hibernation phenomenon were proposed with the development of scientific theory and technology.<sup>15</sup> Nevertheless, an increasing number of studies had focused on myocardial energy metabolism as cells, which seemed unable to maintain proper metabolism in septic patients.<sup>16-18</sup> Consequently, this imbalance led to energy failure and even death.<sup>19</sup> It was demonstrated that cardiomyocyte injury occurred in sepsis-induced cardiac dysfunction, but there was almost no cell death.<sup>20</sup> It is particularly important to find the correct treatment to repair cell function. More specifically, many researchers identified mitochondrial dysfunction as the key pathological change in septic cardiomyopathy.<sup>21,22</sup> Since the heart is one of the organs that is highly dependent on abundant adenosine triphosphate (ATP) levels to maintain its contraction and diastolic function, impaired mitochondrial function is lethally detrimental to the heart. According to cardiac pathophysiology, energy depletion resulting from mitochondrial dysfunction would contribute to significant myocardial damage,<sup>23</sup> such as diabetic cardiomyopathy, ischaemic reperfusion injury and heart failure.<sup>24,25</sup> A series of mitochondria-related mechanisms in septic cardiomyopathy have been explored in order to find a way to restore mitochondrial function. This

review will provide an overview of recent insights into the factors contributing to septic cardiomyopathy.

### **Mechanism of mitochondrial dysfunction in septic cardiomyopathy**

Mitochondrial dysfunction in sepsis can cause continuous damage to cells and organs. Reactive oxygen species (ROS) like superoxide are increased during sepsis and evoke oxidative and nitrosative injury.<sup>26,27</sup> Increased superoxide can result in the inhibition of oxidative phosphorylation complexes, decreased O<sub>2</sub> consumption and mitochondrial membrane potential ( $\Delta\Psi$ ).<sup>28</sup> As a consequence, the release of ROS increases.<sup>28</sup> In addition, the increased levels of uncoupling proteins increase the extent of proton leakage.<sup>29</sup> Mitochondrial permeability transition pores (mPTP) open due to increased Ca<sup>2+</sup>.<sup>30</sup> Hence, oxidative damage happens to the inner mitochondrial membrane. In addition, inappropriate mitochondrial autophagy (mitophagy) leads to the reduction of mitochondrial mass and dysfunctional mitochondria.<sup>31</sup> In summary, ATP regeneration is a complex process, and once the heart lacks energy, damage to cardiac function may follow.

### **Mitochondrial ROS production and oxidative stress**

Complexes I and III of the respiratory chain in mitochondria produce small amounts of ROS physiologically.<sup>32</sup> However, sepsis is a disease accompanied by increased oxidative stress, and large amounts of ROS come from activated neutrophils.<sup>33</sup> In addition, azotized stress leads to the oxidation of xanthine and its reactive nitrite increased in plasma. While in serum, the antioxidant capacity is diluted because of decreased levels of antioxidants like

vitamin C, vitamin E, unconjugated bilirubin, uric acid, and other unknown factors.<sup>34</sup> Intracellularly, the concentration of oxidized glutathione dimers increase while amounts of glutathione drop.<sup>35</sup> A large body of evidence strongly suggests that ROS and reactive nitrogen species led to specific impairments of oxidative phosphorylation in the septic myocardium (e.g. complex I, complex IV, F0F1 dysfunctions),<sup>36,37</sup> especially myocardial cell mitochondria. Research has demonstrated that the activity of inducible mitochondrial nitric oxide synthase was obviously increased in a septic mouse model, which led to the growth of peroxynitrite ONOO<sup>-</sup>.<sup>38</sup> It is clear that nitric oxide (NO) and its derivatives play an indisputable role in the regulation of cardiovascular function and vascular tone.<sup>39</sup> Research has shown that ONOO<sup>-</sup> has a negative influence on myocardial mitochondrial dysfunction in sepsis.<sup>38</sup> Nevertheless, we should realize that there is a casual relationship between NO and heart function, since NO is not only produced by cardiac mitochondria but it is also found in other intracellular locations and it is produced in different cell types. To prevent and alleviate oxidative damage, researchers have been searching for suitable antioxidants, such as mitochondria-targeted vitamin E,<sup>40</sup> mitochondria-targeted antioxidant MitoQ,<sup>41</sup> and vitamin C.<sup>42</sup> These antioxidants represent an attractive treatment for mitochondrial injury. However, in order to gain wider acceptance, more clinical experiments are needed to confirm the practical use of antioxidants.

### **Mitochondrial Ca<sup>2+</sup> handling**

One of the most important steps in regenerating ATP is building up a proton gradient that is dependent on the impermeability of the inner mitochondrial membrane (IMM). Evidence shows that the electron transport chain that pumps protons to maintain the chemiosmotic energy gradient

is based on the impermeability of the IMM. Although mPTP is formed due to a sudden change in the IMM and the influence of other substances, as it is located in the inner mitochondrial membrane it is mainly mediated by  $\text{Ca}^{2+}$ .<sup>43</sup> During sepsis, mitochondrial  $\text{Ca}^{2+}$  content is raised by increased  $\text{Ca}^{2+}$  leaking from the sarcoplasmic reticulum and decreased  $\text{Ca}^{2+}$  uptake into the same organelle.<sup>44</sup> Researchers found that inhibition of nicotinamide adenine dinucleotide phosphate oxidase 2 preserved intracellular calcium handling, mitochondrial function and played a protective role in sepsis-induced cardiomyopathy.<sup>45</sup>  $\text{Ca}^{2+}$  overload can result in the mPTPs opening and becomes the trigger of sequential pathological changes; and for myocardial cells, mPTP opening can lead to activation of caspase proteins, and ultimately to cardiomyocyte contractile dysfunction.<sup>46,47</sup> For the heart in sepsis, the abnormal transport of calcium is an important factor affecting heart function.<sup>48</sup> An animal study showed that cytokines like tumour necrosis factor- $\alpha$  and interleukin-1 $\beta$  were released and affected calcium leakage.<sup>49</sup> This disorder proved that sepsis can weaken cardiomyocyte contractility in isolated rat heart model,<sup>49</sup> and there was no difference in the damage between right and left ventricles.<sup>50</sup> Prevention of mPTP opening through decreased calcium leakage can reduce the activation of cytochrome c release.<sup>51</sup>

### Mitochondrial DNA in sepsis

With the development of molecular biology, researchers now recognize the role of pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) in the occurrence and development of disease. Among the known DAMPs, mitochondrial DNA (mtDNA) has become the focus of considerable research.<sup>52</sup> MtDNA is a circular

molecule that encodes the key proteins involved in the oxidative phosphorylation system.<sup>53</sup> In addition to its coding function, mtDNA is also involved in cellular immune functions.<sup>54</sup> Like bacteria, mtDNA is a component that can be recognized as a DAMP by the immune system and triggers or promotes a series of defence reactions.<sup>55</sup> Multiple *in vivo* and *in vitro* studies have demonstrated that mtDNA can be transferred from mitochondria to the cytosol via mPTPs, and thus any pathological changes leading mPTP opening will increase the leakage of mtDNA.<sup>56,57</sup> In 2013, the first study of mtDNA in ICU patients found that the levels of circulating mtDNA were significantly higher in non-survivors than survivors.<sup>58</sup> Subsequently, another study found that plasma mtDNA levels in patients with sepsis was greater than in healthy controls.<sup>59</sup> Consequently, the authors demonstrated via an *in vivo* experiment that the high concentration of mtDNA was able to increase neutrophil viability.<sup>59</sup> However, delayed neutrophils apoptosis and local accumulation were associated the poor outcome in patients with sepsis.<sup>59</sup>

### Mitochondrial fission and fusion

It is widely known that mitochondria are hyperdynamic organelles and that their morphology is inextricably linked to their function.<sup>60</sup> Fission and fusion are the determinative factors in mitochondrial morphology. Balanced and proper mitochondrial membrane fission and fusion support the reliable production of mitochondria, while abnormal morphology cannot meet the metabolic demands.<sup>61,62</sup> Usually, the changes of structures caused by the fusion/fission processes are observed within 24 h.<sup>63</sup> Very recent research has demonstrated that proper mitochondrial fusion and fission can regulate mitochondrial function and maintain heart development.<sup>64</sup>

Different inner or outer membrane fusion and fission depends on proteins encoded by different genes (outer membrane fusion: mitofusin-1 and mitofusin-2 [*MFN1* and *MFN2* genes], phospholipase D family member 6 [mitoPLD; *PLD6* gene]; inner membrane fusion: mitochondrial dynamin like GTPase [*L-OPA1* gene]; outer membrane fission: death associated protein kinase 2 (*DAPK2* gene, also known as *DRP1*); inner membrane fission: mitochondrial dynamin like GTPase [*S-OPA1* gene], mitochondrial fission process 1 [*MTFP1* gene, also known as *MTP18*). In a septic mouse model, scientists found that thioredoxin 1 overexpression can alter the ultrastructure in the mitochondrial cristae accompanied by increased expression of mitochondrial dynamin like GTPase (*OPA1*) gene and activation of the dynamin 1-like (*DNM1L* gene, also known as *DRP1*) gene.<sup>65</sup> In addition, it has been proved that the inhibition of unbalanced mitochondrial fission through the inhibitor Mdivi-1 can protect organs function in endotoxaemia.<sup>66</sup>

### Mitochondrial biogenesis

In addition to mitochondrial fission and fusion, mitochondrial biogenesis is the other main component of the mitochondrial mass control system. Physiologically, creation of new and healthy mitochondria in terms of biogenesis is important to meet cellular metabolic energy demands.<sup>67</sup> It has been reported that mitochondrial biogenesis may partially counteract mitochondrial protein depletion, helping to maintain functionality and energetic status in the critically ill patients.<sup>68</sup> During sepsis, excess ROS and free radical generation damage mitochondria and result in impaired mitochondrial synthesis, while biogenesis becomes decreased in early sepsis and increased in later sepsis.<sup>32</sup> Endotoxin causes the activation of oestrogen-related receptor alpha,

peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 $\alpha$ ).<sup>69</sup> Mitochondrial synthesis can be affected by regulating the above substances. A study suggested that acetylcholine promoted mitochondria biogenesis via the PGC-1 $\alpha$  pathway and improved mitochondrial function.<sup>70</sup> Some other studies also have demonstrated that increased biogenesis can improve the prognosis in sepsis, and the inhibition of biogenesis can increase mortality.<sup>71,72</sup> However, it appears that redundant biogenesis can aggravate mitochondrial function. For example, a study demonstrated that the overexpression of PGC1- $\alpha$  resulted in an over-dose of biogenesis and led to heart failure.<sup>73</sup> Thus, we need to do further research on biogenesis in order to more clearly understand its effect on mitochondrial and organ function.

### Mitochondrial genes

One of the molecular mechanisms that occurs in mitochondria during trauma or sepsis is mitochondrial gene modification, though there are limited publications in this field. In a mouse model of haemorrhage trauma, the transcriptional profile of mitochondria genes was changed by the trauma and led to worsened heart function.<sup>74</sup> To date, no specific gene changes have been identified in cardiac muscle cells during sepsis. However, in a hepatic model, a mutation in the ATPase subunit-8 partially protected mice against endotoxaemic stress, most probably leading to better hepatic energy status despite elevated oxidative stress.<sup>75</sup> It has become popular to research the circadian rhythms and to some extent clock genes appear to control mitochondrial function. For example, disruption of the clock genes affects the immune response, which in turn induces proinflammatory mediators, leading to bioenergetic decay and formation of ROS.<sup>76</sup> More basic and clinical research is required to determine

the role of mitochondrial and clock genes in the prediction of disease development and prognosis. More importantly, evidence is required to determine whether these two types of genes can work as effective treatments for diseases.

### Mitochondrial autophagy

As discussed above, mitochondrial impairment can be lethal to cells, initiating the necrotic or apoptotic cell death pathways. The body has a series of mechanisms that it can use to correct sepsis-related organ dysfunction caused by abnormal mitochondria. Apart from generation of new and functionally normal mitochondria (biogenesis), removal of dysfunctional mitochondria is another key mechanism of organ recovery.<sup>76</sup> This removal of mitochondria via autophagy is known as mitophagy.<sup>76</sup> Damaged mitochondria are isolated by autophagosomes and ultimately degraded by fusion with lysosomes.<sup>77</sup> Morphological and biochemical evidence indicates that mitophagy is associated with recovery, suggesting that this process has something to do with cardiac recovery from sepsis.<sup>78</sup> Mitochondrial autophagy was activated during sepsis in *PARK2*-deficient mice and *PARK2* exerted additional protective roles in sepsis-induced mitochondrial and cardiac contractile dysfunction.<sup>79</sup> Many studies put mitophagy as a therapeutic target to improve heart function. Current data demonstrate that the hypophosphorylated form of I $\kappa$ B $\beta$  (an inhibitor of nuclear factor  $\kappa$ B) at Ser313 is beneficial to the heart in sepsis through enhancement of autophagy and inhibition of apoptosis.<sup>80</sup> Other research indicates that fasudil prevented lipopolysaccharide-induced heart oxidative stress by inhibiting RhoA/ROCK from activating the autophagic processes.<sup>81</sup> In addition, lysosome reformation mediated by cobalt protoporphyrin IX or transcription factor EB may be involved

in cardioprotection against lipopolysaccharide-induced septic insults, and may be a novel mechanism for protecting the heart against oxidative stress.<sup>82</sup>

### Uncoupling proteins in mitochondria

Mitochondrial uncoupling proteins (UCPs) located in the mitochondrial inner membrane can promote the leakage of protons across the mitochondrial inner membrane.<sup>29</sup> It is an essential regulator of mitochondrial membrane potential, which can disperse the mitochondrial proton gradient by translocating H<sup>+</sup> across the inner membrane, and finally influencing ATP generation.<sup>83,84</sup> Physiologically, uncoupling can decrease mitochondrial ROS production and increase heat generation.<sup>29</sup> UCPs are part of a protein family consisting of five subtypes.<sup>85</sup> The UCP molecule is composed of six hydrophobic membrane-spanning  $\alpha$ -helices, which are responsible for creating the channel within the inner membrane.<sup>86</sup> Furthermore, the  $\alpha$ -helices are arranged into three cassettes; the latter ones being connected by amino, carboxyl termini and two loops.<sup>86</sup> The loops are implicated in the control of access to the channel.<sup>86</sup> UCPs possess a binding site for purine nucleotides in order to inhibit the uncoupling activity physically.<sup>87</sup> The essential function of UCP1 is to produce the heat from brown adipose tissue (BAT) to maintain body temperature.<sup>88</sup> UCP2 through to UCP5 have been found in fungi, plants and animals.<sup>84,85</sup> All five subtypes of UCPs can be expressed in mammalian cells and have different tissue distributions.<sup>88</sup> UCP1 is mainly distributed in BAT, but is also found in other places such as white adipose tissue, pancreatic  $\beta$  cells, retinal cells and skeletal muscle.<sup>88</sup> It plays an important role in glucose metabolism.<sup>88</sup> UCP2 is the most common protein in this family, as it is

found in various tissues, such as the central nerve system, kidney, heart, liver, pancreas, spleen, thymus and macrophages.<sup>89</sup> UCP3 is mostly found in BAT and skeletal muscle, and despite of high sequence similarity with UCP1, it has no thermoregulation properties and its physiological functions remain unknown.<sup>90</sup> The *UCP4* and *UCP5* genes have less sequence similarity with *UCP1*, and they are mostly expressed in the brain.<sup>91</sup> Recently, it was hypothesized that the *UCP4* and *UCP5* genes originate from a common ancestral gene and are probably responsible for ATP transportation.<sup>91</sup>

Uncoupling protein 2 can be regulated at various levels; at the molecular level (gene, mRNA and protein transcriptional, translational, turn-over), proton conductance and by pharmacological regulation.<sup>89</sup> Recent research identified two of the most common gene polymorphisms: the promoter variant -866G>A and the codon 55 missense polymorphism.<sup>92</sup> The former polymorphism can promote higher *UCP2* mRNA expression,<sup>93</sup> while the latter polymorphism can reduce the degree of uncoupling in pathological process.<sup>94</sup> To date, the four most common transcription regulatory proteins and their relative transcription factor binding sites that are involved in the regulation of human *UCP2* transcription are the peroxisomal proliferator-activated receptors,<sup>95,96</sup> PGC-1 $\alpha$ ,<sup>97,98</sup> forkhead box protein A1,<sup>99</sup> and the SMAD family.<sup>100</sup> MicroRNAs and heterogeneous nuclear ribonucleoprotein K induce a totally new layer of protein regulation after transcription.<sup>101,102</sup> It has been found that *UCP2* expression can be modulated by different drugs. For example, adenosine monophosphate-activated protein kinase activator can up-regulate *UCP2* activation.<sup>103,104</sup> However, some chemotherapeutic drugs like doxorubicin and taxol can down-regulate *UCP2* expression and influence the cardiac function.<sup>103-106</sup>

In contrast to the single function of UCP1, UCP2 is more related to organ function as it can be found in several tissue and organs.<sup>89</sup> The wide distribution of UCP2 means that it is involved in regulating metabolism, including ROS production, food intake, glucose control, and immunity; and some pathologies, such as heart failure, diabetes, and cancer.<sup>107</sup> A number of studies highlight the importance of UCP2 in cardiovascular diseases.<sup>108,109</sup> ROS are elevated in some pathological processes, including sepsis-induced cardiomyopathy, cardiac reperfusion injury, and diabetic cardiomyopathy.<sup>110-112</sup> Redundant ROS can stimulate proton leakage, thus leading to decreased UCP2 activity and reduced generation of ROS.<sup>113</sup> UCP2 plays a protective role in the heart via this negative feedback loop.<sup>114</sup> For the human heart, modulation of UCP2 level appears increased in oxidative stress status.<sup>111</sup> Uncoupling of oxidative phosphorylation diminishes superoxide formation by complex I.<sup>115</sup> In addition, uncoupled substances might inhibit superoxide formation by complexes I and III by virtue of their antioxidant ability.<sup>116</sup>

Evidence suggests that UCP2 has a protective effect on myocardial damage and down-regulated UCP2 is associated with a failing heart.<sup>108</sup> UCP2 up-regulation attenuated ROS generation and prevented mitochondrial Ca<sup>2+</sup> overload, significantly suppressing markers of cell death.<sup>117</sup> In a UCP2 gene silencing animal model, UCP2<sup>-/-</sup> mice were more likely to have damaged mitochondrial morphology and function, suggesting UCP2 may play a protective role in cardiomyocytes under septic conditions.<sup>110</sup> Decreased membrane potential and ATP content, depletion of mtDNA and increased ROS were aggravated by silencing of UCP2.<sup>118</sup> In addition, researchers found that UCP2 had a regulatory role in the activation of p38 mitogen-activated protein kinase, nuclear factor kappa B and

the expression of downstream inflammatory mediators in H9C2 cells stimulated with septic serum.<sup>109</sup>

## Conclusions

Mitochondria account for one third of the volume of cardiomyocytes and have an important role to play in the regulation of ROS generation and ATP production, which is important for maintaining heart function and cardiomyocyte survival. Therefore, any modification of mitochondria may contribute to cardiovascular diseases. There is considerable research evidence to show that UCP2 acts as an essential protein in mitochondrial function, by decreasing ROS generation and increasing ATP production. This protective feedback loop helps organs to recover their functions after sustaining damage. More research is needed to develop new drugs or promising therapeutic approaches that could potentially be used to reverse the mitochondrial damage associated with several diseases. Further research about UCP activity and regulation could advance our understanding of myocardial depression. The increasing interest in sepsis will allow novel research tools to be used to develop effective treatments in the future.

## Declaration of conflicting interests

The authors declare that there are no conflicts of interest.

## Funding

The development of this review article was supported by funding from the National Natural Science Foundation of China (grant no. 81671878) and the Beijing Municipal Natural Science Foundation (grant no. 7162158).

## References

1. Singer M, Deutschman CS, Seymour CW, et al. The Third International Consensus

Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016; 315: 801–810.

2. Craft JA, Gordon CJ, Huether SE, et al. *Understanding Pathophysiology – ANZ adaptation*. 2nd ed. Australia: Mosby, 2014.
3. Zhou J, Qian C, Zhao M, et al. Epidemiology and outcome of severe sepsis and septic shock in intensive care units in mainland China. *PLoS One* 2014; 9: e107181.
4. Al-Ostad G, Kezouh A, Spence AR, et al. Incidence and risk factors of sepsis mortality in labor, delivery and after birth: population-based study in the USA. *J Obstet Gynaecol Res* 2015; 41: 1201–1206.
5. Ronsmans C, Graham WJ and Lancet Maternal Survival Series steering group. Maternal mortality: who, when, where, and why. *Lancet* 2006; 368: 1189–1200.
6. Angus DC, Linde-Zwirble WT, Lidicker J, et al. Epidemiology of severe sepsis in the United States: analysis of incidence, outcome, and associated costs of care. *Crit Care Med* 2001; 29: 1303–1310.
7. Martin GS, Mannino DM, Eaton S, et al. The epidemiology of sepsis in the United States from 1979 through 2000. *N Engl J Med* 2003; 348: 1546–1554.
8. Romero-Bermejo FJ, Ruiz-Bailen M, Gil-Cebrian J, et al. Sepsis-induced cardiomyopathy. *Curr Cardiol Rev* 2011; 7: 163–183.
9. Waisbren BA. Bacteremia due to gram-negative bacilli other than the Salmonella; a clinical and therapeutic study. *AMA Arch Intern Med* 1951; 88: 467–488.
10. Rivers E, Nguyen B, Havstad S, et al. Early goal-directed therapy in the treatment of severe sepsis and septic shock. *N Engl J Med* 2001; 345: 1368–1377.
11. Parker MM, Shelhamer JH, Bacharach SL, et al. Profound but reversible myocardial depression in patients with septic shock. *Ann Intern Med* 1984; 100: 483–490.
12. Parrillo JE, Parker MM, Natanson C, et al. Septic shock in humans. Advances in the understanding of pathogenesis, cardiovascular dysfunction, and therapy. *Ann Intern Med* 1990; 113: 227–242.
13. Wiggers CJ. Myocardial depression in shock; a survey of cardiodynamic studies. *Am Heart J* 1947; 33: 633–650.



14. Parrillo JE, Burch C, Shelhamer JH, et al. A circulating myocardial depressant substance in humans with septic shock. Septic shock patients with a reduced ejection fraction have a circulating factor that depresses in vitro myocardial cell performance. *J Clin Invest* 1985; 76: 1539–1553.
15. Cunnion RE, Schaer GL, Parker MM, et al. The coronary circulation in human septic shock. *Circulation* 1986; 73: 637–644.
16. Hotchkiss RS and Karl IE. Reevaluation of the role of cellular hypoxia and bioenergetic failure in sepsis. *JAMA* 1992; 267: 1503–1510.
17. Kantrow SP, Taylor DE, Carraway MS, et al. Oxidative metabolism in rat hepatocytes and mitochondria during sepsis. *Arch Biochem Biophys* 1997; 345: 278–288.
18. Du M, Chang P and Liu Z. Advance in the treatment of sepsis with mitochondria targeted antioxidants. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue* 2015; 27: 148–151 [in Chinese, English Abstract].
19. Kula R, Chylek V, Szturz P, et al. Pathogenesis of severe sepsis – from macrocirculation to mitochondria. *Klin Mikrobiol Infekc Lek* 2006; 12: 143–149 [in Czech, English Abstract].
20. Takasu O, Gaut JP, Watanabe E, et al. Mechanisms of cardiac and renal dysfunction in patients dying of sepsis. *Am J Respir Crit Care Med* 2013; 187: 509–517.
21. Jang DH, Greenwood JC, Spyras MB, et al. Measurement of Mitochondrial Respiration and Motility in Acute Care: Sepsis, Trauma, and Poisoning. *J Intensive Care Med* 2017; 32: 86–94.
22. Matkovich SJ, Al Khiami B, Efimov IR, et al. Widespread Down-Regulation of Cardiac Mitochondrial and Sarcomeric Genes in Patients With Sepsis. *Crit Care Med* 2017; 45: 407–414.
23. Drosatos K, Khan RS, Trent CM, et al. Peroxisome proliferator-activated receptor- $\gamma$  activation prevents sepsis-related cardiac dysfunction and mortality in mice. *Circ Heart Fail* 2013; 6: 550–562.
24. Bugger H and Abel ED. Mitochondria in the diabetic heart. *Cardiovasc Res* 2010; 88: 229–240.
25. Hu L, Wang J, Zhu H, et al. Ischemic post-conditioning protects the heart against ischemia-reperfusion injury via neuronal nitric oxide synthase in the sarcoplasmic reticulum and mitochondria. *Cell Death Dis* 2016; 7: e2222.
26. Potz BA, Sellke FW and Abid MR. Endothelial ROS and impaired myocardial oxygen consumption in sepsis-induced cardiac dysfunction. *J Intensive Crit Care* 2016; 2: pii: 20.
27. Yao X, Carlson D, Sun Y, et al. Mitochondrial ROS Induces Cardiac Inflammation via a Pathway through mtDNA Damage in a Pneumonia-Related Sepsis Model. *PLoS One* 2015; 10: e0139416.
28. Cimolai MC, Alvarez S, Bode C, et al. Mitochondrial Mechanisms in Septic Cardiomyopathy. *Int J Mol Sci* 2015; 16: 17763–17778.
29. Ricquier D and Bouillaud F. Mitochondrial uncoupling proteins: from mitochondria to the regulation of energy balance. *J Physiol* 2000; 529(Pt 1): 3–10.
30. Halestrap AP, McStay GP and Clarke SJ. The permeability transition pore complex: another view. *Biochimie* 2002; 84: 153–166.
31. Choi AM, Ryter SW and Levine B. Autophagy in human health and disease. *N Engl J Med* 2013; 368: 1845–1846.
32. Arulkumaran N, Deutschman CS, Pinsky MR, et al. Mitochondrial Function in Sepsis. *Shock* 2016; 45: 271–281.
33. Boueiz A and Hassoun PM. Regulation of endothelial barrier function by reactive oxygen and nitrogen species. *Microvasc Res* 2009; 77: 26–34.
34. Kayali R, Aydin S and Cakatay U. Effect of gender on main clinical chemistry parameters in aged rats. *Curr Aging Sci* 2009; 2: 67–71.
35. Wendel M and Heller AR. Mitochondrial function and dysfunction in sepsis. *Wien Med Wochenschr* 2010; 160: 118–123.
36. Alvarez S and Evelson PA. Nitric oxide and oxygen metabolism in inflammatory conditions: sepsis and exposition to polluted ambients. *Front Biosci* 2007; 12: 964–974.
37. Kanai AJ, Pearce LL, Clemens PR, et al. Identification of a neuronal nitric oxide

- synthase in isolated cardiac mitochondria using electrochemical detection. *Proc Natl Acad Sci U S A* 2001; 98: 14126–14131.
38. Escames G, Lopez LC, Ortiz F, et al. Attenuation of cardiac mitochondrial dysfunction by melatonin in septic mice. *FEBS J* 2007; 274: 2135–2147.
  39. Beltowski J and Jamroz-Wisniewska A. Hydrogen sulfide and endothelium-dependent vasorelaxation. *Molecules* 2014; 19: 21183–21199.
  40. Zang QS, Martinez B, Yao X, et al. Sepsis-induced cardiac mitochondrial dysfunction involves altered mitochondrial-localization of tyrosine kinase Src and tyrosine phosphatase SHP2. *PLoS One* 2012; 7: e43424.
  41. Dare AJ, Bolton EA, Pettigrew GJ, et al. Protection against renal ischemia-reperfusion injury in vivo by the mitochondria targeted antioxidant MitoQ. *Redox Biol* 2015; 5: 163–168.
  42. Oudemans-van Straaten HM, Spoelstra-de Man AM and de Waard MC. Vitamin C revisited. *Crit Care* 2014; 18: 460.
  43. Bernardi P and Di Lisa F. The mitochondrial permeability transition pore: molecular nature and role as a target in cardioprotection. *J Mol Cell Cardiol* 2015; 78: 100–106.
  44. Hassoun SM, Marechal X, Montaigne D, et al. Prevention of endotoxin-induced sarcoplasmic reticulum calcium leak improves mitochondrial and myocardial dysfunction. *Crit Care Med* 2008; 36: 2590–2596.
  45. Joseph LC, Kokkinaki D, Valenti MC, et al. Inhibition of NADPH oxidase 2 (NOX2) prevents sepsis-induced cardiomyopathy by improving calcium handling and mitochondrial function. *JCI Insight* 2017; 2. pii: 94248.
  46. Shanmuganathan S, Hausenloy DJ, Duchon MR, et al. Mitochondrial permeability transition pore as a target for cardioprotection in the human heart. *Am J Physiol Heart Circ Physiol* 2005; 289: H237–H242.
  47. Halestrap AP. Calcium, mitochondria and reperfusion injury: a pore way to die. *Biochem Soc Trans* 2006; 34: 232–237.
  48. Rudiger A and Singer M. Mechanisms of sepsis-induced cardiac dysfunction. *Crit Care Med* 2007; 35: 1599–1608.
  49. Duncan DJ, Yang Z, Hopkins PM, et al. TNF-alpha and IL-1beta increase Ca<sup>2+</sup> leak from the sarcoplasmic reticulum and susceptibility to arrhythmia in rat ventricular myocytes. *Cell Calcium* 2010; 47: 378–386.
  50. Parker MM, McCarthy KE, Ognibene FP, et al. Right ventricular dysfunction and dilatation, similar to left ventricular changes, characterize the cardiac depression of septic shock in humans. *Chest* 1990; 97: 126–131.
  51. Fauvel H, Marchetti P, Obert G, et al. Protective effects of cyclosporin A from endotoxin-induced myocardial dysfunction and apoptosis in rats. *Am J Respir Crit Care Med* 2002; 165: 449–455.
  52. Galluzzi L, Kepp O, Trojel-Hansen C, et al. Mitochondrial control of cellular life, stress, and death. *Circ Res* 2012; 111: 1198–1207.
  53. Nunnari J and Suomalainen A. Mitochondria: in sickness and in health. *Cell* 2012; 148: 1145–1159.
  54. West AP and Shadel GS. Mitochondrial DNA in innate immune responses and inflammatory pathology. *Nat Rev Immunol* 2017; 17: 363–375.
  55. Nakahira K, Hisata S and Choi AM. The Roles of Mitochondrial Damage-Associated Molecular Patterns in Diseases. *Antioxid Redox Signal* 2015; 23: 1329–1350.
  56. Patrushev M, Kasymov V, Patrusheva V, et al. Mitochondrial permeability transition triggers the release of mtDNA fragments. *Cell Mol Life Sci* 2004; 61: 3100–3103.
  57. Nakahira K, Haspel JA, Rathinam VA, et al. Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat Immunol* 2011; 12: 222–230.
  58. Nakahira K, Kyung SY, Rogers AJ, et al. Circulating mitochondrial DNA in patients in the ICU as a marker of mortality: derivation and validation. *PLoS Med* 2013; 10: e1001577; discussion e1001577.
  59. Bhagirath VC, Dwivedi DJ and Liaw PC. Comparison of the Proinflammatory and Procoagulant Properties of Nuclear,

- Mitochondrial, and Bacterial DNA. *Shock* 2015; 44: 265–271.
60. Mishra P and Chan DC. Metabolic regulation of mitochondrial dynamics. *J Cell Biol* 2016; 212: 379–387.
  61. Wai T and Langer T. Mitochondrial Dynamics and Metabolic Regulation. *Trends Endocrinol Metab* 2016; 27: 105–117.
  62. Liu N, Jiang Z, Liu Y, et al. Human trypsin inhibitor reduces the apoptosis of lipopolysaccharide induced human kidney-2 cells by promoting mitochondrial fusion. *Mol Med Rep* 2017; 16: 2899–2906.
  63. Scott I and Youle RJ. Mitochondrial fission and fusion. *Essays Biochem* 2010; 47: 85–98.
  64. Ishihara T, Ban-Ishihara R, Maeda M, et al. Dynamics of mitochondrial DNA nucleoids regulated by mitochondrial fission is essential for maintenance of homogeneously active mitochondria during neonatal heart development. *Mol Cell Biol* 2015; 35: 211–223.
  65. Sanchez-Villamil JP, D'Annunzio V, Finocchietto P, et al. Cardiac-specific overexpression of thioredoxin 1 attenuates mitochondrial and myocardial dysfunction in septic mice. *Int J Biochem Cell Biol* 2016; 81: 323–334.
  66. Zhan M, Brooks C, Liu F, et al. Mitochondrial dynamics: regulatory mechanisms and emerging role in renal pathophysiology. *Kidney Int* 2013; 83: 568–581.
  67. Exline MC and Crouser ED. Mitochondrial mechanisms of sepsis-induced organ failure. *Front Biosci* 2008; 13: 5030–5041.
  68. Carre JE, Orban JC, Re L, et al. Survival in critical illness is associated with early activation of mitochondrial biogenesis. *Am J Respir Crit Care Med* 2010; 182: 745–751.
  69. Kelly DP and Scarpulla RC. Transcriptional regulatory circuits controlling mitochondrial biogenesis and function. *Genes Dev* 2004; 18: 357–368.
  70. Sun L, Zhao M, Yu XJ, et al. Cardioprotection by acetylcholine: a novel mechanism via mitochondrial biogenesis and function involving the PGC-1 $\alpha$  pathway. *J Cell Physiol* 2013; 228: 1238–1248.
  71. Lancel S, Hassoun SM, Favory R, et al. Carbon monoxide rescues mice from lethal sepsis by supporting mitochondrial energetic metabolism and activating mitochondrial biogenesis. *J Pharmacol Exp Ther* 2009; 329: 641–648.
  72. Reynolds CM, Suliman HB, Hollingsworth JW, et al. Nitric oxide synthase-2 induction optimizes cardiac mitochondrial biogenesis after endotoxemia. *Free Radic Biol Med* 2009; 46: 564–572.
  73. Russell LK, Mansfield CM, Lehman JJ, et al. Cardiac-specific induction of the transcriptional coactivator peroxisome proliferator-activated receptor gamma coactivator-1alpha promotes mitochondrial biogenesis and reversible cardiomyopathy in a developmental stage-dependent manner. *Circ Res* 2004; 94: 525–533.
  74. Jian B, Yang S, Chen D, et al. Aging influences cardiac mitochondrial gene expression and cardiovascular function following hemorrhage injury. *Mol Med* 2011; 17: 542–549.
  75. Eipel C, Hildebrandt A, Scholz B, et al. Mutation of mitochondrial ATP8 gene improves hepatic energy status in a murine model of acute endotoxemic liver failure. *Life Sci* 2011; 88: 343–349.
  76. Sato S, Sakurai T, Ogasawara J, et al. A circadian clock gene, Rev-erb $\alpha$ , modulates the inflammatory function of macrophages through the negative regulation of Ccl2 expression. *J Immunol* 2014; 192: 407–417.
  77. Turdi S, Han X, Huff AF, et al. Cardiac-specific overexpression of catalase attenuates lipopolysaccharide-induced myocardial contractile dysfunction: role of autophagy. *Free Radic Biol Med* 2012; 53: 1327–1338.
  78. Zhao P, Gao J, Jiang J, et al. Myocardial cells and mitochondrial autophagy in sepsis mice induced by lipopolysaccharide. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi* 2016; 32: 177–181 [in Chinese, English Abstract].
  79. Piquereau J, Godin R, Deschenes S, et al. Protective role of PARK2/Parkin in sepsis-induced cardiac contractile and mitochondrial dysfunction. *Autophagy* 2013; 9: 1837–1851.
  80. Wang GQ, Tang T, Wang ZS, et al. Overexpression of Hypo-Phosphorylated I $\kappa$ B $\beta$  at Ser313 Protects the Heart against Sepsis. *PLoS One* 2016; 11: e0160860.

81. Preau S, Delguste F, Yu Y, et al. Endotoxemia Engages the RhoA Kinase Pathway to Impair Cardiac Function By Altering Cytoskeleton, Mitochondrial Fission, and Autophagy. *Antioxid Redox Signal* 2016; 24: 529–542.
82. Unuma K, Aki T, Funakoshi T, et al. Cobalt protoporphyrin accelerates TFEB activation and lysosome reformation during LPS-induced septic insults in the rat heart. *PLoS One* 2013; 8: e56526.
83. Wallace DC. Animal models for mitochondrial disease. *Methods Mol Biol* 2002; 197: 3–54.
84. Yu XX, Barger JL, Boyer BB, et al. Impact of endotoxin on UCP homolog mRNA abundance, thermoregulation, and mitochondrial proton leak kinetics. *Am J Physiol Endocrinol Metab* 2000; 279: E433–E446.
85. Sokolova IM and Sokolov EP. Evolution of mitochondrial uncoupling proteins: novel invertebrate UCP homologues suggest early evolutionary divergence of the UCP family. *FEBS Lett* 2005; 579: 313–317.
86. Liu XQ, Bell AW, Freeman KB, et al. Topogenesis of mitochondrial inner membrane uncoupling protein. Rerouting transmembrane segments to the soluble matrix compartment. *J Cell Biol* 1988; 107: 503–509.
87. Jarmuszkiewicz W and Woyda-Ploszczyca A. Mitochondrial uncoupling proteins: regulation and physiological role. *Postepy Biochem* 2008; 54: 179–187 [in Polish, English Abstract].
88. Arechaga I, Ledesma A and Rial E. The mitochondrial uncoupling protein UCP1: a gated pore. *IUBMB Life* 2001; 52: 165–173.
89. Donadelli M, Dando I, Fiorini C, et al. UCP2, a mitochondrial protein regulated at multiple levels. *Cell Mol Life Sci* 2014; 71: 1171–1190.
90. Barger JL, Barnes BM and Boyer BB. Regulation of UCP1 and UCP3 in arctic ground squirrels and relation with mitochondrial proton leak. *J Appl Physiol (1985)* 2006; 101: 339–347.
91. Ramsden DB, Ho PW, Ho JW, et al. Human neuronal uncoupling proteins 4 and 5 (UCP4 and UCP5): structural properties, regulation, and physiological role in protection against oxidative stress and mitochondrial dysfunction. *Brain Behav* 2012; 2: 468–478.
92. de Souza BM, Assmann TS, Kliemann LM, et al. The presence of the -866A/55Val/Ins haplotype in the uncoupling protein 2 (UCP2) gene is associated with decreased UCP2 gene expression in human retina. *Exp Eye Res* 2012; 94: 49–55.
93. Lapice E, Pinelli M, Pisu E, et al. Uncoupling protein 2 G(-866)A polymorphism: a new gene polymorphism associated with C-reactive protein in type 2 diabetic patients. *Cardiovasc Diabetol* 2010; 9: 68.
94. Lenten KU, Tu N, Chen H, et al. Genomic organization and mutational analysis of the human UCP2 gene, a prime candidate gene for human obesity. *J Recept Signal Transduct Res* 1999; 19: 229–244.
95. Garabuczi E, Sarang Z and Szondy Z. Glucocorticoids enhance prolonged clearance of apoptotic cells by upregulating liver X receptor, peroxisome proliferator-activated receptor- $\delta$  and UCP2. *Biochim Biophys Acta* 2015; 1853: 573–582.
96. Zhang J and Li D. Effect of conjugated linoleic acid on inhibition of prolyl hydroxylase 1 in hearts of mice. *Lipids Health Dis* 2012; 11: 22.
97. Chen SD, Yang DI, Lin TK, et al. Roles of oxidative stress, apoptosis, PGC-1 $\alpha$  and mitochondrial biogenesis in cerebral ischemia. *Int J Mol Sci* 2011; 12: 7199–7215.
98. Han YX, Lin YT, Xu JJ, et al. Status epilepticus stimulates peroxisome proliferator-activated receptor  $\gamma$  coactivator 1-alpha/mitochondrial antioxidant system pathway by a nitric oxide-dependent mechanism. *Neuroscience* 2011; 186: 128–134.
99. Song L, Xu Z, Li L, et al. Forkhead box protein A1 inhibits the expression of uncoupling protein 2 in hydrogen peroxide-induced A549 cell line. *Cell Stress Chaperones* 2014; 19: 53–60.
100. Sayeed A, Meng Z, Luciani G, et al. Negative regulation of UCP2 by TGF $\beta$  signaling characterizes low and intermediate-grade primary breast cancer. *Cell Death Dis* 2010; 1: e53.

101. Jin X, Chen D, Zheng RH, et al. miRNA-133a-UCP2 pathway regulates inflammatory bowel disease progress by influencing inflammation, oxidative stress and energy metabolism. *World J Gastroenterol* 2017; 23: 76–86.
102. Tahir TA, Singh H and Brindle NP. The RNA binding protein hnRNP-K mediates post-transcriptional regulation of uncoupling protein-2 by angiopoietin-1. *Cell Signal* 2014; 26: 1379–1384.
103. Li L, Xiao L, Hou Y, et al. Sestrin2 Silencing Exacerbates Cerebral Ischemia/Reperfusion Injury by Decreasing Mitochondrial Biogenesis through the AMPK/PGC-1 $\alpha$  Pathway in Rats. *Sci Rep* 2016; 6: 30272.
104. Zhang Y, Mi SL, Hu N, et al. Mitochondrial aldehyde dehydrogenase 2 accentuates aging-induced cardiac remodeling and contractile dysfunction: role of AMPK, Sirt1, and mitochondrial function. *Free Radic Biol Med* 2014; 71: 208–220.
105. Yuan H, Zhang Q, Guo J, et al. A PGC-1 $\alpha$ -Mediated Transcriptional Network Maintains Mitochondrial Redox and Bioenergetic Homeostasis against Doxorubicin-Induced Toxicity in Human Cardiomyocytes: Implementation of TT21C. *Toxicol Sci* 2016; 150: 400–417.
106. Alvero AB, Montagna MK, Sumi NJ, et al. Multiple blocks in the engagement of oxidative phosphorylation in putative ovarian cancer stem cells: implication for maintenance therapy with glycolysis inhibitors. *Oncotarget* 2014; 5: 8703–8715.
107. Goncalves-de-Albuquerque CF, Medeiros-de-Moraes IM, Oliveira FM, et al. Omega-9 Oleic Acid Induces Fatty Acid Oxidation and Decreases Organ Dysfunction and Mortality in Experimental Sepsis. *PLoS One* 2016; 11: e0153607.
108. Wang X, Liu D, Chai W, et al. The Role of Uncoupling Protein 2 During Myocardial Dysfunction in a Canine Model of Endotoxin Shock. *Shock* 2015; 43: 292–297.
109. Chen ZJ, Song YB, Wang HL, et al. Effect of UCP2-siRNA on inflammatory response of cardiomyocytes induced by septic serum. *Zhongguo Dang Dai Er Ke Za Zhi* 2014; 16: 851–855 [in Chinese, English Abstract].
110. Zheng G, Lyu J, Liu S, et al. Silencing of uncoupling protein 2 by small interfering RNA aggravates mitochondrial dysfunction in cardiomyocytes under septic conditions. *Int J Mol Med* 2015; 35: 1525–1536.
111. Cheng J, Nanayakkara G, Shao Y, et al. Mitochondrial Proton Leak Plays a Critical Role in Pathogenesis of Cardiovascular Diseases. *Adv Exp Med Biol* 2017; 982: 359–370.
112. Ko TH, Marquez JC, Kim HK, et al. Resistance exercise improves cardiac function and mitochondrial efficiency in diabetic rat hearts. *Pflugers Arch* 2018; 470: 263–275.
113. Dando I, Fiorini C, Pozza ED, et al. UCP2 inhibition triggers ROS-dependent nuclear translocation of GAPDH and autophagic cell death in pancreatic adenocarcinoma cells. *Biochim Biophys Acta* 2013; 1833: 672–679.
114. Akhmedov AT, Rybin V and Marin-Garcia J. Mitochondrial oxidative metabolism and uncoupling proteins in the failing heart. *Heart Fail Rev* 2015; 20: 227–249.
115. Makazan Z, Saini HK and Dhalla NS. Role of oxidative stress in alterations of mitochondrial function in ischemic-reperfused hearts. *Am J Physiol Heart Circ Physiol* 2007; 292: H1986–H1994.
116. Teshima Y, Akao M, Jones SP, et al. Uncoupling protein-2 overexpression inhibits mitochondrial death pathway in cardiomyocytes. *Circ Res* 2003; 93: 192–200.
117. Motloch LJ, Larbig R, Gebing T, et al. By Regulating Mitochondrial Ca<sup>2+</sup>-Uptake UCP2 Modulates Intracellular Ca<sup>2+</sup>. *PLoS One* 2016; 11: e0148359.
118. Huang JD, Chen SL, Lyu JJ, et al. Correlation between uncoupling protein 2 expression and myocardial mitochondrial injury in rats with sepsis induced by lipopolysaccharide. *Zhongguo Dang Dai Er Ke Za Zhi* 2016; 18: 159–164 [in Chinese, English Abstract].