

Review Article



Dual Effect of Hepatic Macrophages on Liver Ischemia and Reperfusion Injury during Liver Transplantation

Tian-Fei Lu^{1,†}, Tai-Hua Yang^{2,†}, Cheng-Peng Zhong^{1,†}, Chuan Shen¹, Wei-Wei Lin³, Guang-Xiang Gu¹, Qiang Xia¹, Ning Xu^{1,*}

¹Department of Liver Surgery, Renji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200127, China

²Department of Gastroenterology, Hepatology, and Endocrinology, Hannover Medicine School, Hannover 30625, Germany

³Department of Laboratory, Renji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200127, China



Received: Feb 6, 2018

Revised: Jun 20, 2018

Accepted: Jun 21, 2018

*Correspondence to

Ning Xu

Department of Liver Surgery, Renji Hospital, Shanghai Jiaotong University School of Medicine, No. 160, Pujian Road, Pudong New District, Shanghai 200127, China.
Email: xuning123@hotmail.com

[†]Tian-Fei Lu, Tai-Hua Yang, and Cheng-Peng Zhong contributed equally to this work.

Copyright © 2018. The Korean Association of Immunologists

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Conflict of Interest

The authors declare no potential conflicts of interest.

Abbreviations

ARE, antioxidant response element; ATP, adenosine-5'-triphosphate; DAMP, damage-associated molecular pattern; DC, dendritic cell; DT, diphtheria toxin; HMGB1, high mobility group box 1; HO, heme oxygenase; ICAM, intercellular adhesion molecule; IFN, interferon; IRI, ischemia-reperfusion injury; KC, Kupffer cell; Keap1, Kelch-like ECH-

ABSTRACT

Ischemia-reperfusion injury (IRI) is a major complication in liver transplantation (LT) and it is closely related to the recovery of grafts' function. Researches has verified that both innate and adaptive immune system are involved in the development of IRI and Kupffer cell (KC), the resident macrophages in the liver, play a pivotal role both in triggering and sustaining the sterile inflammation. Damage-associated molecular patterns (DAMPs), released by the initial dead cell because of the ischemia insult, firstly activate the KC through pattern recognition receptors (PRRs) such as toll-like receptors. Activated KCs is the dominant players in the IRI as it can secret various pro-inflammatory cytokines to exacerbate the injury and recruit other types of immune cells from the circulation. On the other hand, KCs can also serve in a contrary way to ameliorate IRI by upregulating the anti-inflammatory factors. Moreover, new standpoint has been put forward that KCs and macrophages from the circulation may function in different way to influence the inflammation. Managements towards KCs are expected to be the effective way to improve the IRI.

Keywords: Kupffer cells; Hepatic macrophages; Ischemia-reperfusion injury; Liver transplantation; Pattern recognition receptors; Tumor necrosis factor- α

INTRODUCTION

With improvements in surgical procedures, immunosuppression techniques and perioperative care, liver transplantation (LT) has evolved into a definitive treatment for all types of end-stage liver diseases and is performed in over 80 countries (1). Although the 1-year survival rates of patients undergoing LT have exceeded 80% (1), some issues are still troublesome limitations, including ischemia-reperfusion injury (IRI). IRI, occurring during hemorrhagic shock, liver resection and LT, is a biphasic pathophysiological process that includes the initial ischemic liver damage and subsequent reperfusion injury. The ischemia insult characterized with the restriction of the blood supply during organ procurement can cause the cellular metabolic stress due to the hypoxia, adenosine-5'-triphosphate (ATP) and glycogen consumption, mitochondrial dysfunction, which all contributes to the initial

associated protein 1; LPS, lipopolysaccharide; LT, liver transplantation; NF, nuclear factor; NK, natural killer; NKT, natural killer T; NLRP3, nucleotide-binding domain leucine-rich repeat containing family pyrin domain containing 3; Nrf2, nuclear factor erythroid 2-related factor 2; PRR, pattern recognition receptor; ROS, reactive oxygen species; TLR, Toll-like receptor; TIM, T cell, immunoglobulin, mucin-containing molecules; TNF, tumor necrosis factor; WT, wild type

Author Contributions

Conceptualization: Xu N, Xia Q; Data curation: Lu TF, Yang TH, Zhong CP, Shen C, Lin WW, Gu GX; Writing - original draft: Lu TF, Yang TH, Zhong CP.

cell deaths. The reperfusion injury, on the other hand, refers to the phenomenon that after the restoration of the blood flow and reoxygenation, the livers usually suffer from a severer damage, which have a great relationship with not only the metabolic disturbance and oxygen radicals, but also the inflammation reaction cascade involving with various immune cells, complements and cytokines or chemokines they produced. Researches have revealed that many pathological processes are involved in the development of IRI, including the increasing vascular permeability and leakage, activation of cell death programs (apoptosis, autophagy-associated cell death, and necrosis), transcriptional reprogramming, generation of reactive oxygen species (ROS) and nitrogen species, and the activation of the innate and adaptive immune network (2-4). It is of particular note that the hepatocellular damage concerning IRI is crucial to the occurrence of poor early grafts function, even primary nonfunction. Furthermore, IRI may also be responsible for a considerable portion of postoperative acute and chronic rejection. Therefore, grafts with longer ischemia time are usually discarded, which further worsen the organ shortage (2,4). Unfortunately, the mechanisms of IRI remain much unknown and no effective therapy is available in the clinical application despite of many great advancements in basic animal models' researches.

The liver is a unique organ that harbors various types of innate cells, including resident macrophages (Kupffer cells; KCs), dendritic cells (DCs), natural killer (NK) cells, and natural killer T (NKT) cells (5,6), which all play important roles in IRI. In this review, we aimed to summarize the current understanding of KCs that are the predominant players in sterile inflammation and highlighted the activation and roles of these cells in IRI.

QUIESCENT STATE OF KCs

Proper macrophage nomenclature is intensively debated at present, but basically the hepatic macrophages origin from three sources (Fig. 1) (7-9). KCs are a group of hepatic macrophages and they are the largest population of tissue-resident macrophages in the body. They reside within the liver sinusoid and play a pivotal role in the immune response. In the steady state condition, the intimate contact with circulation blood flowing from the portal vein or hepatic artery allows them to phagocytize majority of the pathogens such as microorganisms and their products, as well as those deriving from the gastrointestinal ducts. Besides, they are also in charge of the clearance of the other substance including aged erythrocytes, cell debris, tumor cells, immune complexes (10-13). It is now convinced that the KCs in a healthy liver exhibit the 'tolerogenic' phenotypes, which maintain the immunological tolerance. However, in the diseases state, they may undergo phenotype switch to participant in the immune reaction. Basically, pro-inflammatory cytokines such as tumor necrosis factor (TNF)- α , IL-6, IL-12, IL-1 β , and inducible nitric oxide (NO) synthase are secreted by a group of KCs while another type may work contrarily to increase the expression of anti-inflammatory mediators. Furthermore, they could also interact with other immune cells to facility the response (12,13). Lots of works has been done to elaborate the contributions of KCs to the liver diseases, such as fibrosis (12), alcoholic and nonalcoholic fatty liver diseases (13), hepatotropic virus infection (14), and cholestatic liver injury (15).

In conclusion, in steady state condition, the KCs exhibits the 'tolerogenic' type and are in charge of the clearance of pathogens. In the diseases, they could change phenotype and play a role in the immune reaction.

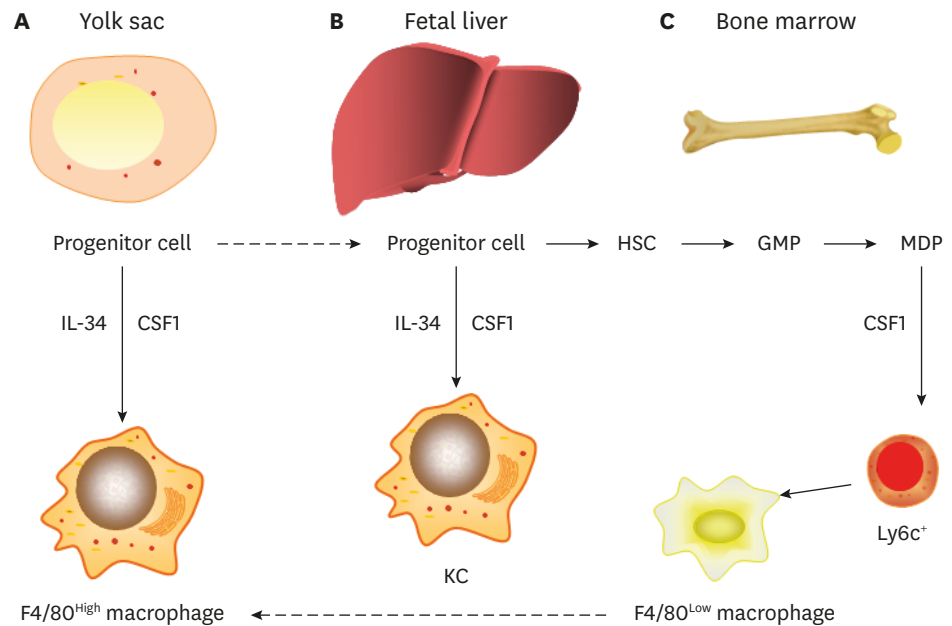


Figure 1. Origins of hepatic macrophages.

The hepatic macrophages in adults derives from at least 3 sources. (A) Yolk sac. It can produce progenitors that populate all tissues and that have progeny that persist throughout life as F4/80 high resident macrophages. These lineages are mainly regulated by CSF1R and its ligands, IL-34 and CSF1. (B) Fetal liver. This is less well defined but seems to contribute to the production of adult KCs. (C) Bone marrow. The source which give rise to circulating monocytes and progeny F4/80 low macrophages. Macrophages that are F4/80 low emanate from Ly6c1 monocytes. The exact role of the patrolling Ly6c negative macrophages, and the contribution of fetal liver to adult tissue macrophages remains unclear.

CSF1, colony-stimulating factor 1; HSC, hepatic stellate cell; GMP, granulocyte-monocyte progenitor; MDP, monocyte dendritic cell progenitor.

ACTIVATION OF KCs

Until now, 2 different stages of IRI have been identified (2,4). In the ischemia injury stage, hypoxia and accompanying metabolic disturbances induce initial cell death, which releases diverse ‘alarmins’ such as damage-associated molecular patterns (DAMPs), ROS, and DNA fragments. In the reperfusion injury stage, in response to stimulation by alarmins, resident immune cells are activated, and these cells facilitate the generation of a pro-inflammatory milieu, which in turn amplifies and sustains immune cascades by recruiting immune cells from the circulation.

Toll-like receptors (TLRs) are a family of pattern recognition receptors (PRRs) that are expressed in various liver cells and can trigger immune inflammation in response to DAMPs and other alarmins (6,16). KCs express TLR2, TLR3, TLR4 and TLR9 on the cell surface or in endosomes/lysosomes (17). TLR4 signaling is a well-studied pathway that is critical not only for nonparenchymal cells, which are involved in the initiation of innate and adaptive immune responses (18), but also for parenchymal cells, which release danger signals such as high mobility group box 1 (HMGB1) (19,20).

HMGB1, the key endogenous DAMP molecule in liver immune activation, is upregulated as early as 1h after reperfusion (21). As a nuclear factor (NF), HMGB1 is actively released by hypoxic hepatocytes during IRI, and the process is promoted by TLR4-dependent ROS production and downstream calcium/calmodulin-dependent protein kinases-mediated

signaling (22). With the binding of TLR4 and its major ligand HMGB1, KCs are activated. Although the inhibition of the activity and release of HMGB1 exerts a protective effect during IRI (21,23), the biological role of HMGB1 seems more complex, as hepatocyte-specific HMGB1 deletion increases nuclear instability and histone release and, consequently, worsens liver IRI through the activation of TLR9 (24). Hence, during liver IRI, HMGB1 may have 2 roles, namely, a beneficial intracellular role and an injurious extracellular role (25).

Innate immune activation involves a complex network consisting of multiple cells and molecules (2,6). In addition to HMGB1, other DAMPs including histones, DNA fragments and ATP, mitochondrial ROS and complement proteins stimulate KCs and other distinctive immune cells via different PRRs, such as TLR9 (recognizing histones) (26), TLR3 (recognizing RNA) (27,28), TLR4 (recognizing heat shock protein [HSP]-70) (29), and nucleotide-binding domain leucine-rich repeat containing family pyrin domain containing 3 (NLRP3) (recognizing ATP) (30-32).

Lipopolysaccharide (LPS) is a gut-derived endotoxin that can translocate into the hepatic circulation during IRI because the occlusion of portal vein blood flow increases the permeability of the intestinal wall. LPS has been reported to bind to LPS-binding proteins, engage CD14 and activate KCs through the mediation of TLR4 signaling (33). However, whether LPS can trigger or sustain the activation of KCs remains to be determined because an increase in LPS was detected in late phases of IRI and because LPS-neutralizing agents failed to protect mouse liver during early phases of IRI (34). It is thus likely that this gut-derived endotoxin sustains immune inflammation (2).

In conclusion, KCs are activated by the DAMPs or other alarmins through PRRs.

DUAL EFFECT OF ACTIVATED KCs IN INNATE AND ADAPTIVE IMMUNE SIGNALING

The response of activated KCs to reperfusion is bifunctional, with the secretion of both pro-inflammatory and anti-inflammatory factors (Fig. 2). TNF- α , produced by KCs, is one of the principal cytokines in IRI. TNF- α exacerbates damage and induces apoptosis by directly binding to the TNF receptor on the surface of hepatocytes and activating the NF- κ B and c-Jun N-terminal kinase (JNK) pathways (35). A recent study also revealed that TNF- α -induced tissue injury can be mediated by the peroxisome proliferator-activated receptor- γ co-activator (PGC)-1 α /mitofusion (Mfn)-2 pathway, which regulates mitochondrial functions and energy metabolism (36). Furthermore, TNF- α can increase the severity of inflammation by upregulating the expression of other inflammatory factors including ROS, intercellular adhesion molecule (ICAM)-1, vascular adhesion molecule (VCAM)-1 and P-selectin and inducing the recruitment of neutrophils (37,38). Notably, KCs and TNF- α also participate in the mediation of hepatic injury after remote organ IRI (39). TNF- α suppression (40) or gene silencing (41) play marked protective roles in animal IRI models; however, improvements have also been observed with TNF- α preconditioning (42), indicating the dual roles played by this molecule (43).

Other quintessential pro-inflammatory cytokines include IL-1 β and IL-18, whose synthesis, activation and release are controlled by caspase-1 (44). NLRP3 is another PRR that is mainly expressed on KCs. NLRP3 can be activated in response to stimulation by DAMPs, and it

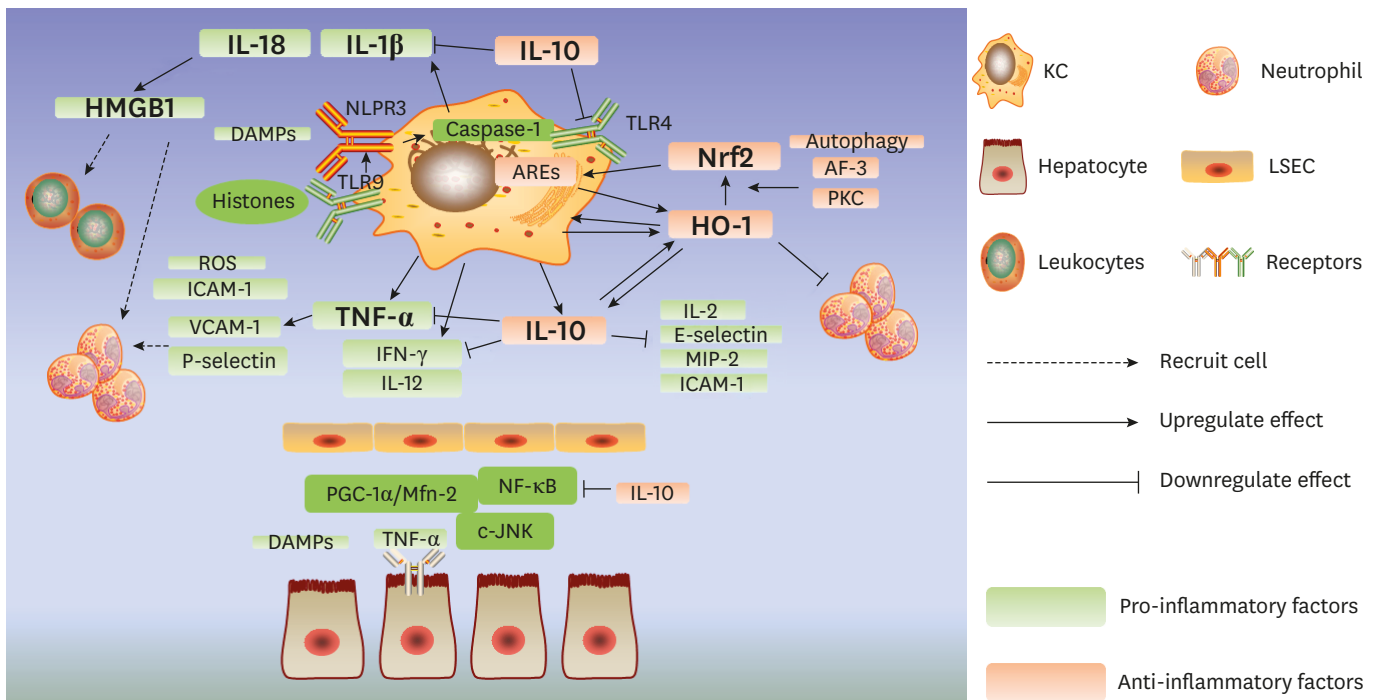


Figure 2. Dual effect of activated KCs in IRI.

The response of activated KCs to reperfusion is bifunctional with the secretion of both pro-inflammatory and anti-inflammatory factors. TNF- α , IL-1 β , IL-18, IFN- γ , and IL-12 secreted by KCs show a pro-inflammatory effect, in contrast, IL-10 and HO-1/Nrf2 pathway contribute to the anti-inflammatory regulation. JNK, c-Jun N-terminal kinase; PGC-1 α , peroxisome proliferator-activated receptor- γ co-activator 1 α ; Mfn-2, mitofusion 2; VCAM-1, vascular adhesion molecule 1; MIP-2, macrophage inflammatory protein 2.

induces the assembly of the NLRP3 inflammasome with apoptosis-associated speck-like protein containing a caspase activation and recruitment domain (ASC) and pro-caspase1. Inflammasomes are multiprotein complexes that can recognize damage signals and directly activate caspase-1 (32,44,45). Studies have demonstrated that endogenous extracellular histones binding to TLR9 can activate the NLRP3 inflammasome, which can then initiate the activation of caspase-1 and the secretion of IL-1 β and IL-18 (46). This promotes the production of HMGB1 (47) and the recruitment of neutrophils and leukocytes from the circulation (37,46). In addition, the absent in melanoma (AIM)-2 inflammasome, activated by double-stranded DNA in KCs, also contributes to caspase-1 activation and propagates the injury (48).

The other pro-inflammatory factors that are completely or partly derived from KCs include ROS, interferon (IFN)- γ and IL-12, which recruit immune cells, cause damage to hepatocytes or sinusoidal endothelial cells and increase the expression of other inflammatory factors and adhesion molecules (37,38).

Although the inhibition of KCs has shown promising outcomes in ameliorating liver IRI in some studies (49-51), KC-depleted mice have also shown an increased level of injury followed with higher mortality (52,53). IL-10 is one of the primary anti-inflammatory cytokines produced by KCs, and it can suppress NF- κ B activation and dramatically inhibit the expression of pro-inflammatory factors such as TNF- α , IL-1 β , IFN- γ , IL-2, E-selectin, macrophage inflammatory protein (MIP)-2, and ICAM-1 (52-56). Moreover, TLR4 suppression and apoptosis inhibition are also observed in animals subjected to adenovirus (Ad)-based viral IL-10 gene transfer (56). Furthermore, IL-10 can reduce inflammation-associated injury

by upregulating the expression of heme oxygenase (HO)-1 (56,57), which plays a profound cytoprotective role in liver IRI. HO-1 is predominantly secreted by KCs and can drive the differentiation of these cells, leading to an increase in the number of 'anti-inflammatory' macrophage populations (58). Both warm ischemia-reperfusion (IR) and orthotopic LT animal models have validated that Ad-based HO-1-transduced macrophages can attenuate liver IRI and improve survival by not only decreasing local neutrophil accumulation and pro-inflammatory factor expression but also increasing the level of IL-10. Cell apoptosis is also ameliorated under these conditions (59,60).

It is well documented that the Kelch-like ECH-associated protein 1 (Keap1)/nuclear factor erythroid 2-related factor 2 (Nrf2)/antioxidant response element (ARE) pathway is responsible for the regulation of HO-1. Under normal conditions, Nrf2 is sequestered in the cytoplasm by Keap1, which facilitates its degradation via the ubiquitin-proteasome pathway. However, under stressful circumstances, in response to specific chemical signals, Nrf2 can dissociate from Keap1 and translocate into the nucleus. The binding of Nrf2 to AREs upregulates the transcription of numerous antioxidants and anti-apoptotic proteins including HO-1 (61,62). Interestingly, another study revealed that nuclear HO-1 can interact with Nrf2 to enhance the stabilization of Nrf2 and, consequently, form a positive loop (63). Upstream factors such as protein kinase C (PKC) and activating transcription factor (ATF)-3 are also found to mediate the Nrf2/HO-1 pathway, and this could be an innovative strategy to manage IRI (64,65). Moreover, autophagy has been shown to be involved in the protection of liver from IRI (66,67) via the Nrf2/HO-1 pathway (68,69).

Though the innate immune response takes the dominant role in hepatic IRI, robust adaptive immune reaction is also indispensable, which mainly depends on the CD4⁺ T cells (2). In the absence of exogenous antigen stimulation, CD4⁺ T cells can be activated by the pro-inflammatory cytokine or chemokines produced by the KCs, neutrophils and hepatocytes and recruited to the post-ischemic livers (37). What is more, the accumulation of CD4⁺ T cells comes rapidly after the IR and facilitate the accumulation of neutrophils via IL-17 (2,37,70). On the other hand, the activated CD4⁺ T cell can in turn activate the innate immune cells including KCs and sustain the inflammation via the CD154/CD40 (2), TIM-1/TIM-4 (2,71). CD154 and TIM-1 are expressed in the CD4⁺ T cells and the corresponding ligands CD40 and TIM-4 are expressed in the macrophage and antigen-presenting cells. In contrast, the PD-1/B7-H1 (72) and TIM-3-galectin-9 pathway play a negatively stimulatory role in promoting IRI (73). That means, the KC and CD4⁺ T cell could interact with each other to regulate the IRI.

In summary, the KCs could play a dual effect in IRI and it could interact with the CD4⁺ T cells in the adaptive immune reaction.

DISTINCTIVE ROLES OF RESIDENT KCs AND INFILTRATING MACROPHAGES

Human macrophages can be classified into three subsets based on the expression of CD11b, CD68 and CD32. In contrast to macrophages in circulation or other organs expressing CD11b, liver-resident KCs are mainly CD68⁺ and/or CD32⁺ (11). In addition, CD32/CD68⁺ KCs can be depleted by clodronate liposomes, whereas the CD11b⁺ cells are sensitive to radiation and diphtheria toxin (DT) (6,11). Depletion of the circulation macrophages with the treatment of DT in the CD11b-DT receptor (DTR) mice resulted in IRI-resistance when

compared with the untreated mouse, which presented a decreased alanine transaminase (ALT) level and preserved architecture and the damage could be restored by the adoptive infusion of macrophages, which demonstrated the pro-inflammatory role of the non-resident macrophages. However, when infusing the TIM-4 deficient macrophages, the restoration was failed, which highlighted the importance of TIM-4 in the hepatic IRI, which could be a potential therapeutic target (74).

Other innate immune cells also function in the hepatic IRI. Studies has demonstrated that both the resident and infiltrating types of DCs are involved in the hepatic IRI. Specifically, the blood recruited DCs may serve as pro-inflammatory cells as the adoptive infusion of wild-type DCs to the Flt3L KO (DC-deficient) or wild type (WT) mice increased IRI injury while the resident DCs may function in a contrary way as the grafts lacking resident DCs (Flt3L KO liver grafts) suffered severer injuries when transplanted into a WT mice compared with the WT livers transplanted into WT mice (75). Besides, those DCs may protect the liver from IRI by secreting IL-10 via TLR9 signals (76). Similar opposing roles has also been observed in NKT cells that type I NKT cells promoted the IRI injury and the type II suppressed it. When activated by the sulfatide, the type II NKT cells would reduce the production of IFN- γ from type I NKT cells and reduce the recruitment of myeloid cells, which could be a novel therapeutic strategy for IRI (77). IFN- γ is also secreted by the CD-39 mediated NK cells. Studies of Beldi (78) has revealed the deletion of CD39 on NK cell could abrogate the level of IFN- γ and attenuates hepatic IRI. Kimura et al. (79) also showed that NK cells diminished quickly in the allogeneic grafts in the rat model of transplantation but not in the syngeneic grafts, which demonstrated its role of alien recognition. Furthermore, depletion of mature NK cells improved the hepatic IRI.

In summary, other innate cells also play significant role in the IRI.

CONCLUSION AND FUTURE DIRECTIONS

In conclusion, IRI is a severe problem during LT, and it activates both the innate and adaptive immune system, leading to sterile inflammatory responses to damage hepatocytes. Liver-resident macrophages or KCs, activated by the interaction between alarmins and PRRs, play a pivotal role during IRI. Activated KCs can function as both a pro-inflammatory and an anti-inflammatory manner.

Studies investigating complex signaling pathways in KCs may aid in the development of effective therapeutic strategies to attenuate inflammation-associated injury. Specifically, inhibition of pro-inflammatory signaling combine with/or stimulation of anti-inflammatory regulators can efficiently ameliorate IRI. NF- κ B is a main pathway activated by TNF- α and its suppression shows an obvious reduction effect of IRI (40,53,80-82). Butyrate as a 4-carbon fatty acid help reduce liver IRI by inhibiting NF- κ B (80). Similarly, down regulation effect of epigallocatechin-3-gallate and peptidyl-prolyl isomerase, Pin1 showed an attenuation of IRI (83,84). HMGB 1 as a key mediator of pro-inflammatory pathway also been investigated to protect IRI (85,86). Tanshinone IIA shows a potential role of prevention of IRI by down-regulation of the HGMB1-TLR-4/NF- κ B pathway in KCs and activation of phosphatase and tensin homolog deleted on chromosome ten (PTEN)/phosphatidylinositol 3 kinase (PI3K)/AKT pathway (85). Glycyrrhizin is a natural anti-inflammatory and antiviral triterpene in clinical use. Ogiku et al. (86) figure out the expression of HGMB1 in serum and production

in HGMB1 in KCs was reduced in the glycyrrhizin group compared with the control. Suppression of pro-inflammatory star mediator TNF- α by glutathione (40) and improvement of anti-inflammatory molecule IL-10 (53) were also verified their controversial effect in the process of IRI development. Functional HO-1 protein conjugated to a cell-penetrating peptide exert significant cytoprotective effect to hepatocytes *in vitro* (87). Some groups approve the positive influence by altering the M2 macrophage polarization (88,89). The small molecules like ROS, NO also can be a power clinical treatment tools for the IRI patients after LT (90,91). Interestingly, non-coding RNA miR-155 deficiency plays an effective role in attenuating liver IRI likely by regulating the activation and inflammatory response, as well as modifying the polarization of KCs (92).

Overall, the efforts to combine the repression of pro-inflammatory mediator with promotion of anti-inflammatory elements in KCs will further improve the prognosis of LT patient suffering from IRI. Recent findings motivate the search for novel therapeutic targets inside KCs to aid in moving personalized health care of IRI patients a step forward.

REFERENCES

- Zarrinpar A, Busuttil RW. Liver transplantation: past, present and future. *Nat Rev Gastroenterol Hepatol* 2013;10:434-440.
[PUBMED](#) | [CROSSREF](#)
- Zhai Y, Petrowsky H, Hong JC, Busuttil RW, Kupiec-Weglinski JW. Ischaemia-reperfusion injury in liver transplantation--from bench to bedside. *Nat Rev Gastroenterol Hepatol* 2013;10:79-89.
[PUBMED](#) | [CROSSREF](#)
- Eltzschig HK, Eckle T. Ischemia and reperfusion--from mechanism to translation. *Nat Med* 2011;17:1391-1401.
[PUBMED](#) | [CROSSREF](#)
- Zhai Y, Busuttil RW, Kupiec-Weglinski JW. Liver ischemia and reperfusion injury: new insights into mechanisms of innate-adaptive immune-mediated tissue inflammation. *Am J Transplant* 2011;11:1563-1569.
[PUBMED](#) | [CROSSREF](#)
- Dong Z, Wei H, Sun R, Tian Z. The roles of innate immune cells in liver injury and regeneration. *Cell Mol Immunol* 2007;4:241-252.
[PUBMED](#)
- Lu L, Zhou H, Ni M, Wang X, Busuttil R, Kupiec-Weglinski J, Zhai Y. Innate immune regulations and liver ischemia-reperfusion injury. *Transplantation* 2016;100:2601-2610.
[PUBMED](#) | [CROSSREF](#)
- Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. *Nature* 2013;496:445-455.
[PUBMED](#) | [CROSSREF](#)
- Naito M, Takahashi K, Nishikawa S. Development, differentiation, and maturation of macrophages in the fetal mouse liver. *J Leukoc Biol* 1990;48:27-37.
[PUBMED](#) | [CROSSREF](#)
- Naito M, Umeda S, Yamamoto T, Moriyama H, Umezumi H, Hasegawa G, Usuda H, Shultz LD, Takahashi K. Development, differentiation, and phenotypic heterogeneity of murine tissue macrophages. *J Leukoc Biol* 1996;59:133-138.
[PUBMED](#) | [CROSSREF](#)
- Davies LC, Jenkins SJ, Allen JE, Taylor PR. Tissue-resident macrophages. *Nat Immunol* 2013;14:986-995.
[PUBMED](#) | [CROSSREF](#)
- Ikarashi M, Nakashima H, Kinoshita M, Sato A, Nakashima M, Miyazaki H, Nishiyama K, Yamamoto J, Seki S. Distinct development and functions of resident and recruited liver Kupffer cells/macrophages. *J Leukoc Biol* 2013;94:1325-1336.
[PUBMED](#) | [CROSSREF](#)
- Tacke F, Zimmermann HW. Macrophage heterogeneity in liver injury and fibrosis. *J Hepatol* 2014;60:1090-1096.
[PUBMED](#) | [CROSSREF](#)

13. Dixon LJ, Barnes M, Tang H, Pritchard MT, Nagy LE. Kupffer cells in the liver. *Compr Physiol* 2013;3:785-797.
[PUBMED](#) | [CROSSREF](#)
14. Boltjes A, Movita D, Boonstra A, Woltman AM. The role of Kupffer cells in hepatitis B and hepatitis C virus infections. *J Hepatol* 2014;61:660-671.
[PUBMED](#) | [CROSSREF](#)
15. Sato K, Hall C, Glaser S, Francis H, Meng F, Alpini G. Pathogenesis of Kupffer cells in cholestatic liver injury. *Am J Pathol* 2016;186:2238-2247.
[PUBMED](#) | [CROSSREF](#)
16. Thaiss CA, Levy M, Itav S, Elinav E. Integration of innate immune signaling. *Trends Immunol* 2016;37:84-101.
[PUBMED](#) | [CROSSREF](#)
17. Chang WJ, Toledo-Pereyra LH. Toll-like receptor signaling in liver ischemia and reperfusion. *J Invest Surg* 2012;25:271-277.
[PUBMED](#) | [CROSSREF](#)
18. Tsung A, Hoffman RA, Izuishi K, Critchlow ND, Nakao A, Chan MH, Lotze MT, Geller DA, Billiar TR. Hepatic ischemia/reperfusion injury involves functional TLR4 signaling in nonparenchymal cells. *J Immunol* 2005;175:7661-7668.
[PUBMED](#) | [CROSSREF](#)
19. Hui W, Jinxiang Z, Heshui W, Zhuoya L, Qichang Z. Bone marrow and non-bone marrow TLR4 regulates hepatic ischemia/reperfusion injury. *Biochem Biophys Res Commun* 2009;389:328-332.
[PUBMED](#) | [CROSSREF](#)
20. Nace GW, Huang H, Klune JR, Eid RE, Rosborough BR, Korff S, Li S, Shapiro RA, Stolz DB, Sodhi CP, et al. Cellular-specific role of toll-like receptor 4 in hepatic ischemia-reperfusion injury in mice. *Hepatology* 2013;58:374-387.
[PUBMED](#) | [CROSSREF](#)
21. Tsung A, Sahai R, Tanaka H, Nakao A, Fink MP, Lotze MT, Yang H, Li J, Tracey KJ, Geller DA, et al. The nuclear factor HMGB1 mediates hepatic injury after murine liver ischemia-reperfusion. *J Exp Med* 2005;201:1135-1143.
[PUBMED](#) | [CROSSREF](#)
22. Tsung A, Klune JR, Zhang X, Jeyabalan G, Cao Z, Peng X, Stolz DB, Geller DA, Rosengart MR, Billiar TR. HMGB1 release induced by liver ischemia involves toll-like receptor 4 dependent reactive oxygen species production and calcium-mediated signaling. *J Exp Med* 2007;204:2913-2923.
[PUBMED](#) | [CROSSREF](#)
23. Zhang W, Li F, Ye Y, Liu Y, Yu S, Cen C, Chen X, Zhou L, Tang X, Yu J, et al. Isoglycyrrhizinate magnesium enhances hepatoprotective effect of FK506 on ischemia-reperfusion injury through HMGB1 inhibition in a rat model of liver transplantation. *Transplantation* 2017;101:2862-2872.
[PUBMED](#) | [CROSSREF](#)
24. Huang H, Nace GW, McDonald KA, Tai S, Klune JR, Rosborough BR, Ding Q, Loughran P, Zhu X, Beer-Stolz D, et al. Hepatocyte-specific high-mobility group box 1 deletion worsens the injury in liver ischemia/reperfusion: a role for intracellular high-mobility group box 1 in cellular protection. *Hepatology* 2014;59:1984-1997.
[PUBMED](#) | [CROSSREF](#)
25. Zhao G, Fu C, Wang L, Zhu L, Yan Y, Xiang Y, Zheng F, Gong F, Chen S, Chen G. Down-regulation of nuclear HMGB1 reduces ischemia-induced HMGB1 translocation and release and protects against liver ischemia-reperfusion injury. *Sci Rep* 2017;7:46272.
[PUBMED](#) | [CROSSREF](#)
26. Huang H, Evankovich J, Yan W, Nace G, Zhang L, Ross M, Liao X, Billiar T, Xu J, Esmon CT, et al. Endogenous histones function as alarmins in sterile inflammatory liver injury through toll-like receptor 9 in mice. *Hepatology* 2011;54:999-1008.
[PUBMED](#) | [CROSSREF](#)
27. Karikó K, Ni H, Capodici J, Lamphier M, Weissman D. mRNA is an endogenous ligand for toll-like receptor 3. *J Biol Chem* 2004;279:12542-12550.
[PUBMED](#) | [CROSSREF](#)
28. Cavassani KA, Ishii M, Wen H, Schaller MA, Lincoln PM, Lukacs NW, Hogaboam CM, Kunkel SL. TLR3 is an endogenous sensor of tissue necrosis during acute inflammatory events. *J Exp Med* 2008;205:2609-2621.
[PUBMED](#) | [CROSSREF](#)
29. Calderwood SK, Gong J, Murshid A. Extracellular HSPs: the complicated roles of extracellular HSPs in immunity. *Front Immunol* 2016;7:159.
[PUBMED](#) | [CROSSREF](#)
30. Nomura J, So A, Tamura M, Busso N. Intracellular ATP decrease mediates NLRP3 inflammasome activation upon nigericin and crystal stimulation. *J Immunol* 2015;195:5718-5724.
[PUBMED](#) | [CROSSREF](#)

31. Baron L, Gombault A, Fanny M, Villeret B, Savigny F, Guillou N, Panek C, Le Bert M, Lagente V, Rassendren F, et al. The NLRP3 inflammasome is activated by nanoparticles through ATP, ADP and adenosine. *Cell Death Dis* 2015;6:e1629.
[PUBMED](#) | [CROSSREF](#)
32. Szabo G, Petrasek J. Inflammasome activation and function in liver disease. *Nat Rev Gastroenterol Hepatol* 2015;12:387-400.
[PUBMED](#) | [CROSSREF](#)
33. Su GL. Lipopolysaccharides in liver injury: molecular mechanisms of Kupffer cell activation. *Am J Physiol Gastrointest Liver Physiol* 2002;283:G256-G265.
[PUBMED](#) | [CROSSREF](#)
34. Zhai Y, Qiao B, Shen XD, Gao F, Busuttill RW, Cheng G, Platt JL, Volk HD, Kupiec-Weglinski JW. Evidence for the pivotal role of endogenous toll-like receptor 4 ligands in liver ischemia and reperfusion injury. *Transplantation* 2008;85:1016-1022.
[PUBMED](#) | [CROSSREF](#)
35. Schwabe RF, Brenner DA. Mechanisms of liver injury. I. TNF- α -induced liver injury: role of IKK, JNK, and ROS pathways. *Am J Physiol Gastrointest Liver Physiol* 2006;290:G583-G589.
[PUBMED](#) | [CROSSREF](#)
36. Li J, Ke W, Zhou Q, Wu Y, Luo H, Zhou H, Yang B, Guo Y, Zheng Q, Zhang Y. Tumour necrosis factor- α promotes liver ischaemia-reperfusion injury through the PGC-1 α /Mfn2 pathway. *J Cell Mol Med* 2014;18:1863-1873.
[PUBMED](#) | [CROSSREF](#)
37. Abu-Amara M, Yang SY, Tapuria N, Fuller B, Davidson B, Seifalian A. Liver ischemia/reperfusion injury: processes in inflammatory networks--a review. *Liver Transpl* 2010;16:1016-1032.
[PUBMED](#) | [CROSSREF](#)
38. Peralta C, Jiménez-Castro MB, Gracia-Sancho J. Hepatic ischemia and reperfusion injury: effects on the liver sinusoidal milieu. *J Hepatol* 2013;59:1094-1106.
[PUBMED](#) | [CROSSREF](#)
39. Brock RW, Lawlor DK, Harris KA, Potter RF. Initiation of remote hepatic injury in the rat: interactions between Kupffer cells, tumor necrosis factor- α , and microvascular perfusion. *Hepatology* 1999;30:137-142.
[PUBMED](#) | [CROSSREF](#)
40. Suyavaran A, Ramamurthy C, Mareeswaran R, Subastri A, Lokeswara Rao P, Thirunavukkarasu C. TNF- α suppression by glutathione preconditioning attenuates hepatic ischemia reperfusion injury in young and aged rats. *Inflamm Res* 2015;64:71-81.
[PUBMED](#) | [CROSSREF](#)
41. Hernandez-Alejandro R, Zhang X, Croome KP, Zheng X, Parfitt J, Chen D, Jevnikar A, Wall W, Min WP, Quan D. Reduction of liver ischemia reperfusion injury by silencing of TNF- α gene with shRNA. *J Surg Res* 2012;176:614-620.
[PUBMED](#) | [CROSSREF](#)
42. Feng M, Wang Q, Wang H, Guan W. Tumor necrosis factor- α preconditioning attenuates liver ischemia/reperfusion injury through preserving sarco/endoplasmic reticulum calcium-ATPase function. *J Surg Res* 2013;184:1109-1113.
[PUBMED](#) | [CROSSREF](#)
43. Teoh N, Field J, Sutton J, Farrell G. Dual role of tumor necrosis factor- α in hepatic ischemia-reperfusion injury: studies in tumor necrosis factor- α gene knockout mice. *Hepatology* 2004;39:412-421.
[PUBMED](#) | [CROSSREF](#)
44. Strowig T, Henao-Mejia J, Elinav E, Flavell R. Inflammasomes in health and disease. *Nature* 2012;481:278-286.
[PUBMED](#) | [CROSSREF](#)
45. Davis BK, Wen H, Ting JP. The inflammasome NLRs in immunity, inflammation, and associated diseases. *Annu Rev Immunol* 2011;29:707-735.
[PUBMED](#) | [CROSSREF](#)
46. Huang H, Chen HW, Evankovich J, Yan W, Rosborough BR, Nace GW, Ding Q, Loughran P, Beer-Stolz D, Billiar TR, et al. Histones activate the NLRP3 inflammasome in Kupffer cells during sterile inflammatory liver injury. *J Immunol* 2013;191:2665-2679.
[PUBMED](#) | [CROSSREF](#)
47. Kamo N, Ke B, Ghaffari AA, Shen XD, Busuttill RW, Cheng G, Kupiec-Weglinski JW. ASC/caspase-1/IL-1 β signaling triggers inflammatory responses by promoting HMGB1 induction in liver ischemia/reperfusion injury. *Hepatology* 2013;58:351-362.
[PUBMED](#) | [CROSSREF](#)

48. Kim HY, Kim SJ, Lee SM. Activation of NLRP3 and AIM2 inflammasomes in Kupffer cells in hepatic ischemia/reperfusion. *FEBS J* 2015;282:259-270.
[PUBMED](#) | [CROSSREF](#)
49. Li JY, Gu X, Zhang WH, Jia S, Zhou Y. GdCl₃ abates hepatic ischemia-reperfusion injury by inhibiting apoptosis in rats. *Hepatobiliary Pancreat Dis Int* 2009;8:518-523.
[PUBMED](#)
50. Tomiyama K, Ikeda A, Ueki S, Nakao A, Stolz DB, Koike Y, Afrazi A, Gandhi C, Tokita D, Geller DA, et al. Inhibition of Kupffer cell-mediated early proinflammatory response with carbon monoxide in transplant-induced hepatic ischemia/reperfusion injury in rats. *Hepatology* 2008;48:1608-1620.
[PUBMED](#) | [CROSSREF](#)
51. Kitagawa T, Yokoyama Y, Kokuryo T, Nagino M. Protective effects of branched-chain amino acids on hepatic ischemia-reperfusion-induced liver injury in rats: a direct attenuation of Kupffer cell activation. *Am J Physiol Gastrointest Liver Physiol* 2013;304:G346-G355.
[PUBMED](#) | [CROSSREF](#)
52. Ellett JD, Atkinson C, Evans ZP, Amani Z, Balish E, Schmidt MG, van Rooijen N, Schnellmann RG, Chavin KD. Murine Kupffer cells are protective in total hepatic ischemia/reperfusion injury with bowel congestion through IL-10. *J Immunol* 2010;184:5849-5858.
[PUBMED](#) | [CROSSREF](#)
53. Sutter AG, Palanisamy AP, Ellett JD, Schmidt MG, Schnellmann RG, Chavin KD. Intereukin-10 and Kupffer cells protect steatotic mice livers from ischemia-reperfusion injury. *Eur Cytokine Netw* 2014;25:69-76.
[PUBMED](#) | [CROSSREF](#)
54. Yoshidome H, Kato A, Edwards MJ, Lentsch AB. Interleukin-10 suppresses hepatic ischemia/reperfusion injury in mice: implications of a central role for nuclear factor κB. *Hepatology* 1999;30:203-208.
[PUBMED](#) | [CROSSREF](#)
55. Si ZZ, Li JQ, Qi HZ, He ZJ, Hu W, Li YN. Recombinant adenovirus vector Ad-hIL-10 protects grafts from cold ischemia-reperfusion injury following orthotopic liver transplantation in rats. *Hepatobiliary Pancreat Dis Int* 2010;9:144-148.
[PUBMED](#)
56. Ke B, Shen XD, Tsuchihashi S, Gao F, Araujo JA, Busuttill RW, Ritter T, Kupiec-Weglinski JW. Viral interleukin-10 gene transfer prevents liver ischemia-reperfusion injury: toll-like receptor-4 and heme oxygenase-1 signaling in innate and adaptive immunity. *Hum Gene Ther* 2007;18:355-366.
[PUBMED](#) | [CROSSREF](#)
57. Gómez-Hurtado I, Zapater P, Bellot P, Pascual S, Pérez-Mateo M, Such J, Francés R. Interleukin-10-mediated heme oxygenase 1-induced underlying mechanism in inflammatory down-regulation by norfloxacin in cirrhosis. *Hepatology* 2011;53:935-944.
[PUBMED](#) | [CROSSREF](#)
58. Devey L, Ferenbach D, Mohr E, Sangster K, Bellamy CO, Hughes J, Wigmore SJ. Tissue-resident macrophages protect the liver from ischemia reperfusion injury via a heme oxygenase-1-dependent mechanism. *Mol Ther* 2009;17:65-72.
[PUBMED](#) | [CROSSREF](#)
59. Shen XD, Ke B, Uchida Y, Ji H, Gao F, Zhai Y, Busuttill RW, Kupiec-Weglinski JW. Native macrophages genetically modified to express heme oxygenase 1 protect rat liver transplants from ischemia/reperfusion injury. *Liver Transpl* 2011;17:201-210.
[PUBMED](#) | [CROSSREF](#)
60. Ke B, Shen XD, Gao F, Ji H, Qiao B, Zhai Y, Farmer DG, Busuttill RW, Kupiec-Weglinski JW. Adoptive transfer of *ex vivo* HO-1 modified bone marrow-derived macrophages prevents liver ischemia and reperfusion injury. *Mol Ther* 2010;18:1019-1025.
[PUBMED](#) | [CROSSREF](#)
61. Kensler TW, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol* 2007;47:89-116.
[PUBMED](#) | [CROSSREF](#)
62. Loboda A, Damulewicz M, Pyza E, Jozkowicz A, Dulak J. Role of Nrf2/HO-1 system in development, oxidative stress response and diseases: an evolutionarily conserved mechanism. *Cell Mol Life Sci* 2016;73:3221-3247.
[PUBMED](#) | [CROSSREF](#)
63. Biswas C, Shah N, Muthu M, La P, Fernando AP, Sengupta S, Yang G, Dennery PA. Nuclear heme oxygenase-1 (HO-1) modulates subcellular distribution and activation of Nrf2, impacting metabolic and anti-oxidant defenses. *J Biol Chem* 2014;289:26882-26894.
[PUBMED](#) | [CROSSREF](#)

64. Mylroie H, Dumont O, Bauer A, Thornton CC, Mackey J, Calay D, Hamdulay SS, Choo JR, Boyle JJ, Samarel AM, et al. PKCε-CREB-Nrf2 signalling induces HO-1 in the vascular endothelium and enhances resistance to inflammation and apoptosis. *Cardiovasc Res* 2015;106:509-519.
[PUBMED](#) | [CROSSREF](#)
65. Rao J, Qian X, Li G, Pan X, Zhang C, Zhang F, Zhai Y, Wang X, Lu L. ATF3-mediated NRF2/HO-1 signaling regulates TLR4 innate immune responses in mouse liver ischemia/reperfusion injury. *Am J Transplant* 2015;15:76-87.
[PUBMED](#) | [CROSSREF](#)
66. Cursio R, Colosetti P, Gugenheim J. Autophagy and liver ischemia-reperfusion injury. *BioMed Res Int* 2015;2015:417590.
[PUBMED](#) | [CROSSREF](#)
67. Lee SC, Kim KH, Kim OH, Lee SK, Kim SJ. Activation of autophagy by everolimus confers hepatoprotection against ischemia-reperfusion injury. *Am J Transplant* 2016;16:2042-2054.
[PUBMED](#) | [CROSSREF](#)
68. Yun N, Cho HI, Lee SM. Impaired autophagy contributes to hepatocellular damage during ischemia/reperfusion: heme oxygenase-1 as a possible regulator. *Free Radic Biol Med* 2014;68:168-177.
[PUBMED](#) | [CROSSREF](#)
69. Xu D, Chen L, Chen X, Wen Y, Yu C, Yao J, Wu H, Wang X, Xia Q, Kong X. The triterpenoid CDDO-imidazolide ameliorates mouse liver ischemia-reperfusion injury through activating the Nrf2/HO-1 pathway enhanced autophagy. *Cell Death Dis* 2017;8:e2983.
[PUBMED](#) | [CROSSREF](#)
70. Caldwell CC, Okaya T, Martignoni A, Husted T, Schuster R, Lentsch AB. Divergent functions of CD4⁺ T lymphocytes in acute liver inflammation and injury after ischemia-reperfusion. *Am J Physiol Gastrointest Liver Physiol* 2005;289:G969-G976.
[PUBMED](#) | [CROSSREF](#)
71. Uchida Y, Ke B, Freitas MC, Ji H, Zhao D, Benjamin ER, Najafian N, Yagita H, Akiba H, Busuttil RW, et al. The emerging role of T cell immunoglobulin mucin-1 in the mechanism of liver ischemia and reperfusion injury in the mouse. *Hepatology* 2010;51:1363-1372.
[PUBMED](#) | [CROSSREF](#)
72. Ji H, Shen X, Gao F, Ke B, Freitas MC, Uchida Y, Busuttil RW, Zhai Y, Kupiec-Weglinski JW. Programmed death-1/B7-H1 negative costimulation protects mouse liver against ischemia and reperfusion injury. *Hepatology* 2010;52:1380-1389.
[PUBMED](#) | [CROSSREF](#)
73. Uchida Y, Ke B, Freitas MC, Yagita H, Akiba H, Busuttil RW, Najafian N, Kupiec-Weglinski JW. T-cell immunoglobulin mucin-3 determines severity of liver ischemia/reperfusion injury in mice in a TLR4-dependent manner. *Gastroenterology* 2010;139:2195-2206.
[PUBMED](#) | [CROSSREF](#)
74. Ji H, Liu Y, Zhang Y, Shen XD, Gao F, Busuttil RW, Kuchroo VK, Kupiec-Weglinski JW. T-cell immunoglobulin and mucin domain 4 (TIM-4) signaling in innate immune-mediated liver ischemia-reperfusion injury. *Hepatology* 2014;60:2052-2064.
[PUBMED](#) | [CROSSREF](#)
75. Zhang M, Ueki S, Kimura S, Yoshida O, Castellaneta A, Ozaki KS, Demetris AJ, Ross M, Vodovotz Y, Thomson AW, et al. Roles of dendritic cells in murine hepatic warm and liver transplantation-induced cold ischemia/reperfusion injury. *Hepatology* 2013;57:1585-1596.
[PUBMED](#) | [CROSSREF](#)
76. Bamboat ZM, Ocuin LM, Balachandran VP, Obaid H, Plitas G, DeMatteo RP. Conventional DCs reduce liver ischemia/reperfusion injury in mice via IL-10 secretion. *J Clin Invest* 2010;120:559-569.
[PUBMED](#) | [CROSSREF](#)
77. Arrenberg P, Maricic I, Kumar V. Sulfatide-mediated activation of type II natural killer T cells prevents hepatic ischemic reperfusion injury in mice. *Gastroenterology* 2011;140:646-655.
[PUBMED](#) | [CROSSREF](#)
78. Beldi G, Banz Y, Kroemer A, Sun X, Wu Y, Graubardt N, Rellstab A, Nowak M, Enjyoji K, Li X, et al. Deletion of CD39 on natural killer cells attenuates hepatic ischemia/reperfusion injury in mice. *Hepatology* 2010;51:1702-1711.
[PUBMED](#) | [CROSSREF](#)
79. Kimura S, Ozaki KS, Ueki S, Zhang M, Yokota S, Stolz DB, Geller DA, Murase N. Contribution of alloantigens to hepatic ischemia/reperfusion injury: roles of natural killer cells and innate immune recognition of nonself. *Liver Transpl* 2016;22:80-90.
[PUBMED](#) | [CROSSREF](#)

80. Qiao YL, Qian JM, Wang FR, Ma ZY, Wang QW. Butyrate protects liver against ischemia reperfusion injury by inhibiting nuclear factor kappa B activation in Kupffer cells. *J Surg Res* 2014;187:653-659.
[PUBMED](#) | [CROSSREF](#)
81. Sakai N, Van Sweringen HL, Schuster R, Blanchard J, Burns JM, Tevar AD, Edwards MJ, Lentsch AB. Receptor activator of nuclear factor- κ B ligand (RANKL) protects against hepatic ischemia/reperfusion injury in mice. *Hepatology* 2012;55:888-897.
[PUBMED](#) | [CROSSREF](#)
82. Li JD, Peng Y, Peng XY, Li QL, Li Q. Suppression of nuclear factor-kappaB activity in Kupffer cells protects rat liver graft from ischemia-reperfusion injury. *Transplant Proc* 2010;42:1582-1586.
[PUBMED](#) | [CROSSREF](#)
83. Giakoustidis DE, Giakoustidis AE, Iliadis S, Koliakou K, Antoniadis N, Kontos N, Papanikolaou V, Papageorgiou G, Kaldrimidou E, Takoudas D. Attenuation of liver ischemia/reperfusion induced apoptosis by epigallocatechin-3-gallate via down-regulation of NF-kappaB and c-Jun expression. *J Surg Res* 2010;159:720-728.
[PUBMED](#) | [CROSSREF](#)
84. Kuboki S, Sakai N, Clarke C, Schuster R, Blanchard J, Edwards MJ, Lentsch AB. The peptidyl-prolyl isomerase, Pin1, facilitates NF- κ B binding in hepatocytes and protects against hepatic ischemia/reperfusion injury. *J Hepatol* 2009;51:296-306.
[PUBMED](#) | [CROSSREF](#)
85. Li X, Wu Y, Zhang W, Gong J, Cheng Y. Pre-conditioning with tanshinone IIA attenuates the ischemia/reperfusion injury caused by liver grafts via regulation of HMGB1 in rat Kupffer cells. *Biomed Pharmacother* 2017;89:1392-1400.
[PUBMED](#) | [CROSSREF](#)
86. Ogiku M, Kono H, Hara M, Tsuchiya M, Fujii H. Glycyrrhizin prevents liver injury by inhibition of high-mobility group box 1 production by Kupffer cells after ischemia-reperfusion in rats. *J Pharmacol Exp Ther* 2011;339:93-98.
[PUBMED](#) | [CROSSREF](#)
87. Venkatachalam AB, Livingstone SM, Hu Q, Ray A, Wood C, Cimen S, Alwayn IP. Delivery of soluble heme oxygenase 1 cell-penetrating peptide into liver cells in *in vitro* and *ex vivo* models of cold ischemia. *Eur Surg Res* 2017;58:51-68.
[PUBMED](#) | [CROSSREF](#)
88. Xie Y, Zhao D, Dong P, Lai L. Macrophage-targeting Fasudil treatment protects liver from the ischemia/reperfusion injury by promoting M2 macrophage polarization. *Biosci Rep*. doi: 10.1042/BSR20171734.
[PUBMED](#) | [CROSSREF](#)
89. Linares I, Farrokhi K, Echeverri J, Kathis JM, Kollmann D, Hamar M, Urbanellis P, Ganesh S, Adeyi OA, Yip P, et al. PPAR-gamma activation is associated with reduced liver ischemia-reperfusion injury and altered tissue-resident macrophages polarization in a mouse model. *PLoS One* 2018;13:e0195212.
[PUBMED](#) | [CROSSREF](#)
90. Guan LY, Fu PY, Li PD, Li ZN, Liu HY, Xin MG, Li W. Mechanisms of hepatic ischemia-reperfusion injury and protective effects of nitric oxide. *World J Gastrointest Surg* 2014;6:122-128.
[PUBMED](#) | [CROSSREF](#)
91. Yu HC, Bai L, Yue SQ, Wang DS, Wang L, Han H, Dou KF. Notch signal protects non-parenchymal cells from ischemia/reperfusion injury *in vitro* by repressing ROS. *Ann Hepatol* 2013;12:815-821.
[PUBMED](#)
92. Li Y, Ma D, Wang Z, Yang J. MicroRNA-155 deficiency in Kupffer cells ameliorates liver ischemia-reperfusion injury in mice. *Transplantation* 2017;101:1600-1608.
[PUBMED](#) | [CROSSREF](#)