

RESEARCH ARTICLE

# Anticancer potential, molecular mechanisms and toxicity of *Euterpe oleracea* extract (açai): A systematic review

Jéssica Alessandra-Perini<sup>1,2</sup>\*, Karina Cristina Rodrigues-Baptista<sup>2,3</sup>, Daniel Escorsim Machado<sup>1,2,4</sup>, Luiz Eurico Nasciutti<sup>1</sup>, Jamila Alessandra Perini<sup>2,3,5</sup>

**1** Morphological Science Program—PCM, Biomedical Sciences Institute, Federal University of Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil, **2** Research Laboratory of Pharmaceutical Sciences—LAPESF, West Zone State University, Rio de Janeiro, Rio de Janeiro, Brazil, **3** Program of Post-graduation in Public Health and Environment—ENSP, National School of Public Health, Oswald Cruz Foundation, Rio de Janeiro, Rio de Janeiro, Brazil, **4** University Center IBMR, Laureate Universities, Rio de Janeiro, Rio de Janeiro, Brazil, **5** Research Division, National Institute of Traumatology and Orthopedics—INTO, Rio de Janeiro, Rio de Janeiro, Brazil

\* These authors contributed equally to this work.

\* [jessicaperini@yahoo.com.br](mailto:jessicaperini@yahoo.com.br)



**OPEN ACCESS**

**Citation:** Alessandra-Perini J, Rodrigues-Baptista KC, Machado DE, Nasciutti LE, Perini JA (2018) Anticancer potential, molecular mechanisms and toxicity of *Euterpe oleracea* extract (açai): A systematic review. PLoS ONE 13(7): e0200101. <https://doi.org/10.1371/journal.pone.0200101>

**Editor:** Siyaram Pandey, University of Windsor, CANADA

**Received:** March 3, 2018

**Accepted:** June 19, 2018

**Published:** July 2, 2018

**Copyright:** © 2018 Alessandra-Perini et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

**Funding:** This study was supported by the Brazilian agency Fundação Carlos Chagas Filho de Amparo à Pesquisa do Estado do Rio de Janeiro – FAPERJ (JAP and DEM), Fundação Ary Frauzino – Oncobiologia (DEM) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – CAPES (JA-P and KCRB).

## Abstract

Cancer is an increasingly frequent malignancy worldwide, and despite the advances in drug development, it is still necessary to develop new plant-derived medicines. *Euterpe oleracea* (açai) is abundant in South and Central America and has health benefits due to its high levels of phytochemicals, including lignans and polyphenols. The aim of this review was to systematically describe the safety and antitumor effects of açai in preclinical models using rodents to provide a more comprehensive assessment of açai for both therapeutic uses and the development of future clinical studies in cancer. Eligible studies were identified using four international databases (PubMed, Medline, Lilacs and SciELO) from their inception date through December 2017. The included studies were analyzed with methodological rigor (QATRS) to enable better quality control for these experimental studies. Sixty publications were identified in the databases, but only 9 articles were eligible: 6 evaluated the pharmacological effects of açai in animal models of cancer (1 model each of esophageal cancer, urothelial cancer, melanoma and Walker-256 tumor and 2 models of colon cancer), and 3 were toxicological assays using preclinical models with rodents. Overall, 747 animals were analyzed. On a QATRS score scale of 0–20, the quality of the studies ranged from 16 to 20 points. Pulp was the main fraction of açai administered, and an oral administration route was most common. The açai dosage administered by gavage ranged from 30 mg/kg to 40,000 mg/kg, and açai fed in the diet accounted for 2.5% to 5% of the diet. The anticarcinogenic and chemopreventive activities of açai were observed in all experimental models of cancer and reduced the incidence, tumor cell proliferation, multiplicity and size of the tumors due to the antiinflammatory, antiproliferative and proapoptotic properties of açai. No genotoxic effects were observed after açai administration. The results of this review suggest that açai is safe and can be used as a chemoprotective agent against cancer development. Açai therapy may be a novel strategy for treating cancer.

**Competing interests:** The authors have declared that no competing interests exist.

## Introduction

The use of natural products as medicines accounts for approximately 30% of the currently available drugs [1], and in some therapeutic areas, the amount of plant-derived medicines reaches 60% [2,3]. Brazil has the greatest amount of biodiversity in the world and plays an important role in the area of natural bioactive compounds by contributing natural products to design new clinical medicines [1,4]. Thus, there has been growing research aimed at establishing the therapeutic potential of natural products against several diseases.

*Euterpe oleracea* Mart. is a member of the family Arecaceae and is a typical palm of the rain-forest in the Amazon region, in the states of the northern region of Brazil, including Guianas, Colombia, Ecuador, and Venezuela [5]. The fruit, popularly known as “açai”, weighs approximately 2 g, and the color of the mature fruit is dark purple [6]. Açai is a traditional food in many regions of Brazil [7,8], and its consumption has increased significantly over the last several years, not only in Brazil but also in Europe and the USA, where the fruit gained popularity after being promoted as a “super fruit” [9]. Currently, due to the health benefits and therapeutic potential of açai, locally grown açai are increasingly exported around the world as energy drinks [6,10], “functional foods” [7,8], cosmetics and pharmaceutical products [9]. Açai pulp is composed of approximately 48% lipids, 13% protein, 8% amino acids, 25% total sugars and minor compounds such as fiber and vitamins (A, B1, B2, B3, C and E) [8,11,12]. Moreover, it is rich in several phytochemicals, including lignans, phenolic compounds (anthocyanins, proanthocyanidins and other flavonoids) and resveratrol, in low concentrations [8,11,12]. The seeds of açai possess the highest concentration of polyphenols (28.3%), followed by the whole fruit (25.5%) and the bark (15.7%) [13].

The pharmacological effects of açai are associated with its chemical composition, particularly the presence of bioactive substances, such as phenolics, flavonoids and anthocyanins [14–17]. To date, açai has been shown to have pharmacological properties including antiinflammatory, antioxidant, cardioprotective and anticancer activities [1,7–9,18,19]. Furthermore, açai was not shown to be genotoxic *in vitro* and *in vivo* studies conducted, in cultured human lymphocytes and hepatoma cell lines [20], in rodents [21] and in humans [22].

The aim of this review was to systematically describe the safety and antitumor effects of açai in preclinical models using rodents, to provide a comprehensive assessment of açai for therapeutic use. Preclinical studies using rodents were evaluated to investigate whether the current knowledge supports cancer clinical trials with açai.

## Methods

### Search strategy

A careful literature search was performed to identify publications that studied the use of *E. oleracea* extract in experimental animal models of cancer and/or evaluated the safety/toxicity of açai in animal models. Studies were identified by searching the electronic databases: PubMed, Medline-Bireme, Lilacs and SciELO from their inception date through December 2017 (S1 Table).

The search terms were as follows: (“*Euterpe oleracea*” AND cancer treatment) OR (“*Euterpe oleracea*” AND cancer animal model) OR (Açai AND cancer treatment) OR (Açai AND cancer animal model) AND (“*Euterpe oleracea*” AND toxicity) OR (Açai AND toxicity). The search was performed without restrictions on the language or year of publication. Two reviewers (KCRB and JA-P) selected the qualified studies independently by browsing the titles, abstracts or full texts based on the eligibility criteria. The duplicates were removed. The eligible articles were separated for analysis of the study methodology and results (S1 Table). Any disagreements were resolved by discussion with two additional reviewers (DEM and JAP).

## Inclusion and exclusion criteria

Articles were included if the following criteria were met: (1) evaluated the pharmacological effect of açai in animal models of cancer and/or (2) performed toxicological analyzes after açai administration in experimental animal models. Articles were excluded if the following criteria were met: (1) were reviews of literature; (2) did not analyze the use of açai *in vivo*; (3) did not use the order *Rodentia*; (4) did not evaluate the toxicological effects of açai administration *in vivo*; and (5) used only *in vitro* experimental models.

## Data extraction

Three investigators (KCRB, JA-P and JAP) independently conducted the extraction of details from each study including the following: (1) basic information, including the publication year, the first author's name, the type of animals, the sex, the *in vivo* model and the experimental interventions; (2) basic information about the açai treatment, including the fraction and origin of *E. oleracea*, dose, administration route, posology, diluents and treatment groups; and (3) outcome measures used to evaluate *E. oleracea* extract, therapeutic indications (pharmacodynamic), açai signaling pathways and safety evaluations. When a single publication included studies with animals, posology or types of interventions that were different, these data were extracted and considered as independent experiments. Any disagreements regarding the extracted data were resolved by discussion with an additional reviewer (DEM).

## Quality assessment

For assessment of quality, two independent reviewers (KCRB and JA-P) used a quality rating scale as an animal/tissue research scale (QATRS). The QATRS is a 20-point scaled evaluation chart that was designed based on randomization, blinding, the similarity of the animal/tissue model to human applications, standardization and the reliability of the measurement techniques, management of study withdrawals, and appropriateness of the statistical methods [23]. Any disagreements were resolved by discussion with two additional reviewers (DEM and JAP).

## Results

### Study selection

A flowchart of the articles that were included in the review is illustrated in Fig 1. A total of 60 publications were identified in the databases; however, 31 were duplicate articles. Among the 29 articles selected, 20 were excluded based on the titles and abstracts because they did not meet the inclusion criteria: 2 were literature reviews [7,24]; 6 did not study açai in an animal model of cancer and/or did not perform a toxicological analysis [25–30]; 10 were *in vitro* studies [13,20,31–38]; and 2 did not use the order *Rodentia* [21,39]. After reading the full texts 9 articles were included for their critical evaluations of the safety and effectiveness of açai in animal experimental models [40–48].

### Characteristics of the experimental models

The articles included were analyzed with a critical appraisal tool (QATRS), which allowed for improved quality control of the experimental studies in animal performed independently (see methods). QATRS scores ranged from 0 to 20, and the quality of the studies ranged from 16 to 20 points (Table 1). Among the 9 studies that were included, 6 evaluated the pharmacological effects of açai in experimental models of cancer, including esophageal [40], urothelial [41], and colon cancer [42,43], and melanoma [44] and Walker-256 tumors [45], and 3 performed

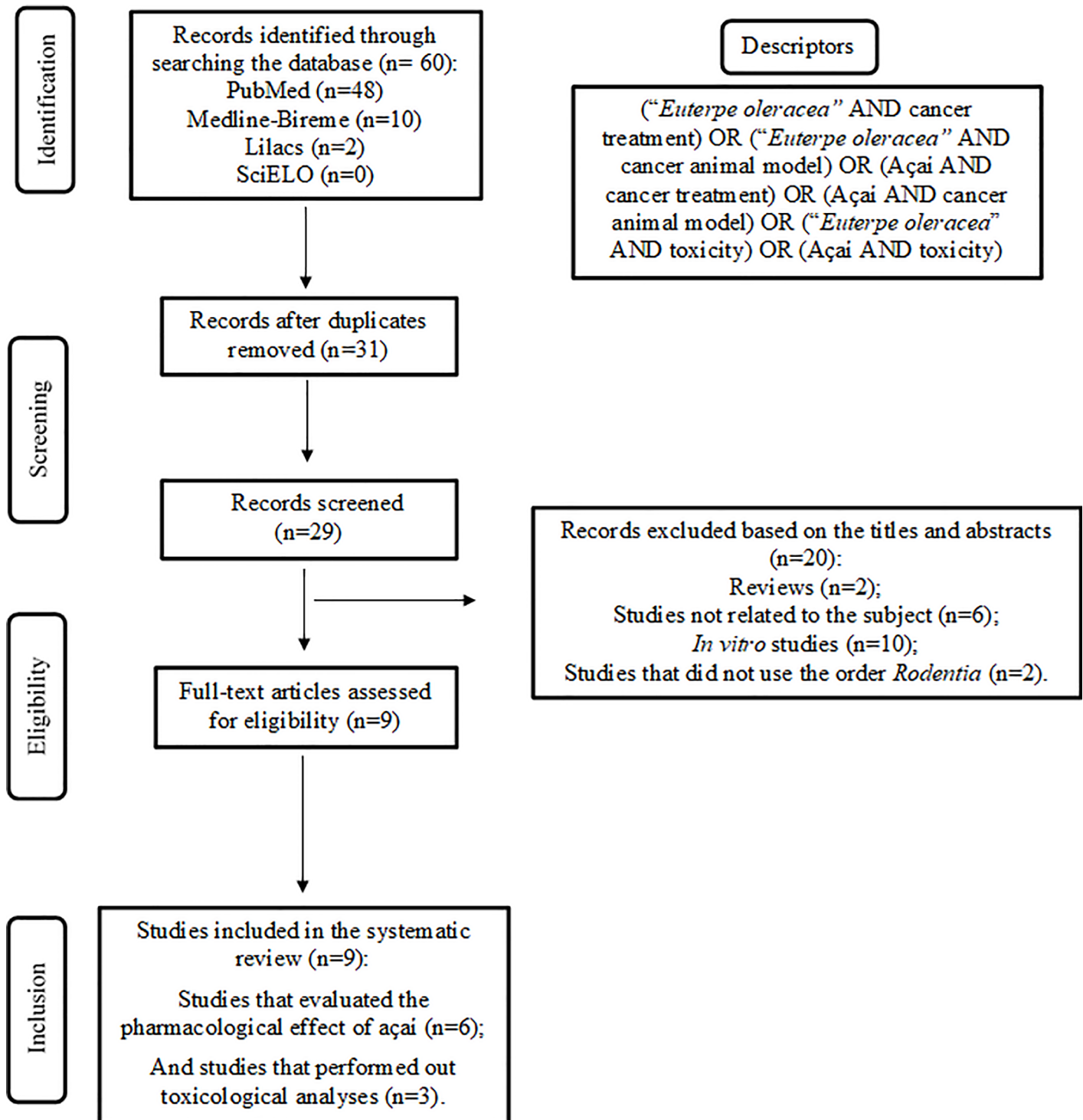


Fig 1. Flowchart of the study selection and inclusion in the review.

<https://doi.org/10.1371/journal.pone.0200101.g001>

toxicological analyses of açai in experimental models [46–48]. For the interventions used in the experimental models, 4 studies used chemically induced cancer models [40,41,43], 2 used inoculation of tumor cells [44,45], and 3 used models with DNA damage induced by a

**Table 1. Basic information on the *in vivo* experimental models used to test the effects of *E. oleracea*.**

Model	Animals	Interventions	References <sup>a</sup>	QATRS
Cancer	Male F344 rats	Esophageal carcinogenesis induced by NMBA	Stoner <i>et al.</i> 2010	16
	Male Swiss mice	Urothelial carcinogenesis induced by BBN and MNU	Fragoso <i>et al.</i> 2012	20
	Male Wistar rats	Colon carcinogenesis induced by DMH	Fragoso <i>et al.</i> 2013	20
	Male Wistar rats	Anorexia-cachexia syndrome induced by Walker-256 tumor	Nascimento <i>et al.</i> 2016	16
	Male ICR mice	Colon carcinogenesis induced by AOM and DSS	Choi <i>et al.</i> 2017	16
	Female C57BL/6 mice	Melanoma induced by transplantation of B16F10 cells	Monge-Fuentes <i>et al.</i> 2017	18
Toxicity	Male Swiss mice	DNA damage induced by doxorubicin	Ribeiro <i>et al.</i> 2010	18
	Male Wistar rats	DNA damage induced by doxorubicin	Marques <i>et al.</i> 2016	18
	BALB/c mice	DNA damage induced by cyclophosphamide	Schauss <i>et al.</i> 2010 <sup>a</sup>	18
	Wistar rats	Acute and subchronic oral toxicity study	Schauss <i>et al.</i> 2010 <sup>a</sup>	18

AOM = azoxymethane; BBN = N-butyl-N-(4-hydroxybutyl)-nitrosamine; B16F10 = melanoma cell lines; DSS = dextran sulfate sodium; DMH = 1,2-dimethylhydrazine; ICR = International Cancer Research; MNU = N-methyl-N-nitrosourea; NMBA = N-nitrosomethylbenzylamine.

<sup>a</sup>A reference can have more than one model of disease.

<https://doi.org/10.1371/journal.pone.0200101.t001>

chemotherapeutic agent [46–48]. The studies involved 2 species and 6 varieties of rodents: C57BL/6 mice [44], F344 rats [40], Wistar rats [42,45,47,48], Swiss mice [41,46], ICR mice [43] and Balb/c mice [48] (Table 1).

### Açai information

Table 2 shows the basic information about the açai extract used in the experimental models. The most commonly used açai fraction was the pulp [40–43,46], followed by the juice [48], oil [44,47] and seeds [45]. Seven studies mentioned the açai origin, and all of the açai extracts were from Brazil [40–43,45–47]. The main administration route of açai was oral; 4 studies administered açai by gavage [45–48], and 4 studies administered açai as part of the diet [40–43]. The dosage ranged from 30 mg/kg to 40,000 mg/kg in studies that administered açai by gavage and was administered as a single dose or as 1 daily dose for 90 consecutive days; in the studies that administered açai as part of the diet 2.5% to 5% açai supplementation was provided in the diet for 10 to 35 weeks (Table 2). In addition, Schauss and colleagues used oral and intraperitoneal administration of açai at a dose of 0.1mg/0.15mL (daily dose during 7 consecutive days) to assess the possible genotoxic effects of açai using BALB/c mice [48], and Monge-Fuentes and colleagues used 50 mg/mL of açai administered intratumorally in an experimental model of melanoma [44]. The results regarding the therapeutic indications, effects and safety of açai in experimental models are summarized in Table 3.

### Safety of açai

The absence of toxicity of açai was reported in 6 studies after testing açai in experimental models [41,42,44,46–48], and no significant differences in animal body weight or food consumption were reported in 4 studies [40–42,48]. DNA damage induced by antitumor medication was evaluated in 3 studies, and no genotoxic effects were observed after açai administration by gavage [46–48] (Table 3).

Using a micronucleus test and a comet assay, Ribeiro and colleagues reported no differences between the control and açai groups in bone marrow and peripheral blood cells polychromatic erythrocytes, and in liver and kidney cells, thus demonstrating the absence of genotoxic effects of açai. In addition, açai reduced DNA damage induced by doxorubicin

**Table 2. Basic information regarding the *E. oleracea* extract used in the *in vivo* experimental models.**

Fraction	Origin of açai	Dosing	Diluent and placebo	Administration	Posology	Reference
Juice <sup>b</sup>	Not mentioned	0.1 mg/0.15mL	Saline	Oral (gavage) and IP	1 daily dose over 7 days	Schauss <i>et al.</i> 2010 <sup>a</sup>
	Not mentioned	5,000 and 20,000 mg/kg	Not mentioned	Oral (gavage)	Single dose	Schauss <i>et al.</i> 2010 <sup>a</sup>
	Not mentioned	10,000; 20,000 and 40,000 mg/kg	Saline	Oral (gavage)	1 daily dose over 90 days	Schauss <i>et al.</i> 2010 <sup>a</sup>
Oil	Brazil (Amapá)	30, 100 and 300 mg/kg	Tween	Oral (gavage)	1 daily dose over 14 days	Marques <i>et al.</i> 2016
	Not mentioned	50 mg/mL	PBS	Intratumoral	Five applications within 15 days (1, 4, 7, 10 and 13 days)	Monge-Fuentes <i>et al.</i> 2017
Pulp	Brazil	5%	AIN diet	Oral (diet)	35 weeks	Stoner <i>et al.</i> 2010
	Brazil (SP)	3,330; 10,000 and 16,670 mg/kg	Saline	Oral (gavage)	Single dose	Ribeiro <i>et al.</i> 2010 <sup>a</sup>
	Brazil (SP)	3,330; 10,000 and 16,670 mg/kg	Distilled water	Oral (gavage)	1 daily dose over 14 days	Ribeiro <i>et al.</i> 2010 <sup>a</sup>
	Brazil (Pará)	2.5% and 5%	Standard diet	Oral (diet)	10 weeks	Fragoso <i>et al.</i> 2012 and 2013 <sup>a</sup>
	Brazil (Pará)	5%	Standard diet	Oral (diet)	20 weeks	Fragoso <i>et al.</i> 2013 <sup>a</sup>
	Brazil (Pará)	2.5% and 5%	Diet formulated <sup>c</sup>	Oral (diet)	14 weeks	Choi <i>et al.</i> 2017
Seed	Brazil	100 and 200 mg/mL	Ethanol-water	Oral (gavage)	1 daily dose over 14 days	Nascimento <i>et al.</i> 2016

AIN = American Institute of Nutrition; IP = intraperitoneal; SP = São Paulo; PBS = Phosphate buffered saline.

<sup>a</sup> A reference can have different methods of administration of açai.

<sup>b</sup>Juice of MonaVie Active® = In addition to açai, contains lesser amounts of 19 fruits and berries.

<sup>c</sup>A cereal-based commercial diet for mice formulated by the Orient Bio Group (Seongnam, Korea).

<https://doi.org/10.1371/journal.pone.0200101.t002>

(DXR), suggesting a protective role in human health [46]. In a study done by Schauss and colleagues, açai did not cause mutagenic effects, as demonstrated by a bacterial reverse mutation assay, a chromosomal aberration assay, a mammalian cell mutation assay and an *in vivo* micronucleus study [48]. In the same way, Marques and colleagues evaluated the genotoxic potential of açai in rat cells. The authors used a comet assay and a micronucleus test and showed that on both cytogenetic tests, no significant genotoxic effects were observed at the three tested dosages of açai [47].

### Antitumoral effects of açai

The anticarcinogenic and chemopreventive activities of açai, as evidenced by reductions in the incidence of tumors, tumor cell proliferation, and multiplicity and size of tumors, were observed in all the experimental models of cancer [40–45] (Table 3).

Stoner and colleagues reported that açai was effective at inhibiting the progression of esophageal tumorigenesis, reducing the levels of the serum cytokines (IL-5 and IL-8), and increasing serum antioxidant capacity and interferon-gamma (IFN $\gamma$ ) levels [40]. By contrast, the esophageal tumor size and serum levels of IL-1 $\beta$ , IL-4, IL-13 and tumor necrosis factor-alpha (TNF- $\alpha$ ) were not significantly affected by adding açai to the diet for 35 weeks [40].

Fragoso and colleagues reported that açai was effective at inhibiting urinary bladder carcinogenesis, reducing DNA damage, and reducing the expression of p63 and proliferating cell nuclear antigen (PCNA) [41]. However, altered cytoplasmatic and nuclear  $\beta$ -catenin were not significantly affected by adding açai to the diet for 10 weeks [41].

**Table 3. Results of cancer treatments and safety evaluations of *E. oleracea* extract in animal models.**

References	Therapeutic indication	Action of açai	Unchanged parameters	Effects of açai
Stoner <i>et al.</i> 2010	Chemopreventive	↓ incidence, multiplicity and inflammatory cytokines; ↑ serum antioxidant capacity and IFN $\gamma$	Body weight, food consumption, pro and antiinflammatory	Inhibits esophageal tumorigenesis progression
Fragoso <i>et al.</i> 2012	Chemopreventive (anticarcinogenic)	↓ incidence, multiplicity, tumor cell proliferation, urothelial preneoplastic lesions, p63 and PCNA expression and DNA damage	Body weight, food consumption, bladder and kidney weight, kidney biochemical markers, cytoplasmatic and nuclear $\beta$ -catenin expression	Inhibits urothelial bladder carcinogenesis
Fragoso <i>et al.</i> 2013	Chemopreventive	↓ invasiveness, multiplicity and growth of tumor, cell proliferation and cleaved caspase-3, number of aberrant crypts	Body weight, food consumption, $\beta$ -catenin expression and toxicity	Inhibits colon carcinogenesis
Nascimento <i>et al.</i> 2016	Anticarcinogenic	↓ tumor, muscle total protein; ↑ oxidative stress in cerebral cortex	Liver protein, oxidative stress in muscle and liver	Reduces Walker-256 tumor
Choi <i>et al.</i> 2017	Anticarcinogenic	↓ incidence, multiplicity and tumor, cell proliferation, proinflammatory cytokines and COX-2; ↑ cleaved-caspase-3 expression.	Not mentioned	Inhibits colon carcinogenesis
Monge-Fuentes <i>et al.</i> 2017	Anticarcinogenic (Photodynamic)	↓ tumor, liver and spleen weight; ↑ necrosis; Differences in body weight	Toxicity of the kidneys and lungs	Reduces melanoma carcinogenesis (photosensitizer)
Ribeiro <i>et al.</i> 2010	Protective effects	↓ MNPCE and DXR-induced genotoxicity in bone marrow or liver and kidney cells	PCE, DNA damage and genotoxic effects	Reduction in DNA damage induced by DXR
Schauss <i>et al.</i> 2010	Not mentioned	Not mentioned	Body weight, food consumption, mortality, organ weights, ophthalmology, urinalysis, hematological and biochemical parameters, and genotoxicity	Negative mutagenic effects
Marques <i>et al.</i> 2016	Not mentioned	↑ cell viability	DNA damage, clastogenic and aneugenic effect	Negative genotoxicity effects

COX-2 = cyclooxygenase 2; DXR = antitumoral agent doxorubicin; IFN $\gamma$  = interferon gamma; MNPCE = number of micronucleated peripheral blood polychromatic erythrocytes cells; PCE = peripheral blood polychromatic erythrocytes cells; PCNA = proliferating cell nuclear antigen.

<https://doi.org/10.1371/journal.pone.0200101.t003>

Two studies reported that açai was effective at inhibiting colon carcinogenesis induced by 1,2-dimethylhydrazine (DMH) in Wistar rats [42] and azoxymethane (AOM) with dextran sulfate sodium (DSS) in ICR mice [43]. Nevertheless, the opposite results were observed with regard to cleaved caspase-3 expression after supplementation with 2.5% and 5% of açai in the diet for 10 [42], 14 [43] or 20 weeks [42]. Despite the discrepancies between these studies, the quality evaluation of the results of the articles showed good quality QATRS (16/20 and 20/20, respectively) [42,43]. Moreover, Choi and colleagues reported that açai treatment down-regulated myeloperoxidase (MPO) and proinflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$  and IL-6), inhibited cyclooxygenase 2 (COX-2), PCNA and Bcl-2, and increased Bad and cleaved caspase-3 expression in an experimental model of cancer colon [43].

Monge-Fuentes and colleagues reported that açai was an effective photosensitizer because it reduced melanoma carcinogenesis by increasing the necrotic tissue per tumor area after 5 applications of intratumoral açai during a period of 15 days [44].

Nascimento and colleagues reported an anticarcinogenic effect (tumor diameter and weight) of açai in anorexia-cachexia syndrome induced by Walker-256 tumors due to the antioxidant activity of açai after 1 daily dose of açai over 14 consecutive days [45].

Finally, based on the results of this review study, we created a schematic representation of the effects of açai in tumor cells (Fig 2). Açai showed antitumoral functions due to its anti-inflammatory, antiproliferative and proapoptotic properties.

## Discussion

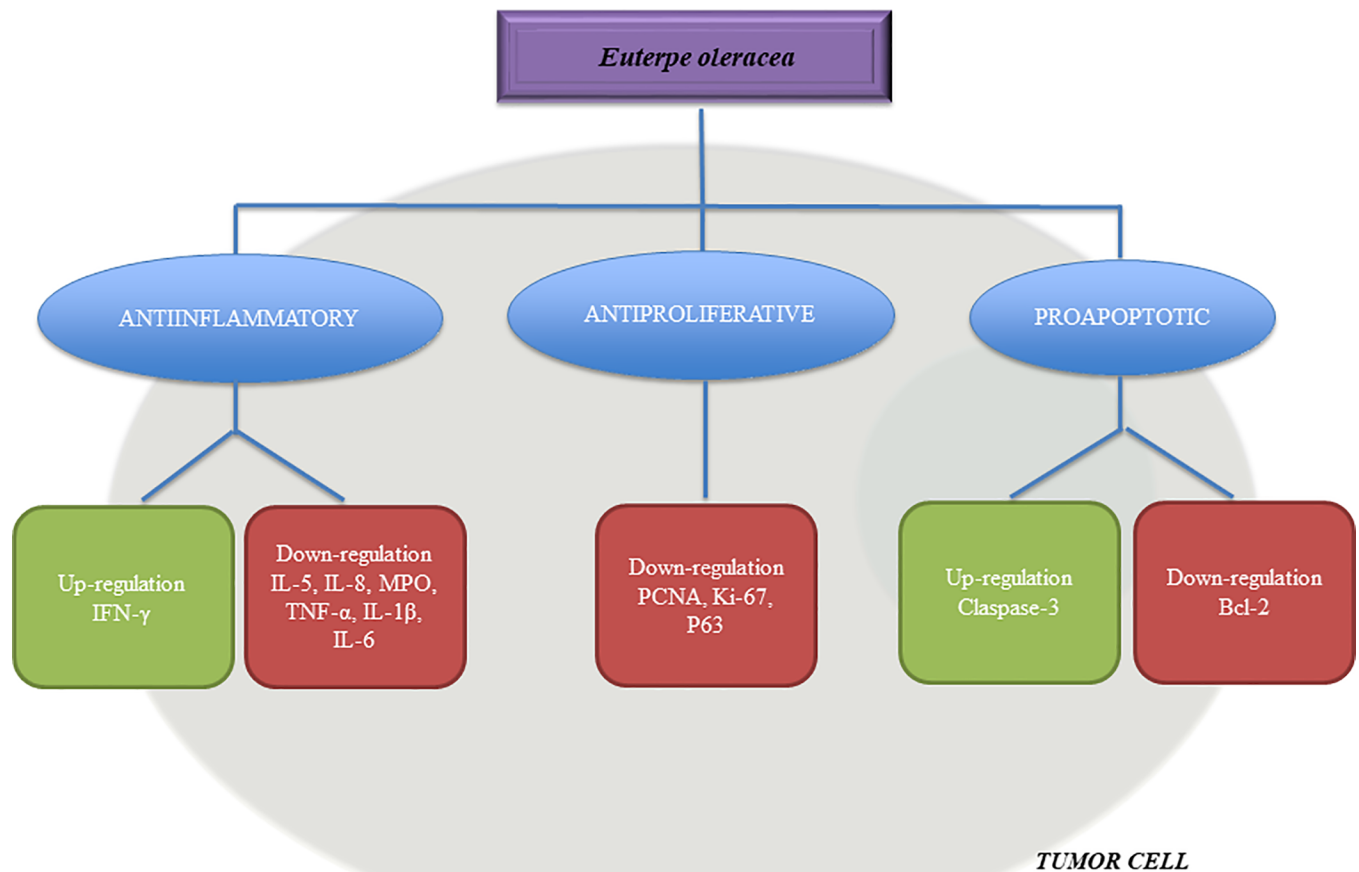
Research on the pharmacological effects of natural products for the treatment of several diseases has significantly increased in the last decades. In this sense, açai has been marketed as a dietary food supplement because of its health benefits due to its high levels of phytochemicals, including lignans and polyphenols. Studies have demonstrated that açai has biological effects, such as antioxidant, antiinflammatory, antiproliferative, antinociceptive and antitumorigenic activities [13,16,17,19,49–55].

To the best of our knowledge, 15 clinical trials with açai was carried [17–19,22,56–66], however none of these studies evaluated the effect of açai in the cancer treatment. The aim of this review was critically to evaluate the existence of scientific data about the safety and antitumor effects of açai in preclinical models using rodents, to support cancer clinical trials. The human health benefits of açai was improvements in antioxidant benefit [17–19,56,61,62,64,65]; cardiovascular health [17,22,56] with beneficial action in hemodynamic [58,60] and metabolic parameters [18,22]; modulation of inflammation [58] and reduction global pain [62]; reduces muscle stress [18,57] and improves effort tolerance in elite athletes [57]; besides to be safe and effective as contrast agent for magnetic resonance imaging [59,63,66]. Due to the important nutritional properties for human benefits and therapeutic potential, the açai became relevant functional foods.

As far as we know, the present work is the first review to focus on the antitumorigenic and toxicological effects of açai in preclinical trials using rodents. Overall, nine studies were included in this review [40–48]. Although we conducted a thorough literature search, using four international databases, one limitation is that our conclusions may be narrow due to the lack of availability of published articles and because all of the included studies were published in English. In spite of the small number of studies found, we assessed them with a range of methodological rigor in accordance with the QATRS, which encompasses various aspects that enable better quality control for these experimental studies [23]. A strong point of our review is that all the included studies had good quality as assessed by the QATRS score (all had a score greater than or equal to 16/20). A total of 747 animals of the order *Rodentia* were analyzed. The results indicated that açai has a chemopreventive effect (anticancer) by inhibiting tumor growth and leads to a reduction in tumor size, suggesting antiproliferative, pro-apoptotic and anti-inflammatory activity [40–45]. In addition, the toxicological studies showed that açai did not cause DNA damage or genotoxic or mutagenic effects in the evaluated animals, suggesting that it is safe for clinical testing [46–48].

Most of the studies found that açai significantly decreased tumor incidence or tumorigenic inhibition and prevented DNA damage without causing genotoxic effects when it was administered orally (in the diet or by gavage). These results suggest that the oral route is a good choice for evaluation of the effects of açai in humans clinical studies since this is an easy and safe route of administration. It should be noted that the significant results found with the oral administration of açai have also been described in other diseases, such as obesity and hepatic steatosis [67], endometriosis [55], renovascular hypertension [68] and neuropathic pain [53]. The articles included in this review described different doses of açai that were administered orally by gavage (range 30 mg/kg to 40,000 mg/kg) [45–48]. However, Marques and colleagues observed that at an açai dose of 300 mg/kg, a few animals showed signs of toxicity (diarrhea and bristling of the hair), which is why they did not test higher doses [47]. Recently, our group reported that a dose of açai of 200 mg/kg administered by gavage for 30 consecutive days had efficacy in suppressing endometriotic lesions in a Sprague-Dawley rat model without any signs of toxicity [55]. Although considered a benign disease, endometriosis frequently presents with characteristics of malignancy [69]. Therefore, we suggest that an açai dose of 200 mg/kg is safe for preclinical testing and is a promising novel pharmacological treatment for cancer due to its anticarcinogenic and chemopreventive effects.





**Fig 2. Schematic representation of the effects of açai on tumor cells.** Açai was shown to have antitumoral functions due its antiinflammatory, antiproliferative and proapoptotic properties.

<https://doi.org/10.1371/journal.pone.0200101.g002>

With regard to the ability of açai to inhibit carcinogenesis, and the incidence and multiplicity of tumors in experimental models of cancer using rodents, *in vitro* studies also showed that açai decreased cell viability, suppressed proliferation and induced apoptosis, suggesting the anticancer and antioxidant activity of açai against C-6 rat brain glioma cells [49], MCF-7 breast cancer cells [13,38] and colon cancer cells [34]. These results suggest that açai contains phytochemicals that can be used as natural chemopreventive agents [13,40,42].

A large number of studies have shown the importance of chronic exposure to proinflammatory cytokines in tumorigenesis [70–72]. The results of this review show that açai acts in the inflammatory processes involved in induced-cancer in animals by decreasing the levels of IL-1β, IL-5, IL-6, IL-8, COX-2, TNF-α and MPO and increasing the levels of IFN-γ [40,43]. An *in vitro* study of polymorphonuclear cells showed a reduction in the IL-8 levels that was associated with the decreasing inflammatory conditions after açai treatment [64]. Xie and colleagues evaluated flavonoids isolated from açai pulp and observed a reduction in serum levels, gene expression and protein levels of both the cytokines TNF-α and IL-6 in the resident macrophages cells [73]. Açai was also able to prevent increases in the levels of IL-1β and TNF-α in the brain tissues of a CCl4 experimental model [27]. In addition, açai reduced the COX-2 expression and PGE<sub>2</sub> levels in an experimental model of endometriosis [55] and reduced the MPO levels in a rat renal ischemia/reperfusion model [74].

As a result of this review, it was possible to identify the antiproliferative pathways by which açai acts by reducing PCNA, Ki-67 and p63 [41–43]. These proteins are involved in tumor development, survival and metastasis of different tumors [75–77]. In addition, the anti-apoptotic proteins Bcl-2 was also reduced after açai treatment in animals with induced-cancer [43], in agreement with a study of human colon cancer cells in which the proapoptotic activities of polyphenolics from açai were described [34]. Polyphenolics may regulate distinct steps of the apoptotic process and/or the expression of regulatory proteins, such as the downregulation of Bcl-2 and cleavage of caspase-3 [78,79]. Açai polyphenolics were previously described to have proapoptotic and antiproliferative activities in leukemia cancer cells through caspase-3 activation [80]. Surprisingly, as a result of this review, it was possible to identify the discrepancies in the levels of cleaved caspase-3 in colon carcinogenesis induced after açai treatment [41,43]. Choi and colleagues observed that açai increased the cleaved caspase-3 levels in the supernatants of colon strips [43], but Fragoso and colleagues described the opposite results using immunohistochemical techniques in colon tumor tissues [41].

Another specie from Brazilian *Euterpe*, o *Euterpe edulis*, has been studied because has important nutritional properties for human health. *E. edulis* Mart., commonly known as juçara or jussara and açai-do-sol, is a native tree of the Atlantic Forest and has similar nutritional properties of açai [81], however açai has twice of the polyphenols concentration [82]. Recently, a review described 25 articles about the phytochemical characterization and biological activities of juçara [81]. Nevertheless, none of these studies evaluated the effect of *E. edulis* in the cancer treatment and two studies described the safety evaluation of *E. edulis*, however with controversial results. Barros Freitas et al, 2017 showed juçara prevent the oxidative damage resulting from the cafeteria diet and no evidenced signs of lipid peroxidation in renal or in cardiac tissue in Wistar rats [82]. On the other hand, Felzenszward et al., 2013, demonstrated *E. edulis* was able to induce mutagenicity and clastogenic/aneugenic effects in Wistar rats [25].

Toxicity data are decisive for evaluating the safety of natural products for clinical treatment because these data investigate the potential for mutagenicity, genotoxicity, clastogenicity and aneugenicity [83]. The toxicological studies included in this review showed that açai is non-toxic [42,44,46–48], has no genotoxic or mutagenic effects, and has a protective effect on DNA damage caused by antitumoral agents [46–48]. Similarly, Santos and colleagues showed that antioxidant compounds prevented the induction of DNA damage induced by DXR [84]. On the other hand, açai showed mutagenic effects when assayed in high concentrations in eukaryotic cells of *Saccharomyces cerevisiae* yeast; however, there is a low mutagenic risk for humans because the tested concentrations were significantly elevated [31]. Since only 3 studies investigated the genetic toxicity of açai in preclinical trials of rodents [46–48], future research is needed to better understand the efficacy of açai because its antimutagenic and antioxidant activities may prevent DNA damage and thus improve human health.

## Conclusions

The results of this review suggest that açai is safe and can be used as a chemoprotective agent against cancer by exhibiting antiinflammatory, antioxidant, antiproliferative, and proapoptotic properties. Further studies on the functional relevance of açai are necessary to build a database that can be used in future clinical investigations aimed at discovering antitumor agents.

## Supporting information

**S1 Table. Complete search on açai in databases.**  
(PDF)

**S2 Table. PRISMA checklist.**  
(PDF)

## Acknowledgments

The authors thank the support of Dr. Roberto Soares de Moura in important intellectual discussion about açai.

## Author Contributions

**Conceptualization:** Jamila Alessandra Perini.

**Data curation:** Jéssica Alessandra-Perini, Karina Cristina Rodrigues-Baptista.

**Formal analysis:** Jéssica Alessandra-Perini, Karina Cristina Rodrigues-Baptista.

**Funding acquisition:** Daniel Escorsim Machado, Jamila Alessandra Perini.

**Investigation:** Daniel Escorsim Machado.

**Methodology:** Jéssica Alessandra-Perini, Karina Cristina Rodrigues-Baptista.

**Project administration:** Jamila Alessandra Perini.

**Supervision:** Jamila Alessandra Perini.

**Validation:** Jéssica Alessandra-Perini.

**Visualization:** Daniel Escorsim Machado.

**Writing – original draft:** Jéssica Alessandra-Perini, Jamila Alessandra Perini.

**Writing – review & editing:** Daniel Escorsim Machado, Luiz Eurico Nasciutti.

## References

1. Dutra RC, Campos MM, Santos AR, Calixto JB. Medicinal plants in Brazil: Pharmacological studies, drug discovery, challenges and perspectives. *Pharmacol Res.* 2016; 112:4–29. <https://doi.org/10.1016/j.phrs.2016.01.021> PMID: 26812486
2. Mishra BB, Tiwari VK. Natural products: an evolving role in future drug discovery. *Eur J Med Chem.* 2011; 46(10):4769–807. <https://doi.org/10.1016/j.ejmech.2011.07.057> PMID: 21889825
3. Newman DJ, Cragg GM. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J Nat Prod.* 2012; 75(3):311–35. <https://doi.org/10.1021/np200906s> PMID: 22316239
4. Valli M, dos Santos RN, Figueira LD, Nakajima CH, Castro-Gamboa I, Andricopulo AD, et al. Development of a natural products database from the biodiversity of Brazil. *J Nat Prod.* 2013; 76(3):439–44. <https://doi.org/10.1021/np3006875> PMID: 23330984
5. E Souza BSF, Carvalho HO, Ferreira IM, da Cunha EL, Barros AS, Taglialegna T, et al. Effect of the treatment with *Euterpe oleracea* Mart. oil in rats with Triton-induced dyslipidemia. *Biomed pharmacother.* 2017; 90:542–7. <https://doi.org/10.1016/j.biopha.2017.04.005> PMID: 28402923
6. de Sousa MO, Souza e Silva L, de Brito Magalhães CL, de Figueiredo BB, Costa DC, Silva ME, et al. The hypocholesterolemic activity of açai (*Euterpe oleracea* Mart.) is mediated by the enhanced expression of the ATP-binding cassette, subfamily G transporters 5 and 8 and low-density lipoprotein receptor genes in the rat. *Nutr Res.* 2012; 32(12):976–84. <https://doi.org/10.1016/j.nutres.2012.10.001> PMID: 23244543
7. Kinghorn AD, Chai HB, Sung CK, Keller WJ. The classical drug discovery approach to defining bioactive constituents of botanicals. *Fitoterapia.* 2011; 82(1):71–9. <https://doi.org/10.1016/j.fitote.2010.08.015> PMID: 20804827
8. Ulbricht C, Brigham A, Burke D, Costa D, Giese N, Iovin R, et al. An evidence-based systematic review of açai (*Euterpe oleracea*) by the natural standard research collaboration. *J Diet Suppl.* 2012; 9(2):128–47. <https://doi.org/10.3109/19390211.2012.686347> PMID: 22607647

9. de Moura RS, Resende ÂC. Cardiovascular and metabolic effects of açai, an amazon plant. *J Cardiovasc Pharmacol*. 2016; 68(1):19–26. <https://doi.org/10.1097/FJC.0000000000000347> PMID: [26657713](https://pubmed.ncbi.nlm.nih.gov/26657713/)
10. Yamaguchi KK, Pereira LF, Lamarão CV, Lima ES, da Veiga-Junior VF. Amazon açai: Chemistry and biological activities: A review. *Food Chem*. 2015; 179:137–51. <https://doi.org/10.1016/j.foodchem.2015.01.055> PMID: [25722148](https://pubmed.ncbi.nlm.nih.gov/25722148/)
11. Schauss AG, Wu X, Prior RL, Ou B, Huang D, Owens J, et al. Antioxidant capacity and other bioactivities of the freeze-dried amazonian palm berry, *Euterpe oleracea* Mart. (Açai). *J Agric Food Chem*. 2006; 54(22):8604–10. <https://doi.org/10.1021/jf0609779> PMID: [17061840](https://pubmed.ncbi.nlm.nih.gov/17061840/)
12. Heinrich M, Dhanji T, Casselman I. Açai (*Euterpe oleracea* Mart.)—A phytochemical and pharmacological assessment of the species' health claims. *Phytochemistry Letters*. 2011; 4(1):10–21. <https://doi.org/10.1016/j.phytol.2010.11.005>
13. Silva DF, Vidal FC, Santos D, Costa MC, Morgado-Díaz JA, do Desterro Soares Brandão Nascimento M, et al. Cytotoxic effects of *Euterpe oleracea* Mart. in malignant cell lines. *BMC Complement Altern Med*. 2014; 14:175. <https://doi.org/10.1186/1472-6882-14-175> PMID: [24886139](https://pubmed.ncbi.nlm.nih.gov/24886139/)
14. Del Pozo-Insfran D, Brenes CH, Talcott ST. Phytochemical composition and pigment stability of açai (*Euterpe oleracea* Mart.). *J Agric Food Chem*. 2004; 52(6):1539–45. <https://doi.org/10.1021/jf035189n> PMID: [15030208](https://pubmed.ncbi.nlm.nih.gov/15030208/)
15. Rodrigues RB, Lichtenthaler R, Zimmermann BF, Papagionopoulos M, Fabricius H, Marx F, et al. Total oxidant scavenging capacity of *Euterpe oleracea* Mart. (Açai) seeds and identification of their polyphenolic compounds. *J Agric Food Chem*. 2006; 54(12):4162–7. <https://doi.org/10.1021/jf058169p> PMID: [16756342](https://pubmed.ncbi.nlm.nih.gov/16756342/)
16. Moura RS, Ferreira TS, Lopes AA, Pires KM, Nesi RT, Resende AC, et al. Effects of *Euterpe oleracea* Mart. (Açai) extract in acute lung inflammation induced by cigarette smoke in the mouse. *Phytomedicine*. 2012; 19(3–4):262–9. <https://doi.org/10.1016/j.phymed.2011.11.004> PMID: [22138278](https://pubmed.ncbi.nlm.nih.gov/22138278/)
17. Alqurashi RM, Galante LA, Rowland IR, Spencer JP, Commane DM. Consumption of a flavonoid-rich açai meal is associated with acute improvements in vascular function and a reduction in total oxidative status in healthy overweight men. *Am J Clin Nutr*. 2016; 104(5):1227–35. <https://doi.org/10.3945/ajcn.115.128728> PMID: [27680990](https://pubmed.ncbi.nlm.nih.gov/27680990/)
18. Sadowska-Krepka E, Klapcinska B, Podgórski T, Szade B, Tyl K, Hadzik A. Effects of supplementation with açai (*Euterpe oleracea* Mart.) berry-based juice blend on the blood antioxidant defence capacity and lipid profile in junior hurdlers. A pilot study. *Biol Sport*. 2015; 32(2):161–8. <https://doi.org/10.5604/20831862.1144419> PMID: [26060341](https://pubmed.ncbi.nlm.nih.gov/26060341/)
19. Barbosa PO, Pala D, Silva CT, de Souza MO, do Amaral JF, Vieira RA, et al. Açai (*Euterpe oleracea* Mart.) pulp dietary intake improves cellular antioxidant enzymes and biomarkers of serum in healthy women. *Nutrition*. 2016; 32(6):674–80. <https://doi.org/10.1016/j.nut.2015.12.030> PMID: [26883870](https://pubmed.ncbi.nlm.nih.gov/26883870/)
20. Marques ES, Tsuboy MSF, Carvalho JCT, Rosa PCP, Perazzo FF, Gaivão IOM, et al. First cytotoxic, genotoxic, and antigenotoxic assessment of *Euterpe oleracea* fruit oil (açai) in cultured human cells. *Genet Mol Res*. 2017; 16(3). <https://doi.org/10.4238/gmr16039700> PMID: [28829893](https://pubmed.ncbi.nlm.nih.gov/28829893/)
21. Caiado RR, Peris CS, Lima-Filho AAS, Urushima JGP, Novais E, Badaró E, et al. Retinal toxicity of açai fruit (*Euterpe Oleracea*) dye concentrations in rabbits: Basic principles of a new dye for chromovitrectomy in humans. *Curr Eye Res*. 2017; 42(8):1185–93. <https://doi.org/10.1080/02713683.2017.1297995> PMID: [28494212](https://pubmed.ncbi.nlm.nih.gov/28494212/)
22. Udani JK, Singh BB, Singh VJ, Barrett ML. Effects of açai (*Euterpe oleracea* Mart.) berry preparation on metabolic parameters in a healthy overweight population: a pilot study. *Nutr J*. 2011; 10:45. <https://doi.org/10.1186/1475-2891-10-45> PMID: [21569436](https://pubmed.ncbi.nlm.nih.gov/21569436/)
23. Bashardoust Tajali S, Macdermid JC, Houghton P, Grewal R. Effects of low power laser irradiation on bone healing in animals: a meta-analysis. *J Orthop Surg Res*. 2010; 5(1):1. <https://doi.org/10.1186/1749-799X-5-1> PMID: [20047683](https://pubmed.ncbi.nlm.nih.gov/20047683/)
24. Schreckinger ME, Lotton J, Lila MA, de Mejia EG. Berries from South America: a comprehensive review on chemistry, health potential, and commercialization. *J Med Food*. 2010; 13(2):233–46. <https://doi.org/10.1089/jmf.2009.0233> PMID: [20170356](https://pubmed.ncbi.nlm.nih.gov/20170356/)
25. Felzenszwalb I, da Costa Marques MR, Mazzei JL, Aiub CA. Toxicological evaluation of *Euterpe edulis*: a potential superfruit to be considered. *Food Chem Toxicol*. 2013; 58:536–44. <https://doi.org/10.1016/j.fct.2013.05.029> PMID: [23712094](https://pubmed.ncbi.nlm.nih.gov/23712094/)
26. Kim YS, Jung H, Zerín T, Song HY. Protein profiling of paraquat-exposed rat lungs following treatment with açai (*Euterpe oleracea* Mart.) berry extract. *Mol Med Rep*. 2013; 7(3):881–6. <https://doi.org/10.3892/mmr.2013.1259> PMID: [23291665](https://pubmed.ncbi.nlm.nih.gov/23291665/)
27. de Souza Machado F, Kuo J, Wohlenberg MF, da Rocha Fruscianté M, Freitas M, Oliveira AS, et al. Subchronic treatment with açai frozen pulp prevents the brain oxidative damage in rats with acute liver

- failure. *Metab Brain Dis.* 2016; 31(6):1427–34. <https://doi.org/10.1007/s11011-016-9873-3> PMID: 27418003
28. Kowar M, Friedrich C, Jacobs AH. [Pregabalin as a rare cause of liver disease]. *Dtsch Med Wochenschr.* 2015; 140(23):1759–60. <https://doi.org/10.1055/s-0041-105987> PMID: 26583821
  29. Leba LJ, Brunschwig C, Saout M, Martial K, Bereau D, Robinson JC. *Oenocarpus bacaba* and *Oenocarpus bataua* leaflets and roots: A new source of antioxidant compounds. *Int J Mol Sci.* 2016; 17(7). <https://doi.org/10.3390/ijms17071014> PMID: 27355943
  30. Brasil A, Rocha FAF, Gomes BD, Oliveira KRM, de Carvalho TS, Batista EJO, et al. Diet enriched with the Amazon fruit açai (*Euterpe oleracea*) prevents electrophysiological deficits and oxidative stress induced by methyl-mercury in the rat retina. *Nutr Neurosci.* 2017; 20(5):265–72. <https://doi.org/10.1080/1028415X.2015.1119378> PMID: 26863909
  31. Spada PD, de Souza GG, Bortolini GV, Henriques JA, Salvador M. Antioxidant, mutagenic, and antimutagenic activity of frozen fruits. *J Med Food.* 2008; 11(1):144–51. <https://doi.org/10.1089/jmf.2007.598> PMID: 18361750
  32. Silva D. Analysis of cytotoxicity of fruit extract juçara (*Euterpe oleracea* mart) of Maranhão in human malignant cells. Tese. Universidade do Estado do Rio de Janeiro, 2013.
  33. Wong DY, Musgrave IF, Harvey BS, Smid SD. Açai (*Euterpe oleracea* Mart.) berry extract exerts neuroprotective effects against  $\beta$ -amyloid exposure *in vitro*. *Neurosci Lett.* 2013; 556:221–6. <https://doi.org/10.1016/j.neulet.2013.10.027> PMID: 24161892
  34. Dias MM, Noratto G, Martino HS, Arbizu S, Peluzio M do C, Talcott S, et al. Pro-apoptotic activities of polyphenolics from açai (*Euterpe oleracea* Martius) in human SW-480 colon cancer cells. *Nutr Cancer.* 2014; 66(8):1394–405. <https://doi.org/10.1080/01635581.2014.956252> PMID: 25329001
  35. Brito C, Stavroullakis AT, Ferreira AC, Li K, Oliveira T, Nogueira-Filho G, et al. Extract of açai-berry inhibits osteoclast differentiation and activity. *Arch Oral Biol.* 2016; 68:29–34. <https://doi.org/10.1016/j.archoralbio.2016.03.016> PMID: 27054700
  36. Machado AK, Andrezza AC, da Silva TM, Boligon AA, do Nascimento V, Scola G, et al. Neuroprotective effects of açai (*Euterpe oleracea* Mart.) against rotenone *in vitro* exposure. *Oxid Med Cell Longev.* 2016; 2016:8940850. <https://doi.org/10.1155/2016/8940850> PMID: 27781077
  37. Brito C, Stavroullakis A, Oliveira T, Prakki A. Cytotoxicity and potential anti-inflammatory activity of velutin on RAW 264.7 cell line differentiation: Implications in periodontal bone loss. *Arch Oral Biol.* 2017; 83:348–56. <https://doi.org/10.1016/j.archoralbio.2017.09.001> PMID: 28898790
  38. Freitas DDS, Morgado-Díaz JA, Gehren AS, Vidal FCB, Fernandes RMT, Romão W, et al. Cytotoxic analysis and chemical characterization of fractions of the hydroalcoholic extract of the *Euterpe oleracea* Mart. seed in the MCF-7 cell line. *J Pharm Pharmacol.* 2017; 69(6):714–21. <https://doi.org/10.1111/jphp.12679> PMID: 28211563
  39. Vrillas-Mortimer A, Gomez R, Dowse H, Sanyal S. A survey of the protective effects of some commercially available antioxidant supplements in genetically and chemically induced models of oxidative stress in *Drosophila melanogaster*. *Exp Gerontol.* 2012; 47(9):712–22. <https://doi.org/10.1016/j.exger.2012.06.016> PMID: 22790021
  40. Stoner GD, Wang LS, Seguin C, Rocha C, Stoner K, Chiu S, et al. Multiple berry types prevent N-nitrosomethylbenzylamine-induced esophageal cancer in rats. *Pharm Res.* 2010; 27(6):1138–45. <https://doi.org/10.1007/s11095-010-0102-1> PMID: 20232121
  41. Fragoso MF, Prado MG, Barbosa L, Rocha NS, Barbisan LF. Inhibition of mouse urinary bladder carcinogenesis by açai fruit (*Euterpe oleracea* Martius) intake. *Plant Foods Hum Nutr.* 2012; 67(3):235–41. <https://doi.org/10.1007/s11130-012-0308-y> PMID: 22961050
  42. Fragoso MF, Romualdo GR, Ribeiro DA, Barbisan LF. Açai (*Euterpe oleracea* Mart.) feeding attenuates dimethylhydrazine-induced rat colon carcinogenesis. *Food Chem Toxicol.* 2013; 58:68–76. <https://doi.org/10.1016/j.fct.2013.04.011> PMID: 23597449
  43. Choi YJ, Choi YJ, Kim N, Nam RH, Lee S, Lee HS, et al. Açai berries inhibit colon tumorigenesis in azoxymethane/dextran sulfate sodium-treated mice. *Gut Liver.* 2017; 11(2):243–52. <https://doi.org/10.5009/gnl16068> PMID: 27965474
  44. Monge-Fuentes V, Muehlmann LA, Longo JP, Silva JR, Fascineli ML, de Souza P, et al. Photodynamic therapy mediated by açai oil (*Euterpe oleracea* Martius) in nanoemulsion: A potential treatment for melanoma. *J Photochem Photobiol.* 2017; 166:301–10. <https://doi.org/10.1016/j.jphotobiol.2016.12.002> PMID: 28024281
  45. Nascimento VH, Lima CD, Paixão JT, Freitas JJ, Kietzer KS. Antioxidant effects of açai seed (*Euterpe oleracea*) in anorexia-cachexia syndrome induced by Walker-256 tumor. *Acta Cir Bras.* 2016; 31(9):597–601. <https://doi.org/10.1590/S0102-865020160090000004> PMID: 27737344

46. Ribeiro JC, Antunes LM, Aissa AF, Darin JD, de Rosso VV, Mercadante AZ, et al. Evaluation of the genotoxic and antigenotoxic effects after acute and subacute treatments with açai pulp (*Euterpe oleracea* Mart.) on mice using the erythrocytes micronucleus test and the comet assay. *Mutat Res.* 2010; 695(1–2):22–8. <https://doi.org/10.1016/j.mrgentox.2009.10.009> PMID: 19892033
47. Marques ES, Froder JG, Carvalho JC, Rosa PC, Perazzo FF, Maistro EL. Evaluation of the genotoxicity of *Euterpe oleraceae* Mart. (Arecaceae) fruit oil (açai), in mammalian cells in vivo. *Food Chem Toxicol.* 2016; 93:13–9. <https://doi.org/10.1016/j.fct.2016.04.018> PMID: 27125964
48. Schauss AG, Clewell A, Balogh L, Szakonyi IP, Financsek I, Horváth J, et al. Safety evaluation of an açai-fortified fruit and berry functional juice beverage (MonaVie Active((R))). *Toxicology.* 2010; 278(1):46–54. <https://doi.org/10.1016/j.tox.2010.04.017> PMID: 20452390
49. Hogan S, Chung H, Zhang L, Li J, Lee Y, Dai Y, et al. Antiproliferative and antioxidant properties of anthocyanin-rich extract from açai. *Food Chem* 2010; 118(2):208–14. <https://doi.org/10.1016/j.foodchem.2009.04.099>.
50. Matheus ME, Oliveira SBF, Silveira CS, Rodrigues VP, de Souza Menezes F, Fernandes PD. Inhibitory effects of *Euterpe oleraceae* Mart. on nitric oxide production and iNOS expression. *J Ethnopharmacol.* 2006; 107(2):291–6. <https://doi.org/10.1016/j.jep.2006.03.010> PMID: 16635558
51. Schauss AG, Wu X, Prior RL, Ou B, Patel D, Huang D, et al. Phytochemical and nutrient composition of the freeze-dried amazonian palm berry, *Euterpe Oleraceae* Mart. (Açai). *J Agric Food Chem.* 2006; 54(22):8598–603. <https://doi.org/10.1021/jf060976g> PMID: 17061839
52. de Moura RS, Pires KM, Santos Ferreira T, Lopes AA, Nesi RT, Resende AC, et al. Addition of açai (*Euterpe oleracea*) to cigarettes has a protective effect against emphysema in mice. *Food Chem Toxicol.* 2011; 49(4):855–63. <https://doi.org/10.1016/j.fct.2010.12.007> PMID: 21147193
53. Sudo RT, Neto ML, Monteiro CE, Amaral RV, Resende ÂC, Souza PJ, et al. Antinociceptive effects of hydroalcoholic extract from *Euterpe oleracea* Mart. (Açai) in a rodent model of acute and neuropathic pain. *BMC Complement Altern Med.* 2015; 15(208). <https://doi.org/10.1186/s12906-015-0724-2> PMID: 26134625
54. Poulouse SM, Fischer DR, Larson J, Bielinski DF, Rimando AM, Carey AN, et al. Anthocyanin-rich açai (*Euterpe oleracea* Mart.) fruit pulp fractions attenuate inflammatory stress signaling in mouse brain BV-2 microglial cells. *J Agric Food Chem.* 2012; 60(4):1084–93. <https://doi.org/10.1021/jf203989k> PMID: 22224493
55. Machado DE, Rodrigues-Baptista KC, Alessandra-Perini J, Soares de Moura R, Santos TA, Pereira KG, et al. *Euterpe Oleracea* extract (Açai) is a promising novel pharmacological therapeutic treatment for experimental endometriosis. *PLoS One.* 2016; 11(11):e0166059. <https://doi.org/10.1371/journal.pone.0166059> PMID: 27851787
56. Pala D, Barbosa PO, Silva CT, de Souza MO, Freitas FR, Volp ACP, et al. Açai (*Euterpe oleracea* Mart.) dietary intake plasma lipids, apolipoproteins, cholesteryl ester transfer to high-density lipoprotein and redox metabolism: A prospective study in women. *Clin Nutri.* 2017; 37(2):618–23. <https://doi.org/10.1016/j.clnu.2017.02.001> PMID: 28249700
57. Carvalho-Peixoto J, Moura MR, Cunha FA, Lollo PC, Monteiro WD, Carvalho LM, et al. Consumption of açai (*Euterpe oleracea* Mart.) functional beverage reduces muscle stress and improves effort tolerance in elite athletes: a randomized controlled intervention study. *Appl Physiol Nutr Metab.* 2015; 40(7):725–33. <https://doi.org/10.1139/apnm-2014-0518> PMID: 26140415
58. Pereira IS, Moreira Cançado Mascarenhas Pontes TC, Lima Vieira RA, de Freitas Folly GA, Cacilda Silva F, Pereira de Oliveira FL, et al. The consumption of açai pulp changes the concentrations of plasminogen activator inhibitor-1 and epidermal growth factor (EGF) in apparently healthy women. *Nutr Hosp.* 2015; 32(2):931–45. <https://doi.org/10.3305/nh.2015.32.2.9135> PMID: 26268131
59. Bittman ME, Callahan MJ. The effective use of açai juice, blueberry juice and pineapple juice as negative contrast agents for magnetic resonance cholangiopancreatography in children. *Pediatr Radiol.* 2014; 44(7):883–7. <https://doi.org/10.1007/s00247-014-2884-5> PMID: 24573534
60. Gale AM, Kaur R, Baker WL. Hemodynamic and electrocardiographic effects of açai berry in healthy volunteers: a randomized controlled trial. *Int J Cardiol.* 2014; 174(2):421–3. <https://doi.org/10.1016/j.ijcard.2014.04.036> PMID: 24767759
61. Ellinger S, Gordon A, Kurten M, Jungfer E, Zimmermann BF, Zur B. Bolus consumption of a specifically designed fruit juice rich in anthocyanins and ascorbic acid did not influence markers of antioxidative defense in healthy humans. *J Agric Food Chem.* 2012; 60(45):11292:300. <https://doi.org/10.1021/jf300719t> PMID: 23072538
62. Jensen GS, Ager DM, Redman KA, Mitzner MA, Benson KF, Schauss AG. Pain reduction and improvement in range of motion after daily consumption of an açai (*Euterpe oleracea* Mart.) pulp-fortified polyphenolic-rich fruit and berry juice blend. *J Med Food.* 2011; 14(7–8):702–11. <https://doi.org/10.1089/jmf.2010.0150> PMID: 21470042

63. Sanchez TA, Elias J Jr, Colnago LA, de Almeida Troncon LE, de Oliveira RB, Baffa O, et al. Clinical feasibility of Açai (*Euterpe oleracea*) pulp as an oral contrast agente for magnetic resonance cholangiopancreatography. *J Comput Assist Tomogr*. 2009; 33(5):666–71. <https://doi.org/10.1097/RCT.0b013e31819012a0> PMID: 19820489
64. Jensen GS, Wu X, Patterson KM, Barnes J, Carter SG, Scherwitz L, et al. *In vitro* and *in vivo* antioxidant and anti-inflammatory capacities of an antioxidant-rich fruit and berry juice blend. Results of a pilot and randomized, double-blinded, placebo-controlled, crossover study. *J Agric Food Chem*. 2008; 56(18):8326–33. <https://doi.org/10.1021/jf8016157> PMID: 18717569
65. Mertens-Talcott SU, Rios J, Jilma-Stohlawetz P, Pacheco-Palencia LA, Meibohm B, Talcott ST. Pharmacokinetics of anthocyanins and antioxidant effects after the consumption of anthocyanin-rich acai juice and pulp (*Euterpe oleracea* Mart.) in human healthy volunteers. *J Agric Food Chem*. 2008; 56(17):7796–802. <https://doi.org/10.1021/jf8007037> PMID: 18693743
66. Córdova-Fraga T, de Araújo DB, Sanchez TA, Elias J Jr, Carneiro AA, Brandt Oliveira R, et al. *Euterpe oleracea* (Açai) as an alternative oral contrast agent in MRI of the gastrointestinal system: preliminary results. *Magn Reson Imaging*. 2004; 22(3):389–93. <https://doi.org/10.1016/j.mri.2004.01.018> PMID: 15062934
67. de Oliveira PR, da Costa CA, de Bem GF, Cordeiro VS, Santos IB, de Carvalho LC, et al. *Euterpe oleracea* Mart.-derived polyphenols protect mice from diet-induced obesity and fatty liver by regulating hepatic lipogenesis and cholesterol excretion. *Plos One*. 2015; 10(12):e0143721. <https://doi.org/10.1371/journal.pone.0143721> PMID: 26630290
68. da Costa CA, Ognibene DT, Cordeiro VSC, de Bem GF, Santos IB, Soares RA, et al. Effect of *Euterpe oleracea* Mart. seeds extract on chronic ischemic renal injury in renovascular hypertensive rats. *J Med Food*. 2017; 20(10):1002–10. <https://doi.org/10.1089/jmf.2017.0011> PMID: 28650699
69. Machado DE, Palumbo AJ, Santos JM, Mattos RM, dos Santos TA, Seabra SH, et al. A GFP endometriosis model reveals important morphological characteristics of the angiogenic process that govern benign and malignant diseases. *Histol Histopathol*. 2014; 29(7):903–12. <https://doi.org/10.14670/HH-29.903> PMID: 24385307
70. Becker C, Fantini MC, Schramm C, Lehr C, Wirtz S, Nikolaev A, et al. TGF-beta suppresses tumor progression in colon cancer by inhibition of IL-6 trans-signaling. *Immunity*. 2004; 21(4):491–501. <https://doi.org/10.1016/j.immuni.2004.07.020> PMID: 15485627
71. Popivanova BK, Kitamura K, Wu Y, Kondo T, Kagaya T, Kaneko S, et al. Blocking TNF- $\alpha$  in mice reduces colorectal carcinogenesis associated with chronic colitis. *J Clin Invest*. 2008; 118(2):560–70. <https://doi.org/10.1172/JCI32453> PMID: 18219394
72. Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, et al. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell*. 2009; 15(2):103–13. <https://doi.org/10.1016/j.ccr.2009.01.001> PMID: 19185845
73. Xie C, Kang J, Li Z, Schauss AG, Badger TM, Nagarajan S, et al. The açai flavonoid velutin is a potent anti-inflammatory agent: blockade of LPS-mediated TNF- $\alpha$  and IL-6 production through inhibiting NF- $\kappa$ B activation and MAPK pathway. *J Nutr Biochem*. 2012; 23(9):1184–91. <https://doi.org/10.1016/j.jnutbio.2011.06.013> PMID: 22137267
74. El Morsy EM, Ahmed MA, Ahmed AA. Attenuation of renal ischemia/reperfusion injury by açai extract preconditioning in a rat model. *Life Sci*. 2015; 123:123–35. <https://doi.org/10.1016/j.lfs.2014.11.013> PMID: 25476829
75. Guzinska-Ustymowicz K, Pryczynicz A, Kemon A, Czyzewska J. Correlation between proliferation markers: PCNA, Ki-67, MCM-2 and antiapoptotic protein Bcl-2 in colorectal cancer. *Anticancer Res*. 2009; 29(8):3049–52. PMID: 19661314
76. Graziano V, de Laurenzi V. Role of p63 in cancer development. *Biochim Biophys Acta*. 2011; 1816(1):57–66. <https://doi.org/10.1016/j.bbcan.2011.04.002> PMID: 21515338
77. Qiu X, Mei J, Yin J, Wang H, Wang J, Xie M. Correlation analysis between expression of PCNA, Ki-67 and COX-2 and X-ray features in mammography in breast cancer. *Oncol Lett*. 2017; 14(3):2912–8. <https://doi.org/10.3892/ol.2017.6516> PMID: 28927045
78. Roy AM, Baliga MS, Katiyar SK. Epigallocatechin-3-gallate induces apoptosis in estrogen receptor-negative human breast carcinoma cells via modulation in protein expression of p53 and Bax and caspase-3 activation. *Mol Cancer Ther*. 2005; 4(1):81–90. PMID: 15657356
79. Forester SC, Gu Y, Lambert JD. Inhibition of starch digestion by the green tea polyphenol, (-)-epigallocatechin-3-gallate. *Mol Nutr Food Res*. 2012; 56(11):1647–54. <https://doi.org/10.1002/mnfr.201200206> PMID: 23038646
80. Del Pozo-Insfran D, Percival SS, Talcott ST. Açai (*Euterpe Oleracea* Mart.) polyphenolics in their glycoside and aglycone forms induce apoptosis of HL-60 leukemia cells. *J Agric Food Chem*. 2006; 54(4):1222–9. <https://doi.org/10.1021/jf052132n> PMID: 16478240

81. Cardoso AL, de Liz S, Rieger DK, Farah ACA, Kunradi Vieira FG, Altenburg de Assis MA, et al. An Update on the Biological Activities of *Euterpe edulis* (Juçara). *Panta Med*. 2018. <https://doi.org/10.1055/s-0044-101624> PMID: 29466809
82. de Barros Freitas R, Melato FA, Oliveira JM, Bastos DS, Cardoso RM, Leite JP, et al. *Euterpe edulis* effects on cardiac and renal tissues of Wistar rats fed with cafeteria diet. *Nutr Hosp*. 2017; 34(1):186–92. <https://doi.org/10.20960/nh.996> PMID: 28244791
83. Rim KT, KIM SJ. A review on mutagenicity testing for hazard classification of chemicals at work: focusing on *in vivo* micronucleus test for allyl chloride. *Saf Health Work*. 2015; 6(3):184–91. <https://doi.org/10.1016/j.shaw.2015.05.005> PMID: 26929826
84. Santos RA, Takahashi CS. Anticlastogenic and antigenotoxic effects of selenomethionine on doxorubicin-induced damage *in vitro* in human lymphocytes. *Food Chem Toxicol*. 2008; 46(2):671–7. <https://doi.org/10.1016/j.fct.2007.09.090> PMID: 17961897