



Contents lists available at ScienceDirect

## Journal of Ginseng Research

journal homepage: <http://www.ginsengres.org>

## Review article

## Gut microbiota-mediated pharmacokinetics of ginseng saponins



Dong-Hyun Kim\*

Department of Life and Nanopharmaceutical Sciences and Department of Pharmacy, College of Pharmacy, Kyung Hee University, Seoul, Republic of Korea

## ARTICLE INFO

## Article history:

Received 21 October 2016

Received in Revised form

26 February 2017

Accepted 18 April 2017

Available online 28 April 2017

## Keywords:

fermentation  
ginseng  
ginsenoside  
gut microbiota  
metabolism

## ABSTRACT

Orally administered ginsengs come in contact with the gut microbiota, and their hydrophilic constituents, such as ginsenosides, are metabolized to hydrophobic compounds by gastric juice and gut microbiota: protopanaxadiol-type ginsenosides are mainly transformed into compound K and ginsenoside Rh2; protopanaxatriol-type ginsenosides to ginsenoside Rh1 and protopanaxatriol, and ocotillol-type ginsenosides to ocotillol. Although this metabolizing activity varies between individuals, the metabolism of ginsenosides to compound K by gut microbiota in individuals treated with ginseng is proportional to the area under the blood concentration curve for compound K in their blood samples. These metabolites such as compound K exhibit potent pharmacological effects, such as antitumor, anti-inflammatory, antidiabetic, antiallergic, and neuroprotective effects compared with the parent ginsenosides, such as Rb1, Rb2, and Re. Therefore, to monitor the potent pharmacological effects of ginseng, a novel probiotic fermentation technology has been developed to produce absorbable and bioactive metabolites. Based on these findings, it is concluded that gut microbiota play an important role in the pharmacological action of orally administered ginseng, and probiotics that can replace gut microbiota can be used in the development of beneficial and bioactive ginsengs.

© 2017 The Korean Society of Ginseng, Published by Elsevier Korea LLC. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

Most herbal medicines are orally administered to humans, and the components inevitably come into contact with the gut microbiota in the gastrointestinal tract where trillions of microbes reside. The gut microbiota exhibits diverse physiological activities including the ability to metabolize orally administered and bile-secreted xenobiotics (e.g., drugs, phytochemicals) [1–3]. Gut microbiota transforms the constituents of orally administered hydrophilic drugs and phytochemicals before absorption by gastrointestinal tract into the blood. Studies on the metabolism of phytochemicals found in natural products, such as ginseng, by the gut microbiota are important in understanding their biological effects [4,5].

This review describes gut microbiota-mediated metabolism of ginsenosides such as protopanaxadiol-type, protopanaxatriol-type, oleanane-type, and ocotillol-type ginsenosides and their bioactive metabolites.

## 2. Gut microbiota

The neonate is born in a germ-free state. Immediately after birth, they are exposed to microbes present within the parturient canal, on the skin of mothers and nurses, and in ambient air. These microbes colonize on body surfaces and the gastrointestinal and vaginal tracts [6,7]. Newer molecular methods have revealed that the gastrointestinal tract hosts over 2,000 species of microbiota in humans. Most species belong to eight dominant phyla: *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, *Actinobacteria*, *Fusobacteria*, *Cyanobacteria*, and *Verrucomicrobia* [8,9]. More than 80% of these microbes belong to the phyla *Firmicutes* and *Bacteroidetes*; *Firmicutes* includes *Clostridia* and *Bacilli*; and *Bacteroidetes* includes *Bacteroides* spp. The highly complex gut ecosystem varies between individuals due to factors, such as diet, genetics, hormones, and drugs [10]. It exhibits various physiological actions: fermentation of carbohydrates and proteins that are not digested in the upper gut, production of vitamins B and K, protection against pathogens,

\* Department of Life and Nanopharmaceutical Sciences and Department of Pharmacy, Kyung Hee University, 26, Kyungheedaero, Dongdaemun-gu, Seoul 02247, Republic of Korea.

E-mail address: [dhkim@khu.ac.kr](mailto:dhkim@khu.ac.kr).

stimulation of innate and adaptive immune responses, and metabolism of orally administered hydrophilic phytochemicals and drugs.

### 3. Ginseng constituents

Ginseng refers to the dried roots of the species *Panax* sp. (Family Araliaceae), including *Panax ginseng* Meyer (Korean ginseng or Asian Ginseng), which has been used as a herbal medicine for more than 2000 years [11], *Panax quinquefolius* L. (American Ginseng), *Panax notoginseng* (Burk.) FH Chen (Notoginseng), and *Panax vietnamensis* Ha et Grushv. (Vietnamese Ginseng) [12–14]. *P. ginseng* is the most commonly used. Garriques prepared the saponin fraction of *P. quinquefolius* [1]. Its constituents were not identified until 1963 [15]. Shibata et al. [16–18] isolated saponins from the root of *P. ginseng* in 1963 and identified their structures. Since then, many researchers have isolated the constituents including ginsenosides. Approximately 200 substances, such as ginsenosides, polysaccharides, and polyacetylenes have been isolated from Korean ginseng [19,20] and more than 100 from American ginseng, notoginseng, and Vietnamese ginseng [12–14].

Shibata et al. [16–18,21] established the chemical structures of main prosapogenins 20S-protopanaxadiol, 20S-protopanaxatriol, protosapogenin, and ginsenoside Rg1 found in the dried root of *P. ginseng*. Kitagawa et al. [22,23] isolated malonyl ginsenoside Rb1, Rb2, Rc, and Rd; Ruan et al. [24] isolated malonyl ginsenoside Ra3; Zhu et al. [25] isolated six protopanaxatriol-type ginsenosides Re1, Re2, Re3, Re4, Re5, and Re6 and 10 known protopanaxatriol ginsenosides including ginsenoside Rg1.

From red ginseng (steamed *P. ginseng*), Matsumura et al. [26] isolated ginsenosides Ra1, Ra2, and Ra3 and notoginsenoside R4. Kasai et al. [27] isolated ginsenosides Ra1, Ra2, Ra3, Rs1, and Rs2, notoginsenoside R1, and quinquenoside R1. Thereafter, ginsenosides Ro, Rb1, Rb2, Tc, Rd, Re, Rf, Rg1, Rg2, Rg3, Rh1, Rh2, 20R-ginsenoside Rh1, 20S-ginsenoside Rg3, and 20R-ginsenoside Rg2 were also isolated by [27]. Ryu et al. [28] isolated ginsenoside Rg6 and 20(E)-ginsenoside F4. Baek et al. [29–31] isolated ginsenoside Rh4 [29] as well as ginsenoside Rs1, Rs2, Rs3, and Rs4, quinquenoside R4; ginsenoside Rg3, Rg5, Rg6, F4, and Rf2 [30,31]. Ginsenosides Rh1, Rg3, and Rg2 were found in large quantities. Under intense steaming or heating, ginsenoside Rg3 can transform into 20S-Rh2 and 20R-Rh2 and subsequently form the aglycone 20S-protopanaxadiol and 20R-protopanaxadiol or even 20-dehydroprotopanaxadiol through chemical degradation [32–34]. Ginsenoside Rk1 and Rg5 can transform into their degradation products, such as Rk2 and Rh3, and Rh1 into aglycone 20S-protopanaxatriol and 20R-protopanaxatriol or even 20-dehydroprotopanaxatriol.

In American ginseng (*P. quinquefolius*), > 60 ginsenosides, including dammarane, ocotillol, and oleanane types, have been isolated: ginsenoside Rb1, Rd, and Re as main constituents, including ocotillol-type ginsenosides (24R-pseudoginsenoside F11, pseudoginsenoside RT5, F-11, 24R-vina-ginsenoside R1) and oleanane-type ginsenosides (ginsenoside Ro, chikusetsusaponin Iva) [4].

In *P. notoginseng*, a total of 56 dammarane-type saponins have been isolated: protopanaxadiol-type and protopanaxatriol ginsenosides, such as ginsenosides Ra3, RK3, Rh4, Rg3, Rk1, Rg5, F2, Rh1, Rg1, Re, Rd, Rb1, and Rb2, 6'-O-acetylginsenoside Rh1, and another group of saponins, notoginsenosides A – N, R1 – R4, R6 – R9, Fa, Fc, and Fe and gypenoside X VII [14,35–39].

From *P. vietnamensis*, Nguyen et al. [14,40,41] isolated ginsenoside Rh1, Rg1, Re, Rd, Rb3, Rb2, and Rb1; pseudoginsenoside RS1; notoginsenosides R1 and Fa; oleanolic acid; and ocotillol-type saponins (pseudoginsenoside-RT4, 24(S)-pseudoginsenoside F11,

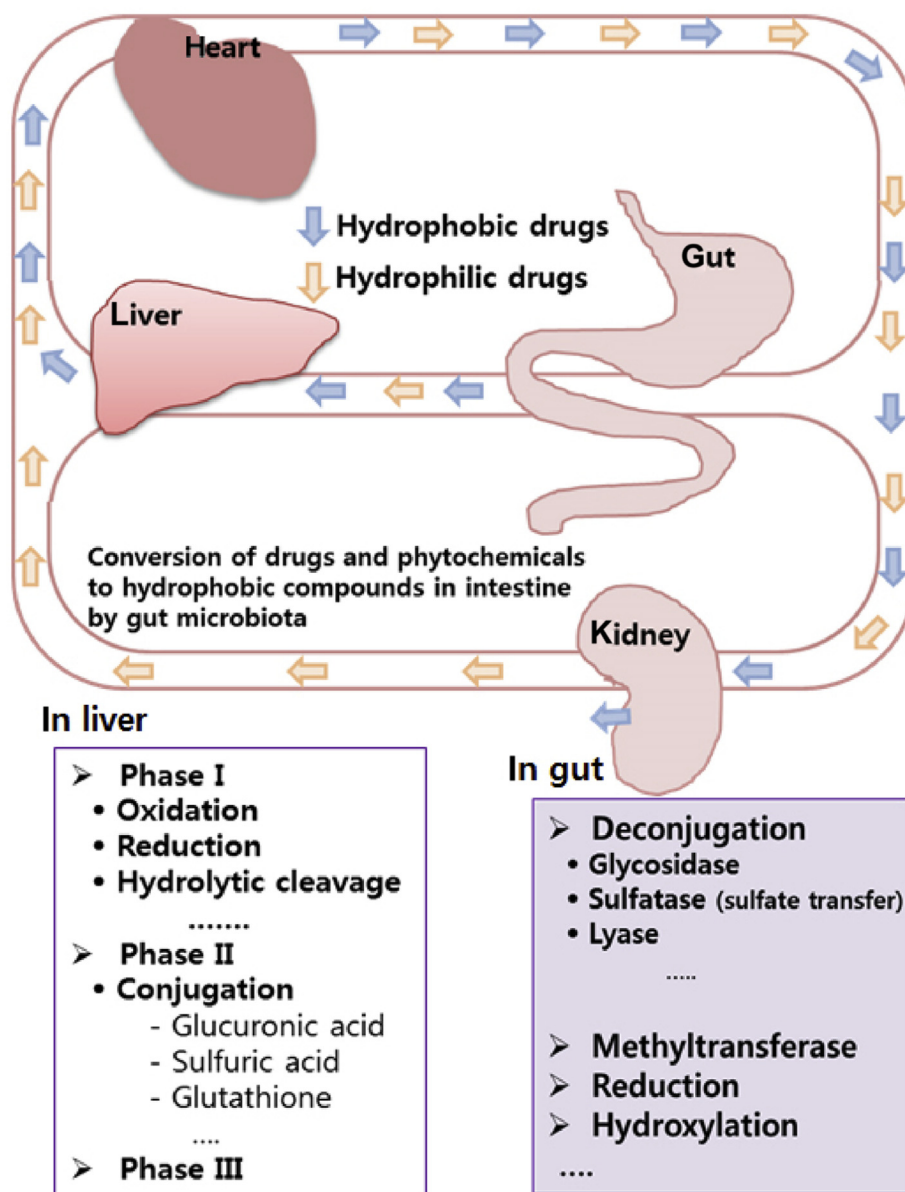
majonoside F1, R1, and R2); vinaginsenoside R3, R4, R5, R6, R7, R8, and R9; 20-gluco-ginsenoside Rf; ginsenoside Rc; notoginsenoside R6; quinquenoside R1; and gypenoside XVII. In addition, Duc et al. [42] isolated 6-O-β-D-glucopyranosyl 20(S),25-epoxydammarane-3β,6α,12β,24α-tetrol, 6-O-β-D-xylopyranosyl-(1->2)-β-D-glucopyranosyl 20(S),25-epoxydammarane-3β,6α,12β,24α-tetrol; 6-O-β-D-glucopyranosyl dammarane-3β,6α,12β,20(S),24 xi,25-hexol; 3-O-[β-D-glucopyranosyl-(1->2)-β-D-glucopyranosyl]-20-O-β-D-glucopyranosyl dammarane-3β,12 β,20(S),24 xi,25-pentol; and 6-O-β-D-xylopyranosyl-(1->2)-β-D-glucopyranosyl 20(S),24(S)-epoxydammarane-3β,6α,12β,25 xi,26-pentol [42].

### 4. Absorption, distribution, metabolism, and excretion of ginseng phytochemicals

The pharmacological effects of ginsengs, particularly their saponins including ginsenosides, may be dependent on their absorption, distribution, metabolism, and excretion (ADME), similar to drugs (Fig. 1). In a pharmacokinetic study, Tawab et al. [43] investigated the parent ginsenosides and their metabolites in the plasma and urine samples of two individuals orally treated with Ginsana extract (ginseng saponin fraction, Pharmaton S.A., Lugano, Switzerland) by liquid chromatography–mass spectrometry/mass spectrometry. The metabolites ginsenosides Rh1, F1, and compound K were detected in the plasma and urine. However, the metabolites were not detected in Ginsana extract. Therefore, these metabolites (hydrolysates) may be produced for parental ginsenosides by gut microbiota or by the liver. Although ginsenoside Rb1 was detected in the plasma and urine of one individual, it was detected at the lower limit of detection. Akao et al. [44,45] conducted a pharmacokinetic study of compound K in germ-free and gnotobiotic rats. They could not detect ginsenoside Rb1 in both rats, but they detected compound K in gnotobiotic rats, not in the germ-free rats. In individuals orally administered ginseng extract, Shibata et al. [46] did not detect ginsenoside Rb1, but detected compound K. In our previous studies, we detected compound K in rats orally treated with 0.2 g/kg ginseng extract. Maximum concentration, time to maximum concentration, and area under the curve (AUC) were  $24.1 \pm 5.5$  ng/mL,  $15.2 \pm 1.8$  h, and  $153.1 \pm 30.6$  ng·h/mL, respectively [47–49]. We found that the absorption of compound K was affected by diets including prebiotic fiber (nutriose). We also performed a pharmacokinetic study of compound K in individuals ( $n = 34$ ) orally treated with ginseng powder [1,49]. Maximum concentration, time to maximum concentration, and AUC were found to be  $27.89 \pm 24.46$  ng/mL,  $10.76 \pm 2.07$  h, and  $221.98 \pm 221.42$  ng h/mL, respectively. These findings suggest that compound K may be the main metabolite produced by intestinal bacteria in humans orally administered ginseng.

### 5. Metabolism of protopanaxadiol-type and protopanaxatriol-type ginsenosides in gastrointestinal tract by gastrointestinal juice and gut microbiota

To understand the metabolism of ginsenosides in the gastrointestinal tract, many experiments were conducted *in vitro* and *in vivo* [44,50–53]. Karikura et al. [33] and Han et al. [51] reported that protopanaxadiol-type ginsenosides Rb1 and Rb2 transformed into ginsenoside Rg3 in diluted hydrochloric acid *in vitro*. In addition, ginsenoside Rb1 transformed into a 25-hydroperoxy-23-ene derivative. Ginsenoside Rb2 transformed into 25-hydroxyl-23-ene, 24-hydroxy-25-ene, 25-hydroperoxy-23-ene, and 24-hydroperoxy-25-ene derivatives. Thus, protopanaxadiol-type ginsenosides hydrolyzed the C-20 glycosyl moiety and hydrated or oxygenated the side chain. Nevertheless, the amount of their metabolites in rat stomach was negligible.



**Fig. 1.** Fates of orally administered drugs and phytochemicals in humans and animals. Orally administered drugs and phytochemicals in humans and animals are converted to hydrophobic compounds in the intestine by enzymes of gut microbiota such as  $\beta$ -D-glucosidase, and delivered into the liver, which metabolizes these hydrophobic metabolites to hydrophilic compounds by enzymes of phase I–III, such as uridine 5'-diphospho-glucuronosyltransferase.

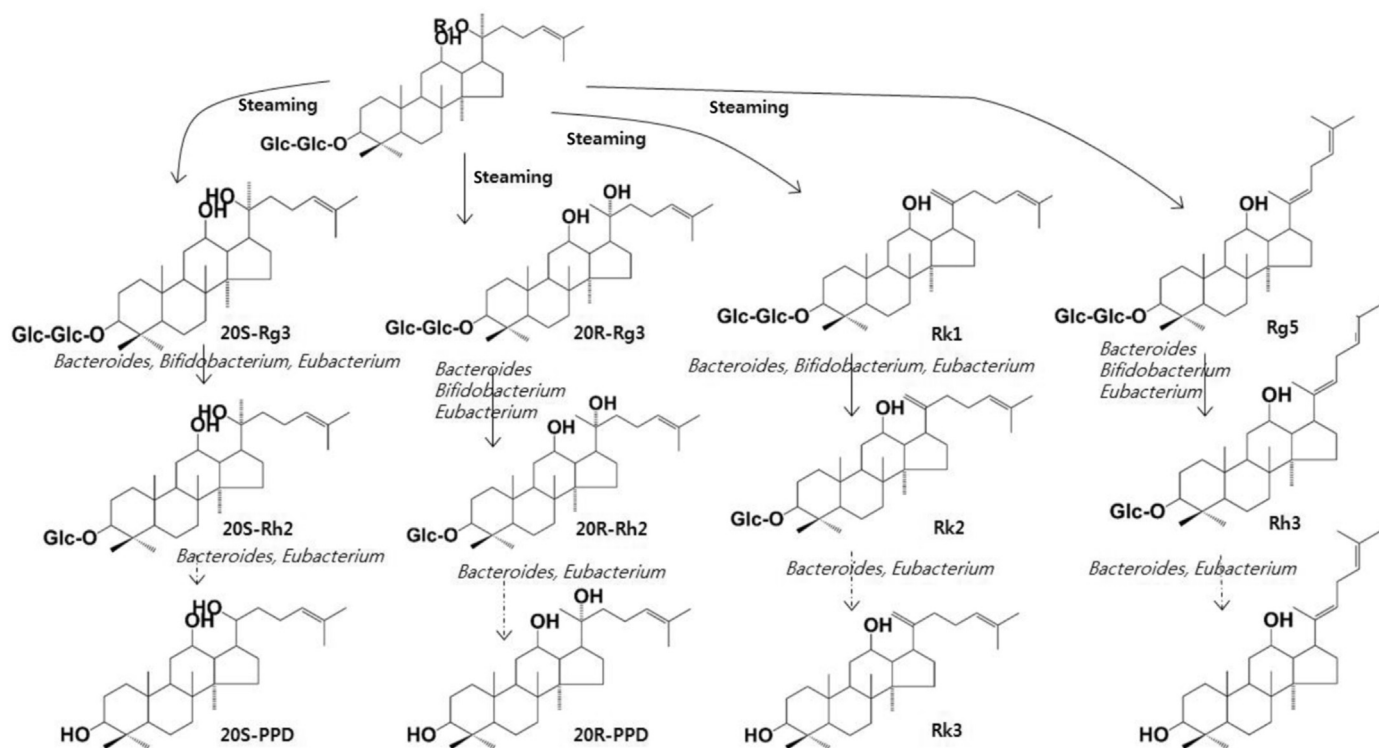
After our incubation in a diluted condition at 60°C or at boiling temperature, protopanaxadiol-type ginsenosides Rb1, Rb2, and Rc transformed into ginsenoside Rg3, Rg5, and Rk1; however, the transformation was negligible at 37°C [32]. These findings suggest that orally administered protopanaxadiol-type ginsenosides may be resistant to the gastric juice, but can be transformed by heat under acidic condition.

Hydrophilic ginsenosides, when orally ingested by humans and animals, come into contact with gut microbiota in the gastrointestinal tract and can be metabolized to hydrophobic metabolites by the gut microbiota [4,5]. Hydrophobic metabolites are easily absorbed from the gastrointestinal tract into the blood compared with the parent ginsenosides. When ginseng was orally administered in humans, compound K (a hydrophobic metabolite of ginsenosides) was detected in the blood as the main component [43,46]; a small quantity of ginsenoside Rb1, but not ginsenoside

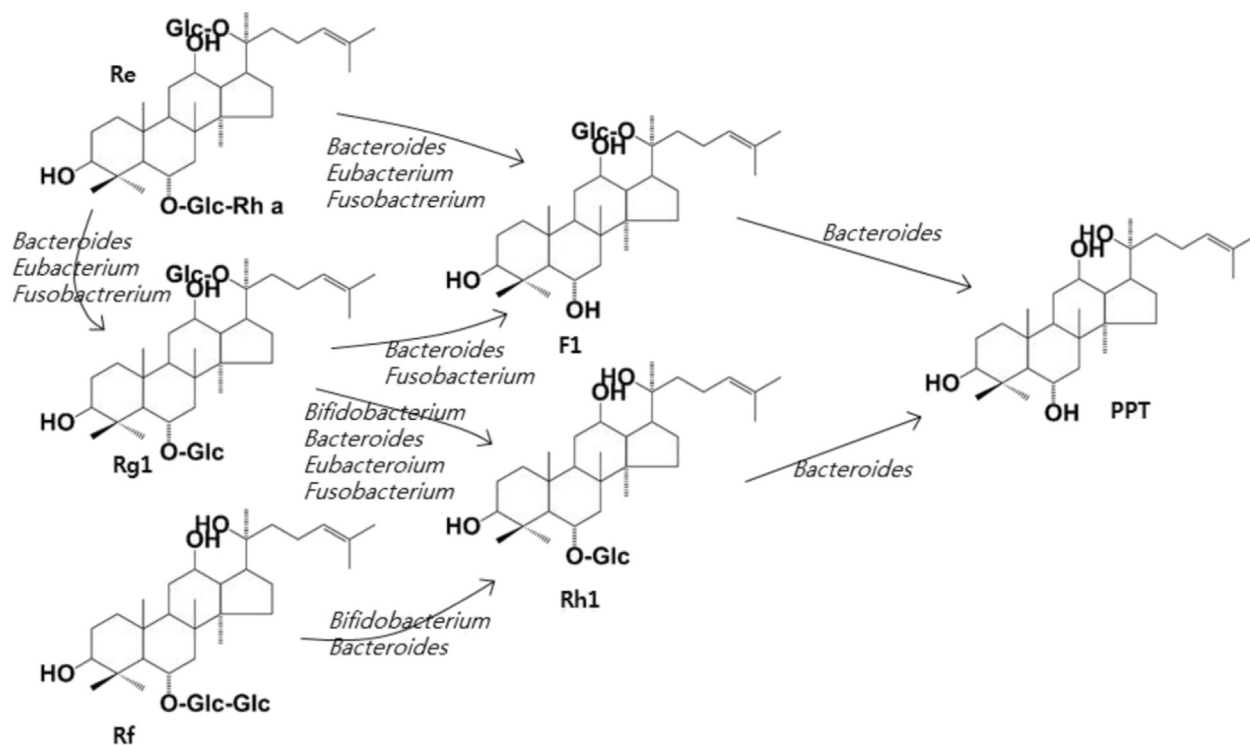
Rb2, Rc, and Re, was detected in the blood of one of two individuals. Akao et al. [44,45] and Park et al. [54] also found compound K in the intestinal content, blood, and urine of conventional and gnotobiotic rats orally treated with ginsenoside Rb1. Moreover, Kato et al. [55] detected compound K in the blood of human following the intake of red ginseng powder. Although ginsenosides Rb1, Rb2, Rc, and Re were detected in some studies [43–45], the levels were insufficient to exhibit show a biological effect.

Protopanaxatriol-type ginsenosides Rg1 and Re are more labile to acidic condition than protopanaxadiol-type ginsenosides. Protopanaxatriol-type ginsenosides are transformed into ginsenosides Rh1 and Rg2 under acidic condition. The metabolite ginsenoside Rh1 was absorbed from the stomach and small intestine. However, ginsenoside Rg2 was not detected in the blood because the rhamnosyl moiety could not be absorbed due to the absence of rhamnose sugar transporter, such as quercetin-4-O-

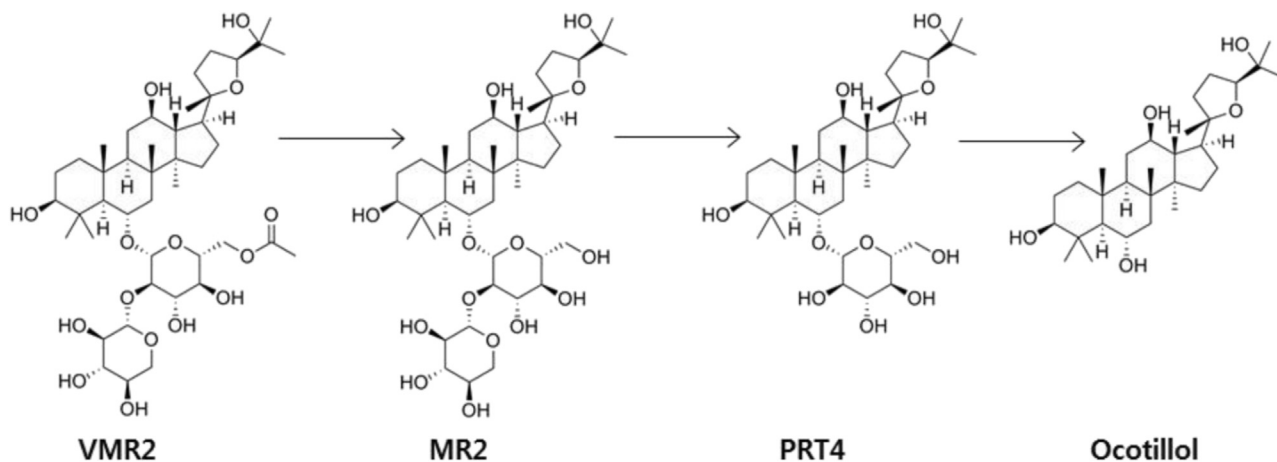




**Fig. 3.** Proposed metabolism of protopanaxadiol-type ginsenoside Rg3, Rg5, and Rk1 from the root of red ginseng by gut microbiota. Ginsenoside Rb1, Rb2, Rc, and Rd present in fresh and dried ginsengs are transformed into ginsenoside Rg3, Rg5, and Rk1 by heating. When the heat-processed ginsengs, such as red ginseng, are orally treated in humans or animals, the transformed compounds ginsenoside Rg3, Rg5, and Rk1 in these ginsengs are metabolized to ginsenoside Rh2, Rk2, and Rh3 or PPD by gut microbiota.



**Fig. 4.** Proposed metabolism of protopanaxatriol ginsenoside Re, Rg1, and Rf from dried ginseng by gut microbiota. When fresh or dried ginsengs are orally administered in humans or animals, ginsenoside Re, Rg1, and Rf in these ginsengs are metabolized to protopanaxatriol, ginsenoside F1 or ginsenoside Rh1 by gut microbiota.

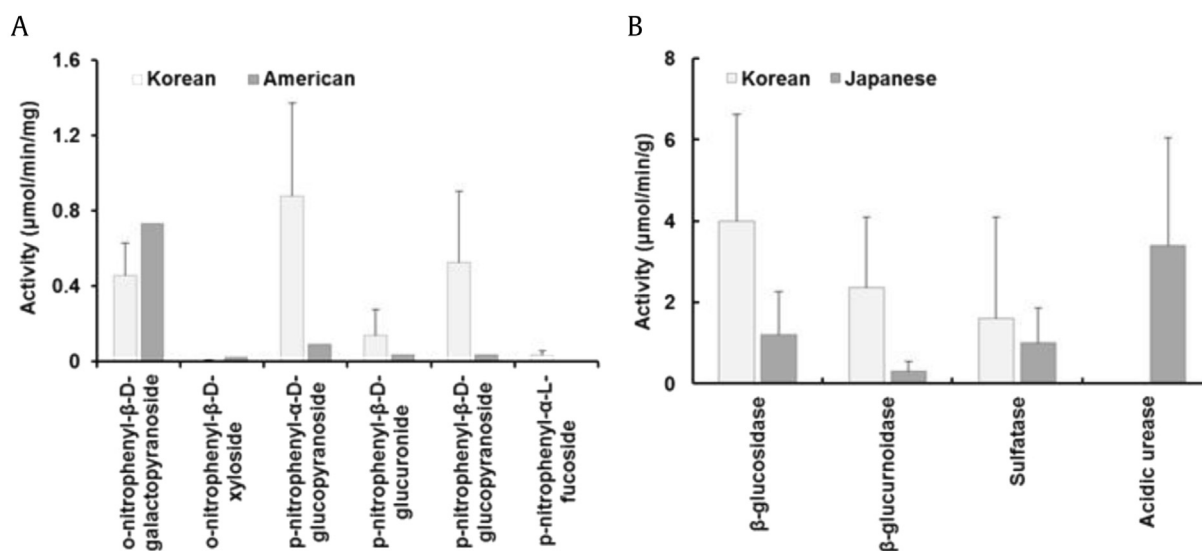


**Fig. 5.** Proposed metabolism of ocotillol-type vinamajonoside R2 and majonoside R2 from Vietnamese ginseng by gut microbiota. When fresh or dried Vietnamese ginseng (VG) is orally administered in humans or animals, vinamajonoside R2 (VMR2) and majonoside R2 (MR2) present in VG are metabolized to ocotillol through pseudoginsenoside RT4 (PRT4) by gut microbiota.

endothelium-independent aorta relaxation [80], and neuro-protective [81,82], adjuvant-like [83], and immunomodulating effects [84]. However, these pharmacological effects of ginsenosides were observed to be different between *in vitro* and *in vivo* studies. Ginsenosides exhibited an antitumor effect in *in vivo* studies, but contradictively a negligible effect in *in vitro* studies [85,86]. In a recent study, orally administered ginsenosides were metabolized to compound K, ginsenoside Rh2 and Rh1, and protopanaxatriol by the gut microbiota; compound K, ginsenoside Rh2, and protopanaxatriol exhibited potent cytotoxicity against tumor cells [45]. When antiallergic activity of the parent molecules and transformed metabolites were evaluated *in vitro*, ginsenoside Rh1 and Rh2, compound K, and protopanaxatriol were found to show potent inhibitory activity [77,87–89]. These results suggest that the pharmacological effects of ginsengs may be dependent on the metabolism of ginsenosides to bioactive compounds by the gut microbiota.

### 9. Difference in gut microbiota-mediated ginsenoside-metabolizing enzyme activity between individuals

Gut microbiota transforms ginsenoside Rb1 to compound K *in vitro* and *in vivo* and the compound K is absorbed into the blood [48]. The metabolism of ginsenosides to compound K by intestinal bacteria was proportional to the amount of compound K absorbed into the blood of volunteers who had been administered ginseng [49]. The compound K-forming activity is significantly different between individuals. Their AUCs are also significantly different. Nevertheless, there was a correlation between the compound K-forming activity and AUC for compound K. Cui et al. [90] determined the total amount of protopanaxatriol and protopanaxadiol in the urine of humans orally administered ginseng preparations. They detected approximately 1.2% of the orally treated dose of protopanaxatriol-type ginsenosides and < 0.2% of the orally treated dose of protopanaxadiol-type ginsenosides. However, Hasegawa



**Fig. 6.** Difference in β-galactosidase, β-xylosidase, β-glucuronidase, β-glucosidase, and α-fucosidase activities in the fecal suspensions of Korean, American, and Japanese. (A) Difference between Korean and American. The metabolic activity of the Korean fecal suspension was measured according to the method described by Tamura et al. [98]. The data of American was obtained from the study conducted by Tamura et al. [98]. (B) Difference between Korean and Japanese. The metabolic activity of the Korean fecal suspension was measured according to the method described by Kobashi et al. [99]. The data of Japanese were obtained from the study conducted by Kobashi et al. [99].

et al. [67] reported that compound K mono-fatty acid esters such as stearyl compound K, accumulated in mouse liver following intravenous administration. These results suggest that the low absorption of ginsenoside metabolites is dependent on the metabolic activity of gut microbiota and due to the low transforming activity of ginsenosides into hydrophobic metabolites.

## 10. Fermented/biotransformed ginseng

When medicinal and functional herbs including ginsengs are orally administered to humans, their hydrophilic constituents come in contact with the gut microbiota in the alimentary tract and are transformed into absorbable hydrophobic ginsenosides before absorption from the gastrointestinal tract. All individuals possess characteristic indigenous strains of gut microbiota and the metabolizing activity was found to be significantly different between individuals (Fig. 6) [63,91–99]. Thus, the metabolic activity of gut microbiota was affected by environmental factors such diet, drugs, and genetics. When the metabolism of ginsenosides Rb1 and Rb2 to active compound K was measured, a significant variation was observed between individuals. Therefore, bioactive and absorbable ginsenoside metabolites are valuable in the therapy of diverse diseases. Therefore, to develop bioactive and well-absorbable ginsenoside metabolites containing ginsengs, we developed the fermented ginsengs, which contained bioactive ginsenoside metabolites, such as compound K and ginsenoside Rh1. These metabolites were transformed from ginsenoside Rb1, Rb2, Rc, and Rd by probiotics [31,100]. However, to use these probiotics, their safety and biotransforming activity should be affirmed. Once their safety and efficacy is confirmed, fermentation biotechnology can be valuable in developing novel ginseng preparations. Gut microbiota play an important role in the pharmacological action of ginseng. Therefore, beneficial and bioactive ginsengs can be developed using probiotic fermentation.

## 11. Conclusion

Hydrophilic components of orally administered ginsengs, such as ginsenosides Rb1 and Rb2, are metabolized to hydrophobic compounds by gastric juice and gut microbiota: the metabolites of protopanadiol-type ginsenosides are mainly compound K and ginsenoside Rh2 and protopanatriol-type ginsenosides are ginsenoside Rh1 and protopanatriol. The absorption of the metabolites, such as compound K, is proportional to fecal gut microbiota-metabolizing activity of ginsenosides to compound K. However, the metabolizing activity varies between individuals. Of these metabolites, absorbable and hydrophobic ones, such as compound K, exhibit potent pharmacological effects, such as antitumor, anti-inflammatory, antidiabetic, antiallergic, and neuroprotective effects, than parent ginsenosides, such as ginsenoside Rb1, Rb2, and Re. These findings suggest that the gut microbiota play an important role in the pharmacological action of orally administered ginseng. Furthermore, some probiotics isolated from human gut microbiota transform hydrophilic ginsenosides to hydrophobic and bioactive ginsenosides, such as compound K, and a novel probiotic fermentation technology has been developed to produce absorbable and bioactive metabolites instead of gut microbiota. These findings suggest that the gut microbiota play an important role in the pharmacological action of orally administered ginseng and probiotics that can replace gut microbiota can be used in the development of beneficial and bioactive ginsengs.

## Conflicts of interest

The authors have no conflicts of interest.

## Acknowledgments

This research was supported by a grant from Ministry of Food and Drug Safety in 2016 [16182MFD416].

## References

- [1] Kim DH. Chemical diversity of *Panax ginseng*, *Panax quinquefolius*, and *Panax notoginseng*. *J Ginseng Res* 2012;36:1–15.
- [2] Mikov M. The metabolism of drugs by the gut flora. *Eur J Drug Metab Pharmacokinet* 1994;19:201–7.
- [3] Sousa T, Paterson R, Moore V, Carlsson A, Abrahamsson B, Basit AW. The gastrointestinal microbiota as a site for the biotransformation of drugs. *Int J Pharm* 2008;363:1–25.
- [4] Kim DH. The possible role of intestinal microflora in pharmacological activities of ginseng. *Int Biomed Pharmacol Sci* 2012;6:90–6.
- [5] Kobashi K, Akao T. Relation of intestinal bacteria to pharmacological effects of glycosides. *Biosci Microflora* 1987;16:1–7.
- [6] Fanaro S, Chierici R, Guerrini P, Vigi V. Intestinal microflora in early infancy: composition and development. *Acta Paediatr Suppl* 2003;91:48–55.
- [7] Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, Knight R. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA* 2010;107:11971–5.
- [8] Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargeant M, Gill SR, Nelson KE, Relman DA. Diversity of the human intestinal microbial flora. *Science* 2005;308:1635–8.
- [9] Lozupone CA, Stombaugh JJ, Gordon JL, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature* 2012;489:220–30.
- [10] Cho I, Blaser MJ. The human microbiome: at the interface of health and disease. *Nat Rev Genet* 2012;13:260–70.
- [11] Li CP, Li RC. An introductory note to ginseng. *Am J Chin Med* 1973;1:249–61.
- [12] Banskota AH, Tezuka Y, Le Tran Q, Kadota S. Chemical constituents and biological activities of Vietnamese medicinal plants. *Curr Top Med Chem* 2003;3:227–48.
- [13] Chen CF, Chiou WF, Zhang JT. Comparison of the pharmacological effects of *Panax ginseng* and *Panax quinquefolium*. *Acta Pharmacol Sin* 2008;29:1103–8.
- [14] Ng TB. Pharmacological activity of *sanchi ginseng* (*Panax notoginseng*). *J Pharm Pharmacol* 2006;58:1007–19.
- [15] Garriques SS. On *panaquilon*, a new vegetable substance. *Ann Chem Pharm* 1954;90:231–4.
- [16] Shibata S, Tanaka O, Nagai M, Ishii T. Studies on the constituents of Japanese and Chinese crude drugs. XII. Panaxadiol, a saponin of ginseng roots. *Chem Pharm Bull* 1963;11:762–5.
- [17] Shibata S, Ando T, Tanaka O, Meguro Y, Sôma K, Iida Y. Saponins and saponinins of *Panax ginseng* C.A. Meyer and some other *Panax* spp. *Yakugaku Zasshi* 1965;85:753–5 [In Japanese].
- [18] Shibata S, Tanaka O, Soma K, Ando T, Iida Y, Nakamura H. Studies on saponins and saponinins of ginseng. The structure of panaxatriol. *Tetrahedron Lett* 1965;42:207–13.
- [19] Baek SH, Bae ON, Park JH. Recent methodology in ginseng analysis. *J Ginseng Res* 2012;36:119–34.
- [20] Ru W, Wang D, Xu Y, He X, Sun YE, Qian L, Zhou X, Qin Y. Chemical constituents and bioactivities of *Panax ginseng* (C.A. Mey.). *Drug Discov Ther* 2015;9:23–32.
- [21] Shibata S, Fujita M, Itokawa H, Tanaka O. Studies on the constituents of Japanese and Chinese crude drugs. XI. Panaxadiol, a saponin of ginseng roots. *Chem Pharm Bull (Tokyo)* 1963;11:759–61.
- [22] Kitagawa I, Taniyama T, Shibuya H, Noda T, Yoshikawa M. Chemical studies on crude drug processing. V. On the constituents of ginseng *radix rubra* (2): Comparison of the constituents of white ginseng and red ginseng prepared from the same *Panax ginseng* root. *Yakugaku Zasshi* 1987;107:495–505 [In Japanese].
- [23] Kitagawa I, Yoshikawa M, Yoshihara M, Hayashi T, Taniyama T. Chemical studies on crude drug processing. I. On the constituents of ginseng *radix rubra* (1). *Yakugaku Zasshi* 1983;103:612–22.
- [24] Ruan CC, Liu Z, Li X, Liu X, Wang LJ, Pan HY, Zheng YN, Sun GZ, Zhang YS, Zhang LX. Isolation and characterization of a new ginsenoside from the fresh root of *Panax ginseng*. *Molecules* 2010;15:2319–25.
- [25] Zhu GY, Li YW, Hau DK, Jiang ZH, Yu ZL, Fong WF. Protopanaxatriol-type ginsenosides from the root of *Panax ginseng*. *Agric Food Chem* 2011;59:200–5.
- [26] Besso H, Kasai R, Saruwatari Y, Fuwa T, Tanaka O. Ginsenoside Ra1 and ginsenoside Ra2, new dammarane-saponins of ginseng roots. *Chem Pharm Bull* 1982;30:2380–5.
- [27] Kasai R, Besso H, Tanaka O, Saruwatari Y, Fuwa T. Saponins of red ginseng. *Chem Pharm Bull* 1983;31:2120–5.
- [28] Ryu JH, Park TH, Kim DH, Sohn JM, Kim HM, Park JH. A genuine dammarane glycoside, (20E)-ginsenoside F4 from Korean red ginseng. *Arch Pharm Res* 1996;19:335–6.
- [29] Baek NI, Kim DS, Lee YH, Park JD, Lee CB, Kim SI. Ginsenoside Rh4, a genuine dammarane glycoside from Korean Red Ginseng. *Planta Med* 1996;62:86–7.

- [30] Anufriev VP, Malinovskaya GV, Denisenko VA, Uvarova NI, Elyakov GB, Kim SI, Baek NI. Synthesis of ginsenoside Rg3, a minor constituent of ginseng *radix*. Carbohydr Res 1997;304:179–82.
- [31] Park JD, Lee YH, Kim SI. Ginsenoside Rf2, a new dammarane glycoside from Korean red ginseng (*Panax ginseng*). Arch Pharm Res 1998;21:615–7.
- [32] Bae EA, Han MJ, Kim EJ, Kim DH. Transformation of ginseng saponins to ginsenoside Rh2 by acids and human intestinal bacteria and biological activities of their transformants. Arch Pharm Res 2004;27:61–7.
- [33] Han BH, Park MH, Han YN, Woo LK, Sankawa U, Yahara S, Tanaka O. Degradation of ginseng saponins under mild acidic conditions. Planta Med 1982;44:146–9.
- [34] Kwon SW, Han SB, Park IH, Kim JM, Park MK, Park JH. Liquid chromatographic determination of less polar ginsenosides in processed ginseng. J Chromatogr A 2001;921:335–9.
- [35] Taniyasu S, Tanaka O, Yang TR, Zhou J. Dammarane saponins of flower buds of *Panax notoginseng* (*sanchi-ginseng*). Planta Med 1982;44:124–5.
- [36] Yu HS, Zhang LJ, Song XB, Liu YX, Zhang J, Cao M, Kang LP, Kang TG, Ma BP. Chemical constituents from processed rhizomes of *Panax notoginseng*. Zhongguo Zhong Yao Za Zhi 2013;38:3910–7 [In Chinese].
- [37] Zeng J, Cui XM, Zhou JM, Jiang ZY, Zhang XM, Chen JJ. Studies on chemical constituents from rhizomes of *Panax notoginseng*. Zhong Yao Cai 2007;30:1388–91.
- [38] Zhou J, Wu MZ, Taniyasu S, Besso H, Tanaka O, Saruwatari Y, Fuwa T. Dammarane-saponins of *sanchi-ginseng*, roots of *Panax notoginseng* (BURK.) F.H. CHEN (Araliaceae): structures of new saponins, notoginsenosides-R1 and -R2, and identification of ginsenosides-Rg2 and -Rh1. Chem Pharm Bull 1981;29:2844–50.
- [39] Zhao P, Liu YQ, Yang CR. Minor dammarane saponins from *Panax notoginseng*. Phytochemistry 1996;41:1419–22.
- [40] Nguyen MD, Nguyen TN, Kasai R, Ito A, Yamasaki K, Tanaka O. Saponins from Vietnamese ginseng, *Panax vietnamensis* Ha et Grushv. Collected in central Vietnam. I. Chem Pharm Bull (Tokyo) 1993;41:2010–4.
- [41] Nguyen MD, Kasai R, Ohtani K, Ito A, Nguyen TN, Yamasaki K, Tanaka O. Saponins from Vietnamese ginseng, *Panax vietnamensis* Ha et Grushv. Collected in central Vietnam. II. Chem Pharm Bull 1994;42:634–40.
- [42] Duc NM, Kasai R, Ohtani K, Ito A, Nham NT, Yamasaki K, Tanaka O. Saponins from Vietnamese ginseng, *Panax vietnamensis* Ha et Grushv. collected in central Vietnam. III. Chem Pharm Bull (Tokyo) 1994;42:115–22.
- [43] Tawab MA, Bahr U, Karas M, Wurglics M, Schubert-Zsilavecz M. Degradation of ginsenosides in humans after oral administration. Drug Metab Disp 2003;31:1065–71.
- [44] Akao T, Kida H, Kanaoka M, Hattori M, Kobashi K. Intestinal bacterial hydrolysis is required for the appearance of compound K in rat plasma after oral administration of ginsenoside Rb1 from *Panax ginseng*. J Pharm Pharmacol 1998;50:1155–60.
- [45] Akao T, Kanaoka M, Kobashi K. Appearance of compound K, a major metabolite of ginsenoside Rb1 by intestinal bacteria, in rat plasma after oral administration—measurement of compound K by enzyme immunoassay. Biol Pharm Bull 1998;21:245–9.
- [46] Shibata S. Chemistry and cancer preventing activities of ginseng saponins and some related triterpenoid compounds. J Kor Med Sci 2001;16(Suppl.):S28–37.
- [47] Kim KA, Yoo HH, Gu W, Yu DH, Jin MJ, Choi HL, Yuan K, Guerin-Deremaux L, Kim DH. Effect of a soluble prebiotic fiber, NUTRIOSE, on the absorption of ginsenoside Rd in rats orally administered ginseng. J Ginseng Res 2014;38:203–7.
- [48] Kim KA, Yoo HH, Gu W, Yu DH, Jin MJ, Choi HL, Yuan K, Guerin-Deremaux L, Kim DH. A prebiotic fiber increases the formation and subsequent absorption of compound K following oral administration of ginseng in rats. J Ginseng Res 2015;39:183–7.
- [49] Lee J, Lee E, Kim DH, Lee J, Yoo J, Koh B. Studies on absorption, distribution and metabolism of ginseng in humans after oral administration. J Ethnopharmacol 2009;122:143–8.
- [50] Hasegawa H, Sung JH, Matsumiya S, Uchiyama M. Main ginseng metabolites formed by intestinal bacteria. Planta Med 1996;62:453–5.
- [51] Karikura M, Miyase T, Tanizawa H, Takino Y, Taniyama T, Hayashi T. Studies on absorption, distribution, excretion and metabolism of ginseng saponins. V. The decomposition products of ginsenoside Rb2 in the large intestine of rats. Chem Pharm Bull 1990;38:2859–61.
- [52] Odani T, Tanizawa H, Takino Y. Studies on the absorption, distribution, excretion and metabolism of ginseng saponins. II. The absorption, distribution and excretion of ginsenoside Rg1 in the rat. Chem Pharm Bull 1983;31:292–8.
- [53] Strömbom J, Sandberg F, Dencker L. Studies on absorption and distribution of ginsenoside Rg1 by whole-body autoradiobiography and chromatography. Acta Pharmaceut Suecica 1985;22:113–22.
- [54] Park EK, Shin YW, Lee HU, Kim SS, Lee YC, Lee BY, Kim DH. Inhibitory effect of ginsenoside Rb1 and compound K on NO and prostaglandin E2 biosyntheses of RAW264.7 cells induced by lipopolysaccharide. Biol Pharm Bull 2005;28:652–6.
- [55] Kato H, Shimada F, Yano S, Kanaoka M. Determination of ginsenoside Rb1 in plasma of human after intake of red ginseng powder. In: Abstract of papers, 11th Symposium of the Medical Society for Red Ginseng Research, Kobe, Japan; 1990. p. 36 [abstract].
- [56] Scalbert A, Williamson G. Dietary intake and bioavailability of polyphenols. J Nutri 2000;130:2073S–85S.
- [57] Bae EA, Park SY, Kim DH. Constitutive beta-glucosidases hydrolyzing ginsenoside Rb1 and Rb2 from human intestinal bacteria. Biol Pharm Bull 2000;23:1481–5.
- [58] Bae EA, Han MJ, Choo MK, Park SY, Kim DH. Metabolism of 20(S)- and 20(R)-ginsenoside Rg3 by human intestinal bacteria and its relation to *in vitro* biological activities. Biol Pharm Bull 2002;25:58–63.
- [59] Bae EA, Choo MK, Park EK, Park SY, Shin HY, Kim DH. Metabolism of ginsenoside Rc and its related anti-allergic activity. Biol Pharm Bull 2002;25:743–7.
- [60] Kim DH. Metabolism of ginsenosides to bioactive compounds by intestinal microflora and its industrial application. J Ginseng Res 2009;33:165–76.
- [61] Park SY, Bae EA, Sung JH, Lee SK, Kim DH. Purification and characterization of ginsenoside Rb1-metabolizing beta-glucosidase from *Fusobacterium* K-60, a human intestinal anaerobic bacterium. Biosci Biotechnol Biochem 2001;65:1163–9.
- [62] Shin HY, Lee JH, Lee JY, Han YO, Han MJ, Kim DH. Purification and characterization of ginsenoside Ra-hydrolyzing beta-D-xylosidase from *Bifidobacterium breve* K-110, a human intestinal anaerobic bacterium. Biol Pharm Bull 2003;26:1170–3.
- [63] Bae EA, Shin JE, Kim DH. Metabolism of ginsenoside Re by human intestinal microflora and its estrogenic effect. Biol Pharm Bull 2005;28:1903–8.
- [64] Jeong JJ, Van Le TH, Lee SY, Eun SH, Nguyen MD, Park JH, Kim DH. Anti-inflammatory effects of vina-ginsenoside R2 and majonoside R2 isolated from *Panax vietnamensis* and their metabolites in lipopolysaccharide-stimulated macrophages. Int Immunopharmacol 2015;28:700–6.
- [65] Lee SY, Jeong JJ, Le TH, Eun SH, Nguyen MD, Park JH, Kim DH. Ocotillo, a majonoside R2 metabolite, ameliorates 2,4,6-trinitrobenzenesulfonic acid-induced colitis in mice by restoring the balance of Th17/Treg cells. J Agric Food Chem 2015;63:7024–31.
- [66] Wakabayashi C, Hasegawa H, Murata J, Saiki I. *In vivo* antimetastatic action of ginseng protopanaxadiol saponins is based on their intestinal bacterial metabolites after oral administration. Oncol Res 1998;9:411–7.
- [67] Hasegawa H, Lee KS, Nagaoka T, Tezuka Y, Uchiyama M, Kadota S, Saiki I. Pharmacokinetics of ginsenoside deglycosylated by intestinal bacteria and its transformation to biologically active fatty acid esters. Biol Pharm Bull 2000;23:298–304.
- [68] Attele AS, Wu JA, Yuan CS. Ginseng pharmacology: multiple constituents and multiple actions. Biochem Pharmacol 1999;58:1685–93.
- [69] Kennedy DO, Scholey AB. Ginseng: potential for the enhancement of cognitive performance and mood. Pharmacol Biochem Behav 2003;75:687–700.
- [70] Scaglione F, Ferrara F, Dugnani S, Falchi M, Santoro G, Frascini F. Immunomodulatory effects of two extracts of *Panax ginseng* C.A. Meyer. Drug Exp Clin Res 1990;16:537–42.
- [71] Singh VK, Agarwal SS, Gupta BM. Immunomodulatory activity of *Panax ginseng* extract. Planta Med 1984;50:462–5.
- [72] Matsuda H, Namba K, Fukuda S, Tani T, Kubo M. Pharmacological study on *Panax ginseng* C.A. Meyer. IV. Effects of red ginseng on experimental disseminated intravascular coagulation. (3). Effect of ginsenoside-Ro on the blood coagulative and fibrinolytic system. Chem Pharm Bull 1986;34:2100–4.
- [73] Yokozawa T, Kobayashi T, Oura H, Kawashima Y. Studies on the mechanism of the hypoglycemic activity of ginsenoside-Rb2 in streptozotocin-diabetic rats. Chem Pharm Bull 1985;33:869–72.
- [74] Xie JT, Mehendale SR, Li X, Quigg R, Wang X, Wang CZ, Wu JA, Aung HH, Rue PA, Bell GI, et al. Anti-diabetic effect of ginsenoside Re in ob/ob mice. Biochim Biophys Acta 2005;1740:319–25.
- [75] Chang YS, Seo EK, Gyllenhaal C, Block KI. *Panax ginseng*: a role in cancer therapy? Integr Cancer Therap 2003;2:13–33.
- [76] Helms S. Cancer prevention and therapeutics: *Panax ginseng*. Altern Med Rev 2004;9:259–74.
- [77] Choo MK, Park EK, Han MJ, Kim DH. Antiallergic activity of ginseng and its ginsenosides. Planta Med 2003;69:518–22.
- [78] Park EK, Choo MK, Kim EJ, Han MJ, Kim DH. Antiallergic activity of ginsenoside Rh2. Biol Pharm Bull 2003;26:1581–4.
- [79] Park EK, Choo MK, Han MJ, Kim DH. Ginsenoside Rh1 possesses antiallergic and anti-inflammatory activities. Int Arch Allergy Immunol 2004;133:113–20.
- [80] Kim ND, Kang SY, Kim MJ, Park JH, Schini-Kerth VB. The ginsenoside Rg3 evokes endothelium-independent relaxation in rat aortic rings: role of K<sup>+</sup> channels. Eur J Pharmacol 1999;367:51–7.
- [81] Park EK, Choo MK, Oh JK, Ryu JH, Kim DH. Ginsenoside Rh2 reduces ischemic brain injury in rats. Biol Pharm Bull 2004;27:433–6.
- [82] Shieh PC, Tsao CW, Li JS, Wu HT, Wen YJ, Kou DH, Cheng JT. Role of pituitary adenylate cyclase-activating polypeptide (PACAP) in the action of ginsenoside Rh2 against beta-amyloid-induced inhibition of rat brain astrocytes. Neurosci Lett 2008;434:1–5.
- [83] Su X, Pei Z, Hu S. Ginsenoside Re as an adjuvant to enhance the immune response to the inactivated rabies virus vaccine in mice. Int Immunopharmacol 2014;20:283–9.
- [84] Lee EJ, Ko E, Lee J, Rho S, Ko S, Shin MK, Min BI, Hong MC, Kim SY, Bae H. Ginsenoside Rg1 enhances CD4(+) T-cell activities and modulates Th1/Th2 differentiation. Int Immunopharmacol 2004;4:235–44.



- [85] Lee SJ, Ko WG, Kim JH, Sung JH, Moon CK, Lee BH. Induction of apoptosis by a novel intestinal metabolite of ginseng saponin via cytochrome c-mediated activation of caspase-3 protease. *Biochem Pharmacol* 2000;60:677–85.
- [86] Tatsuka M, Maeda M, Ota T. Anticarcinogenic effect and enhancement of metastatic potential of BALB/c 3T3 cells by ginsenoside Rh(2). *Jpn J Cancer Res* 2001;92:1184–9.
- [87] Shin YW, Kim DH. Antipruritic effect of ginsenoside rb1 and compound k in scratching behavior mouse models. *J Pharm Sci* 2005;99:83–8.
- [88] Shin YW, Bae EA, Kim SS, Lee YC, Kim DH. Effect of ginsenoside Rb1 and compound K in chronic oxazolone-induced mouse dermatitis. *Int Immunopharmacol* 2005;5:1183–91.
- [89] Choo MK, Sakurai H, Kim DH, Saiki I. A ginseng saponin metabolite suppresses tumor necrosis factor- $\alpha$ -promoted metastasis by suppressing nuclear factor- $\kappa$ B signaling in murine colon cancer cells. *Oncol Rep* 2008;19:595–600.
- [90] Cui JF, Björkhem I, Eneroth P. Gas chromatographic-mass spectrometric determination of 20(S)-protopanaxadiol and 20(S)-protopanaxatriol for study on human urinary excretion of ginsenosides after ingestion of ginseng preparations. *J Chromatogr B Biomed Sci Appl* 1997;689:349–55.
- [91] Bae EA, Hyun YJ, Choo MK, Oh JK, Ryu JH, Kim DH. Protective effect of fermented red ginseng on a transient focal ischemic rats. *Arch Pharm Res* 2004;27:1136–40.
- [92] Kim KA, Jung IH, Park SH, Ahn YT, Huh CS, Kim DH. Comparative analysis of the gut microbiota in people with different levels of ginsenoside Rb1 degradation to compound K. *PLoS One* 2013;8:e62409.
- [93] Lee DS, Kim YS, Ko CN, Cho KH, Bae HS, Lee KS, Kim JJ, Park EK, Kim DH. Fecal metabolic activities of herbal components to bioactive compounds. *Arch Pharm Res* 2002;25:165–9.
- [94] Yim JS, Kim YS, Moon SK, Cho KH, Bae HS, Kim JJ, Park EK, Kim DH. Metabolic activities of ginsenoside Rb1, baicalin, glycyrrhizin and geniposide to their bioactive compounds by human intestinal microflora. *Biol Pharm Bull* 2004;27:1580–3.
- [95] Choi JR, Hong SW, Kim Y, Jang SE, Kim NJ, Han MJ, Kim DH. Metabolic activities of ginseng and its constituents, ginsenoside rb1 and rg1, by human intestinal microflora. *J Ginseng Res* 2011;35:301–7.
- [96] Kim DH. Herbal medicines are activated by intestinal microflora. *Nat Prod Sci* 2002;8:35–43.
- [97] Kim DH. Gut microbiota-mediated drug-antibiotic interactions. *Drug Metab Dispos* 2015;43:1581–9.
- [98] Tamura G, Gold C, Ferro-Luzzi A, Ames BN. Fecalase: a model for activation of dietary glycosides to mutagens by intestinal flora. *Proc Natl Acad Sci USA* 1980;77:4961–5.
- [99] Kobashi K, Nakata H, Takebe H, Terasawa K. Relation of intestinal microflora to Syo. *Wakan-iyaku-kaishi* 1984;1:166–7.
- [100] Trinh HT, Han SJ, Kim SW, Lee YC, Kim DH. *Bifidus* fermentation increases hypolipidemic and hypoglycemic effects of red ginseng. *J Microbiol Biotechnol* 2007;17:1127–33.